

Supporting Information

Structure-based bioisosterism design, synthesis, biological evaluation and *in silico* studies of benzamide analogs as potential anthelmintics.

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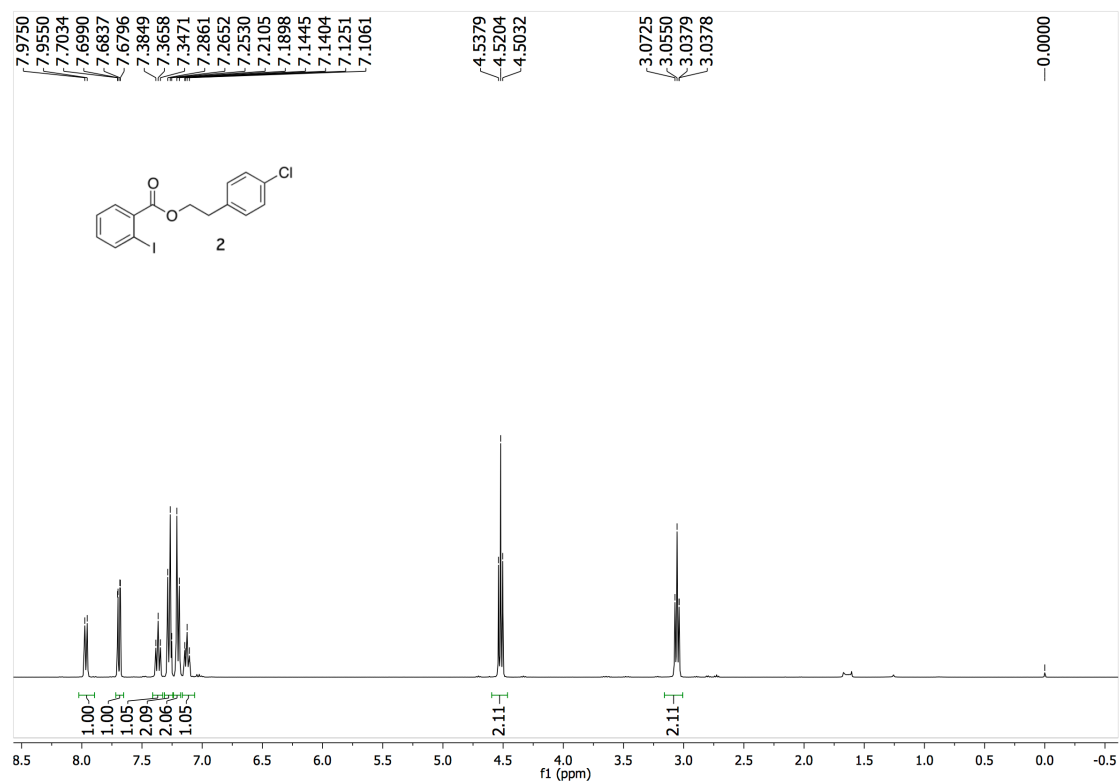
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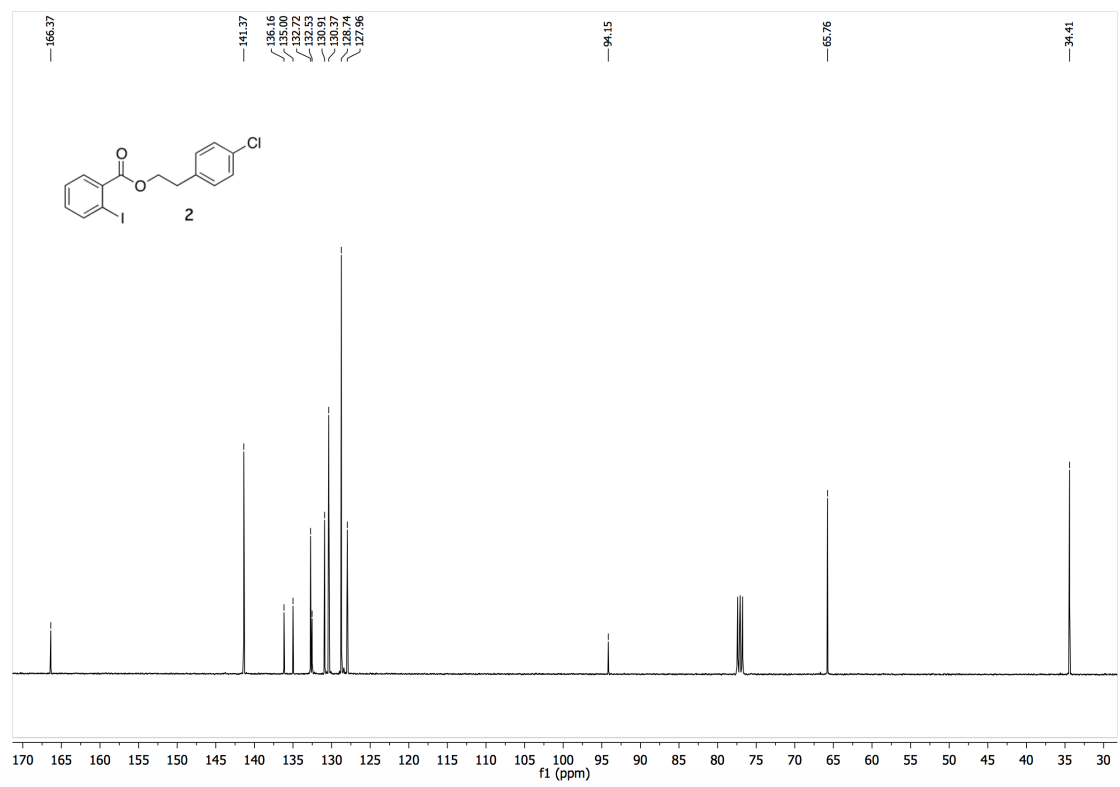
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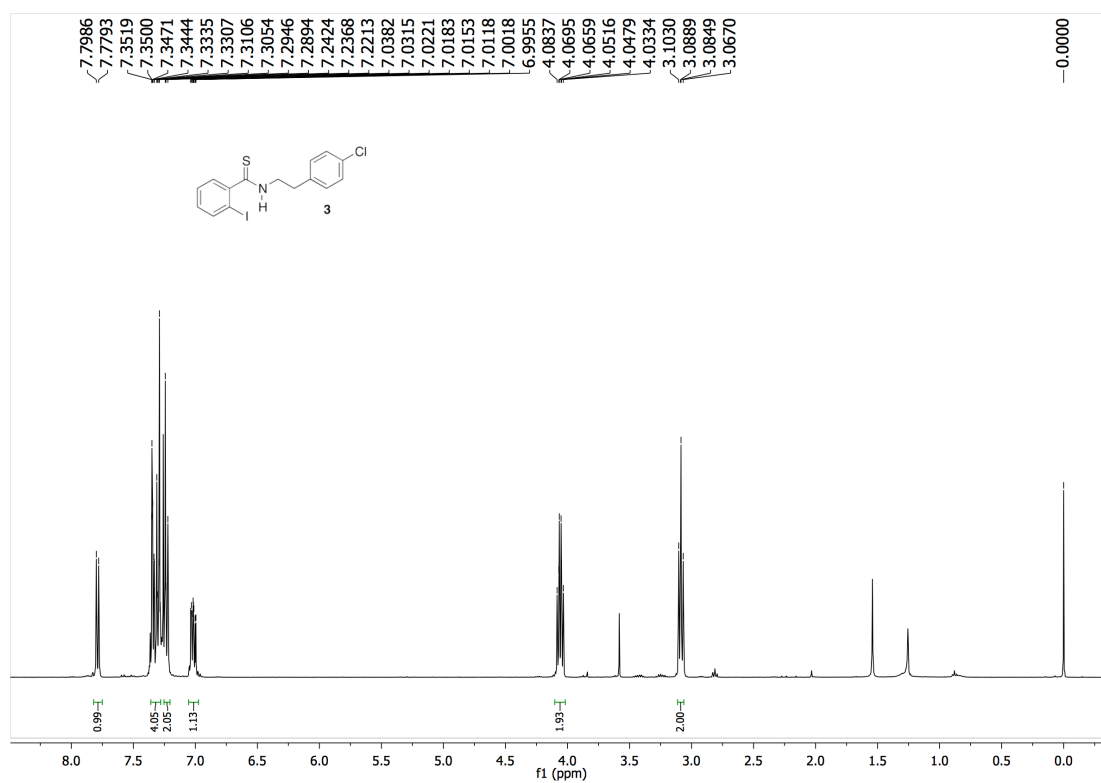
