

Review

Bioactivities from Marine Algae of the Genus *Gracilaria*

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Abstract: Seaweeds are an important source of bioactive metabolites for the pharmaceutical industry in drug development. Many of these compounds are used to treat diseases like cancer, acquired immune-deficiency syndrome (AIDS), inflammation, pain, arthritis, as well as viral, bacterial, and fungal infections. This paper offers a survey of the literature for *Gracilaria* algae extracts with biological activity, and identifies avenues for future research. Nineteen species of this genus that were tested for antibacterial, antiviral, antifungal, antihypertensive, cytotoxic, spermicidal, embriotoxic, and anti-inflammatory activities are cited from the 121 references consulted.

Keywords: *Gracilaria*; macroalgae; seaweed; biological activity; natural product; review

1. Introduction

The ocean environment contains over 80% of world's plant and animal species [1] and with more than 150,000 seaweeds found in the intertidal zones and tropical waters of the oceans, it is a primary source of natural products [2].

Seaweeds are floating and submerged plants of shallow marine meadows. They have salt tolerance because the osmolarity of cytoplasm is adjusted to match the osmolarity of the seawater so that desiccation does not occur. They lack true stems, roots and leaves; however, they possess a blade that is leaf like, a stipe that is stem like, and a holdfast that resembles roots like terrestrial plants. Seaweeds contain photosynthetic pigments and use sunlight to produce food and oxygen from carbon dioxide, and the water [3].

Marine macroalgae are important ecologically and commercially to many regions of the world, especially in Asian countries such as China, Japan and Korea [4]. They are a valuable food resource which contains low calories, and they are rich in vitamins, minerals, proteins, polysaccharides, steroids and dietary fibers [5–7]. Since as early as 3000 BC, they were also considered important as traditional remedies [4]. The Japanese and Chinese use brown algae in the treatment of hyperthyroidism and other glandular disorders [8–11]. The unsaturated lipids afford protection against cardiovascular pathogens [12].

Seaweeds have been one of the richest and most promising sources of bioactive primary and secondary metabolites [13] and their discovery has significantly expanded in the past three decades [4,14,15]. The algae synthesize a variety of compounds such as carotenoids, terpenoids, xanthophylls, chlorophyll, vitamins, saturated and polyunsaturated fatty acids, amino acids, acetogenins, antioxidants such as polyphenols, alkaloids, halogenated compounds and polysaccharides such as agar, carrageenan, proteoglycans, alginate, laminaran, rhamnan sulfate, galactosyl glycerol and fucoidan [16–25].

These compounds probably have diverse simultaneous functions for the seaweeds and can act as allelopathic, antimicrobial, antifouling, and herbivore deterrents, or as ultraviolet-screening agents [26]. They are also used by the pharmaceutical industry in drug development to treat diseases like cancer, acquired immune-deficiency syndrome (AIDS), inflammation, pain, arthritis, infection for virus, bacteria and fungus [27]. Currently, algae represent about 9% of biomedical compounds obtained from the sea [28].

Compounds with cytostatic, antiviral, antihelmintic, antifungal and antibacterial activities have been detected in green, brown and red algae [29,30]. The algae produce pure forms of the fatty acids found in human milk that appear to be building blocks for mental and visual development [31] and have been extensively screened for syntheses of new drugs [32,33].

During the 1970s, Ryther and collaborators evaluated numerous species of red, green and brown macroalgae for their potential growth rates and dry weight yields [34]. They demonstrated that the genus *Gracilaria* was the most attractive candidate because of its ability to achieve high yields and while producing commercially valuable extracts [35].

Gracilaria Greville genus (Gracilariales, Rhodophyta) is a macroalgae group with more than 300 species of which 160 have been accepted taxonomically. These are usually red, green or greenish brown with a three-phase cycle and can be found in tropical and subtropical seas [36,37].

The *Gracilaria* species are important for the industrial and biotechnological uses because they have phycocolloids, the main source of agar α -(1,4)-3,6-anhydro-L-galactose and β -(1,3)-D-galactose with little esterification in cell wall [2,38]. Among the carbohydrates, agar and other polysaccharides are present in *G. confervoides* [39], *G. dura* [40], *G. chilensi* and *G. secundata* [41,42].

These algae also produce important bioactive metabolites like the primary compound with antibiotic activity acrylic acid [43], and the eicosanoids which are derivatives C₂₀ polyunsaturated fatty acid (PUFA) metabolism through oxidative pathways that originate mainly from arachidonic acid and eicosapentaenoic acids, the precursors of prostaglandins (PGs) [44,45]. Species such as *G. asiatica* and *G. lichenoides* contain PGE₂ [46,47]. PGF₂ and 15-keto-PGE₂ were respectively isolated from *G. lichenoides* and *G. asiatica* [45]; *G. verrucosa* contains PGA₂ that appears to be responsible for a gastrointestinal disorder, known as “ogonori” poisoning in Japan [48].

Lipids are abundant in this genus being mainly prostaglandins [49], steroids, such as cholesterol and clinoasterol are present in *G. crassa* and *G. coronopifolia* respectively [50–52], as well as *G. longa* [48,53–57] and *G. dura*. Other steroids such as 3-beta-hydroxy-poriferast-5-en-7-one, 3-beta-7-alpha-diol-poriferast-5-ene and 5-alpha-poriferast-9(11)-en-3-beta-ol are isolated from *G. dura* [50]; cholestane-3-β-5-diol,5-α:24(S)-ethyl [52], poriferastene 8 [50], poriferast-5-ene-3-β-7-β-diol [51] and poriferast-5-ene-3-β-7-α-diol [51] were identified in *G. coronopifolia*; *G. longa* also has a variety of compounds like alpha linolenic acid, gamma linolenic acid [58], glycolipids [59], 5-dehydro avenasterol, fucosterol, myristic acid, desmosterol and 5-alpha-24(S)-ethyl-cholestane-3-beta-6-beta-diol [60]. Phytochemical studies with extracts from fresh thallus of *G. andersoniana* showed the following isolates: oleic acid, linoleic acid, cholesterol, prostaglandin A₂, prostaglandin E₂, leukotriene B₄ and phytol [61–63].

Studies with *G. asiatica* reported the diterpenes cis and trans-phytol [63]. A variety of lactones are present in *Gracilaria* from the Pacific Ocean, such as aplysiatoxin isolated from *G. confervoides* [64,65], polycavernoside B, polycavernoside B₂, and polycavernoside A₂ and A₃ isolated from *G. crassa* [49,66]. Other constituents are also contained in this genus such as proteins r-phycoerythrin from *G. salicornia* [67] and *G. longa* [68], gigartinine from *G. chilensis* [69] and proteoglycan from *G. longa* [70].

The possibility of finding new molecules from natural products is immeasurable. For this reason the plants and their derivatives are major sources of all drugs, affecting about 30% of pharmaceutical market [71]. According to Newman *et al.* (2003), between the years 1981 and 2002, 877 new molecules were introduced into the market, with 49% of substances isolated from natural sources followed by semi-synthetic derivatives or synthesized molecules taking the structures of natural origin as models [29].

The search for new effective medicines remains a challenge for scientists. Therefore around the world, many researchers have focused on natural sources for new molecules with algae among the targets of these studies. So in this study we reviewed the literature related to bioactivities for *Gracilaria* algae.

2. Results and Discussion

In this review, among the 160 species of *Gracilaria* already identified taxonomically, only 19 of them had their extracts and fractions chemically tested for toxicity, cytotoxic, spermicidal, antiimplantation, antibacterial, antiviral, antifungal, antiprotozoa, antihypertensive, antioxidant, anti-inflammatory, analgesic, and spasmolytic effects in gastrointestinal tract (Table 1).

Table 1. Bioactivities of marine algae of the *Gracilaria* genus.

Botanical Name	Part used	Type of extract	Bioassays models, organism, dose or route of administration	Result
Studies of toxicity				
<i>Gracilaria bursa-pastoris</i> (S.G.Gmelin) P.C.Silva	FzDTh	H ₂ O Ext.	Cytotoxic activity-cell culture-10.0 µg/mL	Inactive [72]
	FTh	95% EtOH Ext. or CHCl ₃ Ext.	Cytotoxic activity-cell culture-10.0 µg/mL	Inactive [72]
<i>Gracilaria chorda</i> (Holmes)	FsO	H ₂ O Ext.	Toxicity assessment-mouse-1.2 mg/animal-i.p.	Active [48]
<i>Gracilaria coronopifolia</i> (J. Agardh)	FTh	Plant	Toxic effect-human adult-oral	Active [65]
<i>Gracilaria corticata</i> (J.Agardh) J.Agardh	Th	50% EtOH-H ₂ O Ext.	Toxicity assessment-mouse-DL50 1000 mg/kg-ip	Active [73]
<i>Gracilaria domingensis</i> (Kützing) Sonder ex Dickie	DO	90% EtOH Ext.	Cytotoxicity- <i>Artemia salina</i> L.-200 µg/mL	Active [74]
<i>Gracilaria edulis</i> (S.G.Gmelin) P.C.Silva	DTh	Plant	Toxicity effect (death)-human adult-oral	Active [49]
<i>Gracilaria foliifera</i> (Forsskål) Borgesen	SDTh	90% EtOH Ext.	Toxicity assessment-mouse-DL50 0.825 mg/kg-i.p.	Active [75]
	DEP	(1:1) EtOH-H ₂ O Ext.	Cytotoxic activity-cell culture-dose: dry weight of plant	Active [76]
<i>Gracilaria textorii</i> (Suringar) De Toni	FzDO	MeOH Ext.	Cytotoxic activity-cell culture (CA 9 KB)	Inactive [77]
	FsTh	Hexane Ext. CCl ₄ Ext. CHCl ₃ Ext.	Cytotoxic activity-culture cell (LEUK P 388)-ED 50 > 100 µg/mL Cytotoxic activity-culture cell (LEUK P 388)-ED 50 22.2 µg/mL Cytotoxic activity-culture cell (LEUK P 388)-ED 50 32.2 µg/mL	Equivocal [78] Equivocal [78] Inactive [78]

