

Supplementary Materials

**Sensitive assay for the lactonase activity of serum paraoxonase 1 (PON1) by
harnessing the fluorescence turn-on characteristics of bioorthogonally
synthesized and geometrically controlled chemical probes**

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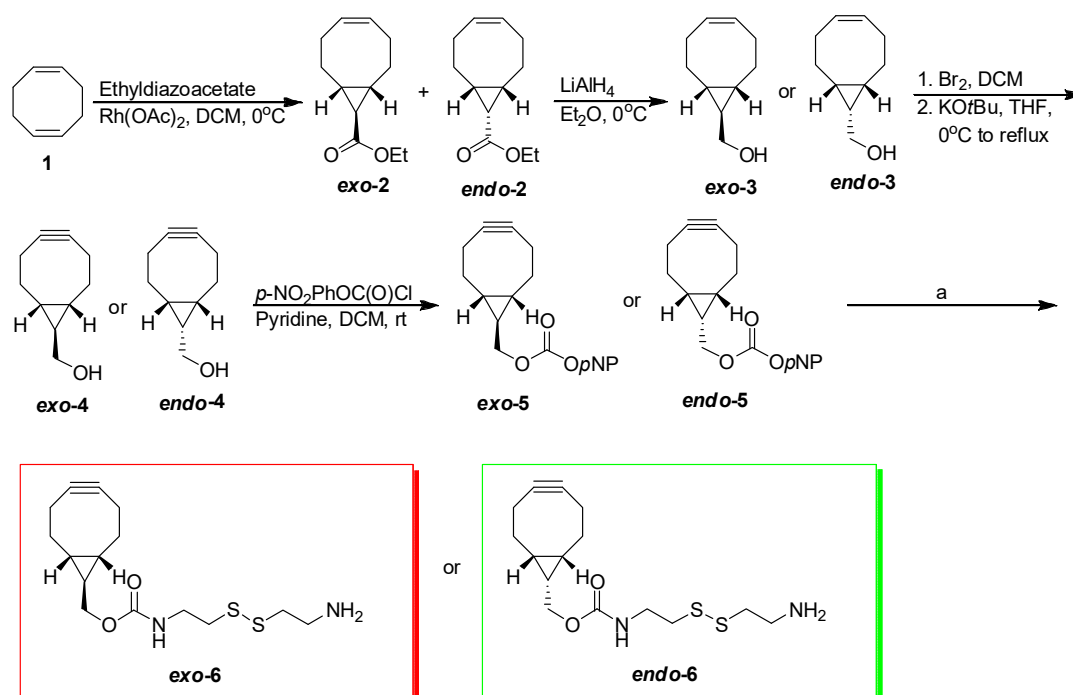
[†]Auhors contributed equally to this study.

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Scheme



Scheme S1. Synthesis of the key bicyclononyne derivatives (**6**). Syntheses of **2** - **5**

were reported previously [1, 2]. a: cystamine, Et₃N, MeOH/DCM, 50°C.

Figures.

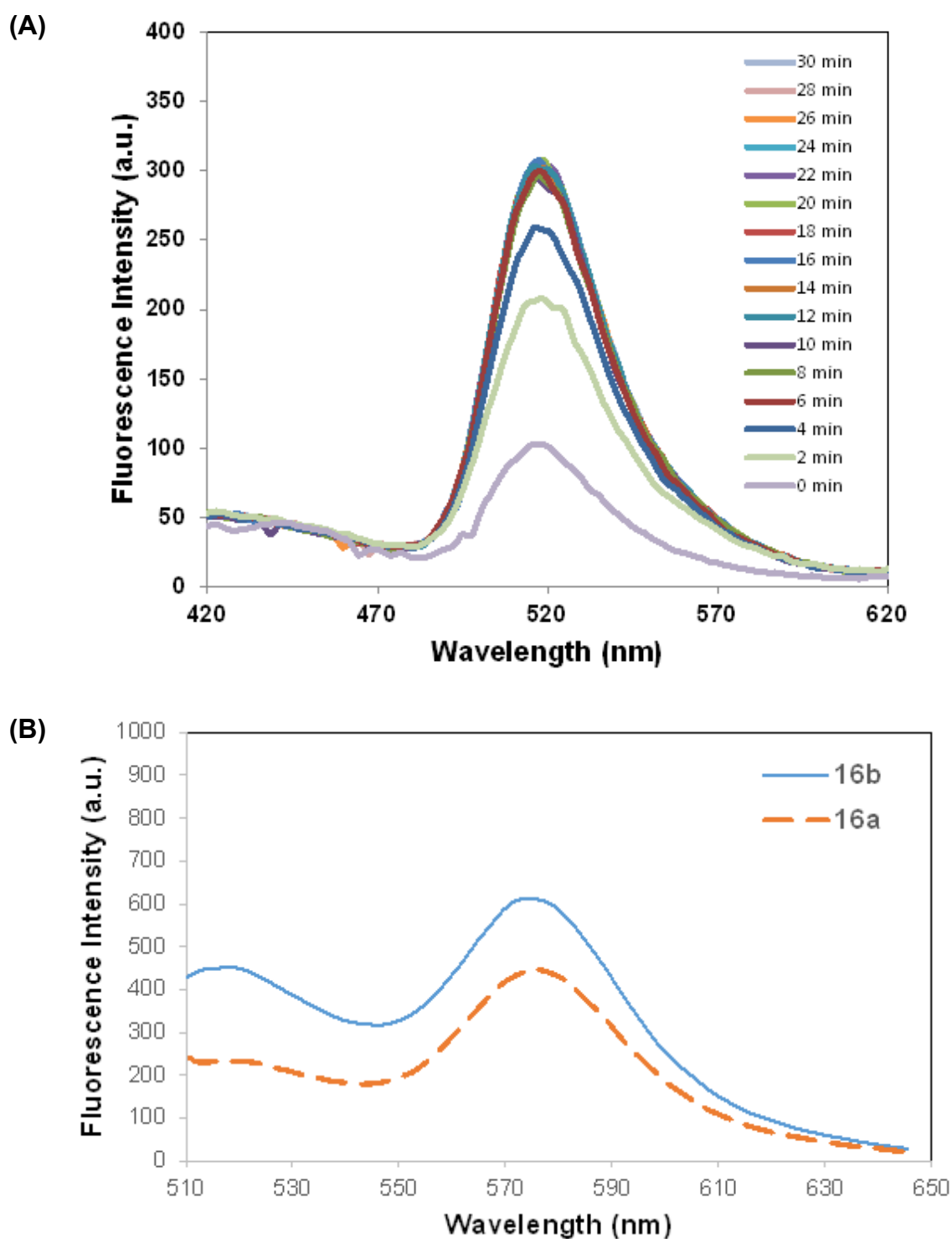


Figure S1. The fluorogenic property of the chemical probe **16a** when reacting with a thiol and the presence of residual FRET effects in **16**. (A) The time-dependent increase of the 5-FAM fluorescence in a reaction of **16a** (1 μ M) with L-cysteine (50 mM) in phosphate buffer (PB; 10% DMF, pH 7.4). (B) The fluorescence spectra of **16** (4 μ M each) in glacial acetic acid containing 0.8% of DMF. The excitation wavelength for **16a** was 494 nm and the excitation wavelength for **16b** was 491 nm.

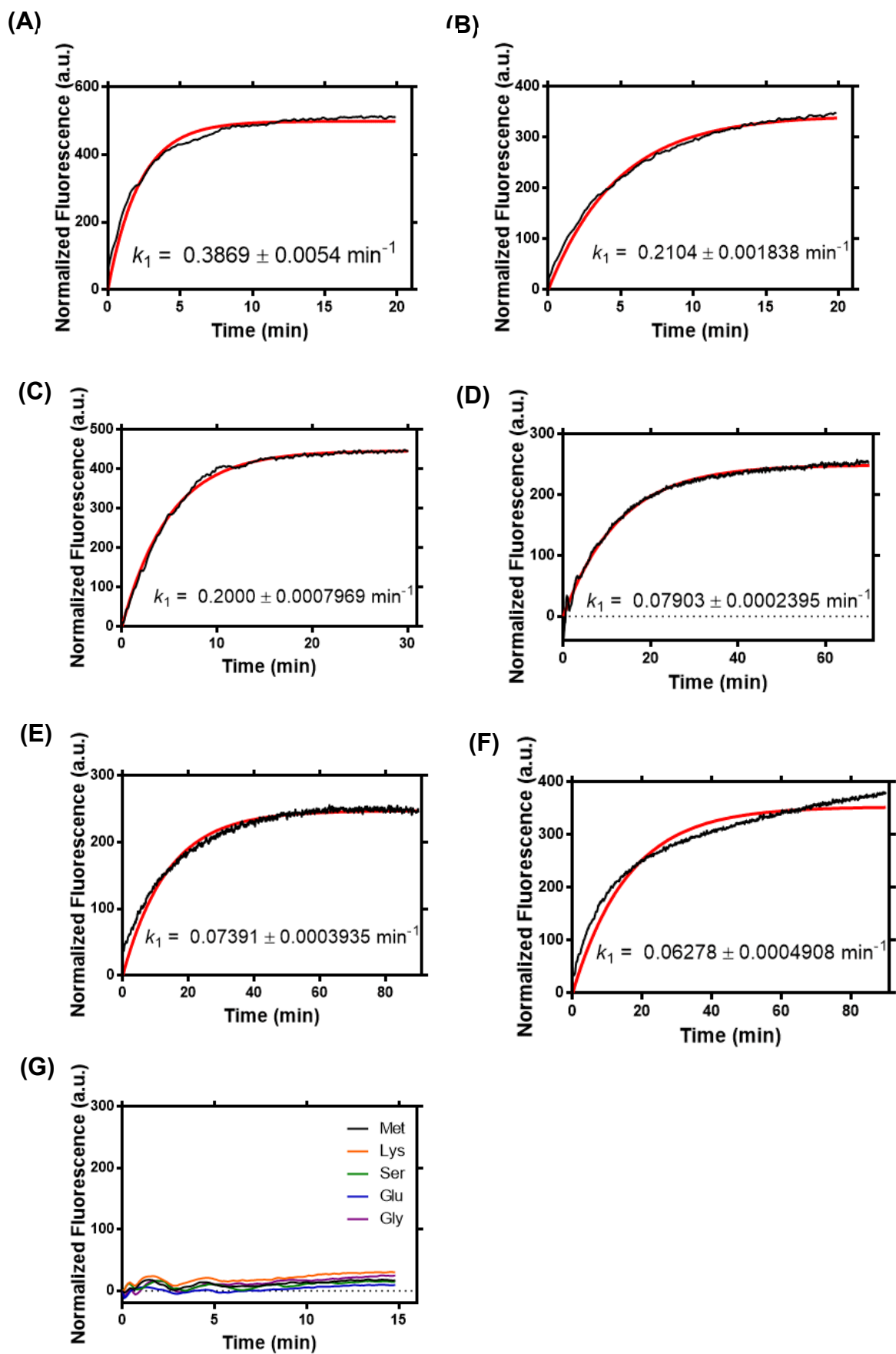
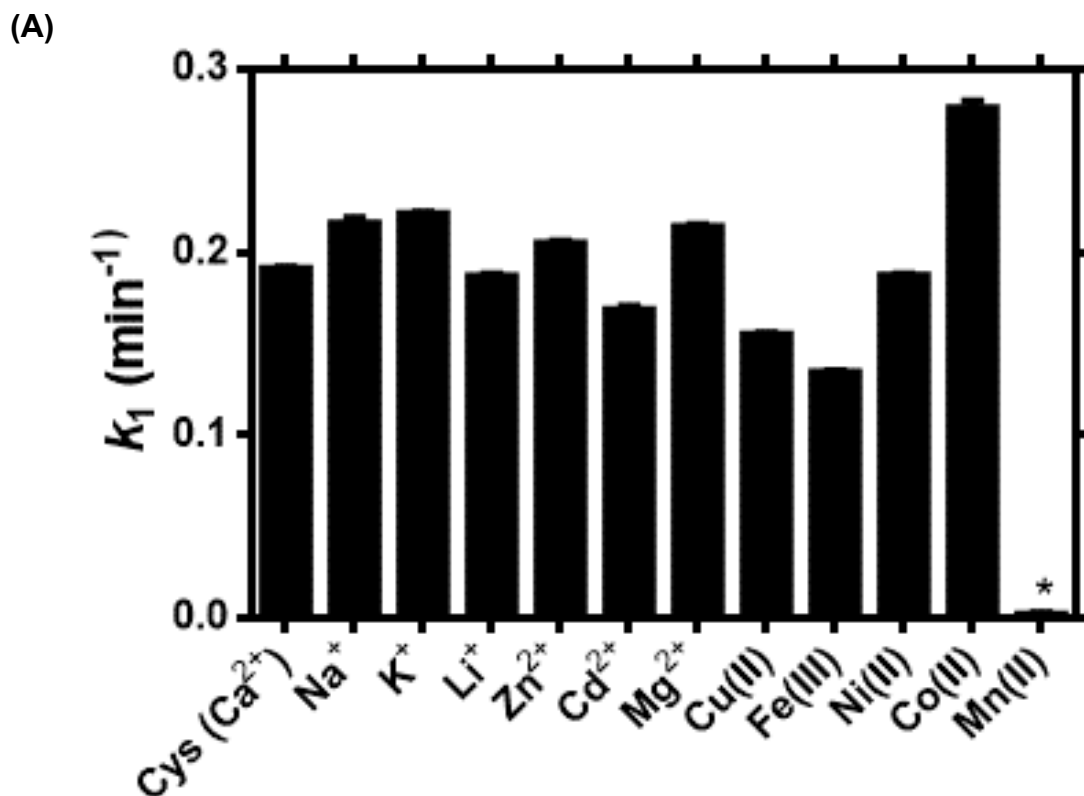
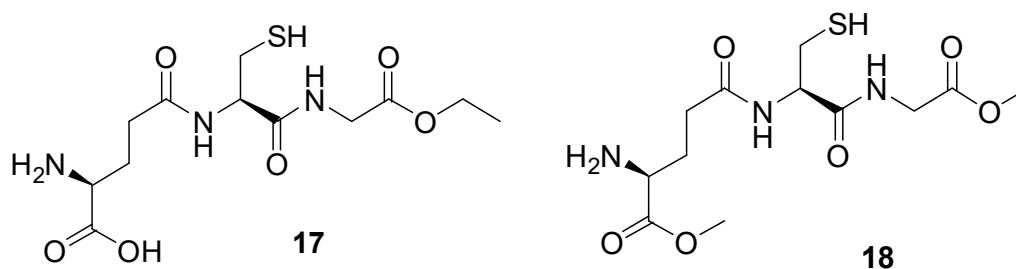