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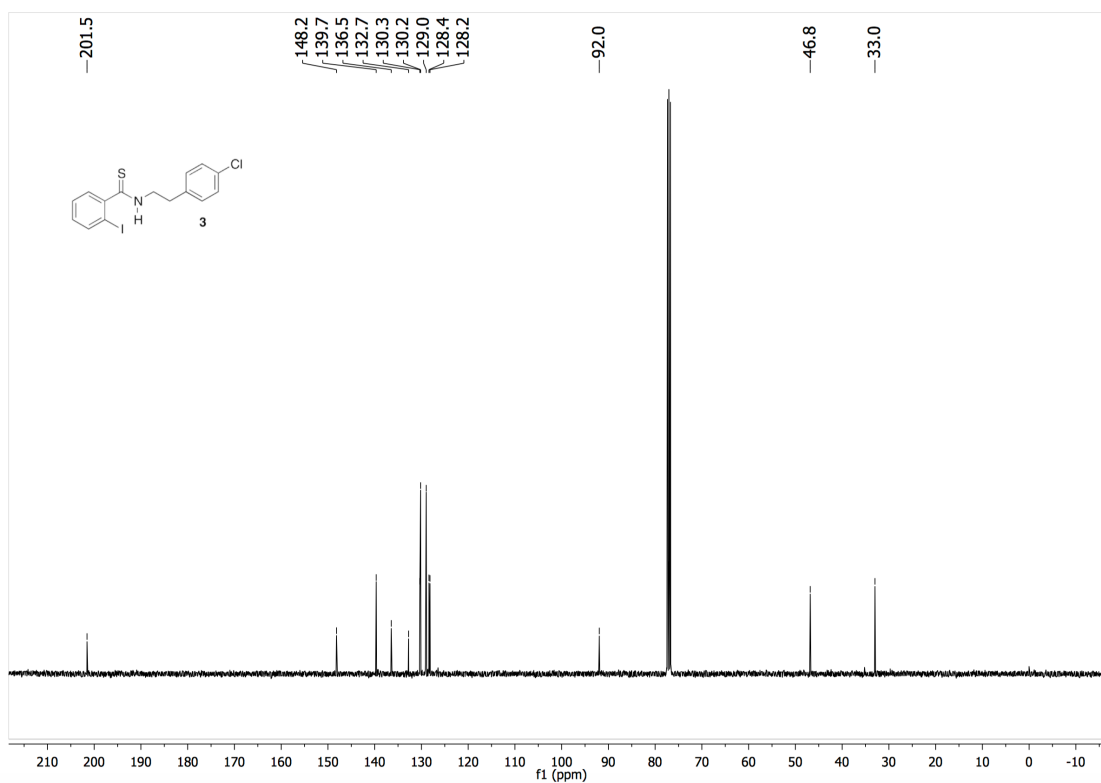
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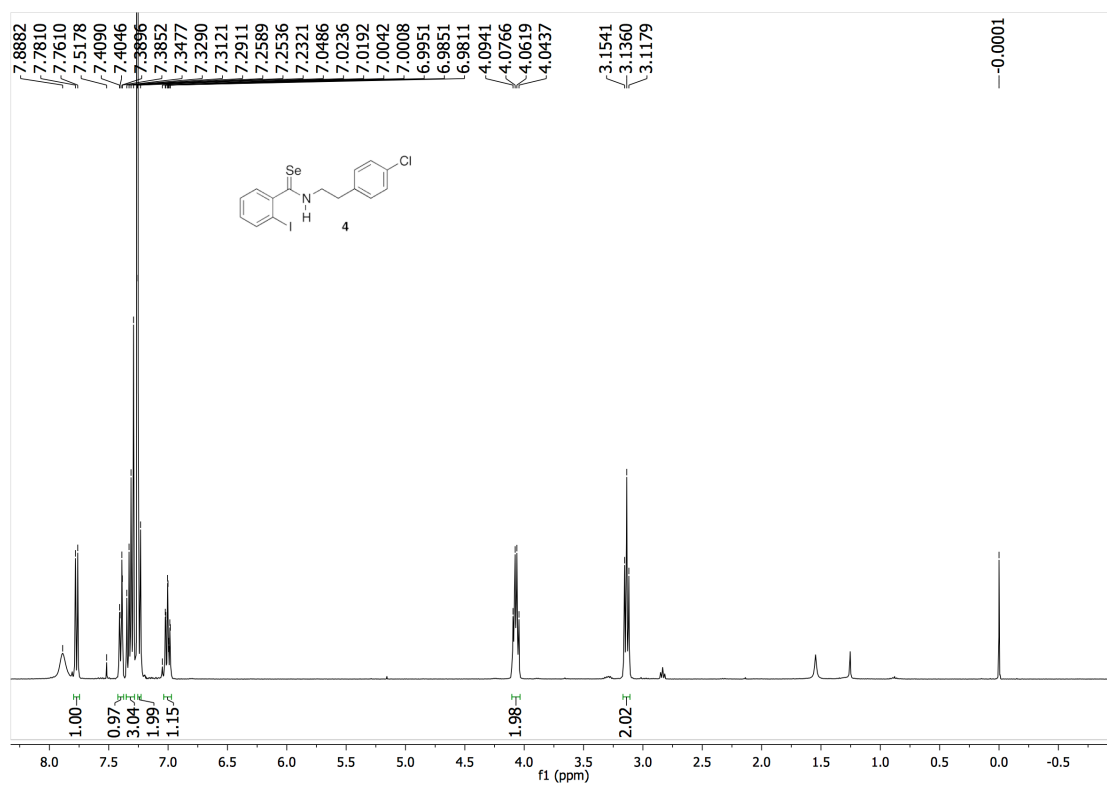
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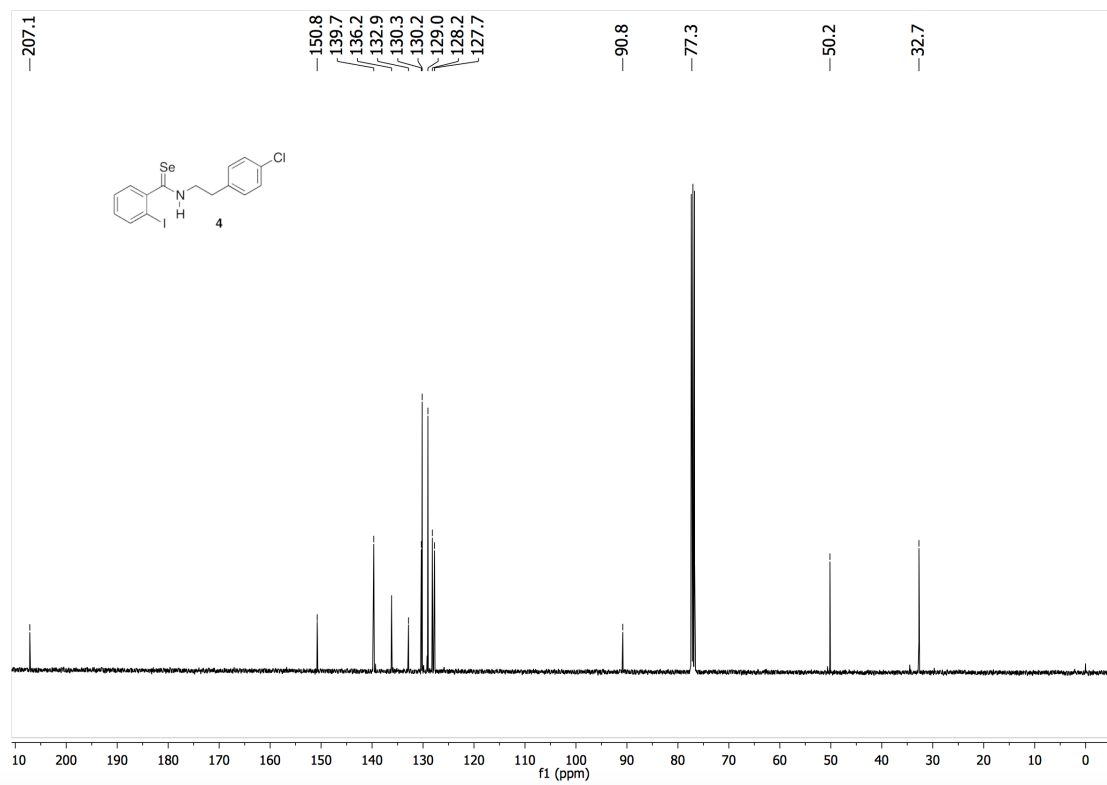
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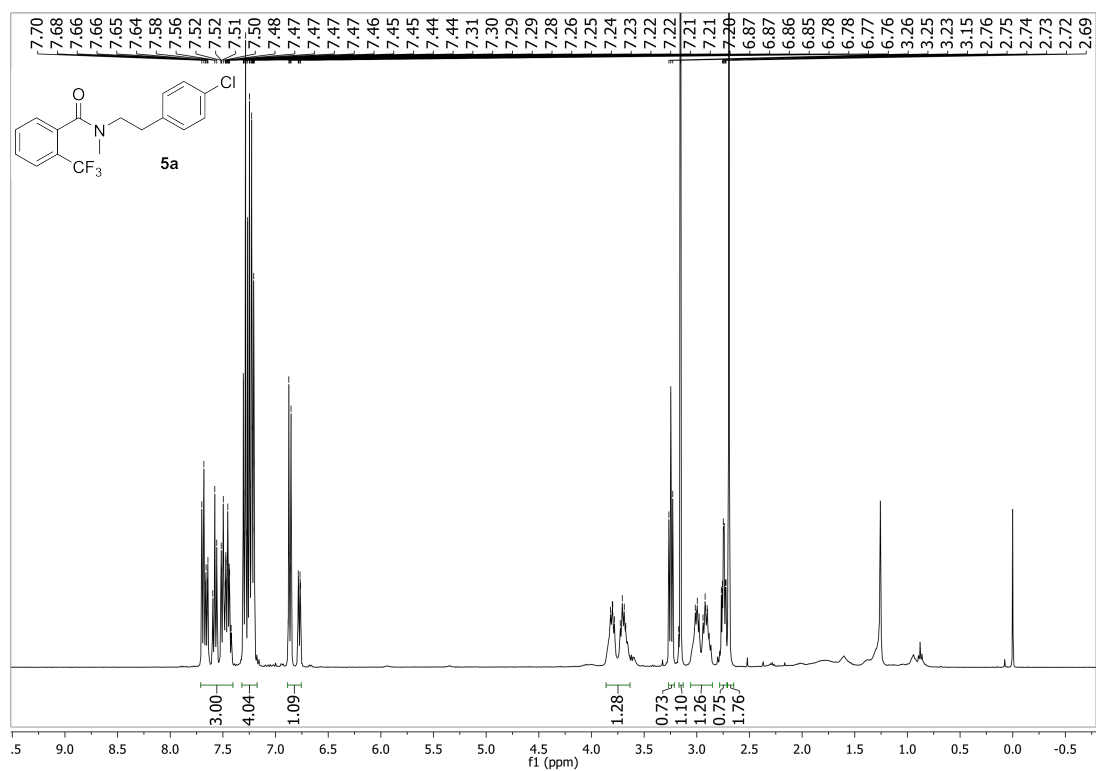
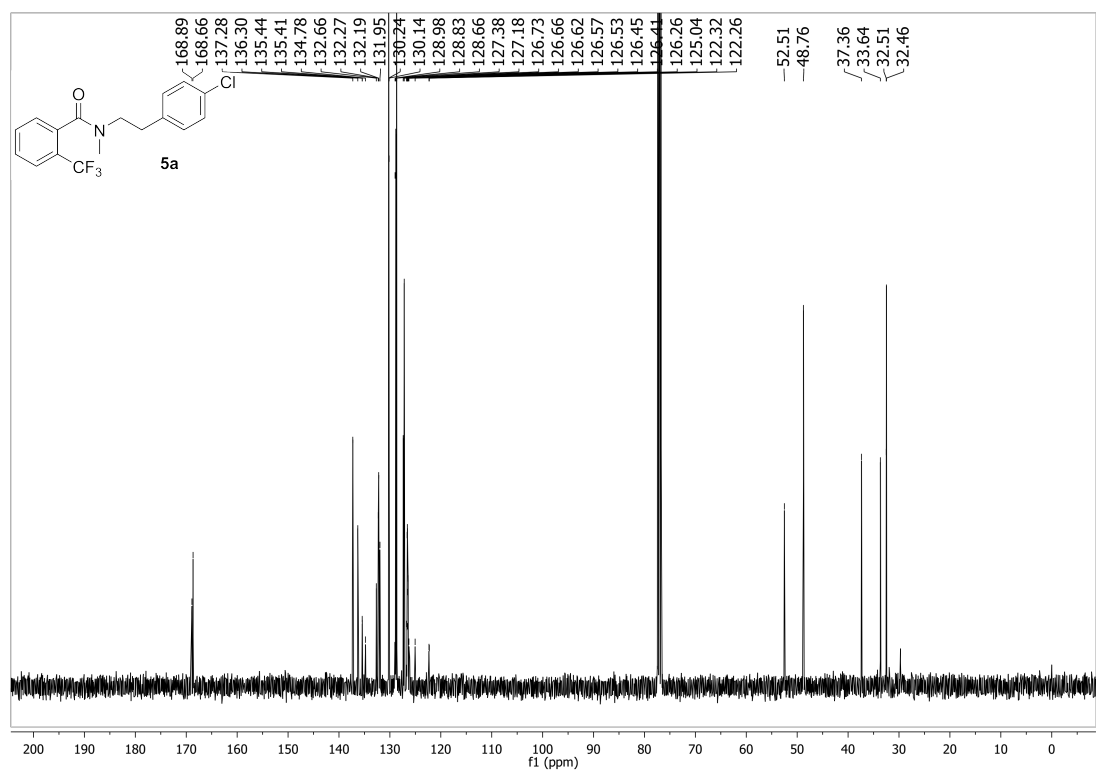


