

## **Supplementary Materials**

### **Rapid full-cycle technique to control adulteration of meat products: Integration of accelerated sample preparation, recombinase polymerase amplification, and test-strip detection**

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#### CONTENT

<b>Section S1. Sequences of DNA amplicons and primers for cytochrome B of chicken and pig.....</b>	<b>2</b>
<b>Section S2. Characterization of primer pairs with real-time PCR.....</b>	<b>4</b>
<b>Section S3. Verification of the developed test with meat samples.....</b>	<b>6</b>
<b>Section S4. Synthesis of gold nanoparticles.....</b>	<b>7</b>

## Section S1. Sequences of DNA amplicons and primers for cytochrome B of chicken and pig

### Cyt B chicken:

ATGGCACCCAACATTCGAAAATCCCACCCCCTACTAAAAATAATTAACAACCTCCCTA  
ATCGACCTCCCAGCCCCATCCAACATCTCTGCTTGATGAAATTTTCGGCTC **CCTATTAG**  
**CAGTCTGCCTCATGACC**CAAATCCTCACCGGCCTACTACTAGCCATGCACTACACAG  
CAGACACATCCCTAGCCTTCTCCTCCGTAGCCCACACTTGCCGGAACGTACAATACG  
GCTGACTCATCC **GGAATCTCCACGCAAACGGCGCCTC**ATTCTTCTTCATCTGTATCTT  
CCT **TCACATCGGACGAGGCCTA TACTAG**GGCTCCTACCTCTACAAGGAAACCTGAAA  
CACAGGAGTAATCCTCCTCCTCACACTCATAGCCACCGCCTTTGTGGGCTATGTTCTC  
CCATGGGGCCAAATATCATTCTGAGGGGCCACCGTTATCACAAACCTATTCTCAGCA  
ATTCCCTACATTGGACACACCCTAGTAGAGTGAGCCTGAGGGGGATTTCAGTCGAC  
AACCCAACCCTTACCCGATTCTTCGCTTTACACTTCCCTCCTCCCCTTTGCAATCGCAG  
GTATTACTATCATCCACCTCACCTCCTACACGAATCAGGCTCAAACAACCCCTAG  
GCATCTCATCCGACTCTGACAAAATTCCATTTACCCATACTACTC **CTTCAAAGACA**  
**TTCTGGGCTTAACTC**TCATACTCACCCATTCCCTAACACTAGCCCTATTCTCCCCAA  
CCTCCTAGGAGACCCAGAAAACCTCACCCAGCAAACCCACTAGTAACCCCCCCAC  
ATATCAAACCAGAATGATATTTTCTATTCGCCTATGCCATCCTACGCTCCATCCCCAA  
CAAACCTGGAGGTGTACTAGCCCTAGCAGCCTCAGTCCTCATCCTCTTCTAATCCC  
CTTCTCCACAAATCTAAACAACGAACAATAACCTTCCGACCACTCTCCCAAACCT  
ATTCTGACTTCTAGTAGCCAACCTTCTTATCCTAACCTGAATCGGAAGCCAACCAGT  
AGAACACCCTTCATCATCATTGGCCAAATAGCATCCCTCTCTTACTTCACCATCCTA  
CTTATCCTCTTCCCC **ACAATCGGAACACTAGAAAACAAAAT**ACTCAACTACTAA

Primer annealing sites marked by: underline for F1c, yellow for F2c, green for F3c, cyan for F4c, double underline for R1c, magenta for R3c, red for R4c