Figure S2. Representative pseudo-first-order reactions of **16b** (0.4 μM) with 5 mM of (A) 2-AET, (B) DTT, (C) L-cysteine, (D) GSH, (E) nBuSH, (F) 2-ThioEtOH, or (G) five non-thiol amino acids (L-methionine, L-lysine, L-serine, L-glutamate, and glycine) in the Tris buffer (50 mM Tris, 1 mM Ca^{2+} , pH 8.0). Progress of each reaction was monitored by measuring fluorescence emission at 525 nm at specific time intervals. Data of normalized 6-FAM fluorescence intensity (a.u.) vs time were fitted to a single-exponential equation for first-order kinetics $F(t) = F_0 + F_{\text{max}}(1 - e^{-k_1 t})$ [$F(t)$, 6-FAM fluorescence at a specific time point t] to provide the values of first-order rate constant k_1 (GraphPad, La Jolla, CA, USA) illustrated in the graphs. In Panels S3A-S3F, the black lines indicate original fluorescence changes in the reaction time courses, and the red curves show the results calculated by the single-exponential equation. The normalized fluorescence intensity data at 525 nm were acquired by subtracting a background fluorescence of **16b** from the original fluorescence intensity measurements. 2-AET, 2-aminoethanethiol; nBuSH, 1-butanethiol; 2-ThioEtOH, 2-mercaptoethanol.



(B)



(C)

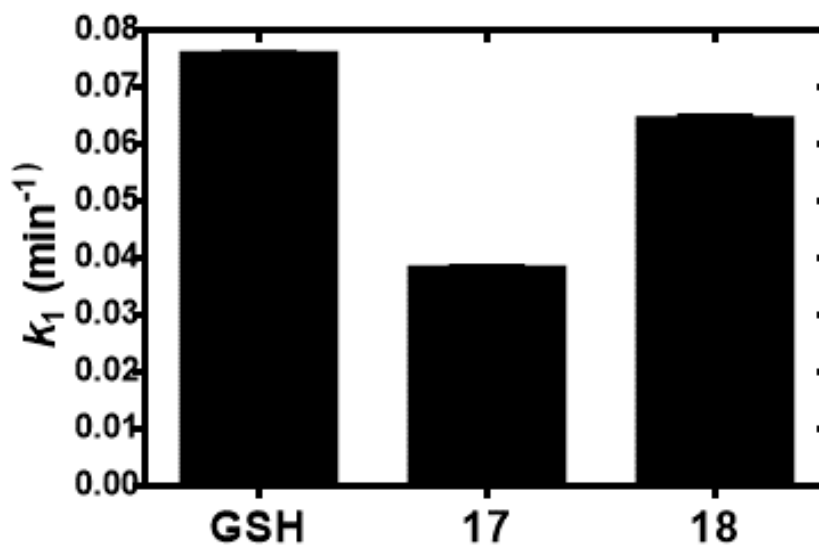


Figure S3. Influence of metal ions and steric constraints of **16b** on the thiol-dependent fluorogenic reaction of **16b**. (A) Effects of metal ion (1 mM) on the pseudo-first-order reactions of **16b** (0.4 μ M) with 5 mM L-cysteine in the Tris buffer. The k_1 values were determined by a method analogous to what was described in Figure S2. The symbol * indicates that **16b** had no detectable 6-FAM fluorescence change in the presence of Mn(II) with which the 6-FAM fluorescence change was so insignificant that the values of averaged k_1 and standard deviation could not be determined. (B) Structures of two GSH derivatives **17** and **18** [2]. (C) Kinetic analysis of the pseudo-first-order reactions of **16b** (0.4 μ M) with 5 mM of one of the structure-modified GSH derivatives **17** and **18** in the Tris buffer. The k_1 values were again determined by a method similar to that described in Figure S2. In addition, each reaction was analyzed at least three times in order to acquire the averaged k_1 values and standard deviation (the error bar).

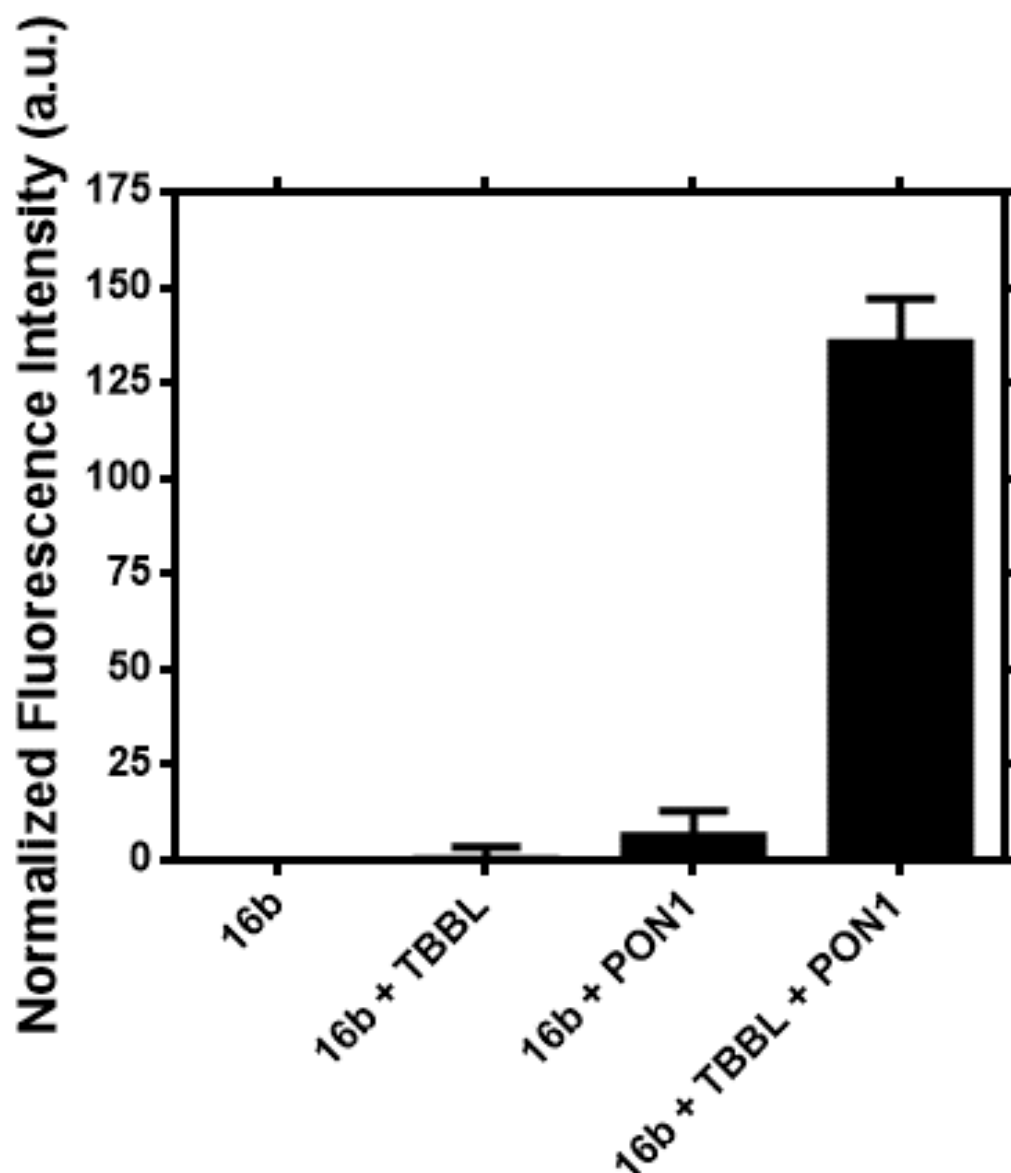


Figure S4. Presence of PON1 and TBBL is required for the fluorogenic property of **16b**. PON1 lactonase activity was determined by measuring the 6-FAM fluorescence released from **16b** in the spectrofluorometer. A **16b** ($0.8 \mu\text{M}$ in DMSO, 1.6%) solution might be supplemented with PON1 (125.6 U L^{-1} , 3.25% glycerol), TBBL (10 mM in ACN, 1%), or both in the Tris buffer. The **16b** reactions were carried out at 25°C for 30 min and immediately determined fluorescence emission afterward. The normalized fluorescence intensity data at 525 nm were acquired by subtracting a background fluorescence of **16b** at the same wavelength from the original fluorescence intensity readings. Each reaction was analyzed three times in order to acquire the averaged fluorescence values and standard deviation (the error bar).