Table 1. *Cont.*

Botanical Name	Part used	Type of extract	Bioassays models, organism, dose or route of administration	Result
<i>Gracilaria verrucosa</i> (Hudson) Papenfuss	DO	H ₂ O Ext.	Toxicity assessment-mouse-1.2 mg/animal-i.p.	Active [48]
	FzDO	MeOH Ext.	Cytotoxic activity-cell culture (CA 9 KB)	Inactive [77]
	FO	30% EtOH Ext. (1:1) CHCl ₃ - MeOH Ext.	Cytotoxic activity-cell culture (CA 9 KB)-10.0 µg/mL Cytotoxic activity-cell culture (CA 9 KB)-1.0 µg/mL	Inactive [79] Equivocal [79]
	FTh	H ₂ O Ext. and 95% EtOH Ext.	Cytotoxic activity-cell culture (LEUK P 388-P 3)-10.0 µg/µL	Inactive [72]
Effects on the nervous system				
<i>Gracilaria corticata</i> J.Agardh	SDTh	90% EtOH Ext.	Autonomic effects-dog-50 mg/kg-iv	Inactive [75]
			CNS effects-mouse	Inactive [75]
			Analgesic activity-mouse	Inactive [75]
			Anticonvulsant activity-mouse	Inactive [75]
<i>Gracilaria edulis</i> (S.G.Gmelin) P.C.Silva	SDTh	90% EtOH Ext.	Autonomic effects-dog-50 mg/kg-iv	Inactive [75]
			CNS effects-mouse	Inactive [75]
			Analgesic activity-mouse	Inactive [75]
			Anticonvulsant activity-mouse	Inactive [75]
<i>Gracilaria verrucosa</i> (Hudson) Papenfuss	SDTh	90% EtOH Ext.	CNS effects-mouse	Inactive [75]
Contraception activity				
<i>Gracilaria corticata</i> J.Agardh	DTh	(1:1) MeOH- CH ₂ Cl ₂ Ext.	Embryotoxic effect-pregnant rat-1.0 mg/kg-intragastric	Inactive [80]
	SDTh	90% EtOH Ext.	Antiimplantation effect-pregnant rat-100.0 mg/kg Spermicidal effect-rat-2.0 %	Inactive [75] Inactive [75]

Table 1. *Cont.*

Botanical Name	Part used	Type of extract	Bioassays models, organism, dose or route of administration	Result
<i>Gracilaria edulis</i> (S.G.Gmelin) P.C.Silva	SDTh	90% EtOH Ext.	Antiimplantation effect-pregnant rat-100.0 mg/kg	Inactive [75]
<i>Gracilaria verrucosa</i> (Hudson) Papenfuss	SDTh	90% EtOH Ext.	Spermicidal effect-rat-2.0% Spermicidal effect-rat-2.0%	Inactive [75] Inactive [75]
Anti-inflammatory activity				
<i>Gracilaria textorii</i> (Suringar) De Toni	EP	H ₂ O Ext.	Platelet aggregation inhibition (adenosine diphosphate; arachidonic acid or collagen stimulation)-100.0 µg/mL Venotonic activity (platelet aggregating factor stimulation)-100.0 µg/mL	Inactive [81] Inactive [81]
<i>Gracilaria verrucosa</i> (Hudson) Papenfuss	DTh	Polysaccharide fraction	Immunostimulant activity-mouse-4.0 mg/animal-i.p. Phagocytosis stimulation-mouse-4.0 mg/animal-i.p.	Active [82] Active [82]
	SDTh	90% EtOH Ext.	Antiinflammatory activity-rat-intragastric	Inactive [75]
Antioxidant activity				
<i>Gracilaria verrucosa</i> (Hudson) Papenfuss	Plant	MeOH Ext.	Radical scavenging effect (DPPH radicals)-IC50 480.0 µg	Active [83]
	DTh	Polysaccharide fraction	Oxygen radical formation induction-mouse-4.0 mg/animal-i.p.	Active [82]
Gastrointestinal effects				
<i>Gracilaria chorda</i> (Holmes)	FsO	H ₂ O Ext.	Mouse-0.5 mg/animal-gastric intubation and dose 0.5 mg/loop-i.p.	Active [48]
<i>Gracilaria verrucosa</i> (Hudson) Papenfuss	DO	H ₂ O Ext.	Mouse-0.5 mg/animal-gastric intubation	Active [48]

Table 1. Cont.

Botanical Name	Part used	Type of extract	Bioassays models, organism, dose or route of administration	Result
Cardiovascular effects				
<i>Gracilaria corticata</i> (J.Agardh) J.Agardh	SDTh	90% EtOH Ext.	Cardiovascular effects-dog-50 mg/kg-iv	Inactive [75]
<i>Gracilaria edulis</i> (S.G.Gmelin) P.C.Silva				
<i>Gracilaria lichenoides</i> (Greville)	EP	H ₂ O Ext.	Diuretic activity-rat-intragastric	Active [75]
	FsTh	H ₂ O Ext.	Antihypertensive activity-rat-iv	Active [84]
<i>Gracilaria verrucosa</i> (Hudson) Papenfuss	SDTh	90%EtOH Ext.	Antihypertensive activity-rat-iv	Active [85]
<i>Gracilaria verrucosa</i> (Hudson) Papenfuss	SDTh	90%EtOH Ext.	Cardiovascular effects-dog-50 mg/kg-iv	Inactive [75]
Hypoglycemic activity				
<i>Gracilaria corticata</i> J.Agardh	SDTh	90% EtOH Ext.	Rat-250 mg/kg – intragastric	Inactive [75]
<i>Gracilaria edulis</i> (S.G.Gmelin) P.C.Silva	SDTh	90% EtOH Ext.	Rat-250.0 mg/kg – intragastric	Inactive [75]
Anti-enzymes activity				
<i>Gracilaria arcuata</i> (Zanardini)	DTh	MeOH Ext.	Tyrosinase inhibition-0.33 mg/mL	Inactive [86]
<i>Gracilaria corticata</i> (J.Agardh) J.Agardh	DO	PET Ether Ext.; CHCl ₃ Ext. or MeOH Ext.	Penicillinase inhibition-1.0 µg/units	Inactive [87]
<i>Gracilaria textorii</i> (Suringar) De Toni	EP	H ₂ O Ext.	Aldose reductase inhibition-10.0 µg/mL	Inactive [81]
<i>Gracilaria verrucosa</i> (Hudson) Papenfuss	FzDO	MeOH Ext.	Cyclic AMP phosphodiesterase inhibition	Inactive [77]
	FzDO	EtOAc Ext. MeOH Ext.	Lipase inhibition Cyclic AMP phosphodiesterase inhibition	Equivocal [88] Inactive [77]

Table 1. Cont.

Botanical Name	Part used	Type of extract	Bioassays models, organism, dose or route of administration	Result
Respiratory effects				
<i>Gracilaria corticata</i> (J.Agardh) J.Agardh	SDTh	90% EtOH Ext.	Respiratory depressant-dog-50 mg/kg-iv	Inactive [75]
<i>Gracilaria edulis</i> (S.G.Gmelin) P.C.Silva				
<i>Gracilaria verrucosa</i> (Hudson) Papenfuss	SDTh	90% EtOH Ext.	Respiratory depressant-dog-50.0 mg/kg-iv	Inactive [75]
Spasmolytic activity				
<i>Gracilaria corticata</i> (J.Agardh) J.Agardh	SDTh	90% EtOH Ext.	Spasmolytic activity-guinea pig	Inactive [75]
<i>Gracilaria edulis</i> (S.G.Gmelin) P.C.Silva	SDTh	90% EtOH Ext.	Negative chronotropic effect-dog-50.0 mg/kg-iv	Inactive [75]
Antibacterial activity				
<i>Gracilaria cervicornis</i> (Turner) J.Agardh	DEP	95% EtOH Ext.	Agar plate- <i>Staphylococcus aureus</i> -5.0 mg/mL	Active [89]
			Agar plate- <i>Proteus vulgaris</i> ; <i>Escherichia coli</i> ; <i>Aspergillus fumigates</i> ; <i>Candida albicans</i> ; <i>Pseudomonas aeruginosa</i> ; <i>Streptococcus pyogenes</i> -50.0 mg/mL	Inactive [89]
<i>Gracilaria corticata</i> (J.Agardh) J.Agardh	DO	PET Ether Ext.; CHCl ₃ Ext. or MeOH Ext.	Agar plate- <i>Staphylococcus aureus</i> ; <i>Escherichia coli</i> -MIC >200 µg/mL	Inactive [90]
	FsO	MeOH Ext.	Agar plate- <i>Escherichia coli</i> ; <i>Salmonella paratyphi A</i> ; <i>Salmonella paratyphi B</i> ; <i>Shigella sonnei</i>	Inactive [91]
			Agar plate- <i>Bacillus subtilis</i> ; <i>Staphylococcus aureus</i> ; <i>Bacillus megaterium</i> ; <i>Streptococcus viridans</i>	Active [91]
	SDTh	90% EtOH Ext.	Agar plate- <i>Klebsiella pneumonia</i> ; <i>Pseudomonas aeruginosa</i> ; <i>Staphylococcus aureus</i> ; <i>Escherichia coli</i> ; <i>Streptococcus faecalis</i>	Inactive [75]

Table 1. Cont.

