

Article

## Novel Polyprenylated Phloroglucinols from *Hypericum sampsonii*

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**Abstract:** *Hypericum sampsonii* Hance (Clusiaceae) is a folk medicine used in Taiwan to treat blood stasis, relieve swelling, and as an anti-hepatitis drug. Two new polyprenylated phloroglucinol derivatives, hypersampsone R (**1**) and hypersampsone S (**2**), and a known prenylated benzophenone, hyperibone K (**3**) were isolated from the aerial parts of *H. sampsonii*. Their structures were determined by extensive 1D and 2D NMR, and MS spectral analyses.

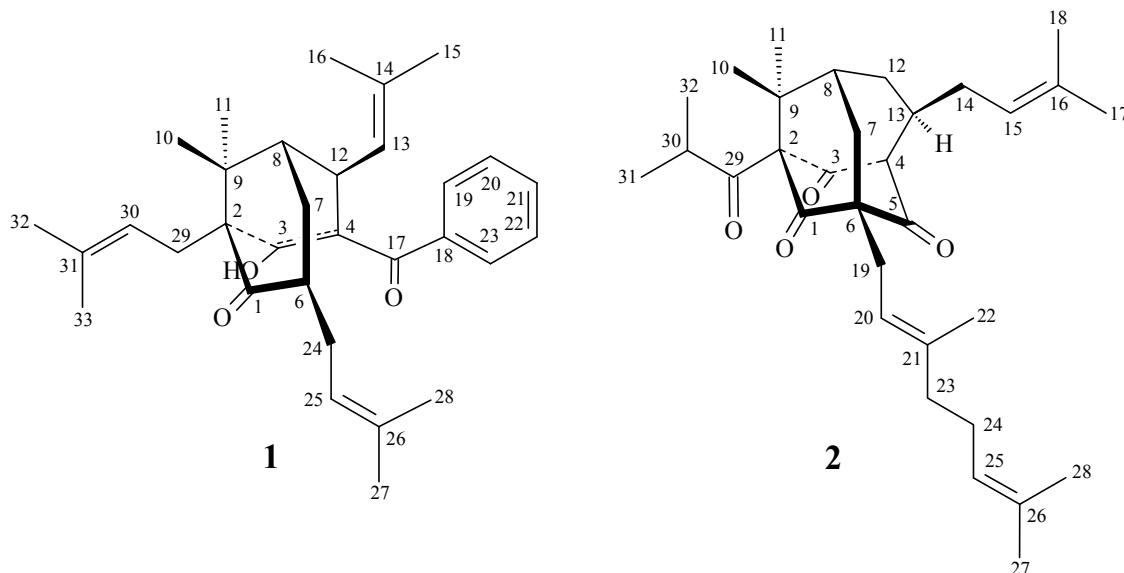
**Keywords:** *Hypericum sampsonii*; Guttiferae; polyprenylated phloroglucinol

### 1. Introduction

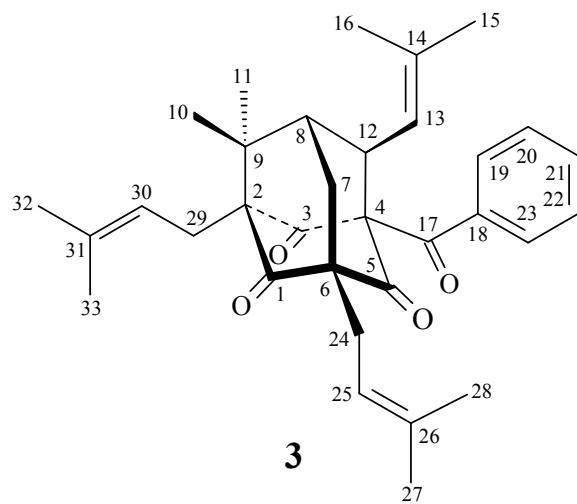
*Hypericum sampsonii* Hance (Guttiferae) is a folk herbal medicine used in Taiwan for treating blood stasis, to relieve swelling, and as an anti-hepatitic drug [1]. Due to not only the biological activities, but also the structural diversity, the chemical constituents of *Hypericum* species have attracted much attention, and different kinds of compounds such as xanthones [2–5], benzophenones [5,6], bisanthraquinones [6,7], and polyprenylated phloroglucinols [8–16] have been isolated. A continuing chemical investigation on the secondary metabolites of this plant resulted in the isolation of a new