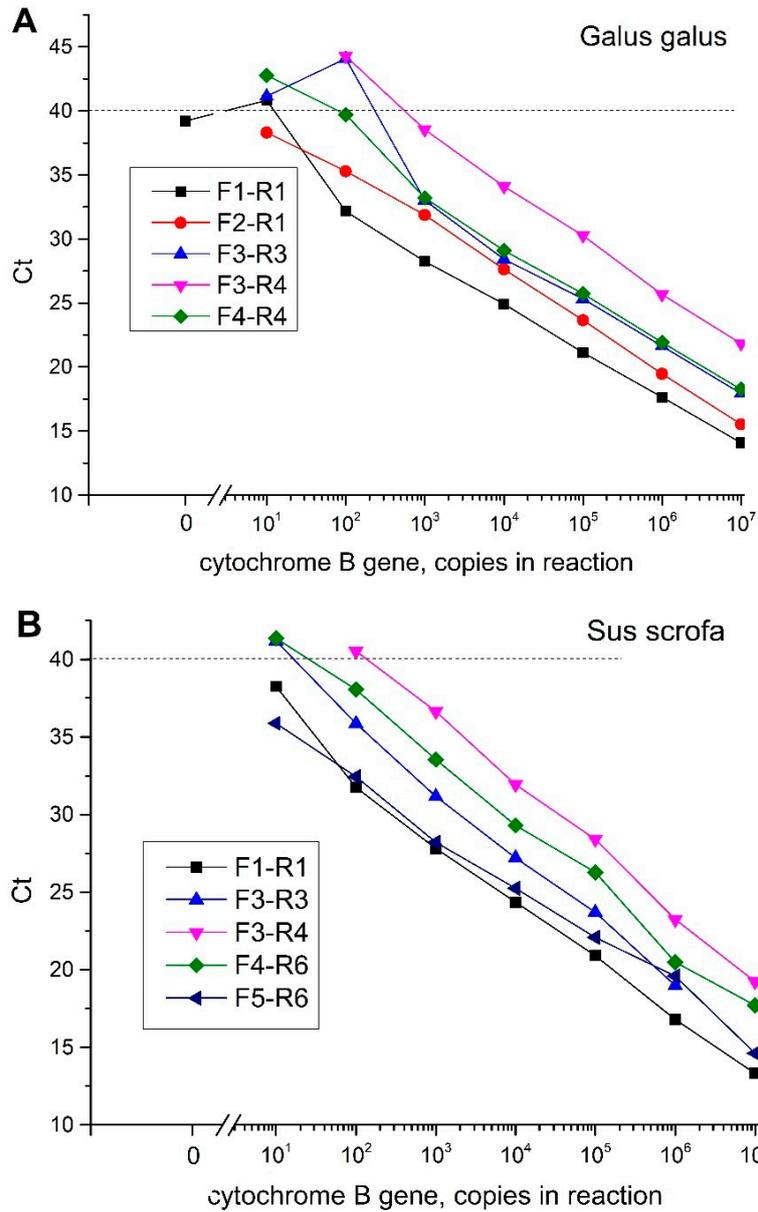
### Cyt B pig:

ATGACCAACATCCGAAAATCACACCCACTAATAAAAATTATCAACAACGCATTTCATT  
**GACCTCCCAGCCCCCTCAAACATCTCATCATGATGAAA**CTTCGGTTCCTCTTAGGC  
ATCTGCCTAATCTTGCAAATCCTAACAGGCCTGTTCTTAGCAATACATTACACATCA  
GACAC **AACAACAGCTTCTCATCAGTTACA**CACATTTGTCGAGACGT **AAATTACGGA**  
**TGAGTTATTCGCTATC**TACATGCAAACGGAGCATCCATATTCTTTATTTGCCTATTCA  
TCCACGTAGGCCGAGGTCTATACTACGGATCCTATATATTCTTAGAAACATGAAACA  
TTGGAGTAGTCTACTATTTACCGTTATAGCAACAGCCTTCATAGGCTACGTCTGC  
CCTGAGGACAAATATCATTCTGAGGAGC **TACGGTCATCACAAATCTACTATCAGC**TA  
TCCCTTATATCGGAACAGACCTCGTAGAATGAATCTGAGGGGGCTTTTCCGTGACAA  
AAGCAACCCTCACACGATTCTTCGCCTTCCACTTTATCCTGCCATTCATCATTACCGC  
CCTCGC **AGCCGTACATCTCCTATTCTGCAC**GAAACCGGATCCAACAACCCTACCGG  
AATCTCATCAGACATAGACAAAATTCCATTTACCCATACTACACTATT **AAAGACAT**  
**TCTAGGAGCCTTATTTA**TAATACTAATCCTACTAATCCTTGTAATTTCTCACCAGAC  
CTACTAGGAGACCCAGACAACCTACACCCAGCAAACCCACTAAACACCCACCCCA  
TATTAACCAAGAAATGATATTTCTTATTCGCCTACGCTATTCTACGTTCAATTCTAAT  
AAACTAGGTGGAGT **GTTGGCCCTAGT** **AGCCTCCATCCTAATCCTAATTTTA**ATGCC  
ATACTACACACATCCAAACAACGAGGCATAATATTTTCGACCACTAAGTCAATGCCTA  
TTCTGAATACTAGTAGCAGACCTCATTACACTAACATGAATTGGAGGACAACCCGTA

