OPEN ACCESS International Journal of Molecular Sciences ISSN 1422-0067 www.mdpi.com/journal/ijms

Review

The Role of Neurotrophins in Multiple Sclerosis—Pathological and Clinical Implications

Alicja Kalinowska-Lyszczarz^{1,2,*} and Jacek Losy^{3,4}

- ¹ Department of Neurochemistry and Neuropathology, Chair of Neurology, Poznan University of Medical Sciences, 49, Przybyszewskiego st., 60-355 Poznan, Poland
- ² Heliodor Swiecicki University Hospital, 60-355 Poznan, Poland
- ³ Department of Clinical Neuroimmunology, Chair of Neurology, Poznan University of Medical Sciences, Poland; E-Mail: jlosy@ump.edu.pl
- ⁴ Neuroimmunological Unit, Institute of Experimental and Clinical Medicine, Polish Academy of Sciences, 60-355 Poznan, Poland
- * Author to whom correspondence should be addressed; E-Mail: akalinowskalyszczarz@ump.edu.pl; Tel.: +48-61-869-1454, Fax: +48-61-869-1697.

Received: 14 August 2012; in revised form: 28 September 2012 / Accepted: 10 October 2012 / Published: 22 October 2012

Abstract: Multiple sclerosis (MS) is a chronic inflammatory demyelinating and neurodegenerative disease of the central nervous system (CNS) with unknown etiology. It was recently suggested that autoimmunity, which had long been considered to be destructive in MS, might also play a protective role in the CNS of MS patients. Neurotrophins are polypeptides belonging to the neurotrophic factor family. While neurotrophins mediate cell survival and proliferation in the nervous system, they are also expressed within peripheral blood mononuclear cells fraction (PBMCs) of immunological system. In MS additional neurotrophic support from PBMCs might compensate relative neurotrophins deficiency in the damaged CNS tissue that needs to be repaired. Failure to produce the adequate neurotrophins concentrations might result in decreased protection of the CNS, consequently leading to increased atrophy, which is the main determinant of MS patients' end-point disability. There are several lines of evidence, both from clinical research and animal models, suggesting that neurotrophins play a pivotal role in neuroprotective and neuroregenerative processes that are often defective in the course of MS. It seems that neuroprotective strategies might be used as potentially valuable add-on therapies, alongside traditional immunomodulatory treatment in multiple sclerosis.

Keywords: multiple sclerosis; neurotrophins; neuroimmunology; peripheral blood mononuclear cells; neuroprotective autoimmunity

1. Introduction

Multiple sclerosis (MS) is a chronic inflammatory disease of the central nervous system (CNS), with yet unknown etiology, leading to formation of disseminated demyelinating lesions and accompanied by axonal degeneration. The disease affects predominantly young women, with the usual 2:1 ratio, as presented in a number of representative studies [1-3]. In its most typical form, namely relapsing-remitting MS (RRMS), it presents as attacks of diverse neurological symptoms, usually—but not necessarily—followed by conversion to the progressive phase and accumulation of disability in patients [4]. Disease severity varies from one patient to another, including benign, monosymptomatic cases, but also malignant and rapidly progressive ones. Median time from disease onset to progression depends mainly on the age of onset: the older the age, the shorter the time to secondary progression [1]. Several therapies are currently approved for MS treatment, and a considerate number is still under clinical trials. Their primary effect is the reduction of the annual relapse rate and of the radiological activity of the disease, as described by the number of new gadolinium-enhancing lesions and new T2-hyperintense lesions demonstrated in subsequent magnetic resonance imaging (MRI) scans; yet, their effect on disease progression is not satisfactory. Ever since the first descriptions of MS, the majority of research in the field has been focusing on its inflammatory component. In the recent years attention has also been drawn to its neurodegenerative aspect. In this context studies on the potentially protective role of neurotrophic factors have emerged in MS research. In this review the role of neurotrophins in multiple sclerosis is presented and potential future therapeutic options are discussed.

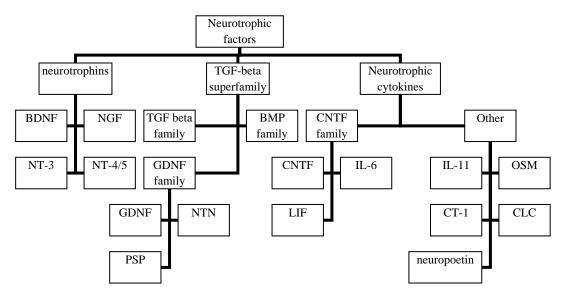
2. Biology of Neurotrophins

Neurotrophins are secretory polypeptides belonging to the neurotrophic factors' family (see Figure 1). During the development of the nervous system, neurotrophin deprivation serves as a physiological mechanism of neuronal elimination [5]. In the adult CNS, neurotrophins play a protective role towards specific neuronal populations. They also facilitate synaptic transmission and plasticity [6–9] that are crucial for memory and regeneration processes.