ring-opened polyisoprenylated benzophenone, hypersampsone R (**1**) and a new polyisoprenylated phloroglucinol, hypersampsone S (**2**) (Figure 1), as well as a known polyisoprenylated benzophenone, hyperibone K (**3**) (Figure 2). This paper describes the structural elucidation of compounds **1** and **2**.

**Figure 1.** The chemical structures of new compounds **1** and **2** isolated from *H. sampsonii*.



**Figure 2.** The chemical structure of known compound **3** isolated from *H. sampsonii*.



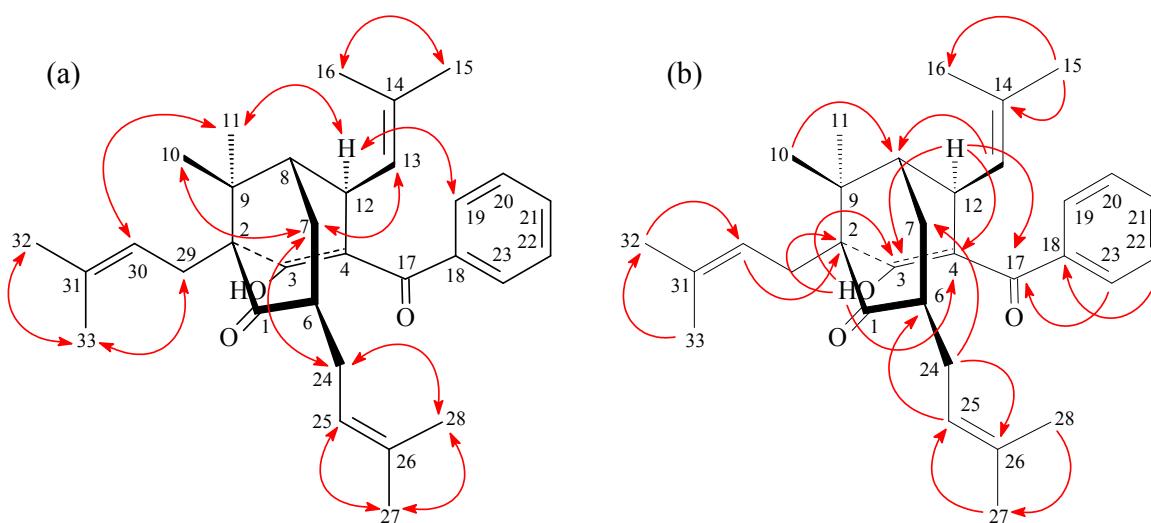
## 2. Results and Discussion

The ethanol extract of the air-dried aerial parts of *H. sampsonii* was successively partitioned with ethyl acetate (EtOAc) and *n*-butanol (*n*-BuOH) to give EtOAc, *n*-BuOH and H<sub>2</sub>O fractions. The EtOAc soluble partition enriched in polyisoprenylated phloroglucinols was subjected to silica gel and RP-18 column chromatography in combination with preparative silica-gel HPLC to yield two new compounds **1** and **2**, together with a known compound **3**.