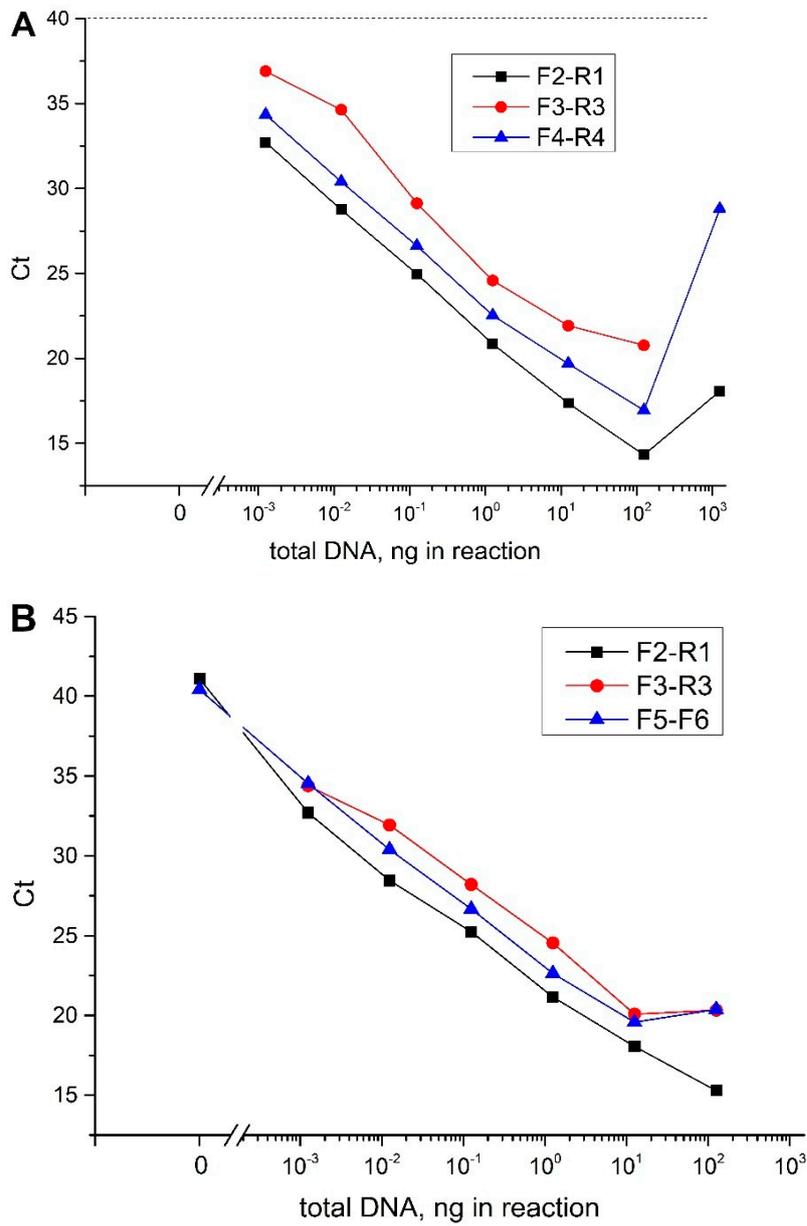
GAACACCCGTTTCATCATCATCGGCCAACTAGCCTCCATCTTATACTTCCTAATCATTC  
TAGTATTGATACCAATCACTAGCATCATCGAAAACAACCTATTAAAATGAAGA

Primer annealing sites marked by: yellow for F1p, green for F2p, cyan for F3p, magenta for F4p,  
red for F5p, olive for R1p, dark grey for R3p, double underline for R4p, brown-green for R6p