Botanical Name	Part used	Type of extract	Bioassays models, organism, dose or route of administration	Result
<i>Gracilaria debilis</i> (Forsskål) Borgesen	DO	95% EtOH Ext.	Agar plate- <i>Escherichia coli</i> ; <i>Staphylococcus aureus</i>	Active [92]
<i>Gracilaria domingensis</i> (Kützing) Sonder ex Dickie	DO	95% EtOH Ext.	Agar plate-Mycobacterium smegmatis Agar plate- <i>Escherichia coli</i> ; <i>Staphylococcus aureus</i>	Inactive [92] Active [92]
<i>Gracilaria edulis</i> (S.G.Gmelin) P.C.Silva	SDTh	Acetone Ext. or Ether Ext. 95% EtOH Ext. or Acetone Ext.	Agar plate- <i>Escherichia coli</i> ; <i>Staphylococcus aureus</i> Agar plate-Mycobacterium smegmatis	Inactive [92] Active [92]
<i>Gracilaria pygmea</i> (Borgesen)	FsO	MeOH Ext.	Agar plate- <i>Bacillus subtilis</i> ; <i>Staphylococcus aureus</i> ; <i>Escherichia coli</i> ; <i>Salmonella paratyphi A</i> ; <i>Streptococcus viridans</i> ; <i>Shigella sonnei</i> ; <i>Salmonella paratyphi B</i> Agar plate- <i>Bacillus megaterium</i>	Inactive [91] Active [91]
<i>Gracilaria sjoestedii</i> (Kylin)	DO	95% EtOH Ext.	Agar plate- <i>Escherichia coli</i> ; <i>Staphylococcus aureus</i>	Active [92]
<i>Gracilaria tikvahiae</i> McLachlan	DEP	CHCl ₃ Ext. or EtOH Ext.	Agar plate-Mycobacterium smegmatis Agar plate - <i>Staphylococcus aureus</i>	Inactive [92] Active [93]
<i>Gracilaria verrucosa</i> (Hudson) Papenfuss	FTh	**	Agar plate- <i>Streptococcus faecalis</i> ; <i>Pseudomonas aeruginosa</i> Agar plate- <i>Vibrio marinofulvis</i> ; <i>Micrococcus imfimus</i> ; <i>Pseudomonas atlantica</i> -40.0 µg/µL	Inactive [93] Inactive [94]
	Th	70% EtOH Ext.	Antiphage activity-agar plate-Bacteriophage T 1; Bacteriophage T 2; Bacteriophage T 4; Bacteriophage T 7; Bacteriophage MS 2; Bacteriophage PHI-CHI 174-0.50 µg/mL	Inactive [95]

Table 1. Cont.

Botanical Name	Part used	Type of extract	Bioassays models, organism, dose or route of administration	Result
Antifungal activity				
<i>Gracilaria corticata</i> (J.Agardh) J.Agardh	FsO	MeOH Ext.	Agar plate- <i>Aspergillus niger</i> ; <i>Fusarium solani</i> ; <i>Alternaria solani</i> ; <i>Penicillium funiculosum</i>	Inactive [91]
	SDTh	90% EtOH Ext.	Agar plate- <i>Sporotrichum schenckii</i> ; <i>Cryptococcun neoformans</i> ; <i>Candida albicans</i> ; <i>Trichophyton mentagrophytes</i> ; <i>Aspergillus fumigates</i>	Inactive [75]
<i>Gracilaria debilis</i> (Forsskål) Borgesen	DO	95% EtOH Ext.	Agar plate- <i>Candida albicans</i>	Active [92]
			Agar plate- <i>Neurospora crassa</i>	Inactive [92]
<i>Gracilaria domingensis</i> (Kützing) Sonder ex Dickie	DO	95% EtOH Ext. and Acetone Ext.	Agar plate- <i>Candida albicans</i> ; <i>Neurospora crassa</i>	Active [92]
		Ether Ext.	Agar plate- <i>Candida albicans</i>	Inactive [92]
<i>Gracilaria edulis</i> (S.G.Gmelin) P.C.Silva	SDTh	90% EtOH Ext.	Agar plate- <i>Sporotrichum schenckii</i> ; <i>Candida albicans</i> ; <i>Cryptococcus neoformans</i> ; <i>Trichophyton mentagrophytes</i> ; <i>Aspergillus fumigates</i>	Inactive [75]
<i>Gracilaria pygmea</i> (Borgesen)	FsO	MeOH Ext.	Agar plate- <i>Aspergillus niger</i> ; <i>Fusarium solani</i> ; <i>Alternaria solani</i> ; <i>Penicillium funiculosum</i>	Inactive [91]
<i>Gracilaria sjoestedii</i> (Kylin)	DO	95% EtOH Ext.	Agar plate- <i>Candida albicans</i>	Active [92]
			Agar plate- <i>Neurospora crassa</i>	Inactive [92]
<i>Gracilaria tikvahiae</i> McLachlan	DEP	CHCl ₃ Ext. and EtOH Ext.	Agar plate- <i>Candida albicans</i>	Active [93]
Antiviral activity				
<i>Gracilaria bursa-pastoris</i> (S.G.Gmelin) P.C.Silva	FzDTh	**	Cell culture- <i>Herpes simplex</i> 1 and HIV Virus	Inactive [96]
<i>Gracilaria corticata</i> (J.Agardh) J.Agardh	Th	50% EtOH-H ₂ O Ext.	Cell culture- <i>Semlicki-forest</i> Virus-0.05 mg/mL	Equivocal [73]
	SDTh	90% EtOH Ext.	Cell culture- <i>Ranikhet</i> and <i>Vaccinia</i> Virus-0.05 mg/mL Cell culture- <i>Ranikhet</i> Virus	Inactive [73] Inactive [75]

Table 1. Cont.

Botanical Name	Part used	Type of extract	Bioassays models, organism, dose or route of administration	Result
<i>Gracilaria edulis</i> (S.G.Gmelin) P.C.Silva	SDTh	90% EtOH Ext.	Cell culture-Semlicki-forest and Ranikhet Virus	Inactive [75]
<i>Gracilaria pacifica</i> (I. A. Abbott)	DO	MeOH Ext.	Cell culture- <i>Herpes simplex</i> 1 Virus-400.0 µg/mL Cell culture- <i>Virus sindbis</i> -200.0 µg/mL	Inactive [97] Active [97]
<i>Gracilaria</i> species	FzDTh	**	Cell culture-Herpes simplex 1 and HIV Virus	Inactive [96]
<i>Gracilaria textorii</i> (Suringar) De Toni	FzDO	MeOH Ext.	Cell culture- <i>Herpes simplex</i> 1 Virus	Inactive [77]
	Th	H ₂ O Ext.	Cell culture-HIV Virus-MIC > 1000 µg/mL	Inactive [98]
	Fresh	MeOH Ext.	Epstein-Barr virus early antigen activation inhibition (telocidin b-4 induced Epstein-Barr virus induced activation)-4.0 µg/mL	** [99]
<i>Gracilaria verrucosa</i> (Hudson) Papenfuss	FzDO	MeOH Ext.	Cell culture- <i>Herpes simplex</i> 1 Virus	Inactive [77]
Antiprotozoal activity				
<i>Gracilaria corticata</i> (J.Agardh) J.Agardh	SDTh	90% EtOH Ext.	Agar plate- <i>Entamoeba histolytica</i> ; <i>Plasmodium berghei</i>	Inactive [75]
<i>Gracilaria edulis</i> (S.G.Gmelin) P.C.Silva	SDTh	90% EtOH Ext.	Agar plate- <i>Entamoeba histolytica</i> ; <i>Plasmodium berghei</i>	Inactive [75]
Allelopathic activity				
<i>Gracilaria compressa</i> (C.Agardh) Greville	DEP	95% EtOH Ext.	Agar plate- <i>Helianthus tuberosus</i> -dose: dry weight of plant	Active [76]
<i>Gracilaria foliifera</i> (Forsskål) Borgesen	DEP	H ₂ O Ext.	Agar plate- <i>Helianthus tuberosus</i> -dose: dry weight of plant	Active [76]
<i>Gracilaria verrucosa</i> (Hudson) Papenfuss	DEP	95% EtOH Ext.	Agar plate- <i>Helianthus tuberosus</i> -dose: dry weight of plant	Active [76]

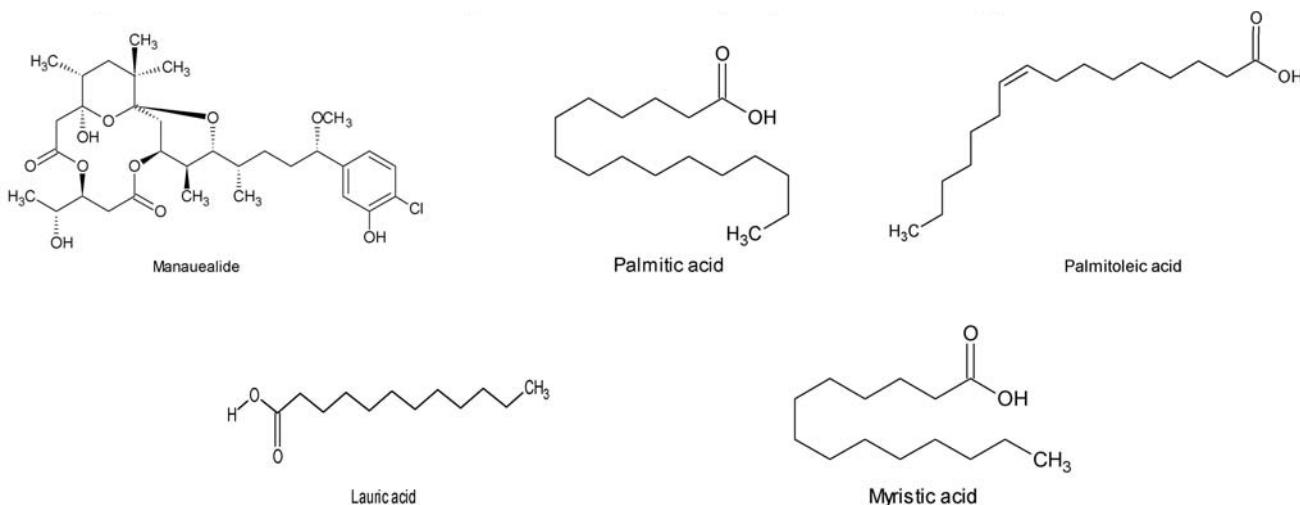
Legend: DEP = Dried entire plant; DO = Dried organism; DTh = Dried thallus; EP = Entire plant; FO = Frozen organism; FsO = Fresh organism; FsTh = Fresh thallus; FTh = Frozen thallus; FzDO = Freeze dried organism; FzDTh = Freeze Dried thallus; SDTh = Shade dried thallus; Th = Thallus; PET Ether); ** = Not stated.