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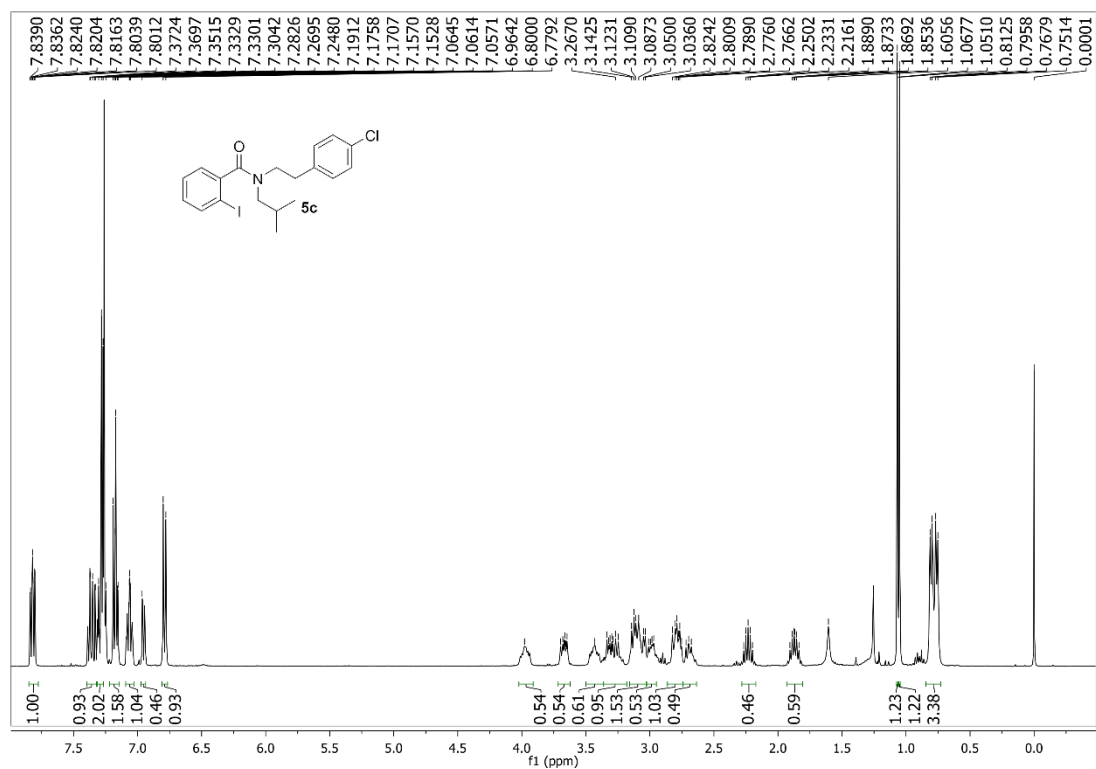


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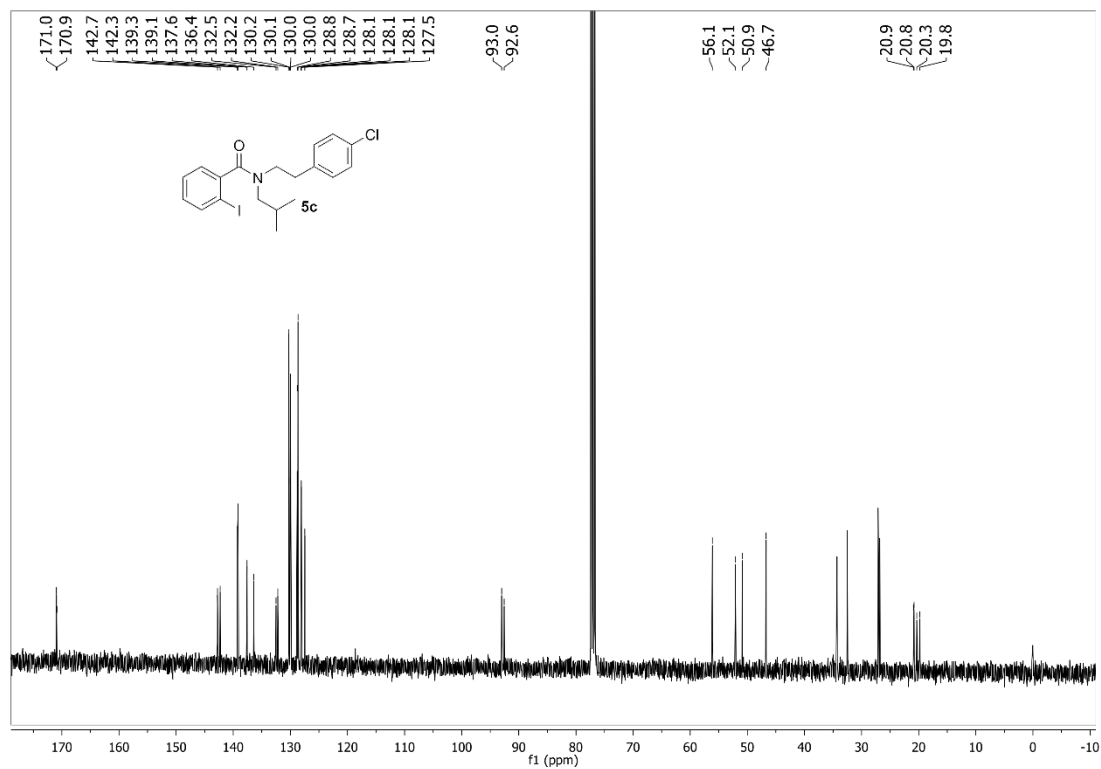