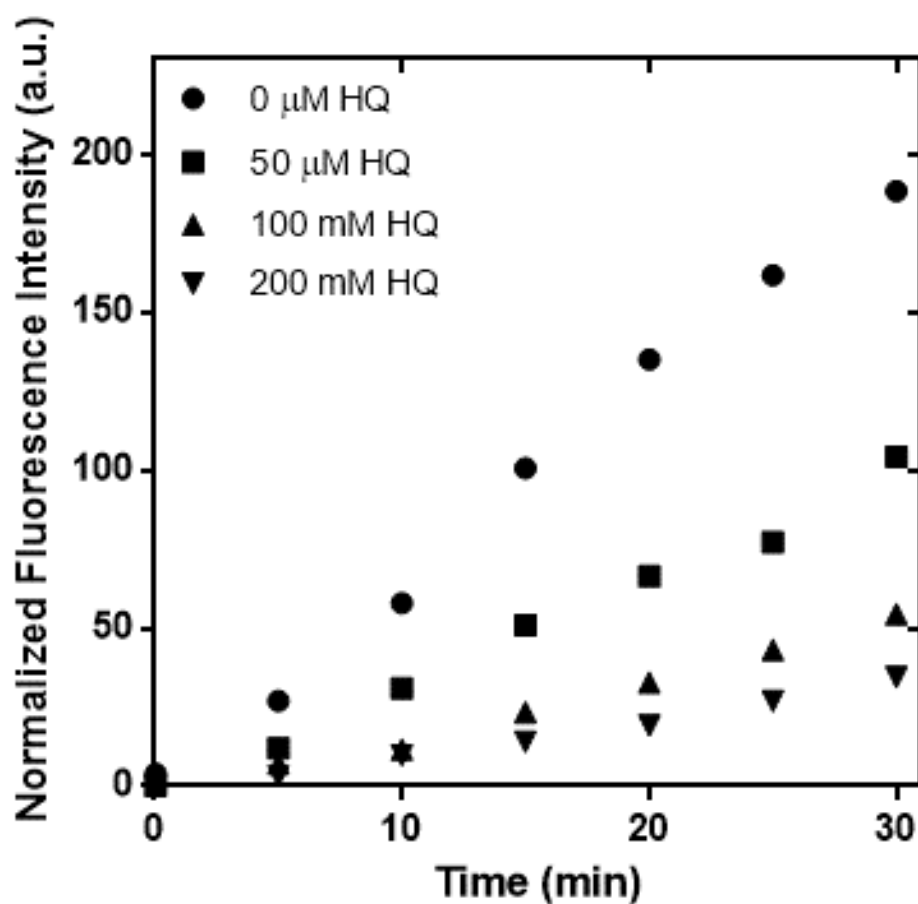


Figure S5. 2-Hydroxyquinoline (HQ) inhibition on PON1 catalysis analyzed by the fluorescence assay based on **16b**. Time-course kinetic analysis of HQ inhibition on PON1 (250.9 U L^{-1} , 1.25% glycerol) catalysis was performed in the presence of TBBL (10 mM in ACN, 1.5%) in the Tris buffer. Each PON1 reaction contained 0, 50, 100 or 200 μM of HQ in 1.5% ACN.

References

- [1] J. Dommerholt, S. Schmidt, R. Temming, L.J.A. Hendriks, F.P.J.T. Rutjes, J.C.M. van Hest, D.J. Lefeber, P. Friedl, F.L. van Delft, Readily Accessible Bicyclononynes for Bioorthogonal Labeling and Three-Dimensional Imaging of Living Cells, *Angew. Chem., Int. Ed.*, 49 (2010) 9422-9425.
- [2] M.-M. Gong, C.-Y. Dai, S. Severance, C.-C. Hwang, B.-K. Fang, H.-B. Lin, C.-H. Huang, C.-W. Ong, J.-J. Wang, P.-L. Lee, T.-P. Wang, A Bioorthogonally Synthesized and Disulfide-Containing Fluorescence Turn-On Chemical Probe for Measurements of Butyrylcholinesterase Activity and Inhibition in the Presence of Physiological Glutathione, *Catalysts*, 10 (2020) 1169.

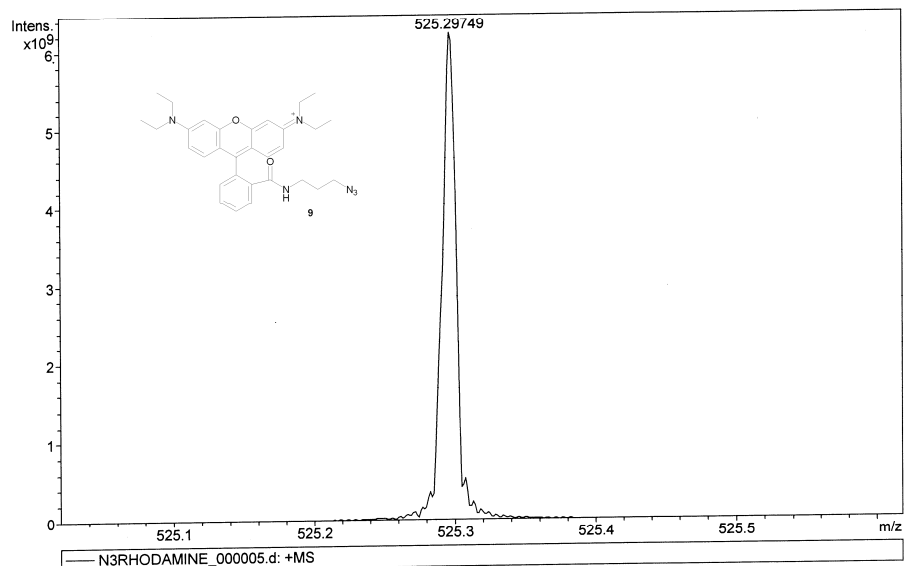
Spectra

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Comment ESI Positive

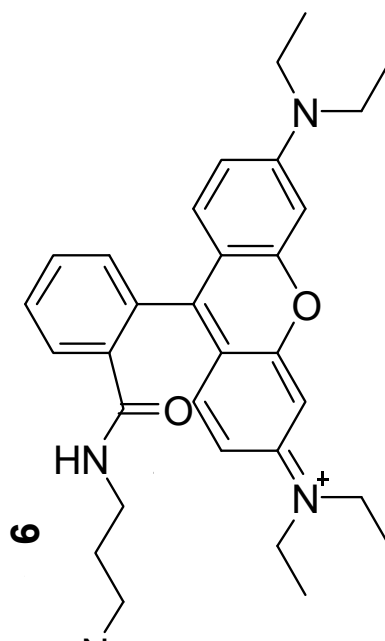
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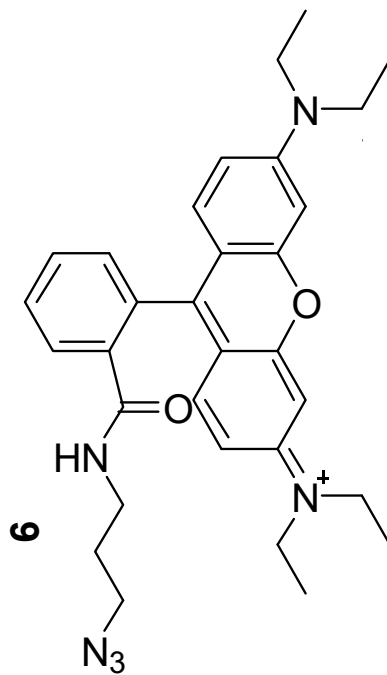
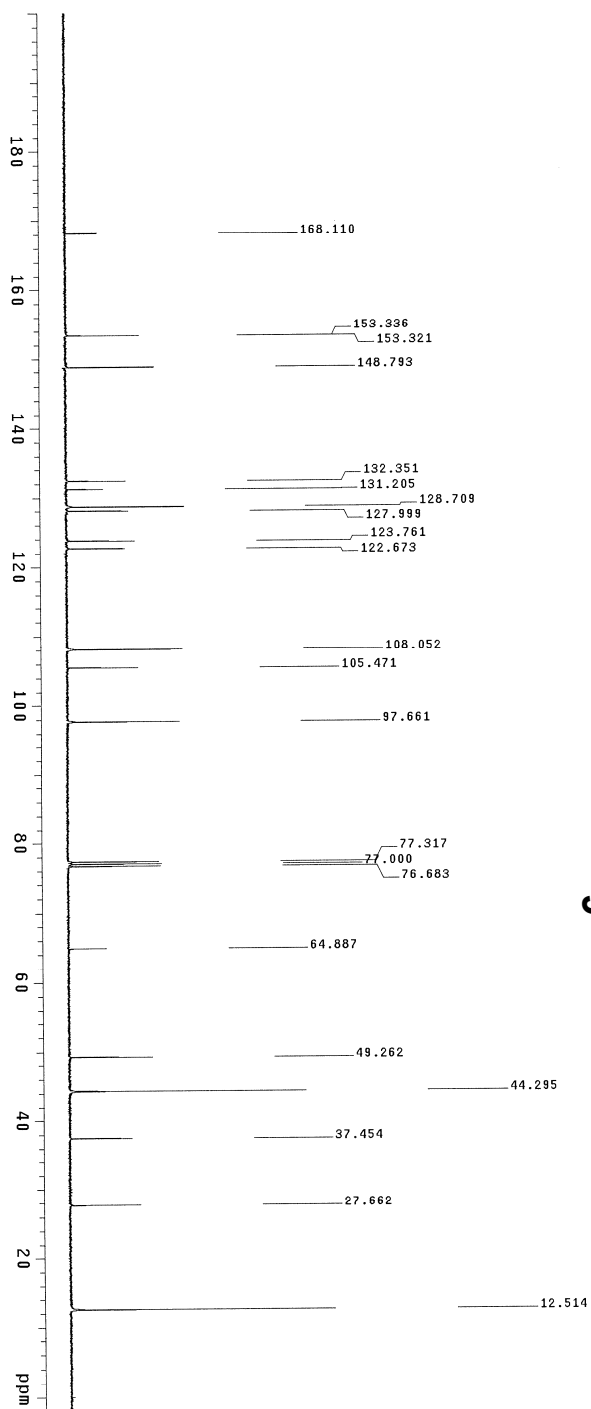


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N3-Rhodamine B

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 Ambient temperature
 Total 32 repetitions