These biological studies were mainly developed in Japan and Brazil. This fact is justified by the extensive coastlines and marine biodiversity and is influenced by several factors for the development of these species, such as temperature, radiation, salinity, metal ions and other chemically fundamental components. Australia and Guam have recently become interested in the study of algae and diverse marine species. The consumption of algae has increased in European countries in recent decades with 15 to 20 species of algae being marketed in Italy, France and Greece. In western countries like Venezuela, USA and Canada, the macroalgae are industrially used as a source of hydrocolloids agar, carrageenan and alginate [100]. Carrageenan has been found to be useful in ulcer therapy and alginates are known to prolong the period of activity of certain drugs [8–11].

2.1. Studies of Toxicity

In France, extract studies with ethanol/water draw up from dried entire plant of *G. foliifera* showed toxicity in humans when treated with oral dose and cytotoxicity studies [75,76]. *G. coronopifolia* and *G. edulis* were also toxic to humans [65,49] (See Table 1). Carbohydrate, heparin [97], agar [101], manaealide A, manaealide B [64], manaealide C [102], palmitic, palmitoleic, oleic, lauric and myristic acids [103], steroids and alkaloids malyngamide [104] were found in these species (Figure 1).

Figure 1. Structure of the compounds found in *G. foliifera*, *G. coronopifolia* and *G. edulis*.

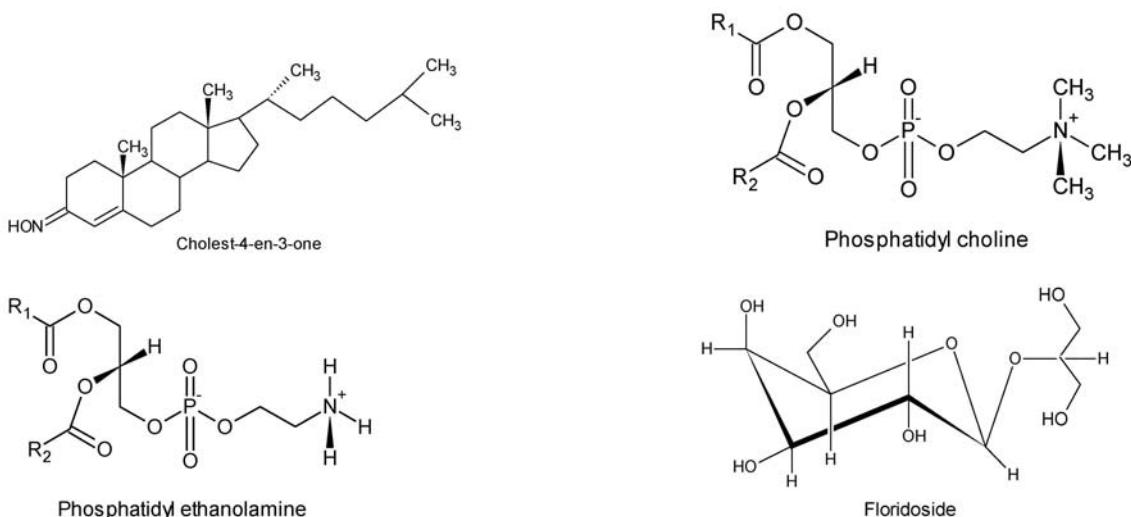


There is currently a tendency to substitute the use of laboratory animals in toxicological tests with alternative methods to reduce their numbers in experiments, or refine the existing methodology in order to minimize pain and stress [105]. A rapid and effective alternative to realize primary toxicity and biological action screening of compounds is the estimation of the 50% lethal concentration (LC_{50}) through brine shrimp assay using *Artemia salina* L. [106]. A 90% ethanol extract of *G. domingensis* had LC_{50} of 200 μ g/mL against *A. salina* [74].

Another method to evaluate toxicity is determining cytotoxic activity. In this context aqueous extract from dried thallus of *G. bursa-pastoris* (10.0 μ g/mL), chloroform and methanol extracts from *G. textorii*, which was isolated steroid cholest-4-en-3-one [107], and ethanol extract from *G. verrucosa* were not toxic in cell culture. However, aqueous extract from *G. verrucosa* at a dose of 1.2 mg/animal showed toxicity to mice [48], according to Table 1. In this seaweed, lipids were indentified,

such as PGF_α [84,85], glycerol, ethanolamine-phosphatidyl [58], choline-phosphatidyl [58,108], ethanol-phosphatidyl [58], floridoside [109], and carbohydrates, such as agar [110–113] (Figure 2).

Figure 2. Structure composed of species of *Gracilaria* tested in cytotoxicity.



2.2. Effects on the Nervous System

Studies related to nervous system are important to understanding and treat complex degenerative and behavioral diseases. 90% ethanol extracts from *G. corticata*, *G. edulis* and *G. verrucosa* did not cause central or periphery effects for mice or dogs (50 mg/kg), and did not show analgesic or anticonvulsant activities for mice [75] (Table 1).

2.3. Contraception Activity

The researchers have also investigated new molecules with anticonceptive action; the post-coital contraceptive action of marine seaweeds was also evaluated in animals. Methanol: methylene chloride (1:1) extract from *G. corticata* was orally administered at 500 or 1000 mg/kg/day to female rats from day 1 to day 7 of their pregnancies. Higher doses produced significant post-coital contraceptive activity due to enhanced pre-implantation without any marked side effects. These findings indicate that red marine algae are a potential source for post-coital contraceptive drugs [80].

90% Ethanol extracts from *G. edulis* (100 mg/kg) and *G. corticata* were inactivated before the antiimplantation effect when they tested in pregnant rats [75,80]. Ethanol extracts from shade dried thallus of *G. edulis* and *G. verrucosa* were inactive in spermicidal bioassays [75]. Extracts from *G. edulis* showed 100% inhibition of sperm motility and this effect was related to disruption of the plasma membrane by spermicidal compounds [3] (Table 1).

2.4. Anti-Inflammatory and Antioxidant Activities

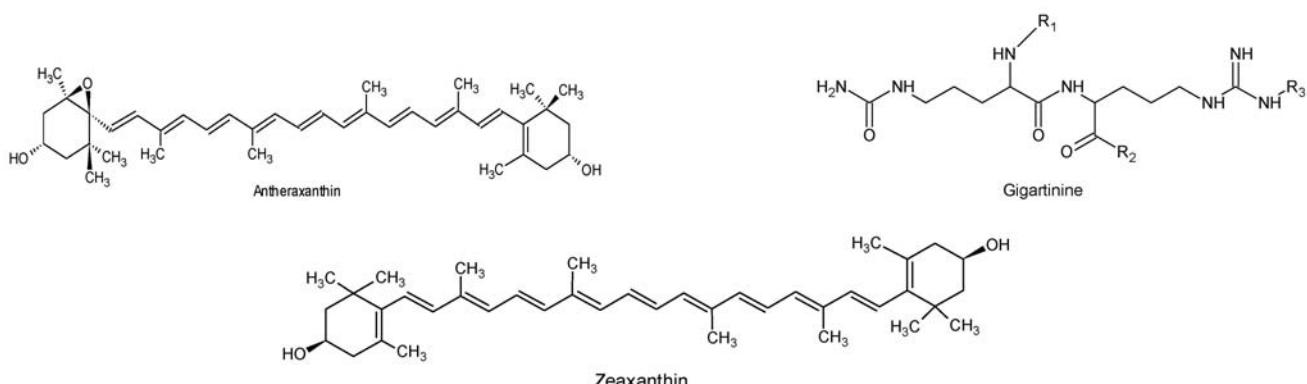
The anti-inflammatory activity of seaweeds has been studied. Polysaccharide fractions from *G. verrucosa* at a dose of 4.0 mg/animal were orally and intraperitoneally administered to mice and showed immunopotentiating activity stimulating phagocytosis [82]. Methanol extract and polysaccharide fractions from *G. verrucosa* were also antioxidant [82,83]. Aqueous extract from

G. textorii at a dose of 100 µg/mL did not inhibit platelet aggregation induced by adenosine diphosphate, arachidonic acid or collagen [81]. *G. verrucosa*, *G. asiatica*, *G. lichenoides* and others species contain PGE₂ [47,85], that have physiological effects including hyperthermia, hypotension, smooth muscle dilatation, hyperalgesia and gastric secretion inhibition [114,115] (Table 1).

2.5. Gastrointestinal Effects

Aqueous extract from dried *G. verrucosa* algae or fresh *G. chorda* algae at a dose of 0.5 mg/animal controlled gastrointestinal disorders in mice [48] (Table 1), resulting from zeaxanthin and antheraxanthin [116], carotenoids, pyrimidine 2-amino-4-carboxy, non-alkaloid nitrogen heterocycle [90], steroids, 5-alpha-poriferastane, 3-beta-6-alpha-diol poriferastane, 5-alpha-3-beta-6-beta-diol [51] and gigatinine [85] (Figure 3).

Figure 3. Structure of the compounds found in *G. verrucosa* and *G. chorda*.