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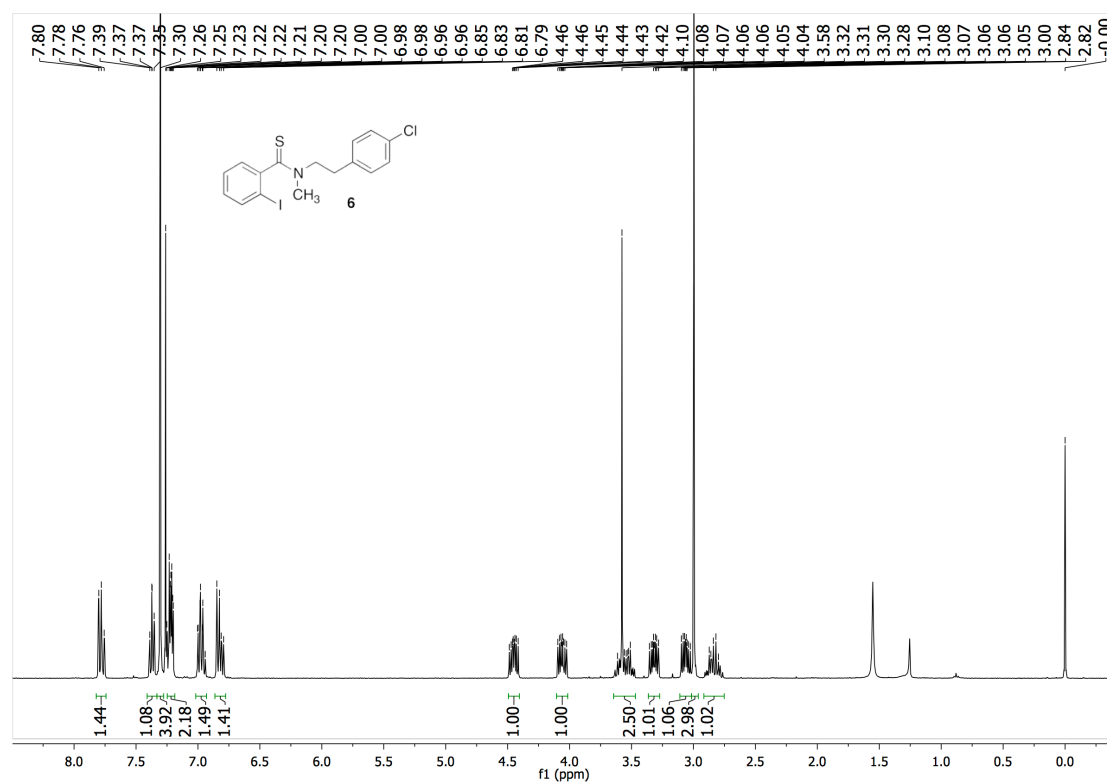
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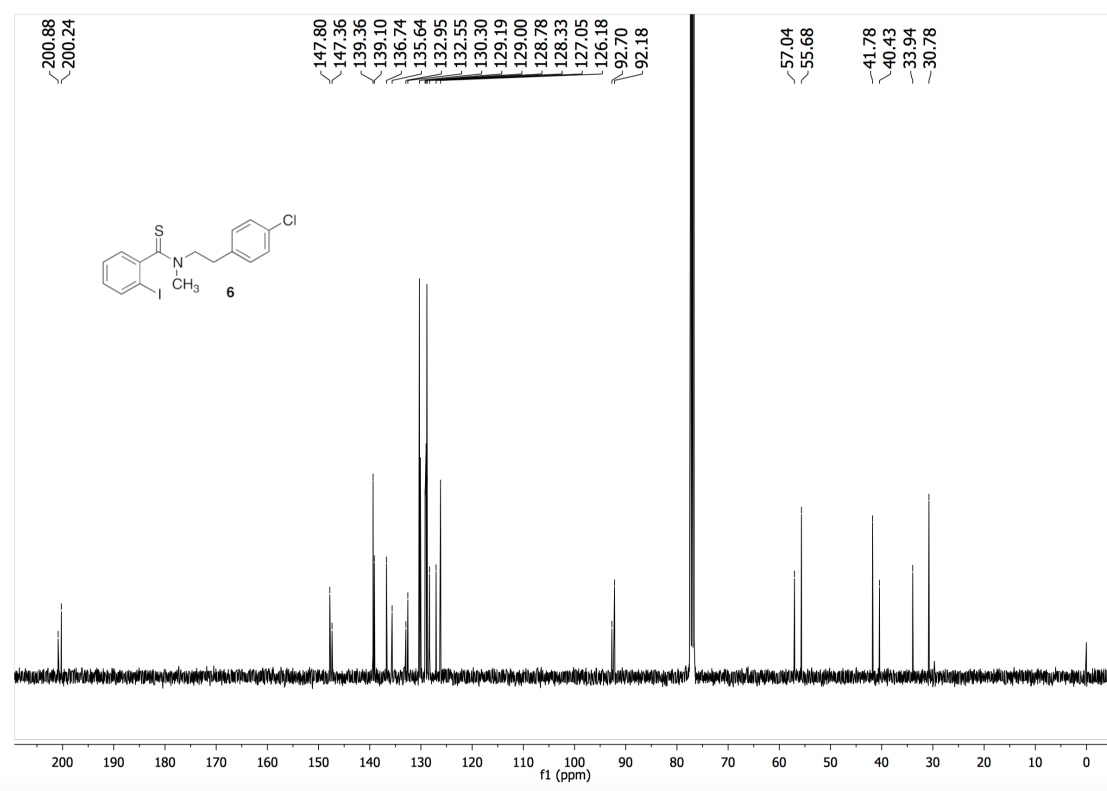
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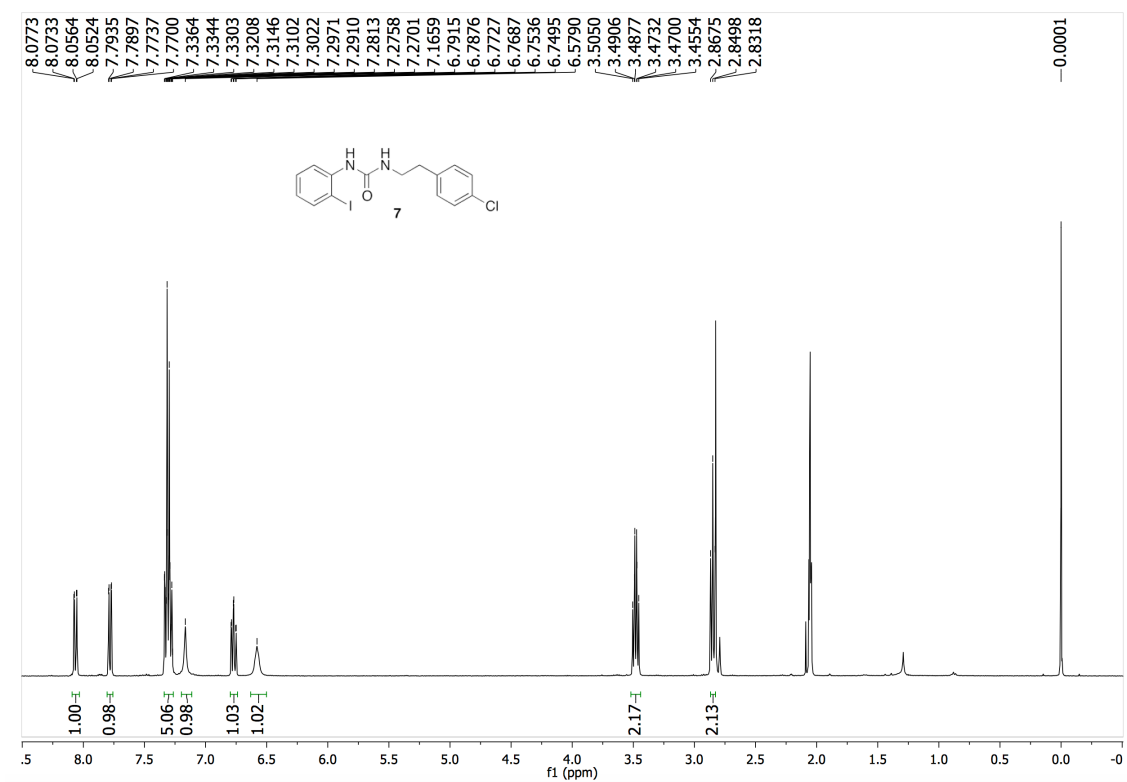
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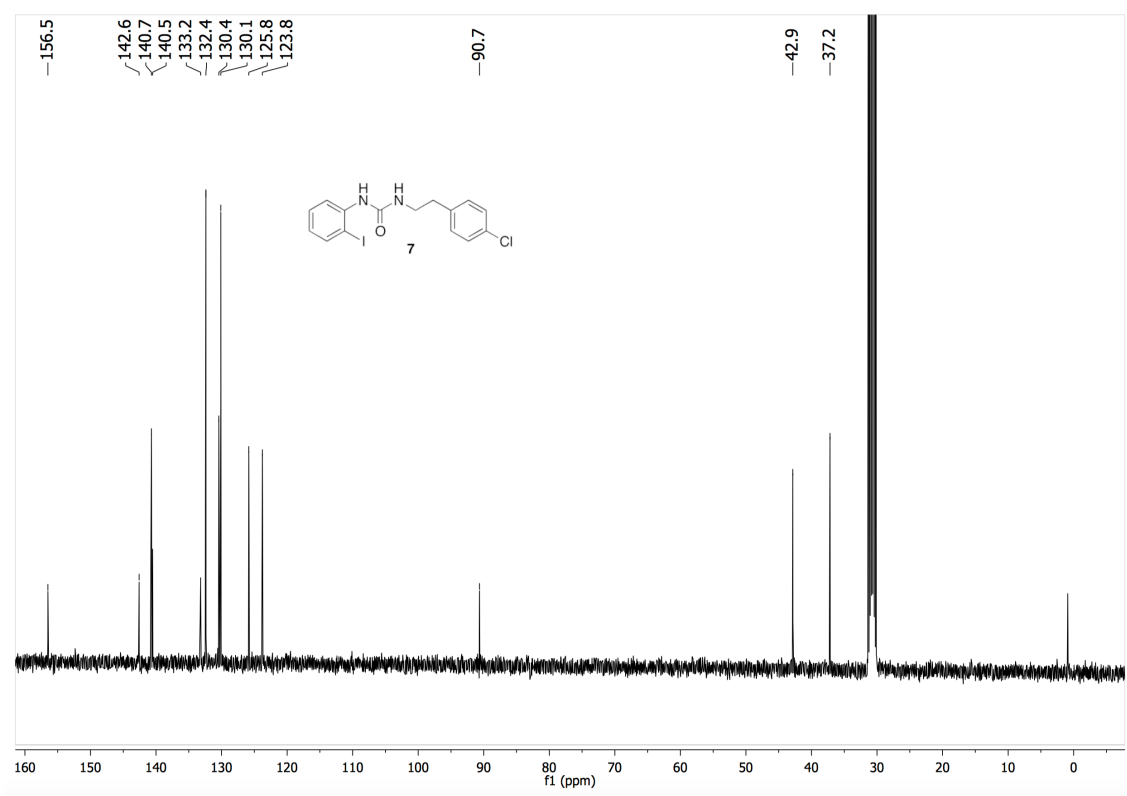
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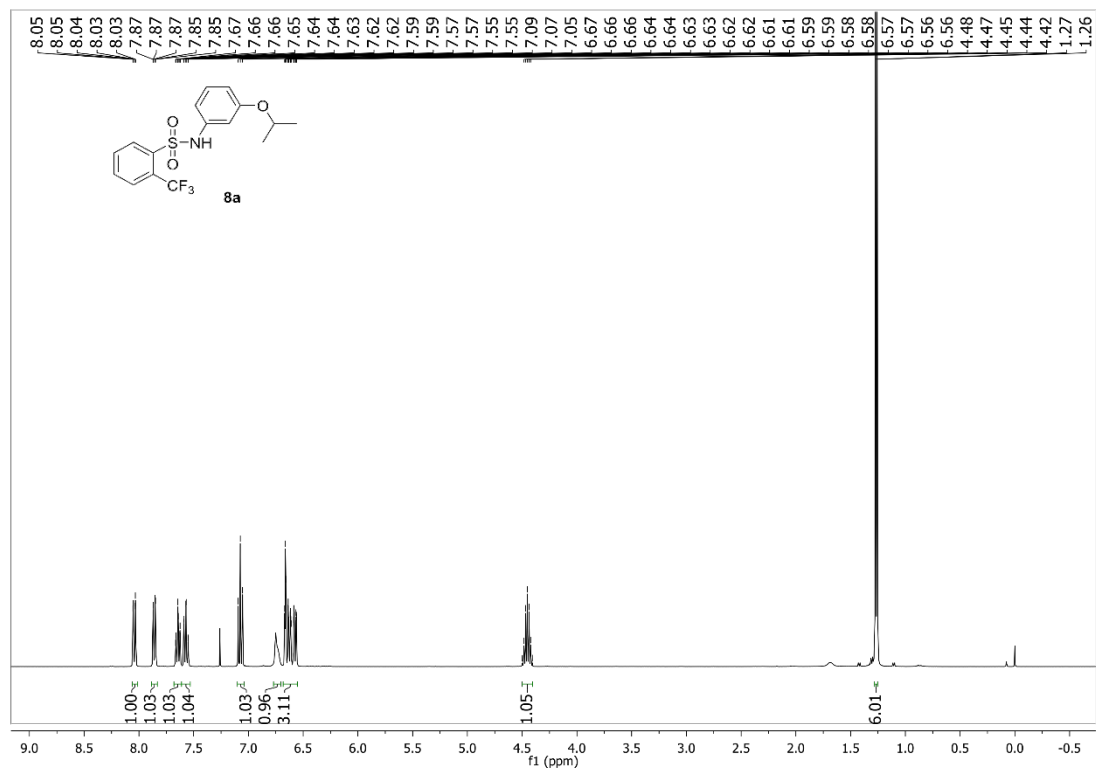