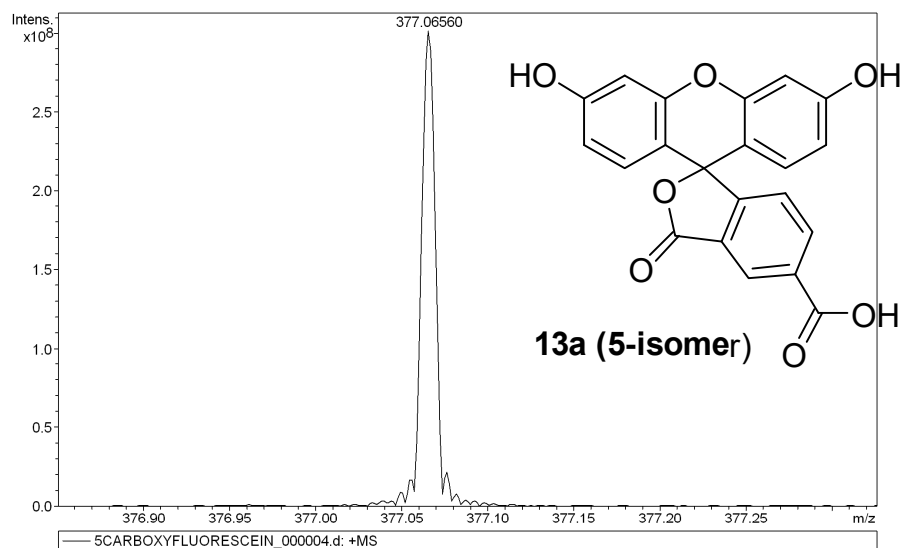


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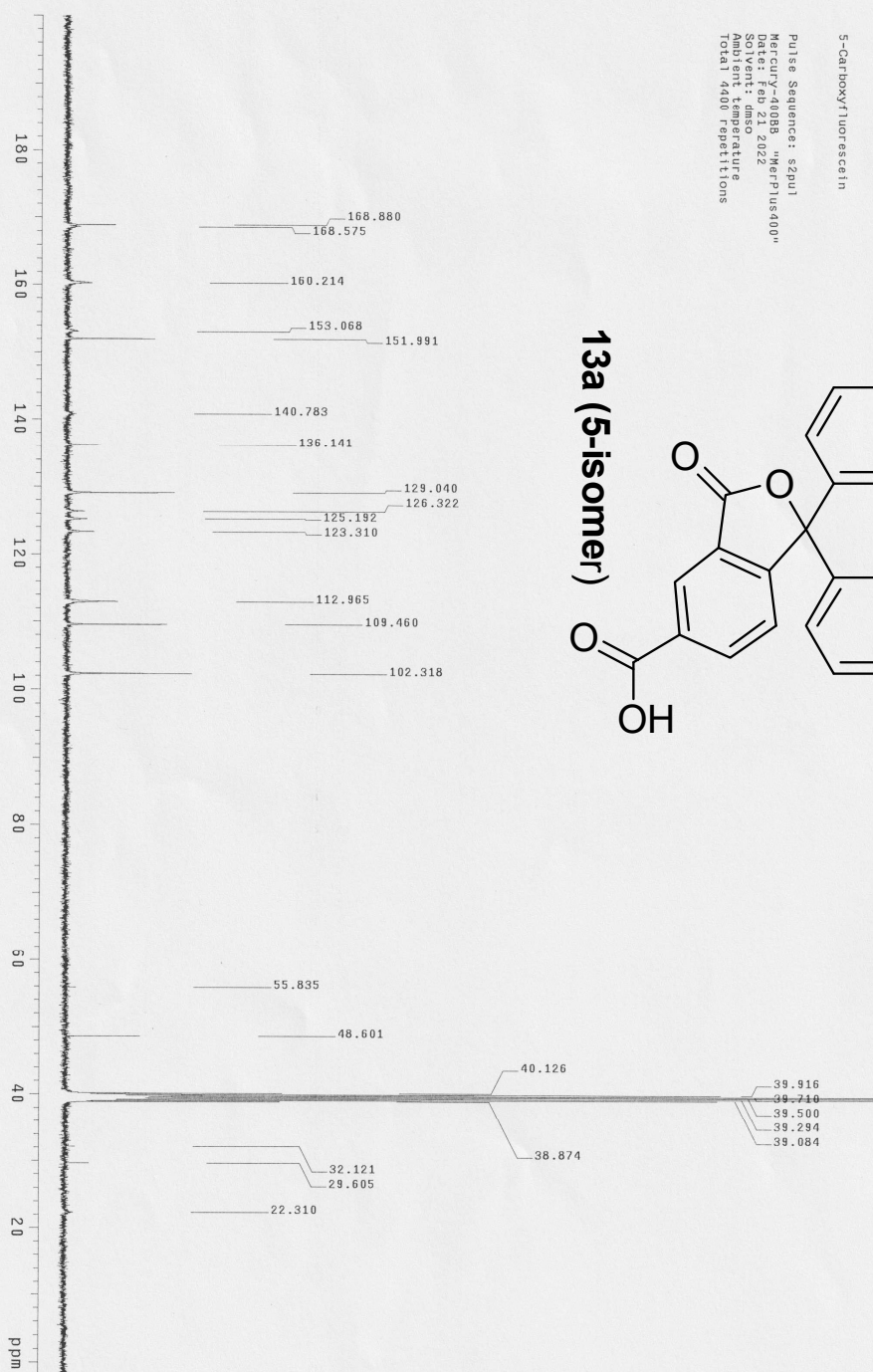
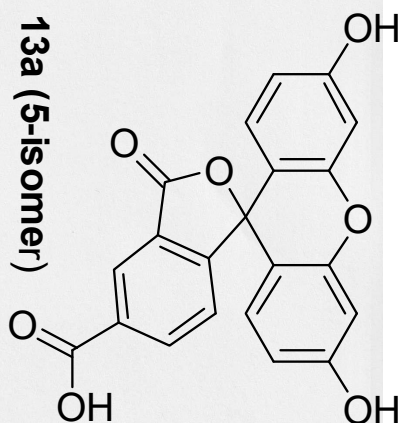
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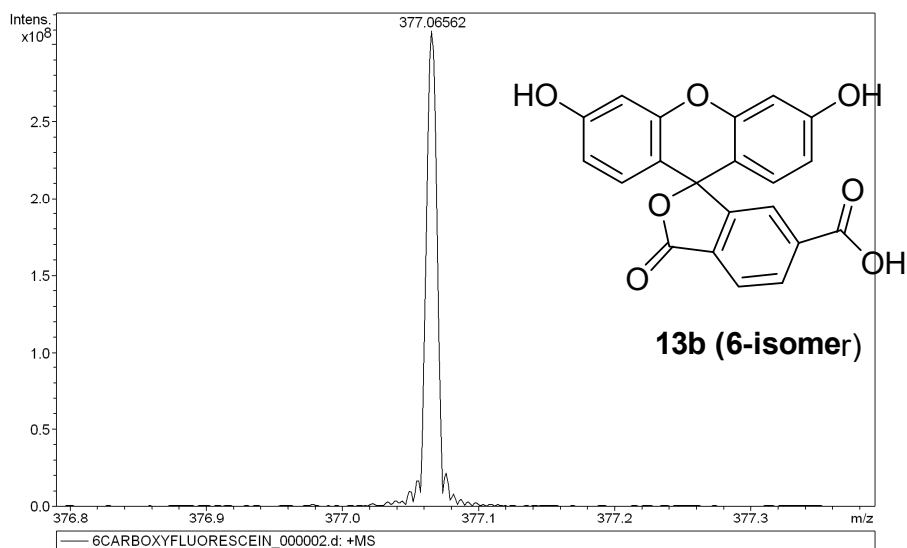


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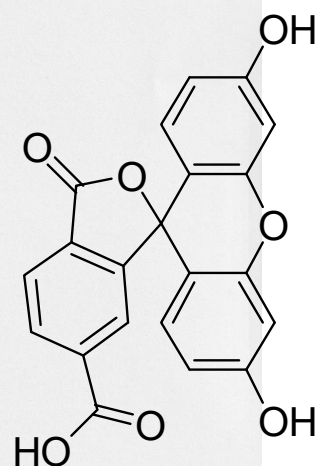
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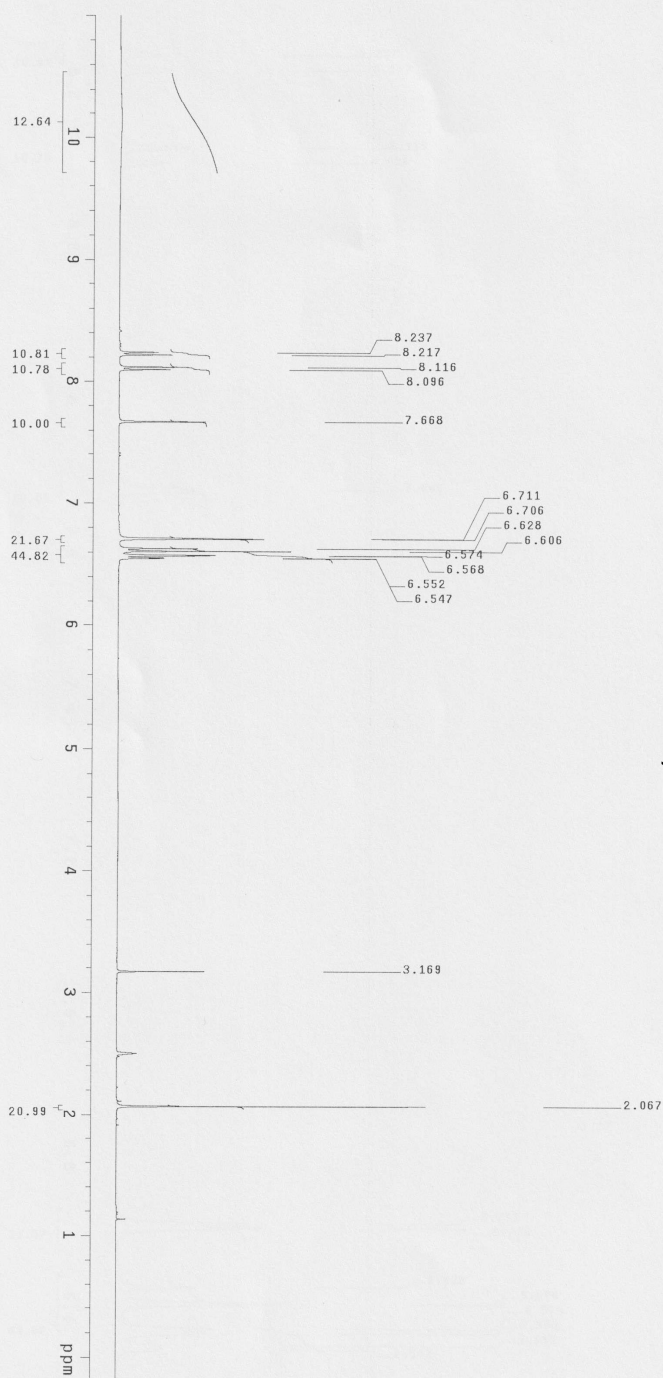


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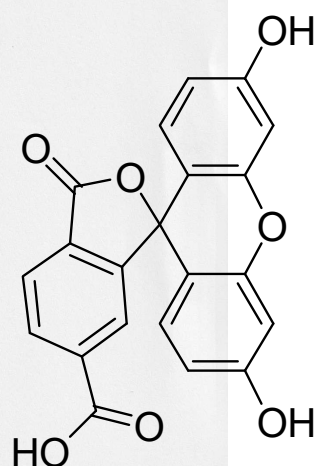