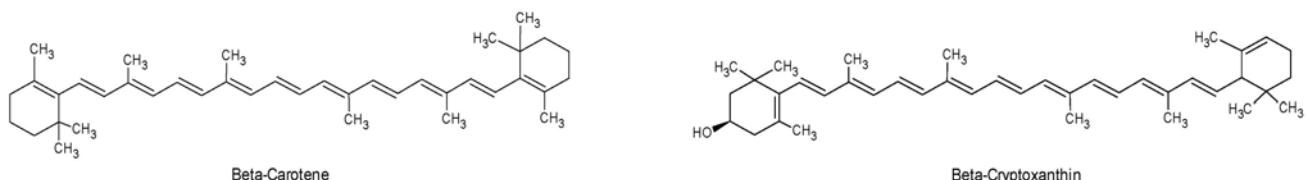


2.6. Cardiovascular Effects

90% Ethanol extracts from *G. corticata*, *G. edulis* and *G. verrucosa* showed no cardiovascular effects in dogs (50 mg/kg) [75]. 90 % ethanol extract from *G. edulis* showed diuretic activity [75]. Aqueous extract from *G. lichenoides* was administered intravenously in rats and it was antihypertensive [84]. Tyrosinase inhibition was not induced by methanol extract from *G. arcuata* [86] and aqueous extract from *G. textorii*, 10 µg/mL, was negligible on aldose reductase [83] (Table 1).

2.7. Antibiotic Activity

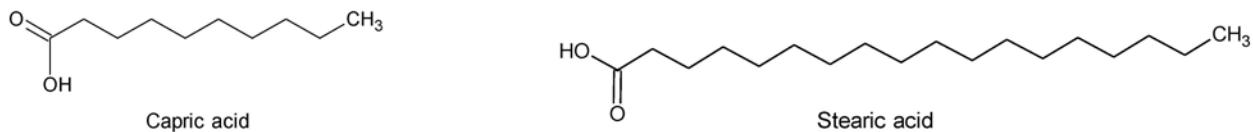
Extracts or ingredients from various algae have shown antibacterial activity *in vitro* against gram-positive and gram-negative bacteria [117]. The agar disc diffusion method for antibacterial susceptibility was used for evaluation and 6 mm discs were impregnated with 20 µL of the extracts and placed in inoculated Muller Hinton agar. Antibacterial activity from chloroform extract of *G. edulis* (Gmelin) Silva was tested against bacterial strains of *Vibrio cholera*, *Staphylococcus aureus*, *Shigella dysenteriae*, *Shigella boylei*, *Salmonella paratyphi*, *Pseudomonas aeruginosa* and *Klebsiella pneumonia* (Table 1). We observed higher activity for *G. edulis* extract than *S. aureus* extract [12]. Yet it was inactive for *Sporotrichum schenckii*, *Candida albicans* and *Cryptococcus neoformans* [75]. In the present investigation, the chemical compounds isolated from the species were steroids (carotenoids, β-cryptoxanthin and β-carotene) [118] and carbohydrates [84,85,119] (Figure 4).

Figure 4. Chemical structure of the steroids isolated from *G. edulis*.

Mahasneh *et al.* (1995) demonstrated activity of organic extracts from algae against multi-resistant bacteria to antibiotics [120]. Ethanol extract from *G. debilis* showed antibacterial activity against *S. aureus* but was inactive against *Mycobacterium smegmatis* [92].

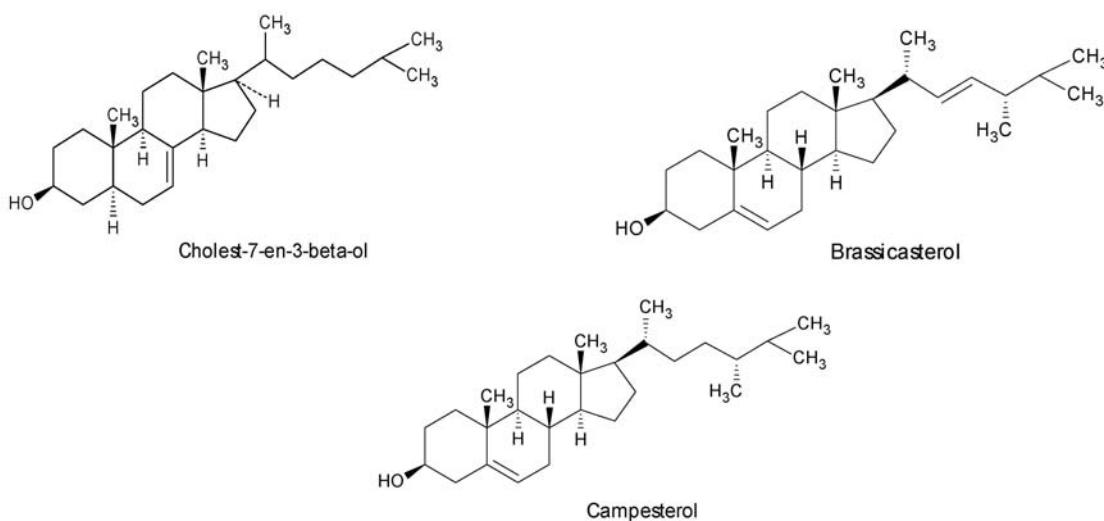
95% ethanol extract from whole dried *G. cervicornis* algae was active against *S. aureus* at a concentration of 5.0 mg/mL [89]. Methanol extract from fresh *G. corticata* was active against *Bacillus subtilis*, *Bacillus megaterium*, *S. aureus* and *Streptococcus viridians* [91].

G. corticata and *G. pygmea* did not inhibit the growth of *Aspergillus niger*, *Fusarium solani*, *Alternaria solani*, or *Penicillium funiculosum* [91]. Petroleum ether, chloroform and methanol extracts from this seaweed at a concentration of 1.0 µg/units proved to be inactive on the inhibition of penicillinase enzyme [87]. From this specie, stearic lipids and capric acids were isolated [121] (Figure 5).

Figure 5. Structure of compounds found in species of *Gracilaria* with antifungal activity.

Ethanol extracts from *G. domingensis* and *G. sjoestedii* showed antibacterial activity against *E. coli* and *S. aureus*. Ethanol extracts from *G. debilis*, *G. domingensis* and *G. sjoestedii* were active against *Candida albicans* shown by agar plate method [92]; Chloroform, ether and methanol extracts from *G. tikvahiae* were inactive [93]. The growth of *Neurospora crassa* was not inhibited by extracts from *G. sjoestedii* and *G. debilis*; ethanol extract from *G. domingensis* was active against *Mycobacterium smegmatis* and *Neurospora crassa* [92]. *G. domingensis* has as chemical constituents, polysaccharide CT-1 [122], palmitic acid and steroids (stigmasterol, sitosterol, campesterol, cholestan-7-en-3-β-ol and brassicasterol) [52] (Figure 6).

Figure 6. Structure of the steroids isolated from *G. domingensis*.

Figure 6. Cont.

Some studies highlighting antiparasitic activity of seaweeds also were verified. 90 % ethanol extract from *G. corticata* and *G. edulis* were tested against *Entamoeba histolytica* and *Plasmodium berghei* and were not active [75].

2.8. Antiviral Activity

Extracts from *G. bursa-pastoris* and *Gracilaria* sp were inactive against the *Herpes simplex* 1 virus (HSV) and the human immunodeficiency virus (HIV) when evaluated in cell cultures [96]. Granin BP and citrullinyl-arginine proteins were isolated from these extracts [123,124]. Methanol extract from dried *G. pacifica* at a concentration of 200.0 µg/mL was active against *Sindbis* virus, but was not effective against *H. simplex* 1 when tested at a concentration of 400 µg/mL. Extracts and compounds obtained from *Gracilaria* sp with anti-HIV activity are also active against other retroviruses such as HSV. However, the pharmacodynamic mechanisms of the antiretroviral activity are still unknown because bioactive compounds from seaweed poorly investigated [9] (Table 1) (Figure 7).

Figure 7. Structure of a compound found in *Gracilaria* sp and *G. bursa-pastoris*.

3. Material and Methods

In this article, some reports about bioactivity of *Gracilaria* algae were reviewed in the specialized literature published up to January 2011. The search was carried out using data banks such as; Biological Abstracts, AlgaeBase, SciFinder Scholar, Pubmed and NAPRALERT (acronym for Natural Products ALERT-University of Illinois in Chicago, USA).

4. Conclusions

Algae are abundant in the oceans and represent a rich source of as yet unknown secondary metabolites. In this review, we found only a few studies with complete chemical profiles and pharmacological potential of the *Gracilaria* species. Most studies raised concerns about antimicrobial activity against *Staphylococcus*, *Streptococcus*, *Candida* and *Herpes* genus. Others referenced the cytotoxicity bioassays in which these algae species were not active, but they produce various types of prostaglandins and others substances that can be toxic to humans such as gastrointestinal disorders and lethality caused by *G. verrucosa* and *G. edulis*, respectively. To research new drugs it is necessary to evaluate other bioassay models to preserve the safety, efficacy and quality of the end products. In Brazil, there is a great need for toxicological, pharmacological, preclinical and clinical studies, as recommended by the RDC 48/2004.

Finally, we conclude that algae of the *Gracilaria* genus are a potential source for synthesis of new natural medicines. It is important to taxonomically classify and standardize extractions, while identifying the active compounds to attenuate possible environmental interference that could undermine the pharmachemical profile, and thus generate different pharmacologic effects. In addition, it is important to sensitize corporate researchers and financial agencies to support this cause.