Hypersamsone R (**1**) was isolated as an optically active ( $[\alpha]_D^{25} = +160^\circ$ ), colorless amorphous powder. The molecular formula was established as C<sub>32</sub>H<sub>42</sub>O<sub>3</sub> on the basis of HR-EI-MS (found m/z 474.3128,

calcd. for  $C_{32}H_{42}O_3$  474.3134) with twelve indices of hydrogen deficiency (IHD). The IR spectrum displayed absorptions of hydroxyl ( $3443\text{ cm}^{-1}$ ) and carbonyl groups ( $1709$  and  $1677\text{ cm}^{-1}$ ). The  $^1\text{H-NMR}$  spectrum of **1** showed three olefinic protons [ $\delta_H$  4.63 (1H, d,  $J = 7.8$  Hz), 4.98 (1H, t,  $J = 6.5$  Hz), and 5.15 (br s)], eight methyls [ $\delta_H$  0.92, 1.18, 1.24, 1.26, 1.54, 1.56, 1.65, and 1.66 (each 3H, s)], a benzoyl [ $\delta_H$  7.36 (3H, m) and 7.39 (2H, m)], and a conjugated hydroxyl group [ $\delta_H$  16.25 (s)]. The  $^{13}\text{C-NMR}$ , DEPT and HMQC spectra indicated the presence of 32 carbons, including two carbonyls ( $\delta_C$  195.2 and 209.4), eight quaternary carbons with one conjugated oxygenated quaternary carbon ( $\delta_C$  185.5), five fully substituted aromatic and olefinic quarternary carbons ( $\delta_C$  111.4, 130.3, 132.7, 133.2, and 139.3) and two quaternary carbons ( $\delta_C$  42.4 and 65.4), eight double-bond methine carbons [ $\delta_C$  121.3, 122.9, 125.7, 126.7, 126.7, 128.0, 128.0 and 130.1], three methine carbons [ $\delta_C$  35.4, 45.3, 45.5], three methylene carbons [ $\delta_C$  26.3, 28.3, 29.9], and eight methyl carbons [ $\delta_C$  17.3, 17.9, 17.9, 25.1, 25.1, 25.7, 26.0, 26.0]. The  $^1\text{H-}^1\text{H COSY}$  indicated the correlations of H-12 ( $\delta_H$  4.00) and H-13 ( $\delta_H$  4.63) and H-8 ( $\delta_H$  1.50); H-6 ( $\delta_H$  2.50) and H-7 ( $\delta_H$  1.44, 2.06) and H-24 ( $\delta_H$  2.32). Comparison of the  $^1\text{H-}$  and  $^{13}\text{C-NMR}$  data (Table 1) of **1** with those of hyperibone K (**3**) [17] suggested that their structures were closely related, except that the C3,C4-double bond, 3-hydroxy group, and the lack of carbonyl group between C-4 and C-6 of **1** replaced the C3,C4-single bond and 3-oxo group of hyperibone K (**3**) [17]. This was supported by HMBC correlations (Figure 3) between OH-3 ( $\delta_H$  16.25) and C-2 ( $\delta_C$  65.4), C-3 ( $\delta_C$  185.5), and C-4 ( $\delta_C$  111.4); between H-12 ( $\delta_H$  4.00) and C-3 ( $\delta_C$  185.5), C-4 ( $\delta_C$  111.4), C-7 ( $\delta_C$  29.9), C-8 ( $\delta_C$  45.3), C-13 ( $\delta_C$  125.7) and C-14 ( $\delta_C$  132.7). The NOESY cross-peaks (Figure 3) between H-7/Me(10), H-7/H-13, H-7/H-24, H <sub>$\alpha$</sub> -12/H-19, H <sub>$\alpha$</sub> -12/Me(11), and H-30/Me(11) suggested that H-12, Me(11), 2-isoprenyl, and the 4-benzoyl groups are  $\alpha$ -oriented, and 6-isoprenyl, Me(10), and C-12 2-methylprop-1-enyl groups are  $\beta$ -oriented. The full assignment of  $^1\text{H-}$  and  $^{13}\text{C-NMR}$  resonances was supported by  $^1\text{H-}^1\text{H COSY}$ , DEPT, HMQC, NOESY (Figure 3), and HMBC (Figure 3) spectral analyses. According to the above data, hypersamsone R was identified as structure **1**.