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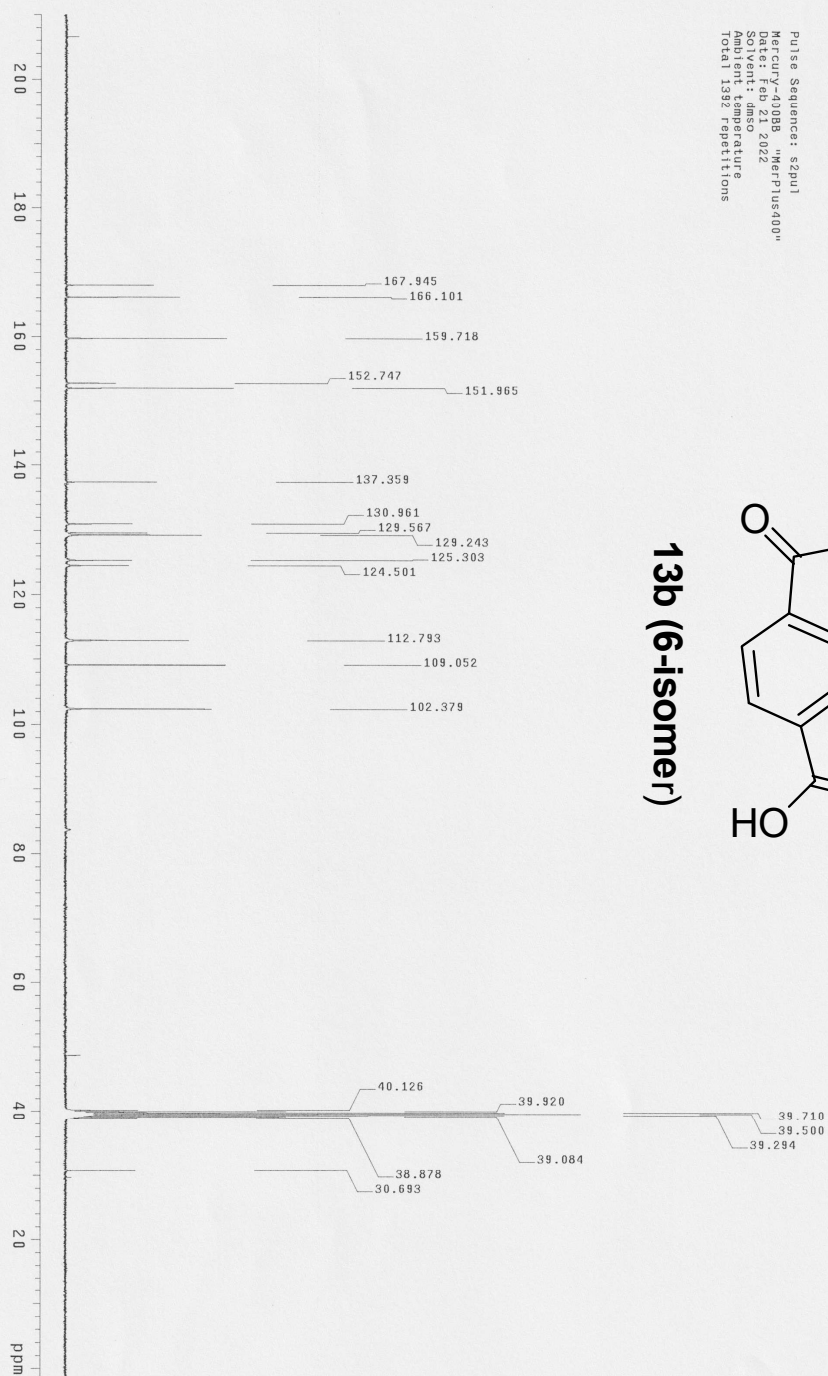
13b (6-isomer)



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 Solvent: DMSO-d6
 Ambient temperature
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13b (6-isomer)



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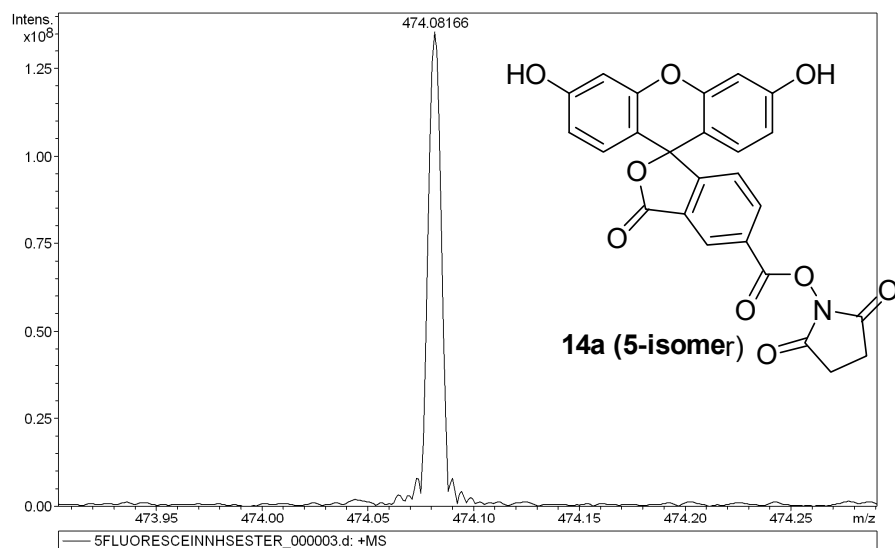
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Comment ESI Positive

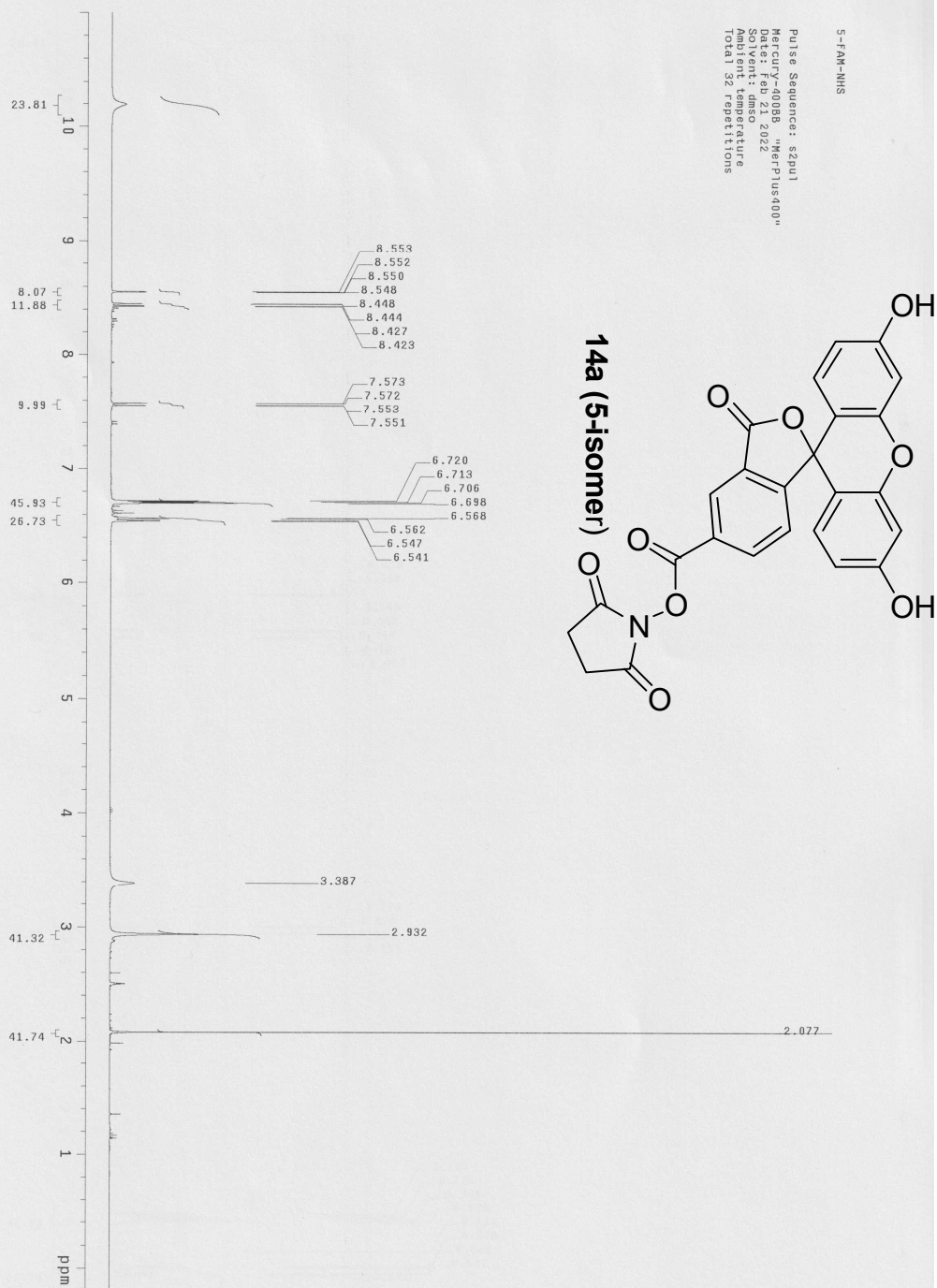
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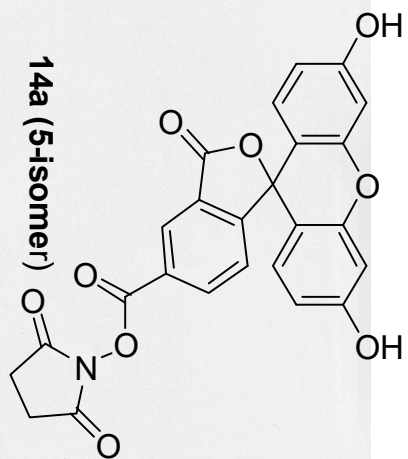
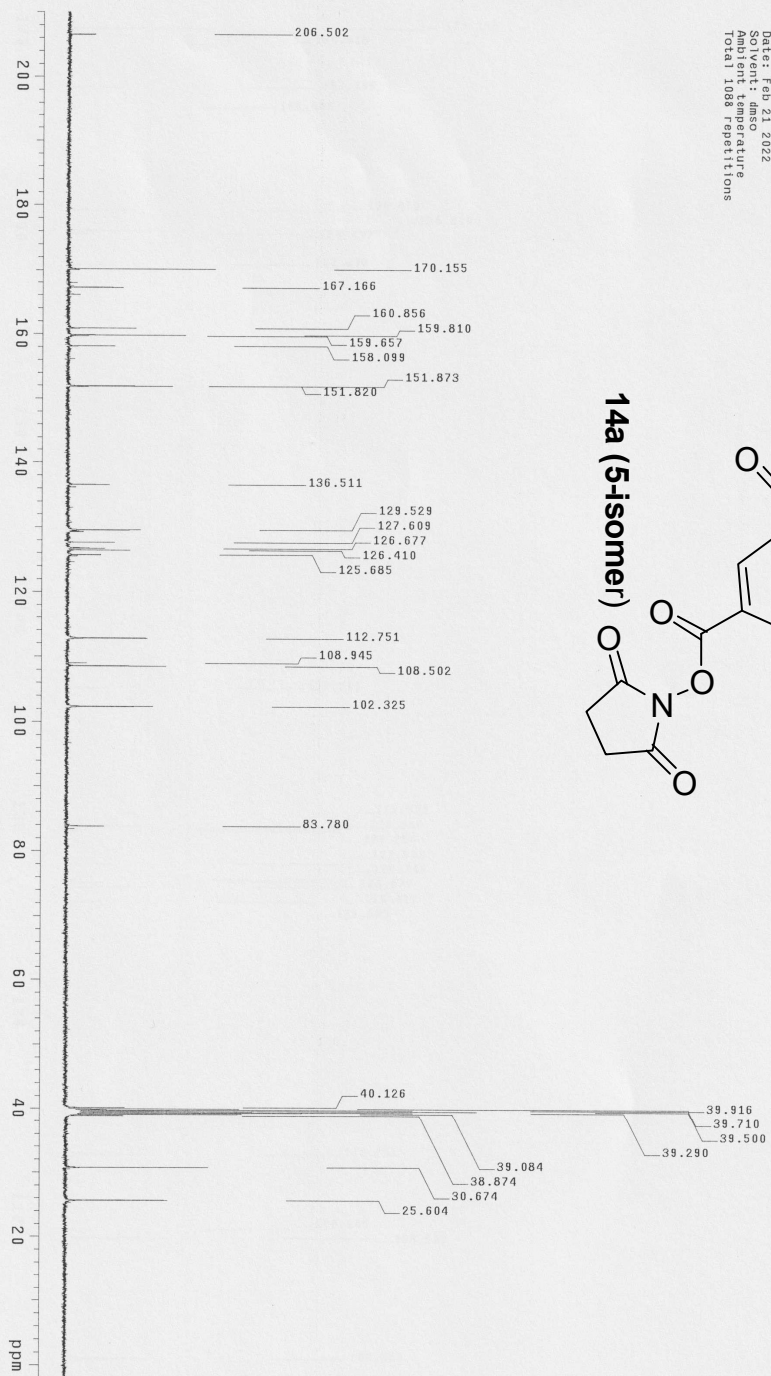
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Mass Spectrum SmartFormula Report