## Section S2. Characterization of primer pairs with real-time PCR



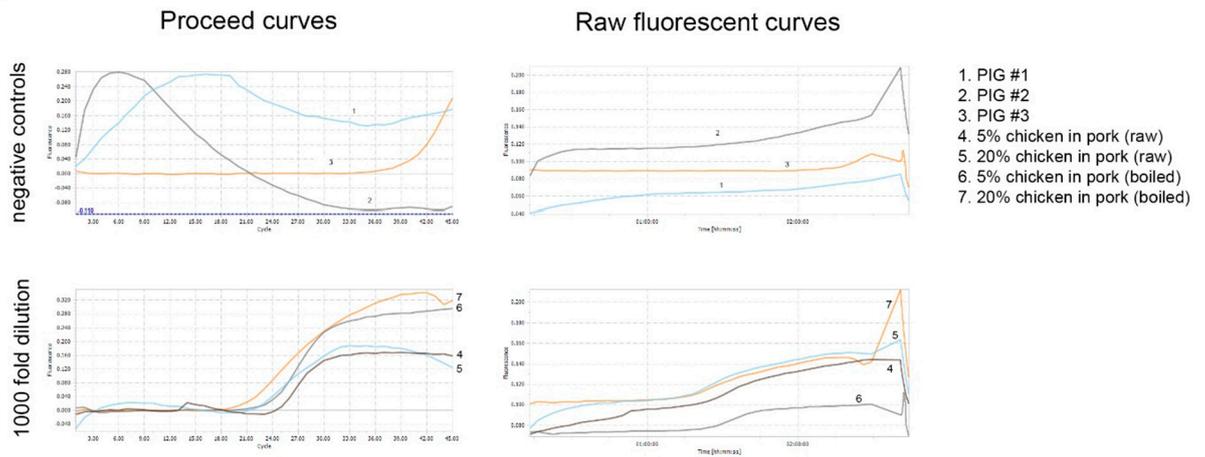
**Figure S1.** Sensitivity of qPCR with different primers for detection of purified cytB gene of chicken (A) and pig (B).



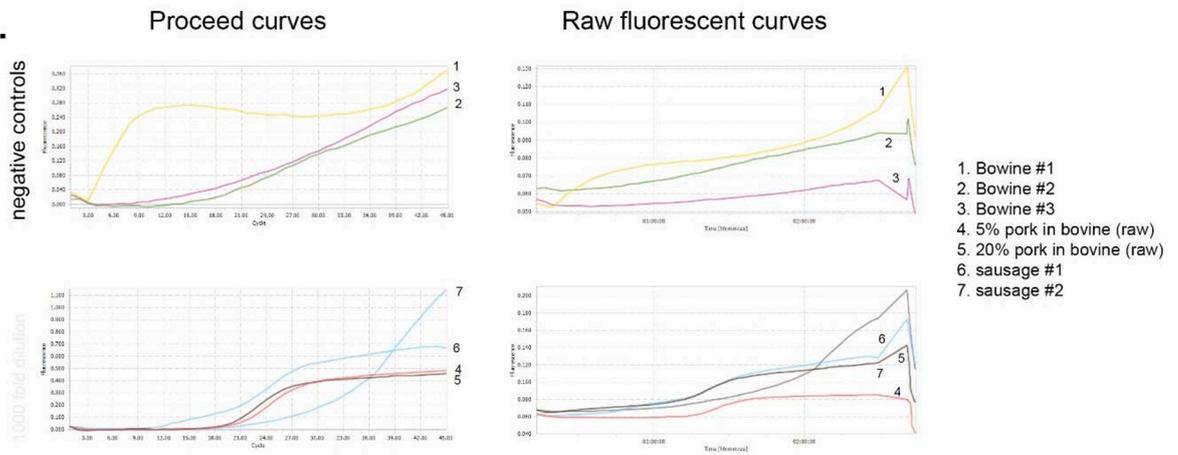
**Figure S2.** Sensitivity of qPCR with different primers for detection of total DNA extracted from chicken (A) and pig (B) meat.

## Section S3. Verification of the developed test with meat samples

A.



B.



**Figure S3.** Detection of chicken (A) or pig (B) DNA in meat samples (after accurate DNA extraction) by qPCR.

#### **Section S4. Synthesis of gold nanoparticles**

We added 1 mL of 1% HAuCl<sub>4</sub> to 95 mL of deionized water and heated to boiling point, then 4 mL of 1% sodium citrate was added while the mixture was stirred. The mixture was boiled for 25 min then cooled and stored at 4°C.

The size and shape of the synthesized GNP were analyzed by dynamic laser scattering (DLS) and imaging with transmission electron microscopy (TEM). DLS was performed by ZetaSizer Nano (Malvern, USA) and showed average Z-size of the GNPs as 20 nm. TEM was performed by JEM CX-100 electron microscope (Jeol, Japan) and confirmed the size of the GNPs. Concentration of GNPs was estimated by optical absorption at 520 nm with Lybra Biochrome spectrometer (USA).