**Figure 3.** Key NOESY (a) and HMBC (b) correlations of **1**.



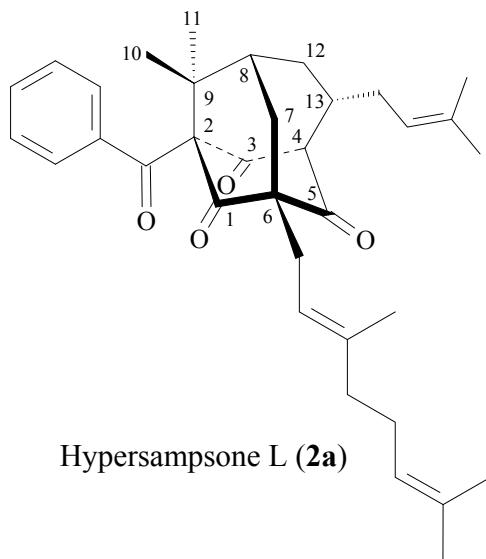
**Table 1.**  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR data of **1** and **2**.

Position	$\delta_{\text{H}}$			
	<b>1</b> <sup>a</sup>		<b>2</b> <sup>a</sup>	
	$^1\text{H}$	$^{13}\text{C}$	$^1\text{H}$	$^{13}\text{C}$
1		209.4 s		202.2 s
2		65.4 s		88.8 s
3		185.5 s		205.4 s
4		111.4 s	2.97 m	50.0 d
5				204.7 s
6	2.50 m	45.5 d		69.2 s
7	1.44 m 2.06 m	29.9 t 45.3 d	1.87 d (10.5) 2.50 m	36.2 t 41.4 d
8	1.50 m		1.75 m	
9		42.4 s		50.0 s
10	1.26 s	26.0 q	1.20 s	24.8 q
11	0.92 s	25.1 q	1.18 s	24.2 q
12	4.00 dd (7.8, 7.0)	35.4 d	1.84 d (7.8) 2.74 m	35.4 d
13	4.63 d (7.8)	125.7 d	2.81 m	58.1 d
14		132.7 s	2.06 m 2.30 m	31.1 t
15	1.24 s	25.1 q	4.97 t (6.5)	119.9 d
16	1.18 s	17.3 q		135.8 s
17		195.2 s	1.67 s	25.8 q
18		139.3 s	1.56 s	17.9 q
19	7.36 m	128.0 d	2.50 m 2.66 dd (11.5, 7.0)	32.3 t
20	7.39 m	126.7 d	4.99 t (7.0)	119.3 d
21	7.36 m	130.1 d		139.6 s
22	7.39 m	126.7 d	1.61 s	16.4 q
23	7.36 m	128.0 d	2.01 m	40.1 t
24	1.88 m 2.32 m	28.3 t	2.03 m	26.6 t
25	4.98 t (6.5)	121.3 d	5.00 t (6.5)	124.1 d
26		133.2 s		131.5 s
27	1.65 s	26.0 q	1.64 s	25.7 q
28	1.54 s	17.9 q	1.56 s	17.6 q
29	2.57 dd (13.0, 7.0) 2.89 br d (13.0)	26.3 t		211.5 s
30	5.15 br s	122.9 d	2.45 hepta (7.0)	42.8 d
31		130.3 s	1.19 d (7.0)	21.0 q
32	1.66 s	25.7 q	1.21 d (7.0)	21.0 q
33	1.56 s	17.9 q		
OH	16.25 s			

<sup>a</sup> Recorded in  $\text{CDCl}_3$  at 500 MHz ( $^1\text{H}$ ) and 125 MHz ( $^{13}\text{C}$ ). Values in ppm ( $\delta$ ).  $J$  (in Hz) in parentheses.