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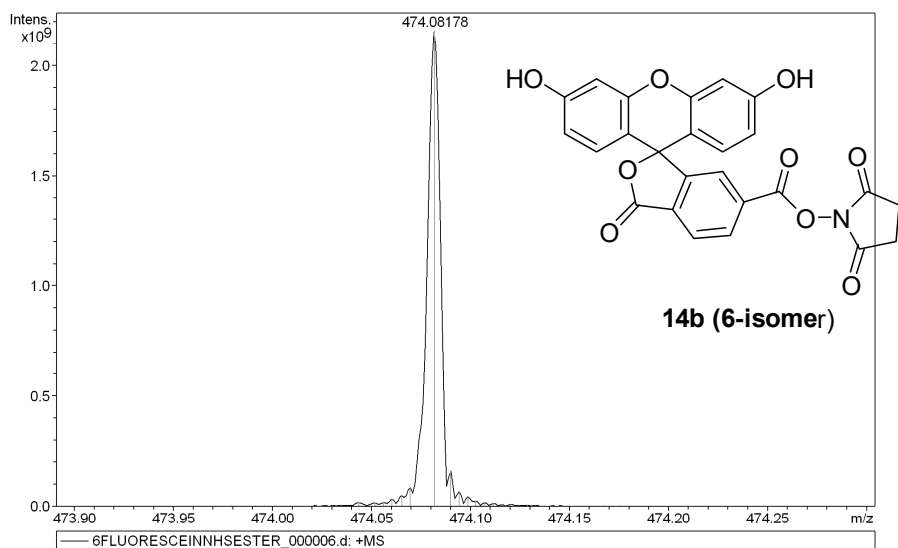
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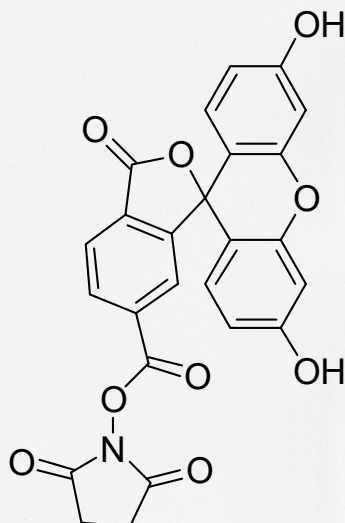
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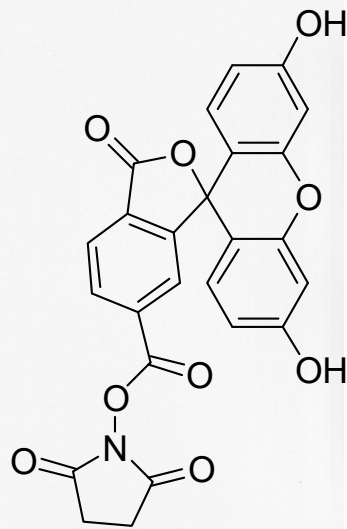
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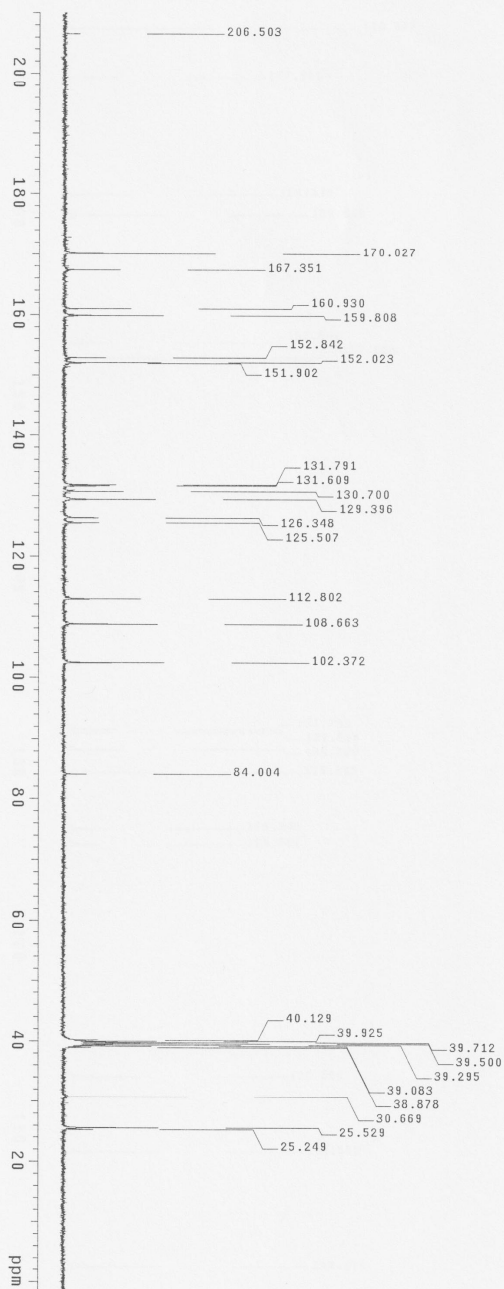
14b (6-isomer)



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14b (6-isomer)

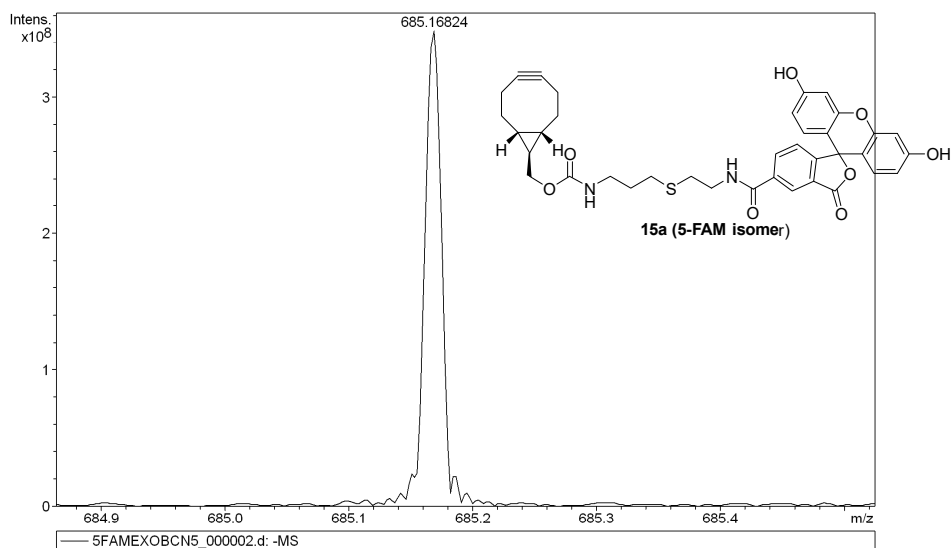


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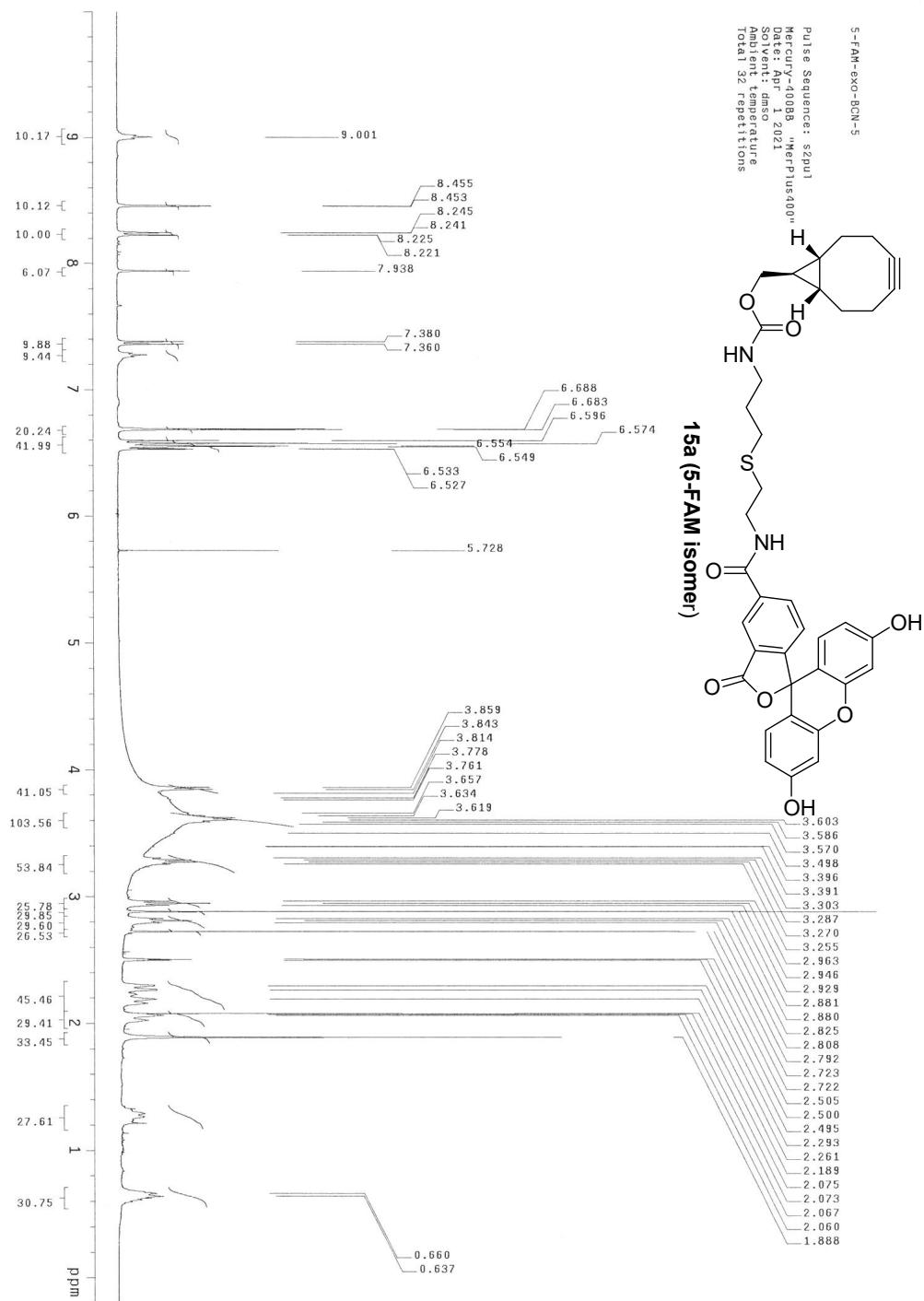
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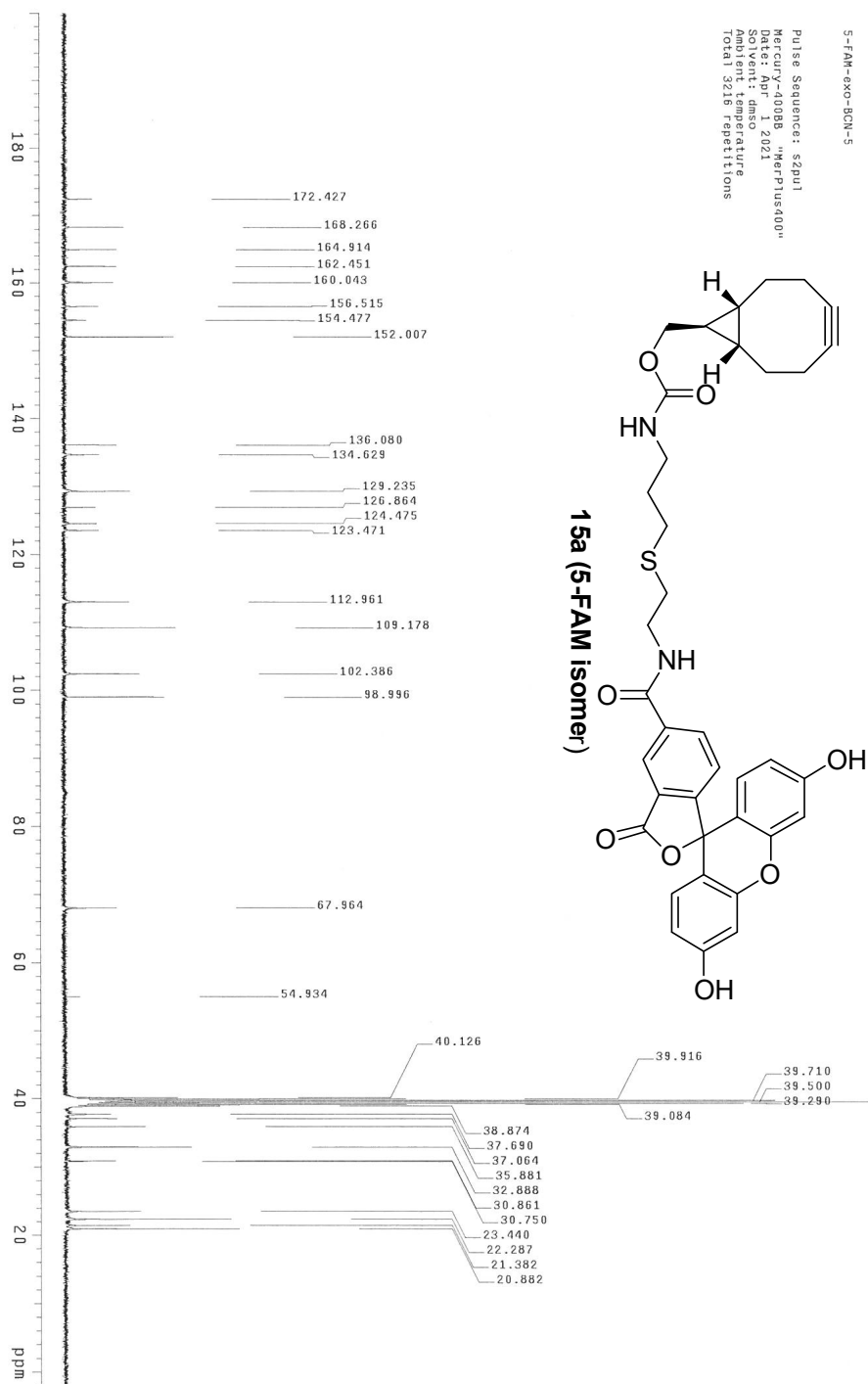
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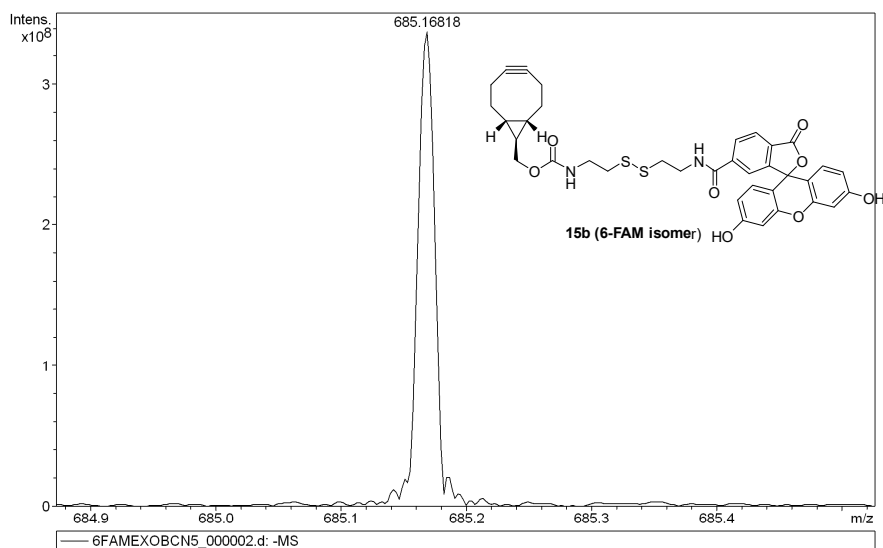


