Systematically characterize the anti-Alzheimer's disease mechanism of lignans in *S. chinensis* based on in-vivo ingredients analysis and target-network pharmacology strategy by UHPLC-Q-TOF-MS

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1 Preparation of lignans purification isolated from S. chinensis

The 50 g crude herbs were soaked overnight with 750 mL water. And then the extraction method of drug sediments were described below: 80% ethanol of extraction solvent, homogenate extraction time and voltage of 100 s and100 v, liquid to solid ratio of 19 : 1. This supernatant was purified with AB-8 macroreticular adsorption resin to get the lignans purification from the extrait of *S. chinensis*. The purity of lignans from *S. chinensis* could reach to more than 84%.

2 Identification of lignans isolated from S. chinensis in purification

The column temperature was kept at 40 °C and the injection volume was 5 μ L. The mobile phase consisted of 0.1 % formic acid (v/v) (A) and methyl alcohol (B) at a flow rate of 0.35 mL/min. The gradient elution in positive mode was performed as follows: 5 % - 55 % B at 0 -0.5 min, 55 % - 88 % B at 0.5 - 8 min, 88 % - 100 % B at 8 - 10 min, 100 % - 5 % B at 10 - 10.5 min, and 5 % B at 10.5 - 15 min. The electrospray ionization (ESI) source with a scanning mass-to-charge (m/z) range from 50 to 1000 Da was used for the experiment. The source temperature was 120 °C. Nitrogen was used as cone and desolvation gas. The flow rates were 50 L/h and 700 L/h, respectively and the solvent temperature was 350 °C. The cone and extraction cone voltages were 40 V and 5.0 V, respectively. The capillary voltage was set to 3.0 kV. Quality standard curves were established using sodium formate. Leucine enkephalin was used as reference mass. The MSE data acquisition was performed using He as collision gas respectively with a low collision energy of 5 eV and a high collision energy of 10 - 30 eV. The database of lignans in S. chinensis based on mass spectrum database (PubMed, Mass Bank, Chemspider, and METLIN) and references was built in UNIFI TM (Waters Corp. Milford, MA, USA). The database of lignans in *S. chinensis* included compound name, molecular formula, structural formula and fragment ion. Then MassFragment TM and UNIFI TM were used to analysis raw data of lignans purification isolated from S. chinensis.



Figure S1. Total ion current chromatogram of lignans isolated from *S. chinensis* in positive mode

The base peak ion chromatogram (BPI) of lignans isolated from *S. chinensis* was showed in Figure S1. Those unknown compounds were identified using MS or MS/MS spectral databases. And some of those were also compared with reference standards. What follows was the description of the identification procedure taking schisandrin as examples. The ion exhibited a retention time of 2.57 min and the [M + H] + ion at m/z 433.2043 in positive mode. Via searching UNIFI databases, the compound may be identified as schisandrin. Then we confirmed that its retention times and accurate masses were consistent with reference standards. In MS/MS analysis, fragment ions of schisandrin in lignans purification were also the same as the reference standards (Figure S2). Finally the fragment ions were identified to be schisandrin. And then fourteen lignans were identified which details were listed in Table S1, including schisandrin, gomisin D, gomisin J, benzoylgomisin H, angeloylgomisin Q, schisandrol B, gomisin G, schisantherin A, gomisin K, tigloylgomisin H, gomisin E, deoxyschizandrin, schisandrin B amd schisandrin C.



Compounds	RT (min)	Measured mass	Formlar	Mass error(ppm)	Adducts	MS ²
Schisandrin ^a	2.57	455.2043	C24H32O7	0.7	+Na, +H	415, 400, 385, 384, 373, 369, 359, 354, 353, 338, 322
Gomisin D	2.97	553.2061	C28H34O10	3	+Na, +H	485, 401, 383, 371, 369, 353, 351
Gomisin J	3.13	389.1960	C22H28O6	0.5	+H, +Na	374, 358, 342, 319, 288
Benzoylgomisin H	3.34	523.2305	C30H34O8	-4.1	+H	505, 401
Angeloylgomisin Q	3.43	553.2403	C29H38O9	-0.9	+Na	431, 413, 401, 399, 389, 387, 357
Schisandrol B ^a	3.45	439.1907	C23H28O7	1	+Na, +H	399, 384, 369, 368, 357, 353, 343, 341, 337, 338, 295
Gomisin G	4.37	537.2109	C30H32O9	-1.9	+H, +Na	415, 397, 384, 373, 371, 356, 341, 340
Schisantherin A ^a	4.52	537.5765	C30H32O9	0.4	+H, +Na	415, 397, 385, 373, 371, 367, 356, 340, 325, 310, 297, 295, 267
Gomisin K	4.71	403.2120	C23H30O6	1.1	+H, +Na	388, 371, 340, 333, 302, 301
Tigloylgomisin H	4.76	523.2296	C28H36O8	-1.2	+Na	483, 401, 357
Schisantherin B ^a	4.84	537.2098	C28H34O9	0.6	+Na	415, 397, 385, 371
Gomisin E	5.17	515.2288	C28H34O9	2.3	+H, +Na	469, 385, 367, 355, 354, 353, 343, 337, 329, 323
Deoxyschizandrin ^a	5.36	417.2276	C24H32O6	1	+H, +Na	402, 386, 370, 371, 355, 347, 332, 316
Schisandrin B ^a	6.33	401.1962	C23H28O6	0.8	+H, +Na	386, 371, 370, 355, 354, 340, 339, 331, 329, 316, 300, 285, 273, 270
Schisandrin C ^a	6.96	385.1642	C22H24O6	-0.8	+H, +Na	370, 355, 325, 315, 300, 285, 257, 227

Table S1. Identification of compounds detected in positive mode

^a Compounds were compared with reference compounds.







Figure S3. The MS/MS spectrum of 10 prototype compounds in dosed plasma (A), lignans purification (B) and standard (C)