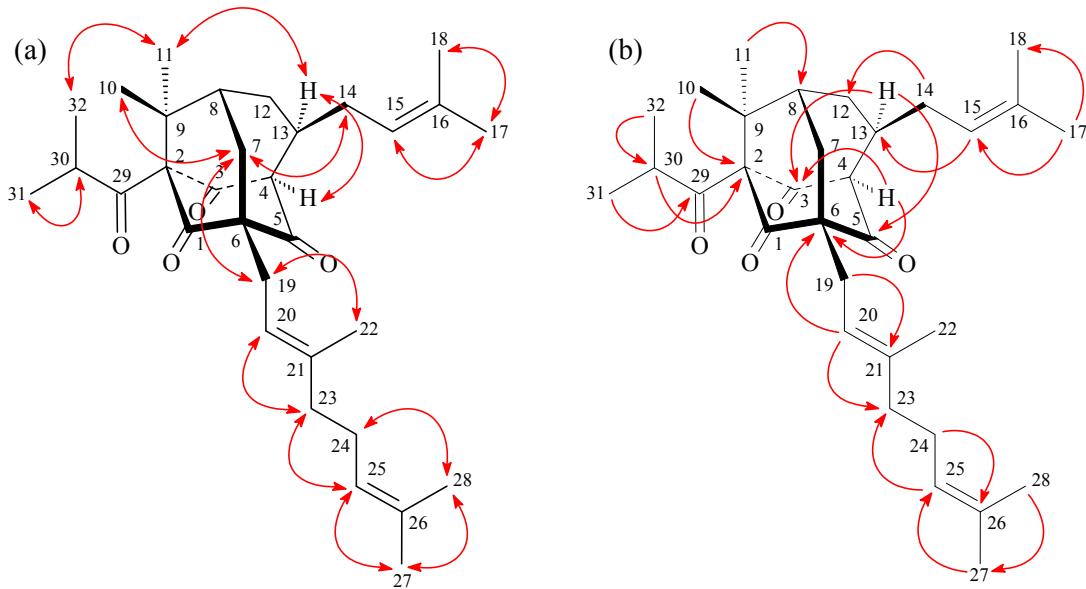
Hypersamsone S (2) was obtained as a colorless amorphous powder. The molecular formula was determined to be C<sub>32</sub>H<sub>46</sub>O<sub>4</sub> on the basis of HR-EI-MS (found *m/z* 494.3396, calcd. for C<sub>32</sub>H<sub>46</sub>O<sub>4</sub> 494.3394) with nine IHD. The <sup>13</sup>C-NMR, DEPT and HMQC spectra indicated ten quaternary carbons [including four carbonyl ( $\delta_c$  202.2, 204.7, 205.4, and 211.5), three olefinic quaternary ( $\delta_c$  131.5, 135.8, and 139.6), and three other quaternary carbons ( $\delta_c$  50.0, 69.2, and 88.8)], seven tertiary carbons [including three olefinic ( $\delta_c$  119.3, 119.9, and 124.1) and four other tertiary carbons ( $\delta_c$  41.4, 42.8, 50.0, and 58.1)], and six methylene carbons ( $\delta_c$  26.6, 31.1, 32.3, 35.4, 36.2, and 40.1), and nine methyl carbons ( $\delta_c$  16.4, 17.6, 17.9, 21.0, 21.0, 24.2, 24.8, 25.7, and 25.8). Comparison of the <sup>1</sup>H- and <sup>13</sup>C-NMR data (Table 1) of 2 with those of hypersampsone L (2a) [18] suggested that their structures were closely related, except that 2-isobutyryl and 13 $\beta$ -isoprenyl groups of 2 replaced 2-benzoyl and 13 $\alpha$ -isoprenyl groups of hypersampsone L (2a, Figure 4) [18]. This was supported by HMBC correlations (Figure 5) between H-15 ( $\delta_H$  4.97) and C-13 ( $\delta_c$  58.1); between H-20 ( $\delta_H$  4.99) and C-6 ( $\delta_c$  69.2); and between H-30 ( $\delta_H$  2.45) and C-2 ( $\delta_c$  88.8). The NOESY cross-peaks (Figure 5) between H <sub>$\alpha$</sub> -4/ H <sub>$\alpha$</sub> -13, H <sub>$\alpha$</sub> -13/Me(11), H-32/Me(11), H-7/Me(10), H-7/H-14, H-7/H-19, suggested that H <sub>$\alpha$</sub> -4, H <sub>$\alpha$</sub> -13, Me(11), and the 2-isobutyryl groups are  $\alpha$ -oriented, and 6-geranyl, Me(10), and 13-isoprenyl groups are  $\beta$ -oriented. On the basis of the evidence above, the structure of hypersamsone S was elucidated as 2, which was further substantiated through 2D-experiments, including HMQC, <sup>1</sup>H-<sup>1</sup>H COSY, HMBC (Figure 5), and NOESY (Figure 5) spectra.