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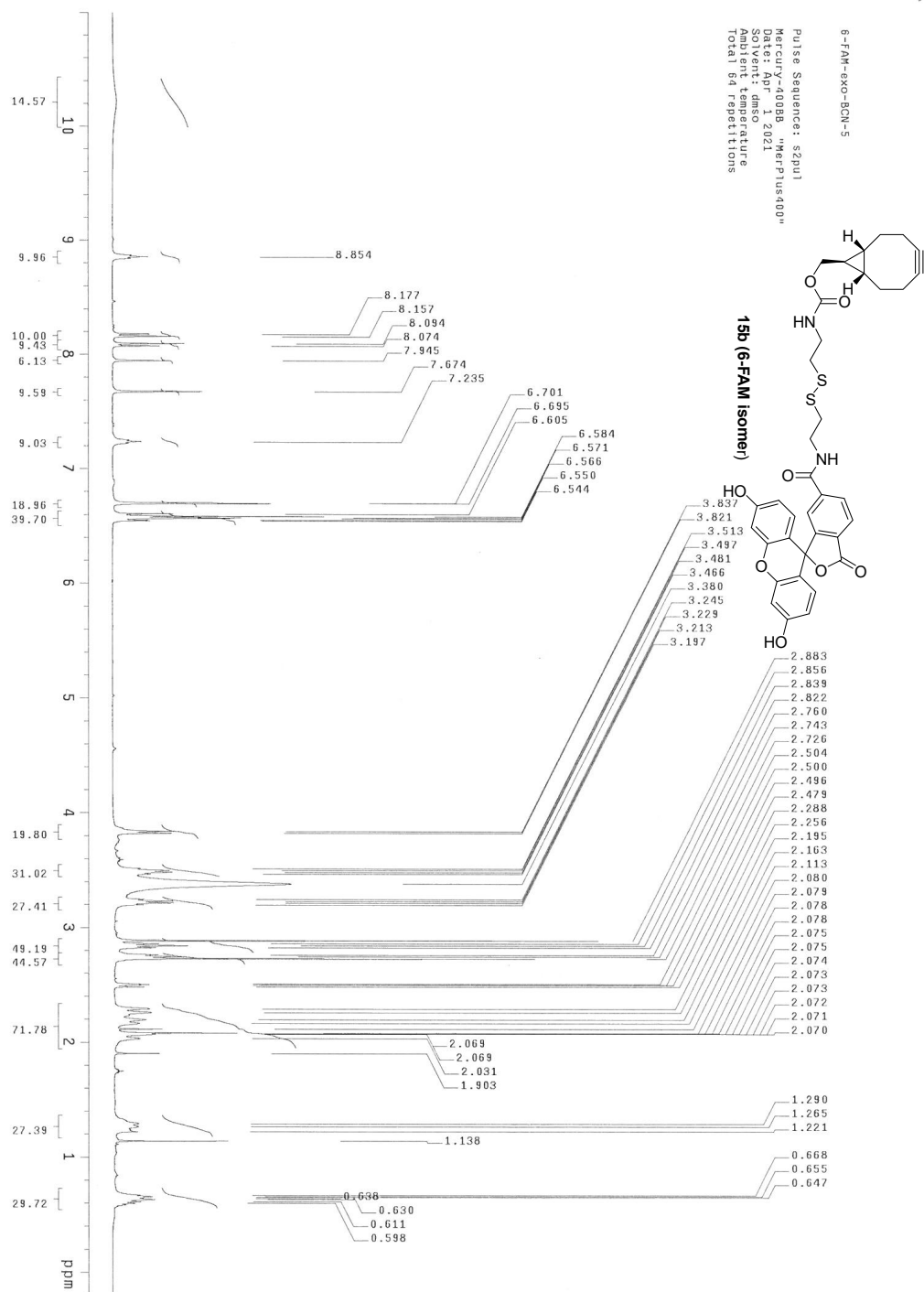
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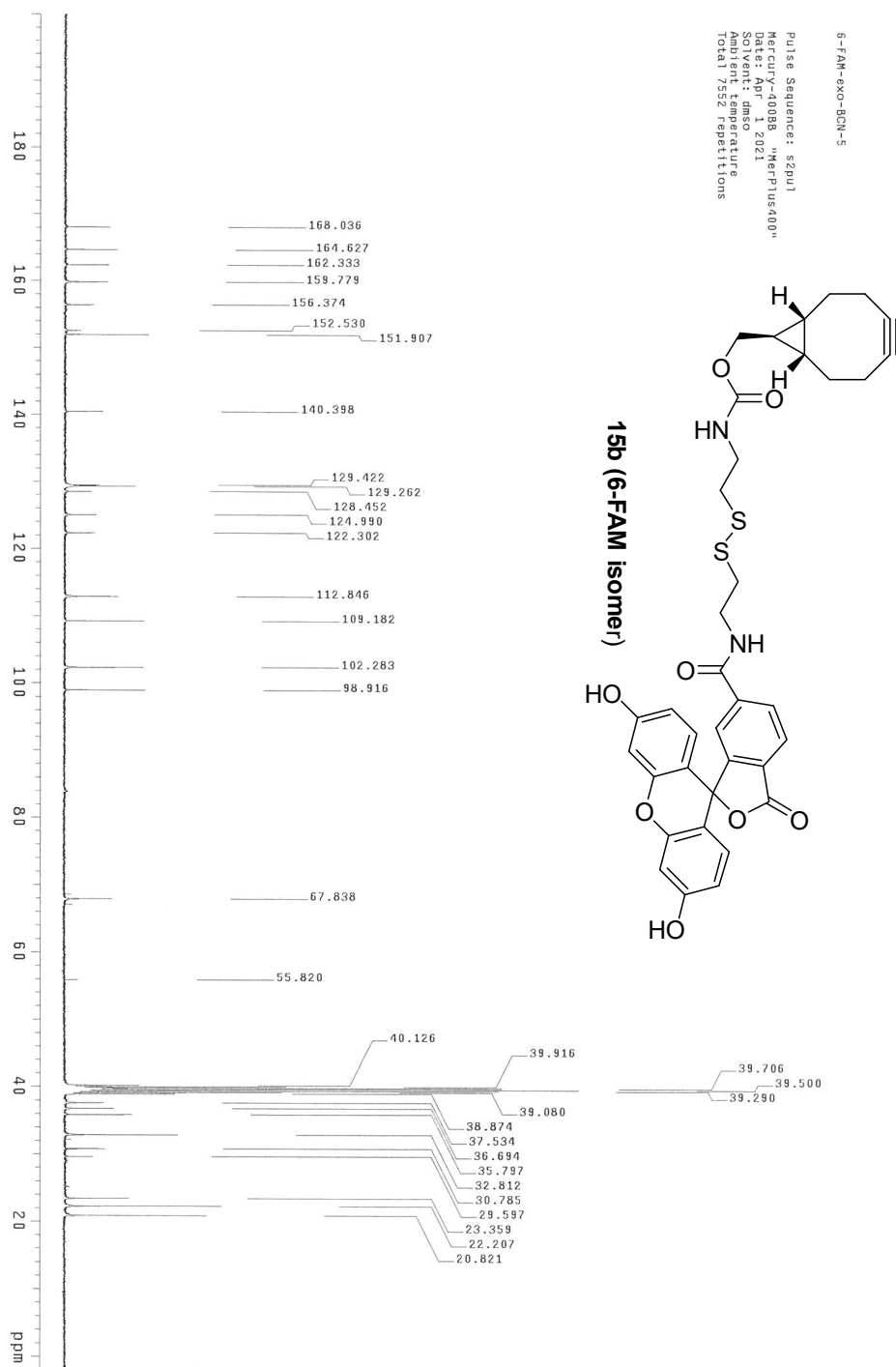
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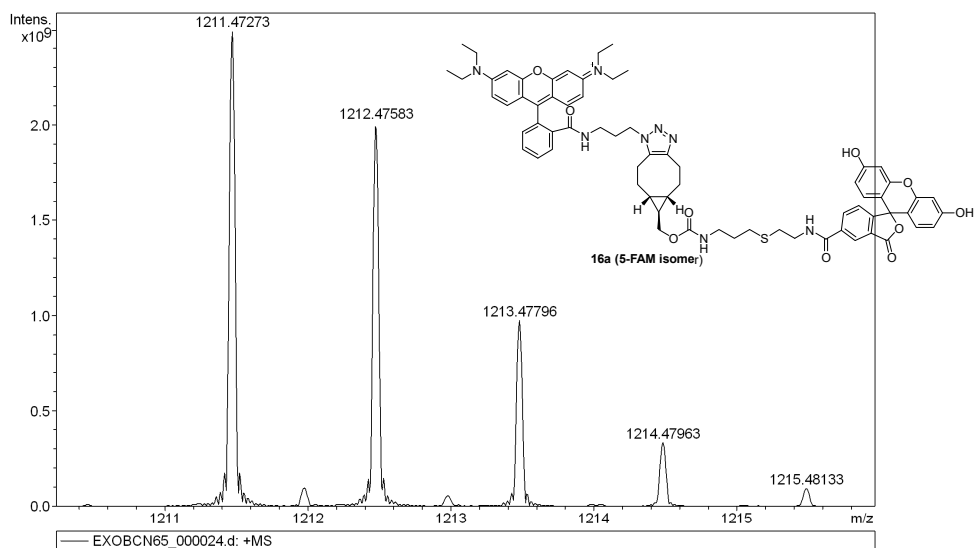