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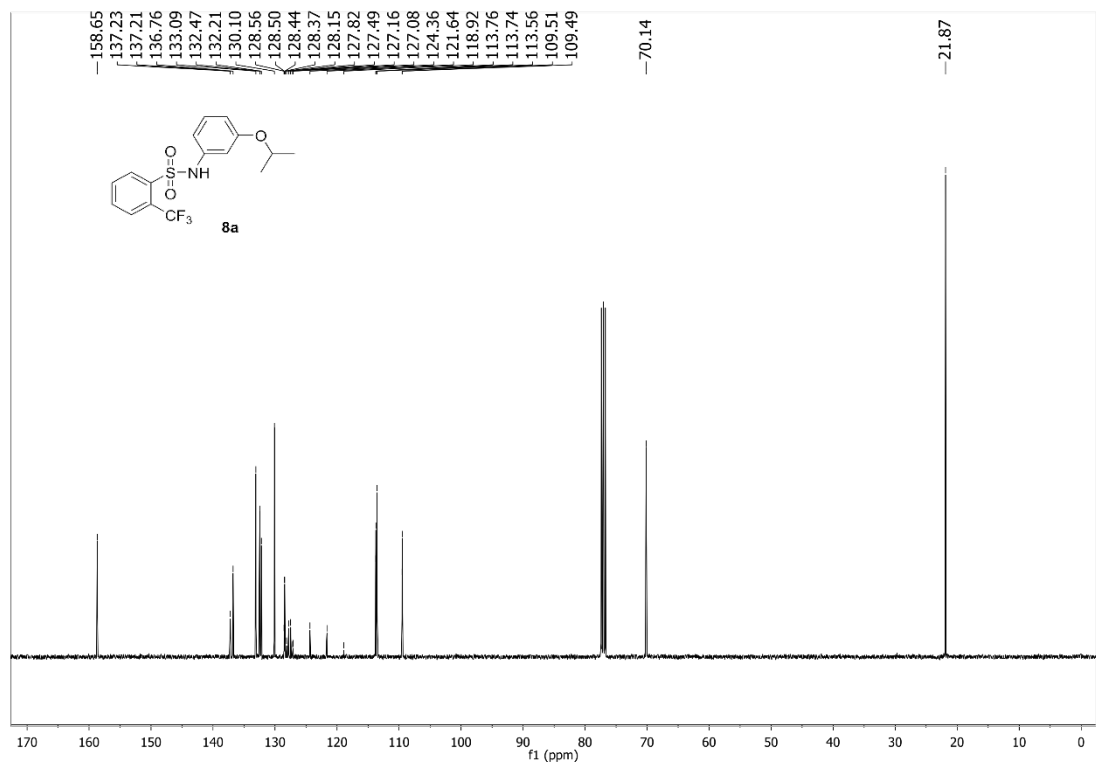
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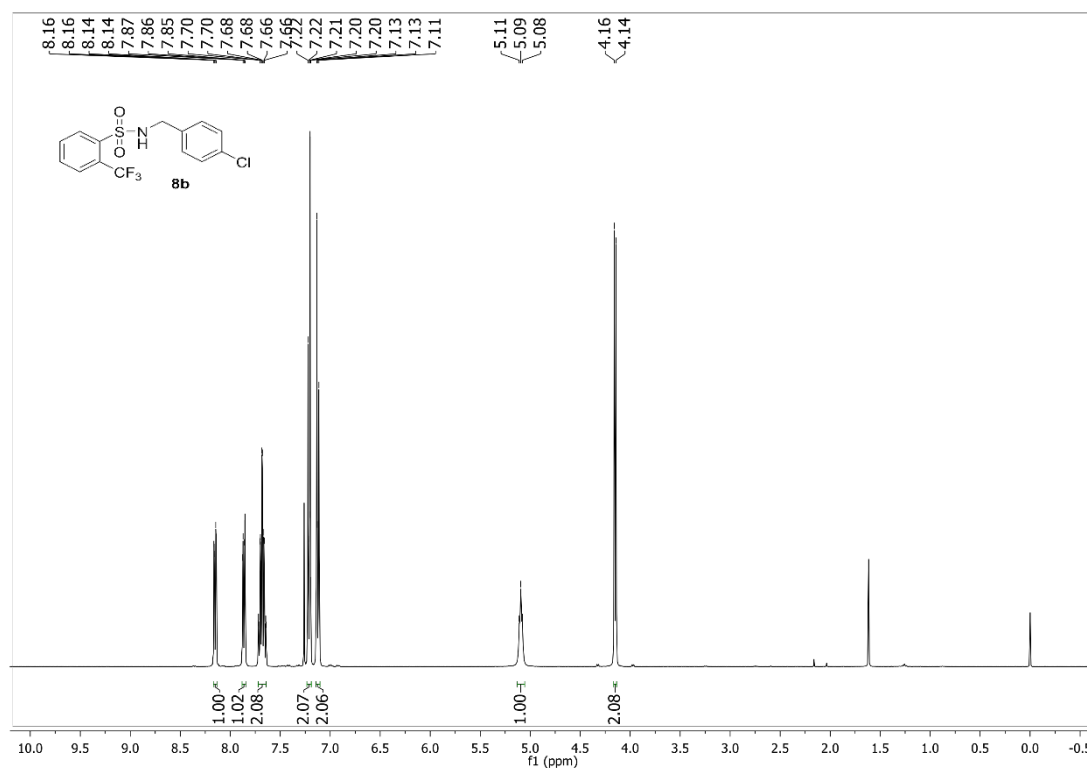
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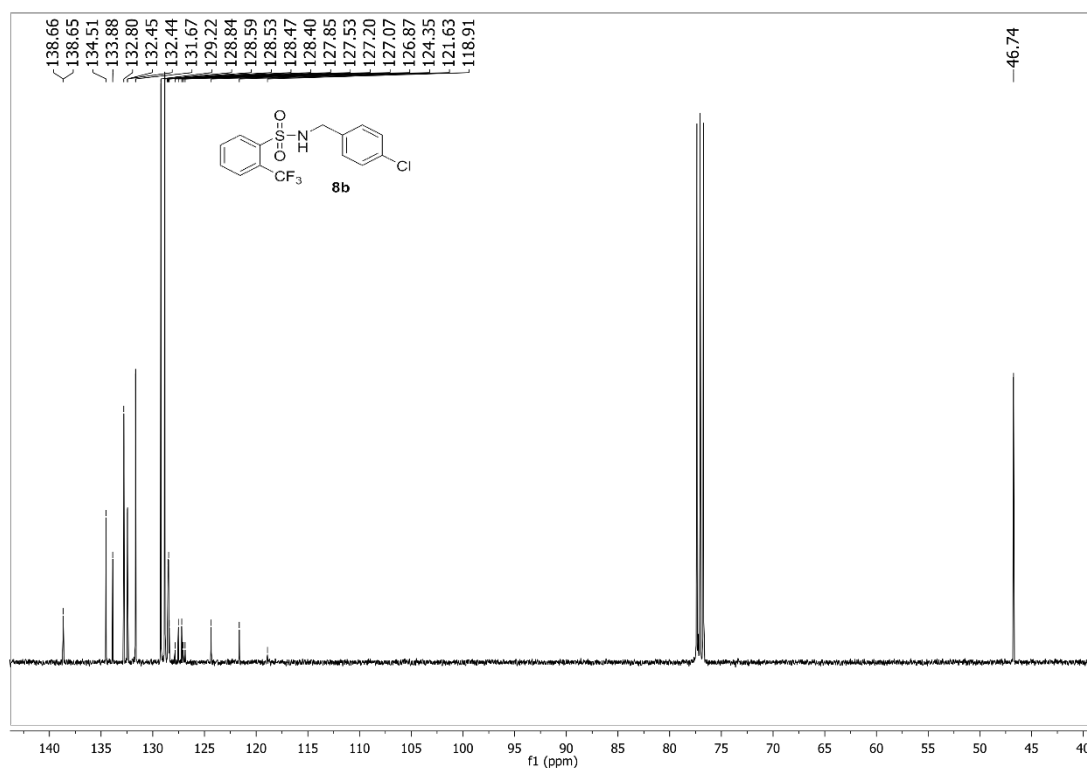
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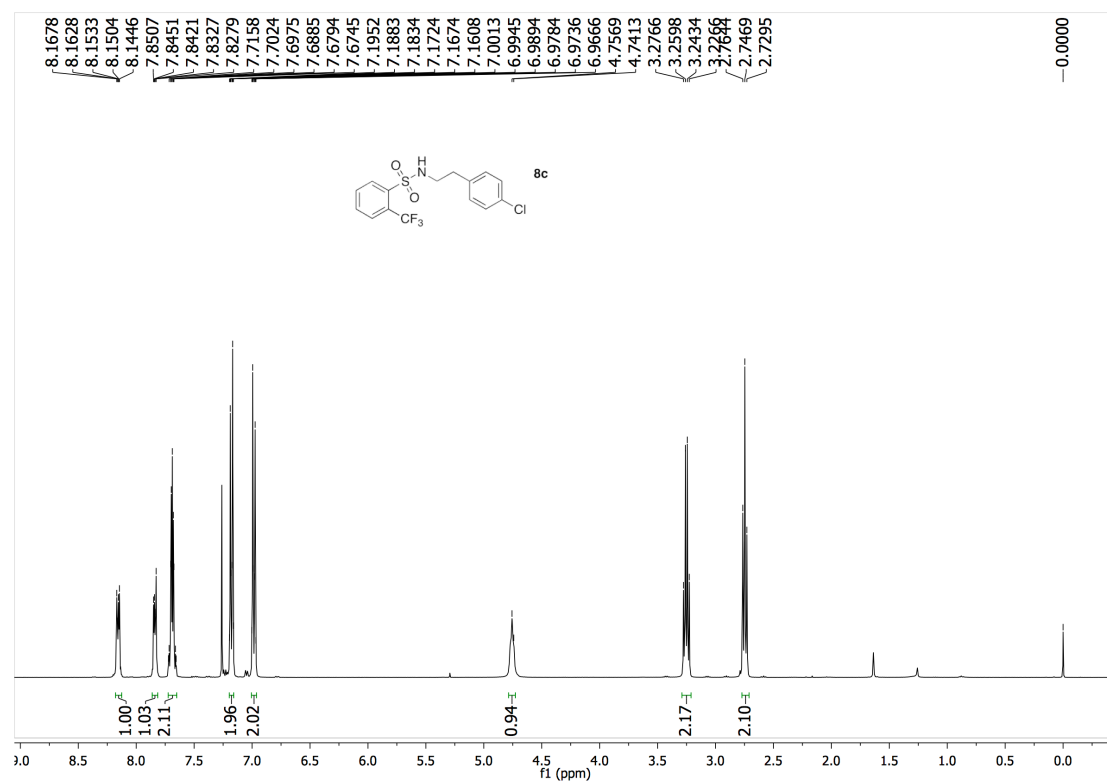
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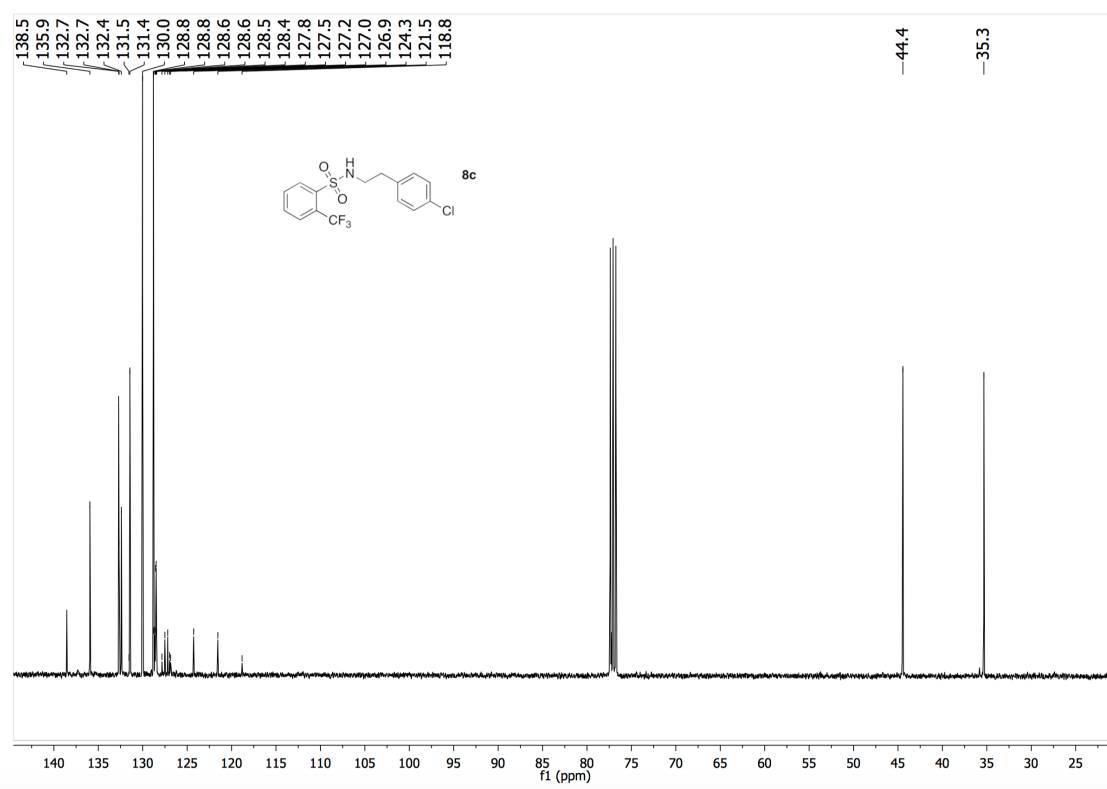