**Figure 4.** The chemical structure of hypersampsone L (2a).



The known isolate, hyperibone K (3) was readily identified by a comparison of physical and spectroscopic data (IR, <sup>1</sup>H-NMR,  $[\alpha]_D$ , and MS) with the literature values [17].

**Figure 5.** Key NOESY (**a**) and HMBC (**b**) correlations of **2**.



### 3. Experimental Section

### *3.1. General Experimental Procedures*

Optical rotations were measured using a Jasco P-2000 polarimeter in CHCl<sub>3</sub>. Infrared (IR) spectra (KBr) were recorded on a Nicolet Avatar 320 FT-IR spectrophotometer. Nuclear magnetic resonance (NMR) spectra, including correlation spectroscopy (COSY), nuclear Overhauser effect spectroscopy (NOESY), heteronuclear multiple-bond correlation (HMBC), and heteronuclear multiple quantum coherence (HMQC) experiments, were acquired using a Varian Inova 500 spectrometer operating at 500 MHz (<sup>1</sup>H) and 125 MHz (<sup>13</sup>C), respectively, with chemical shifts given in ppm ( $\delta$ ) using CDCl<sub>3</sub> as solvent. Chemical shifts were referenced to the residual solvent peaks ( $\delta_H$  7.24 and  $\delta_C$  77.0). Mass spectra (EIMS and HREIMS) were recorded on a Finnigan LCQ and JEOL Finnigan MAT 95S Mass Spectrometer, respectively. Column chromatography was performed using silica gel (70-230 mesh, Merck, Darmstadt, Germany) and Sephadex<sup>TM</sup> LH-20 (Amersham Biosciences, Uppsala, Sweden). Preparative HPLC was conducted using a L-2130 pump (Hitachi, Tokyo, Japan) and a LiChrosorb Si-60 column (Merck).

### *3.2. Plant Material*

The aerial parts of *Hypericum sampsonii* Hance were collected from Chia-Yi county in June 2007. The plant was identified by Mr. Jun-Chih Ou, former associate research fellow of National Research Institute of Chinese Medicine, and compared with a voucher specimen which was deposited in the Herbarium of the Institute of Ecology and Evolutionary Biology, National Taiwan University, Taipei, Taiwan (No.077152).

### 3.3. Extraction and Isolation

The dried aerial parts of *H. sampsonii* (12.0 kg) were extracted overnight with 95% ethanol (EtOH) at 60 °C three times (80 L each). The EtOH extracts were concentrated under reduced pressure, and the residue (215 g) was partitioned successively with ethyl acetate (EtOAc) and *n*-butanol (BuOH), respectively. The EtOAc fraction (103 g) was subjected to silica gel column chromatography (8 × 80 cm) and eluted with a EtOAc/hexane gradient. Fractions of 10%–15% EtOAc eluate were collected and rechromatographed over silica gel and RP-18 (MeOH) columns in combination with silica-gel preparative HPLC (15% or 20% EtOAc/Hex) to give hypersampsone R (**1**) (15 mg), hypersampsone S (**2**) (12 mg), and hyperibone K (**3**) (25 mg).