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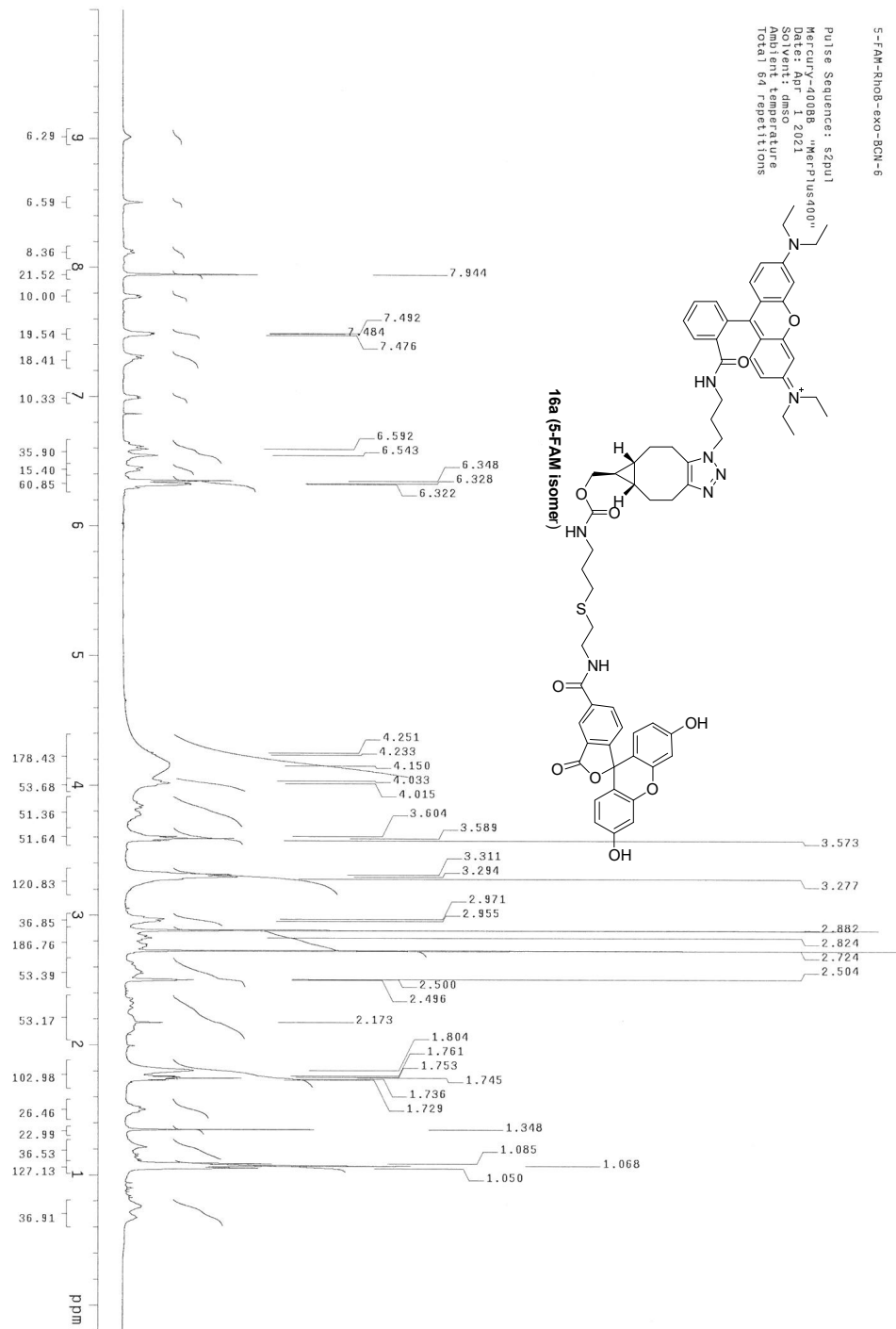
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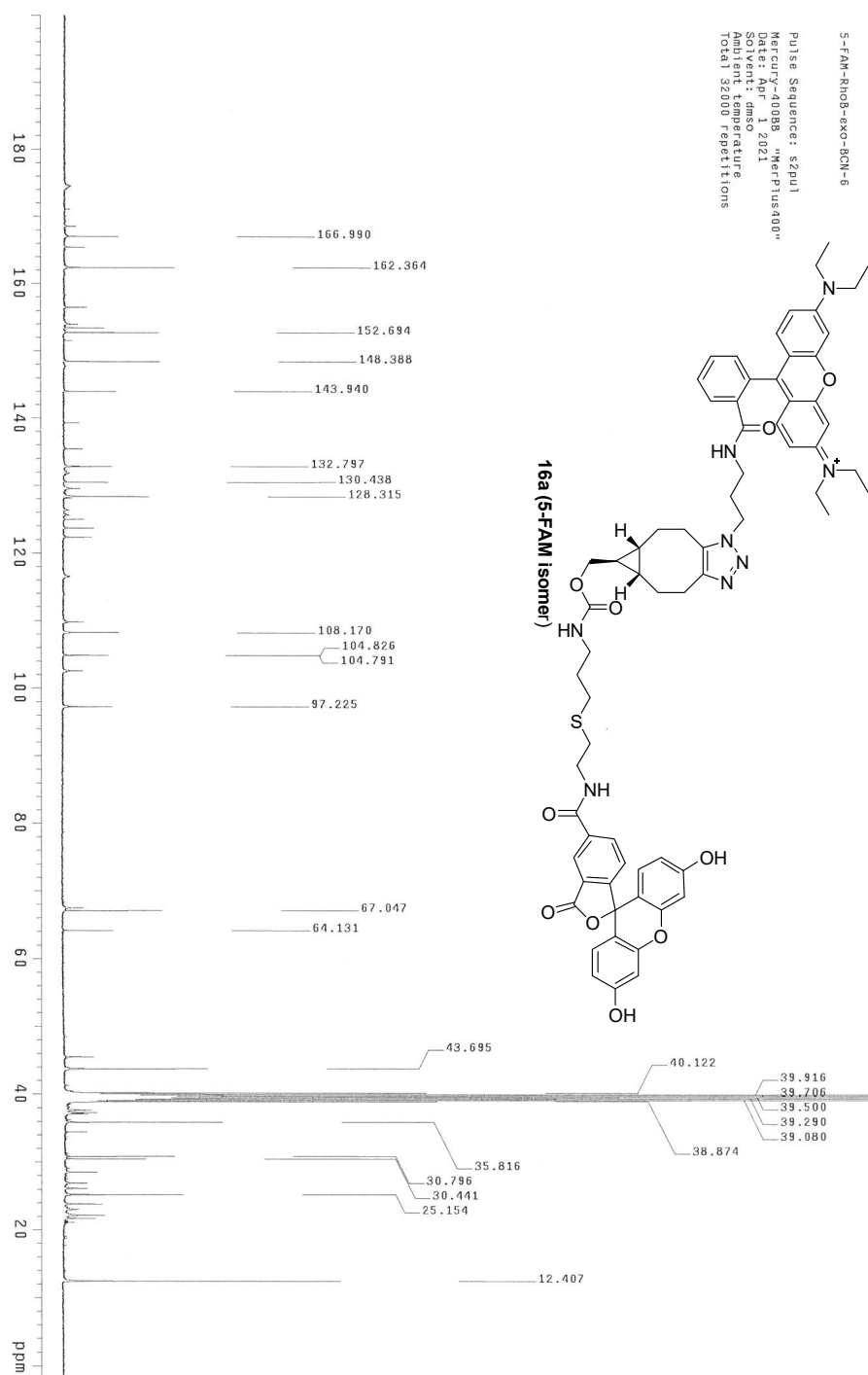
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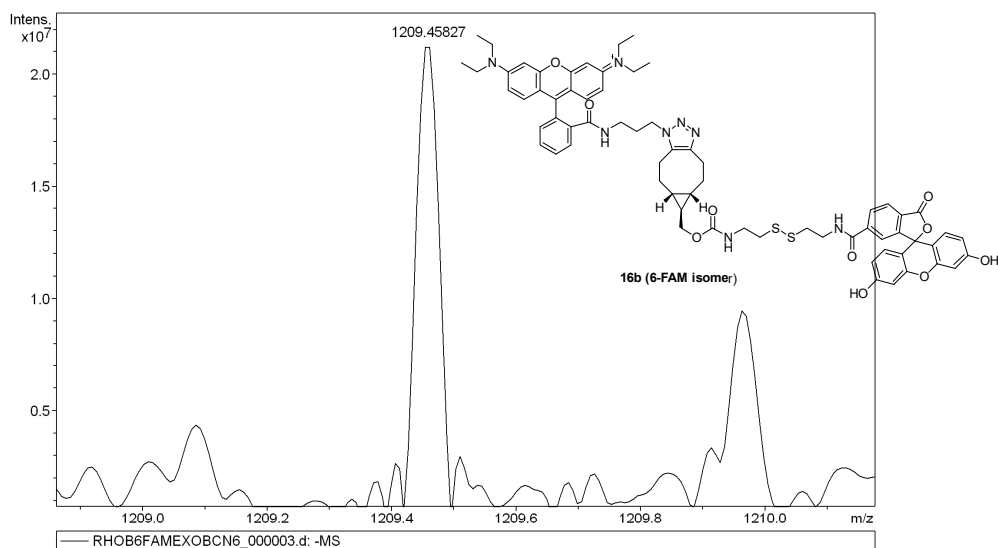


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