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References

1. Jha, R.K.; Zi-rong, X. Biomedical compounds from marine organisms. *Mar. Drugs* **2004**, *2*, 123–146.
2. Falcão, V.R. Aspectos moleculares de nitrato redutase da macroalga marinha *Gracilaria tenuistipitata* (Rhodophyta): Seqüenciamento do gene e estudo da expressão do RNA mensageiro. PhD Thesis, Institute of Chemical, University of São Paulo: São Paulo, Brazil, 2006; pp. 1–187.
3. Babuselvam, M.; Ravikumar, S. Screening of Male Anti-Fertility Compounds from Marine Seaweed Macro Algae. Division of Marine Microbiology and Medicine, Manonmaniam Sundaranar University: Rajakkamangalam, India, 1993; pp. 1–14. Available online: http://www.scisoc.or.th/stt/32/sec_h/paper/stt32_H_H0001.pdf (accessed on 14 May 2011).
4. Smit, A.J. Medicinal and pharmaceutical uses of seaweed natural products: A review. *J. Appl. Phycol.* **2004**, *16*, 245–262.
5. Ito, K.; Hori, K. Seaweed: Chemical composition and potential uses. *Food Rev. Int.* **1989**, *5*, 101–1144.
6. Darcy-Vrillon, B. Nutritional aspects of the developing use of marine macroalgae for the human food industry. *Int. J. Food Sci. Nutr.* **1993**, *44*, 523–535.
7. Lahaye, M. Marine algae as a source of dietary fibers: Determination of soluble and insoluble dietary fiber contents in some ‘sea vegetable’. *J. Sci. Food Agric.* **1993**, *54*, 523–535.

8. Elena, M.; Francisco, Y.; Erickson, K.L. Mailiohydrin, a cytotoxic chamigrene dibromohydrin from a Phillipine *Laurencia* species. *J. Nat. Prod.* **2001**, *64*, 790–791.
9. Kim, J.B.; Hudson, A.M.; Huang, K.; Bannistes, A.; Jin, T.J.; Choi, G.H.N.; Towers, Y.K.; Wrede, R.E. Biological activity of seaweed extracts from British, Colombia, Canada and Korea. I. Antiviral activity. *Can. J. Bot.* **1997**, *75*, 1656–1660.
10. Okai, Y.; Highasi, O.K.; Ishizaka, S.; Yamashita, U. Enhancing effect of polysaccharides from a edible brown algae, *Hijikia furiform* (Hijki) on release of tumour necrosis factor alpha from macrophages of endotoxin non responder C3H/HCl mice. *Nutr. Cancer* **1997**, *27*, 381–386.
11. Premila, J.C.; Raviraja, N.S.; Sridhar, K.R. Antimicrobial activity of some marine algae of south-west coast of India. *Indian J. Mar. Sci.* **1996**, *26*, 201–205.
12. Vallinayagam, K.; Arumugan, R.; Ragupathi Raja Kannan, R.R.R.; Thirumaram, G.; Anantharaman, P. Antibacterial activity of some selected seaweeds from Pudumadam coastal regions. *Glob. J. Pharmacol.* **2009**, *3*, 50–52.
13. Faulkner, D.J. Marine natural products. *Nat. Prod. Rep.* **2002**, *19*, 1–48.
14. Cardozo, K.H.M.; Guaratini, T.; Barros, M.P.; Falcão, V.R.; Tonon, A.P.; Lopes, N.P.; Campos, S.; Torres, M.A.; Souza, A.O.; Colepicolo, P.; et al. Metabolites from algae with economical impact. *Comp. Biochem. Physiol. C: Comp. Pharmacol.* **2006**, *146*, 60–78.
15. O’Sullivan, L.; Murphy, B.; McLoughlin, P.; Duggan, P.; Lawlor, P.G.; Hughes, H.; Gardiner, G.E. Prebiotics from marine macroalgae for human and animal health application. *Mar. Drugs* **2010**, *8*, 2038–2064.
16. Paniagua-Michel, J.; Capa-Robles, W.; Olmos-Soto, J.; Gutierrez-Millan, L.E. The carotenogenesis pathway via the isoprenoid-beta-carotene interference approach in a new strain of *Dunaliella salina* isolated from Baja California Mexico. *Mar. Drugs* **2009**, *7*, 45–56.
17. Cen-Pacheco, F.; Nordstrom, L.; Souto, M.L.; Martin, M.N.; Fernandez, J.J.; Daranas, A.H. Studies on polyethers produced by red algae. *Mar. Drugs* **2010**, *8*, 1178–1188.
18. Klisch, M.; Hader, D.P. Mycosporine-like amino acids and marine toxins—The common and the different. *Mar. Drugs* **2008**, *6*, 147–163.
19. Pallela, R.; Na-Young, Y.; Kim, S.K. Anti-photoaging and photoprotective compounds derived from marine organisms. *Mar. Drugs* **2010**, *8*, 1189–1202.
20. D’Ayala, G.G.; Malinconico, M.; Laurienzo, P. Marine derived polysaccharides for biomedical applications: Chemical modification approaches. *Molecules* **2008**, *13*, 2069–2106.
21. Kellmann, R.; Stuken, A.; Orr, R.J.S.; Svendsen, H.M.; Jakobsen, K.S. Biosynthesis and molecular genetics of polyketides in marine Dinoflagellates. *Mar. Drugs* **2010**, *8*, 1011–1048.
22. Souza, E.T.; Lira, D.P.; Queiroz, A.C.; Silva, D.J.C.; Aquino, A.B.; Mella, E.A.C.; Lorenzo, V.P.; Miranda, G.E.C.; Araújo-Júnior, J.X.; Chaves, M.C.O.; et al. The antinociceptive and anti-inflammatory activities of caulerpin, a bisindole alkaloid isolated from seaweeds of the genus *Caulerpa*. *Mar. Drugs* **2009**, *7*, 689–704.
23. Guven, K.C.; Percot, A.; Sezik, E. Alkaloids in marine algae. *Mar. Drugs* **2010**, *8*, 269–284.
24. Cabrita, M.T.; Vale, C.; Rauter, A.P. Halogenated compounds from marine algae. *Mar. Drugs* **2010**, *8*, 2301–2317.

25. La Barre, S.; Potin, P.; Leblanc, C.; Delage, L. The halogenated metabolism of brown algae (Phaeophyta), its biological importance and its environmental significance. *Mar. Drugs* **2010**, *8*, 988–1010.
26. Ianora, A.; Boersma, M.; Casotti, R.; Fontana, A.; Harder, J.; Hoffmann F.; Pavia, H.; Potin, P.; Poulet, S.A.; Toth, G. New trends in marine chemical ecology. *Estuaries Coasts* **2006**, *29*, 531–551.
27. Deig, E.F.; Ehresmann, D.W.; Hatch, M.T.; Riedlinger, D.J. Inhibition of herpesvirus replication by marine algae extracts. *Antimicrob. Agents Chemother.* **1974**, *6*, 524–525.
28. Jha, R.K.; Zi-rong, X. Biomedical compounds from marine organisms. *Mar. Drugs* **2004**, *2*, 123–146.
29. Newman, D.J.; Cragg, G.M.; Snader, K.M. Natural products as sources of new drugs over the period 1981–2002. *J. Nat. Prod.* **2003**, *66*, 1022–1037.
30. Lindequist, U.; Schweder, T. Marine biotechnology, In *Biotechnology*; Rehm, H.J., Reed, G., Eds.; Wley-VHC: Weinheim, Germany, 2001; Volume 10, pp. 441–484.
31. Linsert, P. Revolution in infant formula brewing in Herman’s calves. DNA algae. *Genet. Eng. Biotechnol. Monit.* **1994**, *1*, 45–46.
32. Khotimchenko, S.V.; Vaskovsky, V.E.; Titlyanavo, T.V. Fatty acids of marine algae from the 12 substances from marine algae Puerto Rico. *Antimicrob. Agents Chemother.* **1963**, *161*, 68–72.
34. Ryther, J.H.; Goldman, J.C.; Gifford, C.E. Physical models of integrated waste recycling marine polyculture systems. *Aquaculture* **1975**, *5*, 163–177.
35. Capo, T.R.; Jaramillo, J.C.; Boyd, A.E.; Lapointe, B.E.; Serafy, J.E. Sustained high yields of *Gracilaria* (Rodophyta) grown in intensive large-scale culture. *J. Appl. Phycol.* **1999**, *11*, 143–147.
36. Guiry, M.D. AlgaeBase. Martin Ryan Institute, National University of Ireland: Galway, Ireland, 1996–2011. Available online: <http://www.algaebase.org> (accessed on 14 May 2011).
37. Skriptsova, A.V.; Titlyanova, T.V.; Titlyanov, E.A. Red algae of the genus *Gracilaria* in south of the Russian far east. *Russ. J. Mar. Biol.* **2001**, *27*, S38–S52.
38. Kain, J.M.; Destombe, C. A review of the life history, reproduction and phenology of *Gracilaria*. *J. Appl. Phycol.* **1995**, *7*, 69–281.
39. Misawa, M. Production of natural substances by plant cell cultures described in Japanese patents. In *Plant Tissue Culture Its Bio-Technol*; Barz, W., Reinhard, E., Zenk, M.H., Eds.; Springer-Verlag: Berlin, Germany, 1977; pp. 17–26.
40. Murano, E.; Toffanin, R.; Paoletti, S.; Rizzo, R. Pyruvate-rich agarose from the red alga *Gracilaria dura*. *Planta Med.* **1992**, *58*, A588–A589.
41. Hemmingson, J.A.; Furneaux, R.X.; Murray-brown, V.H. Biosynthesis of agar polysaccharides in *Gracilaria chilensis* bird. *Carbohydr. Res.* **1996**, *287*, 101–115.
42. Brasch, D.J.; Chuah, C.T.; Melton, L.D. Marine algal polysaccharides, Part 2. The agar-type polysaccharide from the red alga *Gracilaria secundata*. *Carbohydr. Res.* **1983**, *115*, 191–198.
43. Glombitzka, K.W. *Marine Algae in Pharmaceutical Science*; Hoppe, H.A., Levring, T., Eds.; Walter de Gruyter: New York, NY, USA, 1979; Volume 1, pp. 303–342.
44. Glickman, M. Utilisation of seaweed hydrocolloids in the food industry. *Hydrobiologia* **1987**, *151/152*, 31–47.