*Hypersamsone R* (**1**). Colorless amorphous powder.  $[\alpha]_D^{25} : +160$  (*c* 0.1, CHCl<sub>3</sub>). IR (KBr):  $\nu_{\text{max}} = 3443, 2965, 2928, 1709, 1677, 1598, 1257, 1115$ , and 747 cm<sup>-1</sup>. <sup>1</sup>H- and <sup>13</sup>C-NMR spectroscopic data, see Table 1. Key COSY correlations: H-6/H-7; H-6/H-24; H-7/H-8; H-8/H-12; H-12/H-13; H-24/H-25; H-29/H-30. Key NOESY correlations: H-7/H-10; H-7/H-13; H-7/H-24; H-11/H-12; H-11/H-30; H-12/H-19. Key HMBC correlations: H-6/C-1, -7, -24; H-10/C-2, -8, -9; H-12/C-3, -4, -7, -8, -13, -14, -17; H-13/C-15, -16; H-19/C-17, -18, -20, -21; H-24/C-6, -7, -25, -26; H-29/C-2, -3, -9, -30, -31; H-32 (H-33)/C-30, -31. EI-MS: *m/z* (%) = 474 [M]<sup>+</sup> (8), 446 (15), 128 (23), 105 (100). HR-EI-MS: *m/z* = 474.3128 [M]<sup>+</sup> (calcd for C<sub>32</sub>H<sub>42</sub>O<sub>3</sub>: 474.3134).

*Hypersamsone S* (**2**). Colorless amorphous powder.  $[\alpha]_D^{25} : +33$  (*c* 0.3, CHCl<sub>3</sub>). IR (KBr):  $\nu_{\text{max}} = 2969, 2927, 1718, 1703, 1441, 1136, 1067$ , and 826 cm<sup>-1</sup>. <sup>1</sup>H- and <sup>13</sup>C-NMR spectroscopic data, see Table 1. Key COSY correlations: H-8/H-7, -12; H-13/H-4, -12, -14; H-24/H-23, -25; H-30/H-31, -32. Key NOESY correlations: H-4/H-13; H-7/H-10, H-14, H-19; H-8/H-7; H-11/H-13, -32; H-15/H-14, -17, -18; H-25/H-23, H-24, -27, -28; H-30/H-31, -32. Key HMBC correlations: H-4/C-2, -3, -6, -12, -14; H-7/C-1, -5, -6, -8, -9, -12; H-12/C-4, -7, -8; H-13/C-4, -5, -12, -14; H-14/C-4, -12, -13, -15, -16; H-15/C-13, -14, -16, -17, -18; H-17/C-15, -16, -18; H-19/C-1, -5, -6, -7, -20, -21; H-31/C-29, -30, -32. EI-MS: *m/z* (%) = 494 [M]<sup>+</sup> (10), 466 (16), 397 (25), 355 (35), 189 (38), 135 (45), 109 (52), 91 (55), 71 (100). HR-EI-MS: *m/z* = 494.3396 [M]<sup>+</sup> (calcd for C<sub>32</sub>H<sub>46</sub>O<sub>4</sub>: 494.3394).

## 4. Conclusions

Three compounds, including two new compounds **1** and **2**, were isolated from the aerial parts of *H. sampsonii*. The structures of these compounds were established on the basis of spectroscopic data. The discovery of new compounds from the genus *Hypericum* may not only provide more spectroscopic data about these isolates, but may also contribute to enhancing our understanding of the taxonomy and evolution of the genus *Hypericum*.

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## Author Contributions

Yun-Lian Lin designed the whole experiment and contributed to manuscript preparation; Jih-Jung Chen analyzed the data and wrote the manuscript; Hong-Jhang Chen contributed to the data collection.

## Conflicts of Interest

The authors declare no conflict of interest.