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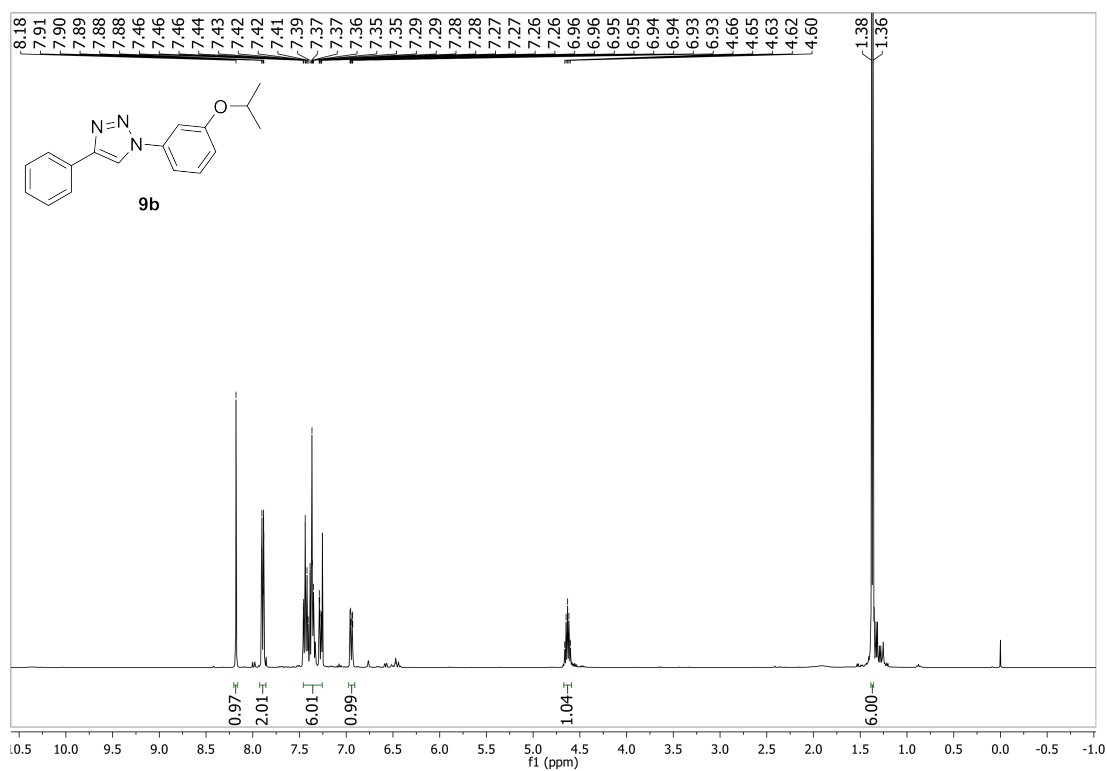
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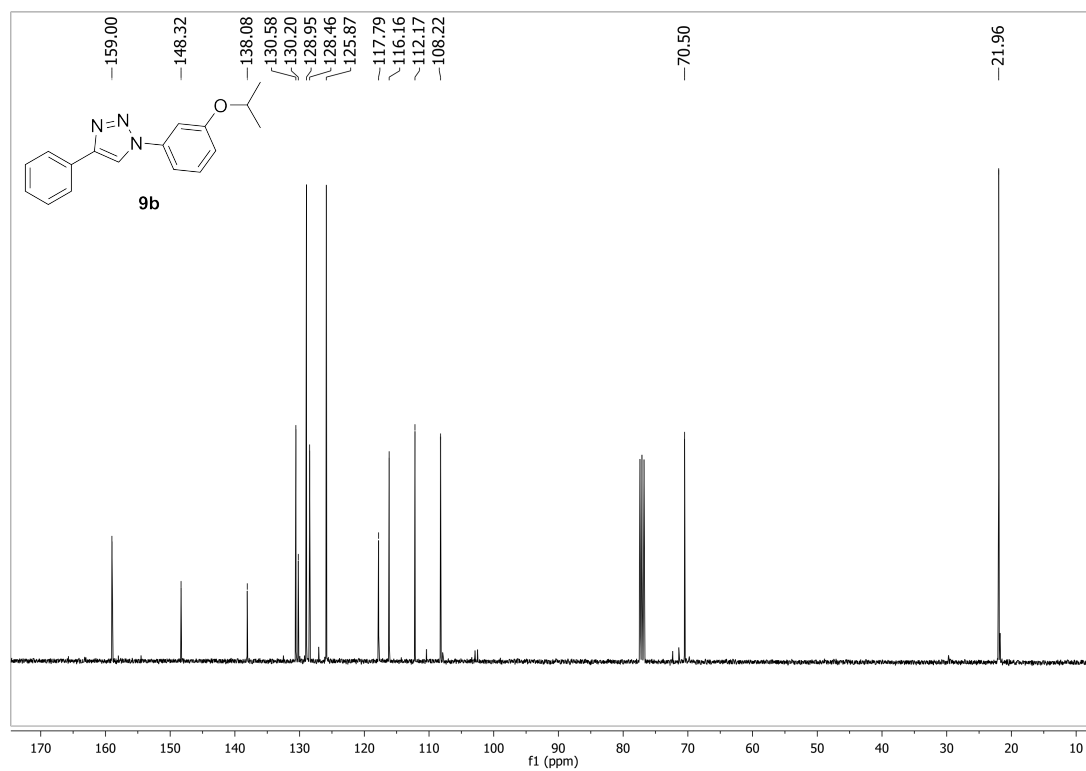
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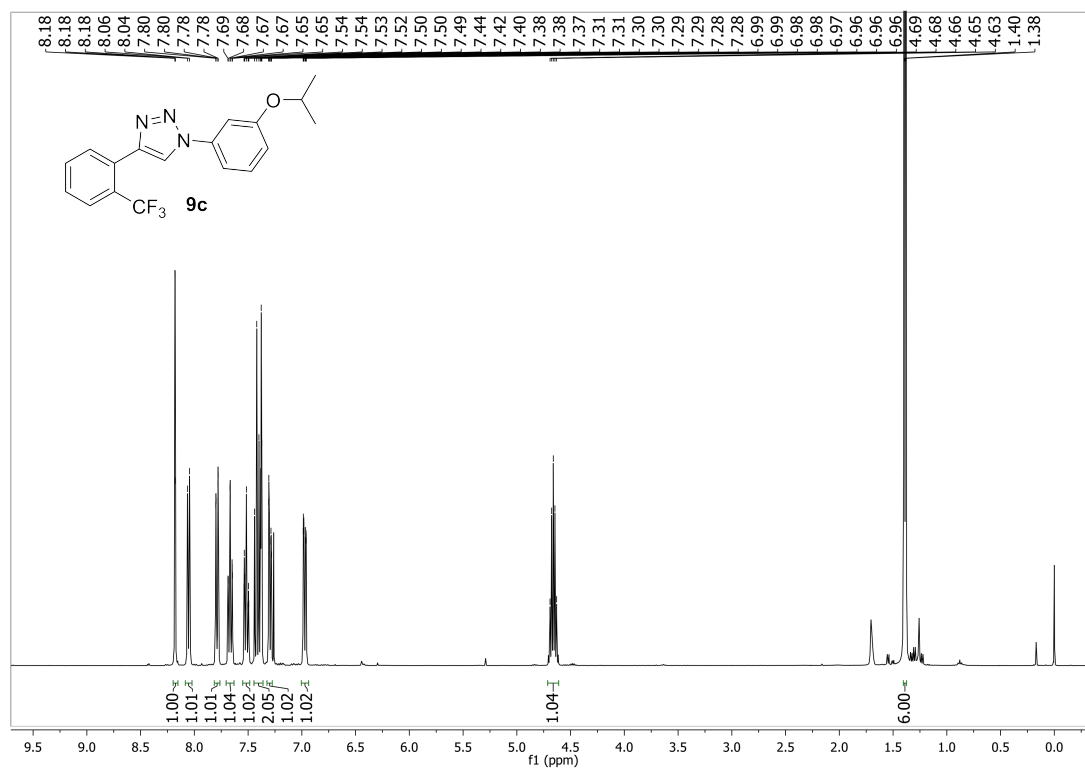
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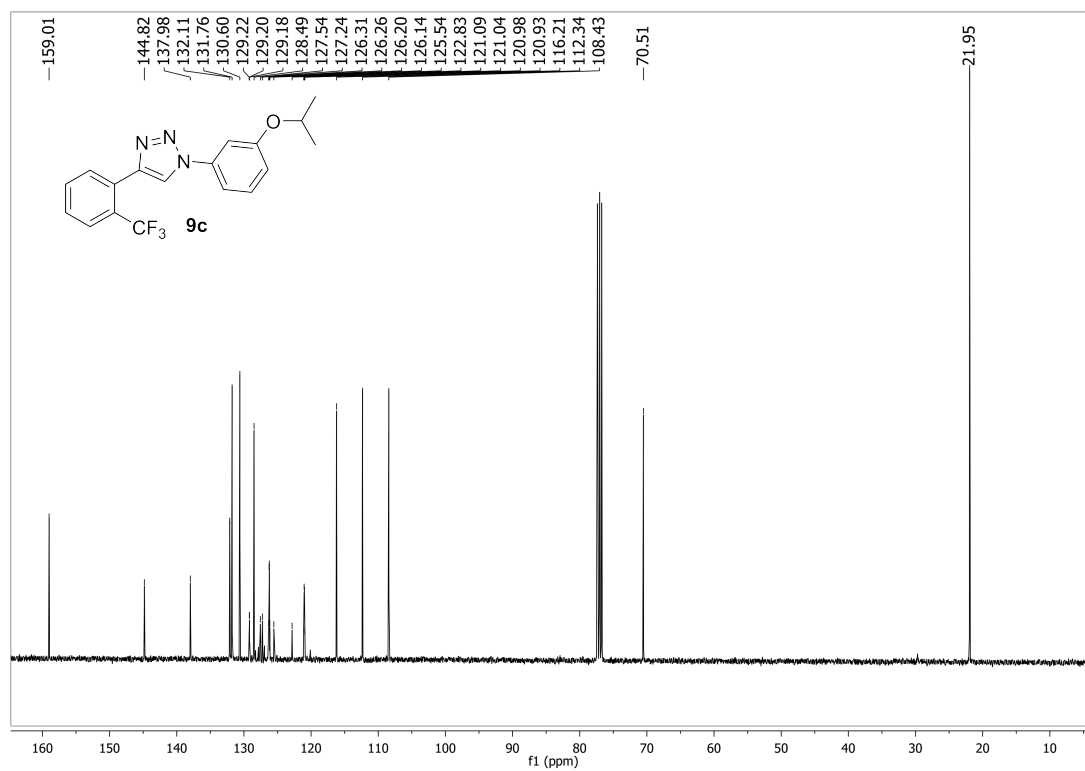
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$^1\text{H-NMR}$ (CDCl_3 with 0.05% TMS, 400 MHz)