45. Imbs, A.B.; Vologodskaya, A.V.; Nevshupova, N.V.; Khotimchenko, S.V.; Titlyanov, E.A. Response of prostaglandin content in the red alga *Gracilaria verrucosa* to season and solar irradiance. *Phytochemistry* **2001**, *58*, 1067–1072.
46. Chapman, V.J.; Chapman, D.J. *Seaweeds and Their Uses*; Springer-Verlag: Berlin, Germany, 1980.
47. Sajiki, J. Effect of acetic acid treatment on the concentrations of arachidonic acid and prostaglandin E2 in the red algae, *Gracilaria asiatica* and *G. rhodocaudata*. *Fish Sci.* **1997**, *63*, 128–131.
48. Fusetani, N.; Hashimoto, K. Prostaglandin E2: A candidate for causative agent of “Ogonori” poisoning. *Bull. Jpn. Soc. Sci. Fish.* **1984**, *50*, 465–469.
49. Yotsu-yamashita, M.; Haddock, R.L.; Yasumoto, T. Polycavernoside A: A novel glycosidic macrolide from the red alga *Polycavernosa tsudai* (*Gracilaria edulis*). *J. Am. Chem. Soc.* **1993**, *115*, 1147–1148.
50. Das, B.; Srinivas, K.V.N.S. Minor C29-steroids from the marine red alga, *Gracilaria edulis*. *Phytochemistry* **1992**, *31*, 2427–2429.
51. Das, B.; Srinivas, K.V.N.S. Dihydroxysterols from the marine red alga, *Gracilaria edulis*. *Phytochemistry* **1992**, *31*, 4731–4373.
52. Combres, A.; Bianchini, J.P.; Gaydou, E.M. Fatty acid and sterol composition of red algae of the Indian ocean. *Oceanol. Acta* **1986**, *9*, 339–342.
53. Toffanin, R.; Murano, E.; Modricky, C.; Kvam, B.J.; Paoletti, S.; Rizzo, R.; Pollesello P. Free and acylated cholesterol in the red alga *Gracilaria longa*: detection and quantification by 1H- and 13C-NMR on lipid extracts. *Planta Med.* **1992**, *58*, A589–A590.
54. Castedo, L.; Quintela, J.M.; Vilalta, R. Sterols from red and brown algae from the Galician coast. *An. Quim. Ser. C* **1985**, *8*, 113–115.
55. Vilalta, R.; Quintela, J.M.; Riguera, R.; Castedo, L. Natural marine products from algae and cnidaria of the Galician estuaries. *Cuad. Area Cienc. Mar.* **1984**, *2*, 617–625.
56. Henriquez, P.; Trucco, R.; Silva, M.; Sammes, P.G. Cholesterol in *Iridaea laminariooides* and *Gracilaria verrucosa*. *Phytochemistry* **1972**, *11*, 1171.
57. Anon. Prostaglandin E-2. *Patent-Japan Kokai Tokkyo Koho* **1982**, *59*, 73–565.
58. Araki, S.; Sakurai, T.; Oohusa, T.; Kayama, M. Component fatty acid of lipid from *Gracilaria verrucosa*. *Bull. Jpn. Soc. Sci. Fish.* **1986**, *52*, 1871–1874.
59. Son, B.W. Glycolipids from *Gracilaria verrucosa*. *Phytochemistry* **1990**, *29*, 307–309.
60. Das, B.; Srinivas, K.V.N.S. Two new sterols from the marine red alga *Gracilaria edulis*. *Planta Med.* **1993**, *59*, 572–573.
61. Sajiki, J.; Hakimi, H. Identification of eicosanoids in the red algae, *Gracilaria asiatica*, using high-performance liquid chromatography and electrospray ionization mass spectrometry. *J. Chromatogr.* **1998**, *795*, 227–237.
62. Ravi, B. N.; Faulkner, D.J. Acyclic diterpenes from the marine sponge *Didiscus* sp. *J. Org. Chem.* **1979**, *44*, 968–970.
63. Sims, J.J.; Pettus, J.A., Jr. Isolation of free *cis* and *trans*-phytol from the red alga *Gracilaria andersoniana*. *Phytochemistry* **1976**, *15*, 1076–1077.

64. Nagai, H.; Yasumoto, T.; Hokama, Y. Manaealides, some of the causative agents of a red alga *Gracilaria coronopifolia* poisoning in Hawaii. *J. Nat. Prod.* **1997**, *60*, 925–928.
65. Nagai, H.; Yasumoto, T.; Hokama, Y. Aplysiatoxin and debromoaplysiatoxin as the causative agents of a red alga *Gracilaria coronopifolia* poisoning in Hawaii. *Toxicon* **1996**, *34*, 753–761.
66. Yotsu-yamashita, M.; Seki, T.; Paul, V.J.; Naoki, H.; Yasumoto, T. Four new analogs of polycavernoside A. *Tetrahedron Lett.* **1995**, *36*, 5563–5566.
67. D’agnolo, E.; Rizzo, R.; Paoletti, S.; Murano, E. R-phycoerythrin from the red alga *Gracilaria longa*. *Phytochemistry* **1994**, *35*, 693–696.
68. Talarico, L.; Kosovel, V. Properties and ultrastructures of r-phycoerythrin from *Gracilaria verrucosa*. *Photosynthetica* **1978**, *12*, 369–374.
69. Wilcox, S.J.; Bloor, S.J.; Hemmingson, J.A.; Furneaux, R.H.; Nelson, W.A. The presence of gigartinine in a New Zealand *Gracilaria* species. *J. Appl. Phycol.* **2001**, *13*, 409–413.
70. Kanoh, H.; Kitamura, T.; Kaboyashi, Y. A sulfated proteoglycan from the red alga *Gracilaria verrucosa* is a hemagglutinin. *Comp. Biochem. Physiol. B* **1992**, *102*, 445–449.
71. Kirkpatrick, P. Antibacterial drugs. Stitching together naturally. *Nat. Rev. Drug Discovery* **2002**, *1*, 748–748.
72. Kosovel, V.; Avanzini, A.; Scarcia, V.; Furlani, A. Algae as possible sources of antitumoural agents: Preliminary evaluation of the “*in vitro*” cytostatic activity of crude extracts. *Pharmacol. Res. Commun.* **1988**, *20*, 27–31.
73. Kamat, S.Y.; Wahidulla, S.; D’Souza, L.; Naik, C.G.; Ambiye, V.; Bhakuni, D.S.; Goel, A.K.; Garg, H.S.; Srimal, R.C. Bioactivity of marine organisms. VI. Antiviral evaluation of marine algal extracts from the Indian Coast. *Bot. Mar.* **1992**, *35*, 161–164.
74. Lhullier, C.; Horta, P.A.; Falkenberg, M. Avaliação de extratos de macroalgas bênticas do litoral catarinense utilizando o teste de letalidade para *Artemia salina*. *Rev. Bras. Farmacogn.* **2006**, *16*, 158–163.
75. Bhakuni, D.S.; Dhawan, B.N.; Garg, H.S.; Goel, A.K.; Mehrotra, B.N.; Srimal, R.C.; Srivastava, M.N. Bioactivity of marine organisms: Part VI-Screening of some marine flora from Indian coasts. *Indian J. Exp. Biol.* **1992**, *30*, 512–517.
76. Chenieux, J.C.; Verbist, J.F.; Biard, J.F.; Clement, E.; Le Boterff, J.; Maupas, P.; Lecocq, M. Seaweeds of French atlantic coast with antimitotic activity. *Planta Med.* **1980**, *40*, 152–162.
77. Arisawa, M.; Hayashi, K.; Nikaido, T.; Koike, K.; Fujita, D.; Nunomura, N.; Tanaka, M.; Sasaki, T. Screening of some marine organism extracts for camp phosphodiesterase inhibition, cytotoxicity, and antiviral activity against HSV-1. *Int. J. Pharmacogn.* **1997**, *35*, 6–11.
78. Numata, A.; Kanbara, S.; Takahashi, C.; Fujiki, R.; Yoneda, M.; Fujita, E.; Nabeshima, Y. Cytotoxic activity of marine algae and a cytotoxic principle of the brown alga *Sargassum tortile*. *Chem. Pharm. Bull.* **1991**, *39*, 2129–2131.
79. Avanzini, A.; Kosovel, V.; Scarcia, V.; Furlani, A.; Ravalico, L. Green, red and brown algae from North Adriatic sea as source of possible cytotoxic products. *Fitoterapia* **1987**, *58*, 391–394.
80. Ratnasooriya, W.D.; Premakumara, G.A.S.; Tillekeratne, L.M.V. Post-coital contraceptive activity of crude extracts of Sri Lankan marine red algae. *Contraception* **1994**, *50*, 291–299.