## References

1. Chiu, N.Y.; Chang, K.H. *The Illustrated Medicinal Plants of Taiwan*; SMC Publishing Inc.: Taipei, Taiwan, 1986; Volume II, p. 126.
2. Chen, M.T.; Chen, C.M. Xanthones from *Hypericum sampsonii*. *Heterocycles* **1985**, *23*, 2543–2548.
3. Hu, L.H.; Yip, S.C.; Sim, K.Y. Xanthones from *Hypericum ascyron*. *Phytochemistry* **1999**, *52*, 1371–1373.
4. Hong, D.; Yin, F.; Hu, L.H.; Lu, P. Sulfonated xanthones from *Hypericum sampsonii*. *Phytochemistry* **2004**, *65*, 2595–2598.
5. Don, M.J.; Huang, Y.J.; Huang, R.L.; Lin, Y.L. New Phenolic principles from *Hypericum sampsonii*. *Chem. Pharm. Bull.* **2004**, *52*, 866–869.
6. Kitanov, G.M.; Nedialkov, P.T. Benzophenone O-glucoside, a biogenic precursor of 1,3,7-trioxygenated xanthones in *Hypericum annulatum*. *Phytochemistry* **2001**, *57*, 1237–1243.
7. Wirz, A.; Simmen, U.; Heilmann, J.; Calis, I.; Meier, B.; Sticher, O. Bisanthraquinone glycosides of *Hypericum perforatum* with binding inhibition to CRH-1 receptors. *Phytochemistry* **2000**, *55*, 941–947.
8. Hu, L.H.; Sim, K.Y. Prenylated phloroglucinol derivatives from *Hypericum sampsonii*. *Tetrahedron Lett.* **1998**, *39*, 7999–8002.
9. Hu, L.H.; Sim, K.Y. Complex caged polyisoprenylated benzophenone derivatives, sampsoniones A and B, from *Hypericum sampsonii*. *Tetrahedron Lett.* **1999**, *40*, 759–762.
10. Hu, L.H.; Sim, K.Y. Cytotoxic polyprenylated benzoylphloroglucinol derivatives with an unusual adamantly skeleton from *Hypericum sampsonii* (Guttiferae). *Org. Lett.* **1999**, *1*, 879–882.
11. Hu, L.H.; Sim, K.Y. Sampsoniones A–M, a unique family of caged polyprenylated benzoylphloroglucinol derivatives, from *Hypericum sampsonii*. *Tetrahedron* **2000**, *56*, 1379–1386.
12. Shan, M.D.; Hu, L.H.; Chen, Z.L. Three new hyperforin analogues from *Hypericum perforatum*. *J. Nat. Prod.* **2001**, *64*, 127–130.
13. Matsuhisa, M.; Shikishima, Y.; Takaishi, Y.; Honda, G.; Ito, M.; Takeda, Y.; Shibata, H.; Higuti, T.; Kodzhimatov, O.K.; Ashurmrtov, O. Benzoylphloroglucinol derivatives from *Hypericum scabrum*. *J. Nat. Prod.* **2002**, *65*, 290–294.
14. Lin, Y.L.; Wu, Y.S. Polyprenylated phloroglucinol derivatives from *Hypericum sampsonii*. *Helv. Chim. Acta* **2003**, *86*, 2156–2163.
15. Xiao, Z.Y.; Mu, Q.; Shiu, W.K.P.; Zeng, Y.H.; Gibbons, S. Polyisoprenylated benzoyl-phloroglucinol derivatives from *Hypericum sampsonii*. *J. Nat. Prod.* **2007**, *70*, 1779–1782.

16. Xin, W.B.; Man, X.H.; Zheng, C.J.; Jia, M.; Jiang, Y.P.; Zhao, X.X.; Jin, G.L.; Mao, Z.J.; Huang, H.Q.; Qin, L.P. Prenylated phloroglucinol derivatives from *Hypericum sampsonii*. *Fitoterapia* **2012**, *83*, 1540–1547.
17. Tanaka, N.; Takaishi, Y.; Shikishima, Y.; Nakanishi, Y.; Bastow, K.; Lee, K.H.; Honda, G.; Ito, M.; Takeda, Y.; Kodzhimatov, O.K.; *et al.* Prenylated benzophenones and xanthones from *Hypericum scabrum*. *J. Nat. Prod.* **2004**, *67*, 1870–1875.
18. Zeng, Y.H.; Osman, K.; Xiao, Z.Y.; Gibbons, S.; Mua, Q. Four geranyl-bearing polyisoprene-nylated benzoylphloroglucinol derivatives from *Hypericum sampsonii*. *Phytochem. Lett.* **2012**, *5*, 200–205.

*Sample Availability:* Samples of the all compounds are available from the authors.

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