$^{13}\text{C-NMR}$ (CDCl_3 with 0.05% TMS, 100 MHz)



2. Synthesis and characterization of compounds: 9a, 10b, 11a, 11b and 11c.

(1-(4-chlorobenzyl)-4-phenyl-1H-1,2,3-triazole) (9a). To a solution of **11b** (200 mg, 1.08 mmol) in a *t*-BuOH/H₂O 1:1 mixture (4 mL), phenylacetylene (120 mg, 1.08 mmol), ascorbic acid (40 mg, 0.2 mmol) and CuSO₄·5H₂O (5 mg, 0.02 mmol) were added. The mixture was stirred for 24 h and then poured into water (10 mL) and extracted with CH₂Cl₂ (3 x 20 mL). Combined organic layers were dried, filtered and removed under reduced pressure. The residue was purified by flash column chromatography on silica gel (EtOAc) to yield **9a** (270 mg, 93%) as a white solid. ¹H NMR (CDCl₃): δ 5.53 (s, 2H), 7.23 (m, 2H), 7.35 (m, 5H), 7.67 (s, 1H), 7.79 (m, 2H). ¹³C NMR (CDCl₃): δ 53.5, 119.5, 125.7, 128.3, 128.9, 129.4, 130.4, 133.2, 134.9, 148.4. Spectroscopic data is in accordance with that previously reported [58].

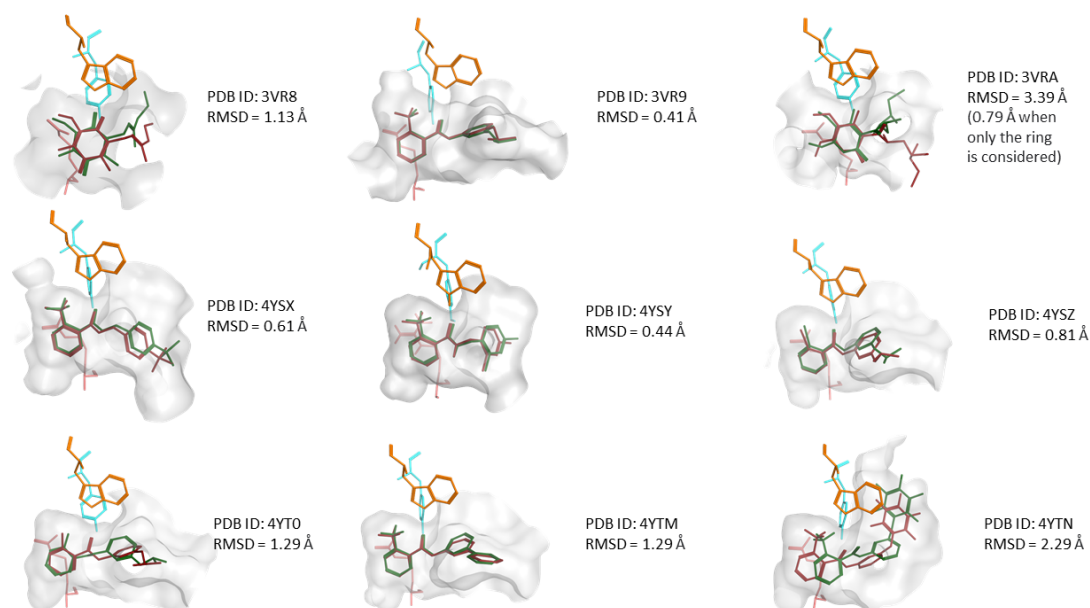
Trimethyl((2-(trifluoromethyl)phenyl)ethynyl)silane) (10b). Under argon atmosphere, Et₃N was degassed by bubbling argon for 10 minutes. Ethynyl trimethylsilane (508 mg, 2.2 mmol) and CuI (4 mg, 0.2 mmol) were added and the mixture was stirred for 5 min. 1-iodo-2-(trifluoromethyl)benzene (500 mg, 1.8 mmol) and Pd[P(Ph)₃]₄ (42 mg, 0.04 mmol) were added and stirred for 18 h. The mixture was poured into water, acidified to pH 2 with HCl 5% and extracted with EtOAc (3 x 20 mL). The combined organic layers were dried with Na₂SO₄, filtered and the solvent was removed under reduced pressure. The residue was purified by flash column chromatography on silica gel (Hx/EtOAc (100:1)) yielding **10b** (270 mg, 61%) as a colourless oil. R_f (Hx) = 0.7. ¹H NMR (CDCl₃): δ 0.26 (s, 9H), 7.37 (dddd, *J* = 8.4, 7.7, 1.5, 0.8 Hz, 1H), 7.45 (tdd, *J* = 7.6, 1.5, 0.7 Hz, 1H), 7.60 (quint, *J*_{CF} = 7.7, 1.4). ¹³C NMR (CDCl₃): δ 101.0, 101.4, 121.9 (q, *J* = 2.2 Hz), 123.9 (d, *J*_{CF} = 273.4 Hz) (CF₃), 126.2 (q, *J*_{CF} = 5.1 Hz), 128.6, 131.7, 132.6 (q, *J* = 30.5 Hz), 134.6 [59].

1-(azidomethyl)-4-chlorobenzene. (11a) To a solution of *p*-chlorobenzylamine (500 mg, 3.5 mmol) in 10 mL of MeOH, K₂CO₃ (880 mg, 1.28 mmol), CuSO₄·5H₂O (10 mg, 0.05 mmol) and 1H-imidazole-1-sulfonyl azide hydrochloride (878 mg, 4.2 mmol) were added. The mixture was stirred overnight and then poured into water, acidified to pH 2 with HCl 5% and extracted with CH₂Cl₂ (3 x 20 mL). The combined organic layers were dried with Na₂SO₄, filtered and removed under reduced pressure. The crude was purified by flash column chromatography on silica gel (Hx/EtOAc (8:1)), yielding **11a** (490 mg, 83%) as a colourless oil. ¹H NMR (CDCl₃): δ 4.29 (s, 2H), 7.23 (m, 2H), 7.34 (m, 2H). ¹³C NMR (CDCl₃): δ 54.1, 129.1, 129.5, 133.9, 134.2 [60].

1-(2-azidoethyl)-4-chlorobenzene (11b) To a solution of 2-(4-chlorophenyl)ethan-1-amine (100 mg, 0.64 mmol) in MeOH (2 mL) 1H-imidazole-1-sulfonyl azide hydrochloride (160 mg, 0.74 mmol), CuSO₄·5H₂O (2.2 mg, 0.01 mmol) and anhydrous K₂CO₃ (176 mg, 1.28 mmol) were added. The mixture was stirred at room temperature overnight. The solvent was removed at reduced pressure and the residue was poured into water, acidified to pH 2 with HCl 5% and extracted with CH₂Cl₂ (3 x 20 mL). The combined organic layers were dried with Na₂SO₄, filtered and the solvent was removed under reduced pressure. The crude was purified by flash column chromatography on silica gel (Hx/EtOAc (8:1)), yielding **11b** (102 mg, 88%) as a colourless oil. ¹H NMR (CDCl₃): δ 2.84 (t, *J* = 7.1 Hz, 2H), 3.47 (t, *J* = 7.1 Hz, 2H), 7.14 (m, 2H), 7.28 (m, 2H). ¹³C NMR (CDCl₃): δ 34.7, 52.3, 128.8, 130.1, 132.7, 136.6 [40-62].