81. Okada, Y.; Miyauchi, N.; Suzuki, K.; Kobayashi, T.; Tsutsui, C.; Mayuzumi, K.; Okuyama, T. Search for naturally occurring substances for prevention against the complications of diabetes; inhibitory effect on aldose reductase and platelet aggregation. *Nat. Med.* **1994**, *48*, 324–329.
82. Yoshizawa, Y.; Tsunehiro, J.; Nomura, K.; Itoh, M.; Fukui, F.; Ametani, A.; Kaminogawa, S. *In vivo* macrophage-stimulation activity of the enzyme-degraded water-soluble polysaccharide fraction from a marine alga (*Gracilaria verrucosa*). *Biosci. Biotechnol. Biochem.* **1996**, *60*, 1667–1671.
83. Choi, J.S.; Lee, J.H.; Park, H.J.; Kim, H.G.; Young, H.S.; Mun, S.I. Screening for antioxidant activity of plants and marine algae and its active principles from *Prunus davidiana*. *Korean J. Pharmacogn.* **1993**, *24*, 299–303.
84. Funayama, S.; Hikino, H. Hypotensive principles from plants. *Heterocycles* **1981**, *15*, 1239–1256.
85. Gregson, R.P.; Marwood, J.F.; Quinn, R.J. The occurrence of prostaglandins PGE2 and PGF2 α in a plant—the red alga *Gracilaria lichenoides*. *Tetrahedron Lett.* **1979**, *20*, 4505–4506.
86. Choi, B.W.; Lee, B.H.; Kang, K.J.; Lee, E.S.; Lee, N.H. Screening of the tyrosinase inhibitors from marine algae and medicinal plants. *Korean J. Pharmacogn.* **1998**, *29*, 237–242.
87. Sridhar, P.; Lakshmi, V.V.; Polasa, H.; Reddy, V.S.; Rao, C.H.P.; Srimannarayana, G. Biological activity of some marine algal extracts. *Indian J. Mar. Sci.* **1984**, *13*, 90–91.
88. Bitou, N.; Inomiya, M.; Tsujita, T.; Okuda, H. Screening of lipase inhibitors from marine algae. *Lipids* **1999**, *34*, 441–445.
89. Perez, R.M.; Avila, J.G.; Perez, S.; Martinez, A.; Martinez, G. Antimicrobial activity of some American algae. *J. Ethnopharmacol.* **1990**, *29*, 111–116.
90. Salleh, A.; Wati Haron, N.; Mahmud, N.; Mohammad, J. Distribution of pyrimidine derivatives in algae. *Biochem. Syst. Ecol.* **1994**, *22*, 860.
91. Usmanghani, K.; Shameel, M.; Sualeh, M.; Khan, K.H.; Mahmood, Z.A. Antibacterial and antifungal activities of marine algae from Karachi seashore of Pakistan. *Fitoterapia* **1984**, *55*, 73–77.
92. Albuquerque, M.R.; Campos-Takaki, Koenig, M.L. Detection of antimicrobial activity in marine seaweeds. *Rev. Inst. Antibiot. Univ. Fed. Pernambuco Recife* **1983**, *21*, 127–138.
93. Oranday, M.A.; Verde, M.S.J.; Martínez-Lozano, N.H.; Waksman, J. Active fractions from four species of marine algae. *Phyton* **2004**, *73*, 165–170.
94. Lustigman, B.; Lee, L.H.; Thees, N.; Masucci, J. Production of antibacterial substances by macroalgae of the New York/New Jersey coast, USA. *Bull. Environ. Contam. Toxicol.* **1992**, *49*, 743–749.
95. Couladis, M.; Vagias, C.; Roussis, V.; Verykokidou, E.; Skaltsa, H. Antiphage properties of some Greek algae extracts. *Phytomedicine* **1998**, *5*, 479–483.
96. Hayashi, K.; Hamada, J.; Hayashi, T. A screening strategy for selection of anti-HSV-1 and anti-HIV extracts from algae. *Phytother. Res.* **1996**, *10*, 233–237.
97. Kamat, S.Y.; Wahidulla, S.; D'Souza, L.; Naik, C.G.; Ambiye, V.; Bhakuni, D.S.; Goel, A.K.; Garg, H.S.; Srimal, R.C. Bioactivity of marine organisms. VI. Antiviral evaluation of marine algal extracts from the Indian Coast. *Bot. Mar.* **1992**, *35*, 161–164.

98. Nakamura, H.; Ohnuki, N.; Sadamasu, K.; Sekine, H.; Tanaka, J.; Okada, Y.; Okuyama, T. Anti-human immunodeficiency virus (HIV) activities of aqueous extracts from marine algae. *Nat. Med.* **1994**, *48*, 173–179.
99. Ohigashi, H.; Sakai, Y.; Yamaguchi, K.; Umezaki, I.; Koshimizu, K. Possible anti-tumor promoting properties of marine algae and *in vivo* activity of wakame seaweed extract. *Biosci. Biotechnol. Biochem.* **1992**, *56*, 994–995.
100. Hsu, B.-Y.; Tsao, C.-Y.; Chiou, T.-K.; Hwang, D.-F. Factors affecting PGE2 production in seaweed *Gracilaria tenuistipitata*. *J. Food Drug Anal.* **2008**, *16*, 59–65.
101. Laserna, E.C.; Veroy, R.L.; Luistro, A.H.; Cajipe, G.J.B. Extracts from some red and brown seaweeds of the Philippines. In *Tenth International Seaweed Symposium*; Levring, T., Ed.; Walter de Gruyter & Co.: Berlin, Germany, 1980; pp. 443–448.
102. Nagai, H.; Kan, Y.; Fujita, T.; Sakamoto, B.; Kokama, Y. Manauealide C and anhydrodebromoaplysiatoxin, toxic constituents of the hawaiian red alga, *Gracilaria coronopifolia*. *Biosci. Biotechnol. Biochem.* **1998**, *62*, 1011–1013.
103. Parekh, K.S.; Parekh, H.H.; Rao, P.S. Fatty acid content of some Indian marine algae. *Indian J. Mar. Sci.* **1984**, *13*, 45–46.
104. Kan, Y.; Fujita, T.; Nagai, H.; Sakamoto, B.; Hokama, Y. Malyngamides M and N from the hawaiian red alga *Gracilaria coronopifolia*. *J. Nat. Prod.* **1998**, *61*, 152–155.
105. Parra, A.L.; Yhebra, R.S.; Sardiñas, I.G.; Buela, L.I. Comparative study of the assay of *Artemia salina* L. and the estimate of the medium lethal dose (LD₅₀ value) in mice, to determine oral acute toxicity of plant extracts. *Phytomedicine* **2001**, *8*, 395–400.
106. Meyer, B.N.; Ferrigni, N.R.; Putnam, J.E.; Jacobsen, L.B.; Nichols, D.E.; McLaughlin, J.L. Brine shrimp: A convenient general bioassay for active plant constituents. *Planta Med.* **1982**, *45*, 31–34.
107. Kanazawa, A.; Yoshioka, M. Occurrence of Cholest-4-en-3-one in the Red Alga *Gracilaria textorii*. In *Proceedings of the Seventh International Seaweed Symposium*, Sapporo, Japan, 1–12 August 1971; Wiley: New York, NY, USA, 1972; Volume 7, pp. 502–505.
108. Kunzler, K.; Eichenberger, W. Betaine lipids and zwitterionic phospholipids in plants and fungi. *Phytochemistry* **1997**, *46*, 883–892.
109. Roh, Y.S.; Son, W.; Im, K.S.; Choi, H.D. Structure of floridoside, a glycerol glycoside from the marine red alga *Gracilaria verrucosa*. *Korean J. Pharmacogn.* **1994**, *25*, 117–120.
110. Laserna, E.C.; Veroy, R.L.; Luistro, A.H.; Cajipe, G.J.B. Extracts from some red and brown seaweeds of the Philippines. In *Tenth International Seaweed Symposium*; Levring, T., Ed.; Walter de Gruyter & Co.: Berlin, Germany, 1980; pp. 443–448.
111. Matsuhashi, T. Effects of the freezing and drying method and the mechanical dehydration method by pressure on gel properties of dried agar. *Reito* **1974**, *49*, 756–760.
112. Shi, S.Y.; Zhang, Y.X.; Liu, W.Q.; Li, Z. Seasonal variation in yield, physical properties and chemical composition of agar from *Gracilaria verrucosa*. *Oceanol. Limnol. Sin.* **1983**, *14*, 272–278.
113. Friedlander, M.; Lipkin, Y.; Yaphe, W. Composition of agars from *Gracilaria verrucosa* and *Pterocladia capillacea*. *Bot. Mar.* **1981**, *24*, 595–598.

114. Minghetti, L.; Levi, G. Microglia as effector cells in brain damage and repair: Focus on prostanoids and nitric oxide. *Prog. Neurobiol.* **1998**, *54*, 99–125.
115. Nishihara, I.; Minami, T.; Uda, R.; Ito, S.; Hyodo, M.; Hayaishi, O. Effect of NMDA receptor antagonists on prostaglandin E2-induced hyperalgesia in conscious mice. *Brain Res.* **1995**, *677*, 138–144.
116. Aihara, M.S.; Yamamoto, H. Occurrence of antheraxanthin in two *Rhodophyceae Acanthophora spicifera* and *Gracilaria lichenoides*. *Phytochemistry* **1968**, *7*, 497–499.
117. Tuney, I.; Çadirci, B.H.; Unal, D.; Sukatar, A. Antimicrobial activities of the extracts of marine algae from the coast of Urla. *Turk. J. Biol.* **2006**, *30*, 171–175.
118. Aihara, M.S.; Yamamoto, H. Occurrence of antheraxanthin in two *Rhodophyceae Acanthophora spicifera* and *Gracilaria lichenoides*. *Phytochemistry* **1968**, *7*, 497–499.
119. Panossian, A.G. Search of prostaglandins and related compounds in plants: A review of the occurrence of prostaglandins and prostaglandin-like compounds in plants. *prostaglandins* **1987**, *33*, 363–381.
120. Mahasneh, I.; Jamal, M.; Kashashneh, M.; Zibdeh, M. Antibiotic-activity of marine-algae against multi-antibiotic resistant-bacteria. *Microbios* **1995**, *83*, 23–26.
121. Parekh, K.S.; Parekh, H.H.; Rao, P.S. Fatty acid content of some Indian marine algae. *Indian J. Mar. Sci.* **1984**, *13*, 45–46.
122. Fernandez, L.E.; Valiente, O.G.; Mainardi, V.; Bello, J.L.; Velez, H.; Rosado, A. Isolation and characterization of an antitumor active agar-type polysaccharide of *Gracilaria dominguensis*. *Carbohydr. Res.* **1989**, *190*, 77–83.
123. Okamoto, R.; Hori, K.; Miyazawa, K.; Ito, K. Isolation and charcterization of a new hemagglutinin from the red alga *Gracilaria bursa-pastoris*. *Experientia* **1990**, *46*, 975–977.
124. Laycock, M.V.; Craigie, J.S. The occurrence and seasonal variation of gigartinine and l-citrullinyl-l-arginine in *Chondrus crispus* stackh. *Can. J. Biochem.* **1977**, *55*, 27–30.

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