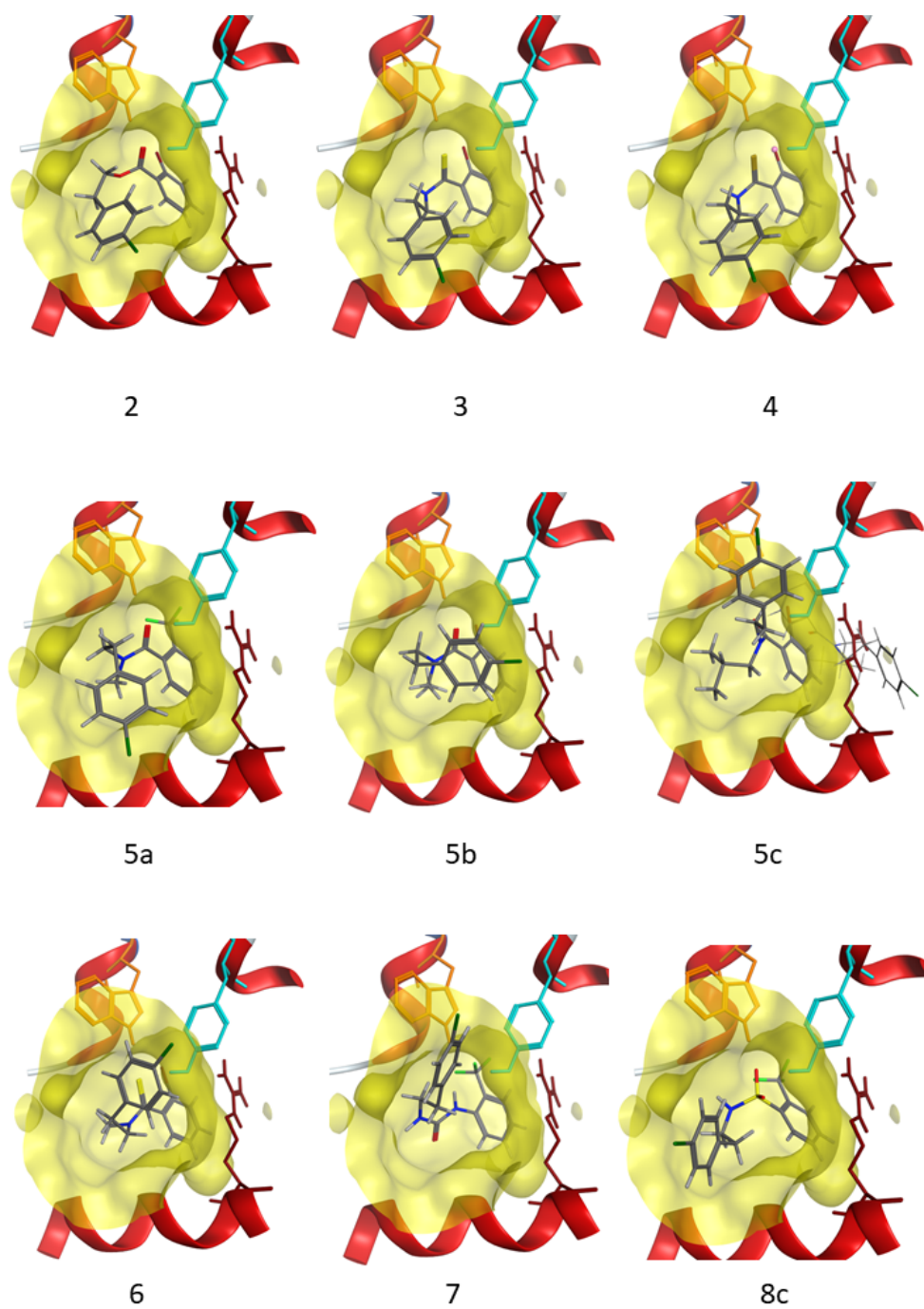
1-azide-3-isopropoxybenzene. (11c) To a solution of 3-isopropoxyaniline (200 mg, 1.32 mmol) in HCl 6 N (5 mL), NaNO₂ (91 mg, 1.32 mmol) was added in ice bath. The mixture was stirred at room temperature for 10 min and NaN₃ (103 mg, 1.6 mmol) was added. After 2 h, the mixture was poured into water, neutralized with NaHCO₃ 10% and extracted with EtOAc (3 x 15 mL). The combined organic layers were dried with Na₂SO₄, filtered and removed under reduced pressure. The reaction crude was used in the next step without further purification [63].

3. Docking optimization



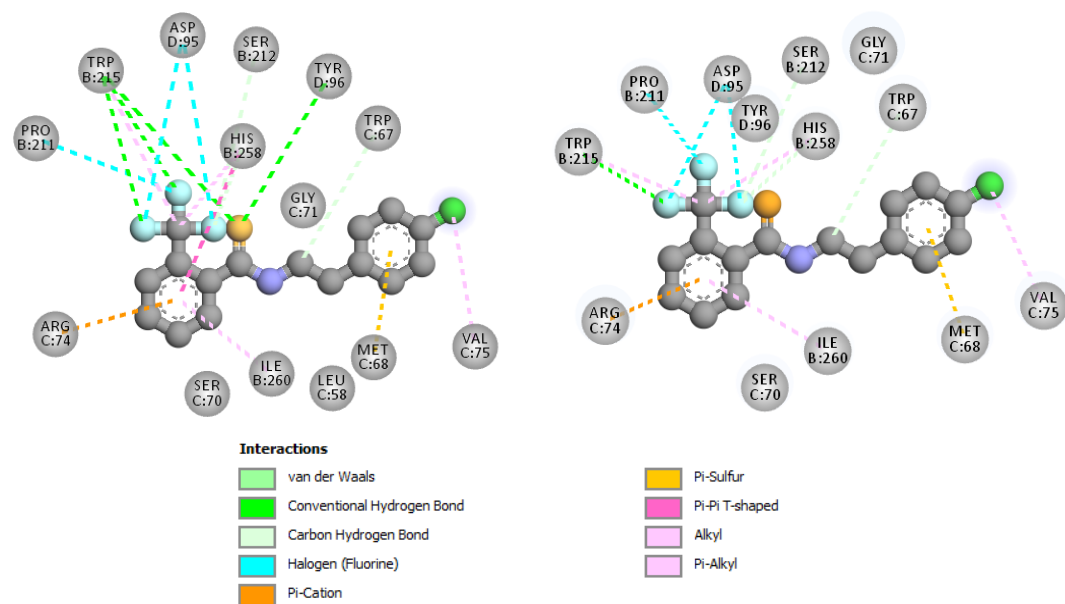
Supplementary Figure S1. Docking optimization. *A.suum* Complex II structures deposited in PDB were retrieved to check the ability of the docking protocol to replicate experimental binding modes of complex II Q-site ligands. For each ligand, the crystallographic pose is shown in green superimposed with the docking pose with best S score, depicted in red. Receptor molecular surface and relevant interacting residues (TRP251B in orange, TYR96D in cyan and ARG76C in pink) are shown. In all cases the docking pose is in agreement with the crystallographic one and except for 4YTN, all satisfy the RMSD criteria. Cross docking experiments yielded similar results.

4. Docking poses obtained for Wact-11 bioisostere derivatives:



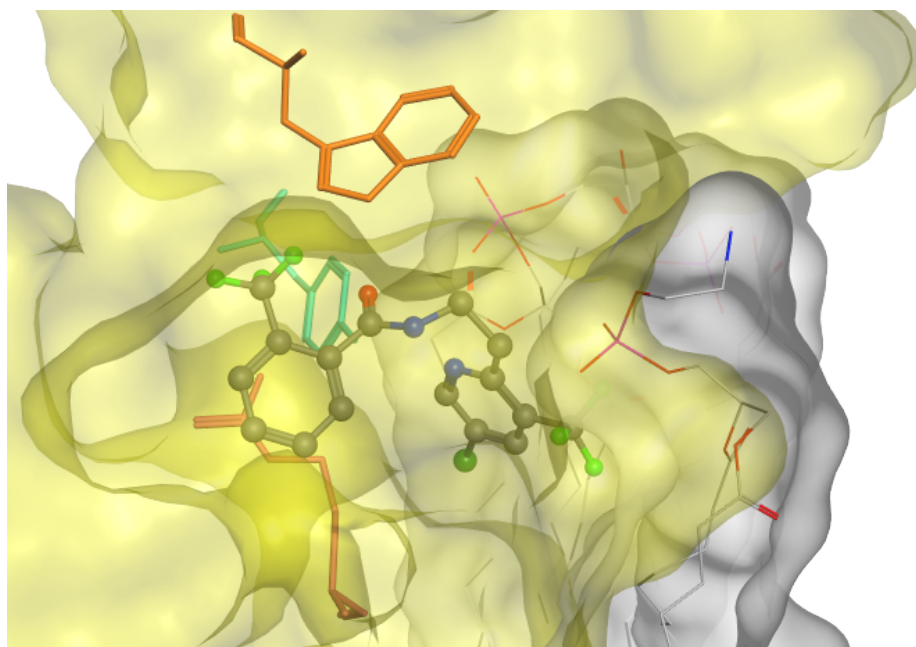
Supplementary Figure S2. Docking poses obtained for Wact-11 bioisostere derivatives 2-8c.

5. Interaction diagrams for compounds 3 and 4.



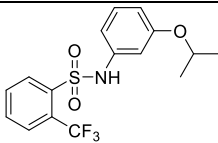
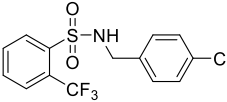
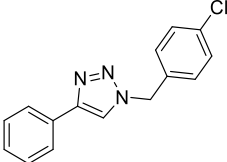
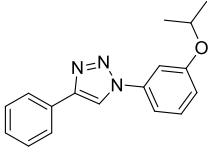
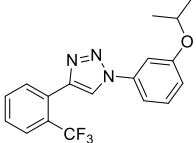
Supplementary Figure S3. Interaction diagram for 3 (left) and 4 (right) docked in *C. elegans* complex II, generated in Discovery Studio Visualizer 16, Biovia Corp.

6. Simulated mitochondrial membrane with Fluopyram.



Supplementary Figure S4. Fluopyram docked in *C. elegans* complex II. Main interacting residues are shown (Trp215 of subunit B in orange, Tyr96 of subunit D in cyan and Arg74 of subunit C in pink). The pose is representative of that obtained for all benzamides.

7. *C. elegans* L1 motility assay results for compounds 8a, 8c, 9a-c.

Entry	Compound	Structure	% <i>C. elegans</i> motility reduction (10 μ M) ^a
1	8a		0
2	8b		2
3	9a		0
4	9b		0
5	9c		4

- a. Calculated compared to vehicle motility (DMSO 1%) taken as 0% motility reduction. Compounds were evaluated at 10 μ M after 20 h.

Supplementary Table S1. Library screening in *C. elegans* L1 of non-Wact11 analogous compounds.