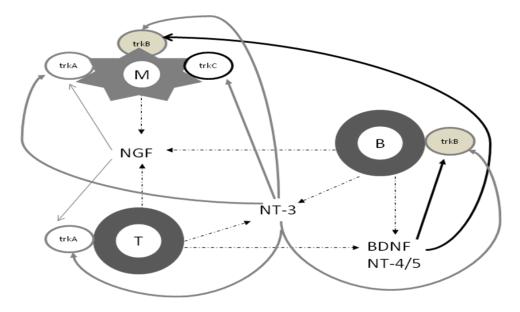
Nerve growth factor (NGF) was the first neurotrophin to be identified [10]. The family also includes brain-derived neurotrophic factor (BDNF), neurotrophin-3 (NT-3) and neurotrophin 4/5 (NT-4/5), otherwise called neurotrophin-4 (NT-4) or neurotrophin-5 (NT-5), with so far not identified in mammals neurotrophin-6 (NT-6) and neurotrophin-7 (NT-7) [11]. All neurotrophins are synthesized as precursor proteins and become biologically active by metalloproteinase cleavage [12]. Neurotrophins bind with high affinity to tropomyosin related kinase (trk) receptors [13], and—with low affinity—to p75^{NTR} receptors that belong to tumor necrosis factor (TNF) receptor superfamily [14]. Different neurotrophins bind preferentially to respective trk receptors [15], see Figure 2, which results in trk dimerization, trans-autophosphorylation and subsequent activation of such intracellular signaling

pathways as Ras/ERK and Akt/PI3K, both promoting cell survival [11,13]. On the other hand, the $p75^{NTR}$ molecule can act as a co-receptor for trk receptors, or it can induce an independent signaling pathway, promoting either survival (via NF- κ B), or apoptosis (Rac1/Jun N-terminal kinase, JNK) [16,17]. The $p75^{NTR}$ receptor is otherwise known as TNF receptor type 2 (TNFR-2). It is a known fact that disruption of TNF signaling pathway occurs in a number of autoimmune diseases, including MS. Interestingly, TNFR-2 pathway is an attractive drug target for autoimmune diseases, since its expression is restricted only to certain immune-cell populations, which could mean less side-effects than with the use of TNF-alpha alone, that acts on both, ubiquitously expressed TNFR-1 and more limited TNFR-2 [18]. The link between neurotrophins and TNF pathway obviously requires special attention. The biological results exerted by neurotrophins depend on their concentrations, receptor affinity and on their structure (pro-neurotrophins have higher affinity towards $p75^{NTR}$ and promote apoptosis) [12,19]. Neurotrophins can also co-operate with other receptors or ion channels. Interactions between trk receptors and voltage-gated potassium channels or NMDA receptors have been described [20,21]. The $p75^{NTR}$ can act as a co-receptor for the Nogo receptor that is known to inhibit axonal growth by interactions with MAG protein [22,23].

Figure 1. Neurotrophic factor family classification. NGF: nerve growth factor, BDNF: brain-derived neurotrophic factor, NT-3: neurotrophin 3, NT-4/5: neurotrophin 4/5, TGF beta: transforming growth factor beta, GDNF: glial derived neurotrophic factor, BMP: bone morphogenic proteins, NTN: neurturin, PSP: persephin, CNTF: ciliary neurotrophic factor, IL-6: interleukin 6, LIF: leukemia inhibitory factor, IL-11: interleukin 11, OSM: oncostatin M, CT-1: kardiotrophin-1, CLC: cardiothrophin-like cytokine.



neurotrophin 4/5, trk: tropomyosin related kinase receptor, M: macrophage, T: T lymphocyte, B: B lymphocyte. Dotted lines indicate secretion. Straight lines indicate affinity towards receptors.



3. Neurotrophins and the Immune System

The nervous and the immune systems are interconnected at several levels. Neurotrophins may serve as a fine example of this complicated relation. Under physiological conditions the main source of neurotrophins within the CNS is the neuronal population, and so is their main target. However, neurotrophins are also expressed within the cells of the immunological system, namely peripheral blood mononuclear cells (PBMCs), as presented in Figure 2 [24–26]. Immune cells also express neurotrophin receptors: p75NTR and trkA are expressed on T cells and macrophages, trkB—on B-cells and macrophages, and trkC on macrophages only [24]. The ability of immune cells to secrete neurotrophins at the injury site within the CNS could be used as a potential therapeutic regimen. Under pathological conditions, as in MS, the additional neurotrophic support from PBMCs could compensate the relative neurotrophin deficiency in the damaged brain [27,28]. In MS PBMC-derived neurotrophins could play a neuroprotective role directly, binding to trk receptors on neurons within the CNS and promoting their survival, or indirectly by interacting with the neurotrophin receptors on the immune cells, regulating their functions (proliferation, immunoglobulin production) and determining their survival or apoptosis [15,29,30].

4. Neuroinflammation, Neurodegeneration and Neuroprotective Autoimmunity in MS

Traditionally, MS is considered to be caused by the autoreactive T-cells that become activated in the periphery, cross the blood-brain barrier (BBB) into the CNS and become re-activated by antigen presenting cells (APCs), which leads to a cascade of destructive inflammatory reactions [31]. Consequently, most of the therapies approved or tested for MS are based on their anti-inflammatory properties, and these are only partially effective. Neurodegeneration accompanies the inflammatory

process in MS and it is still debated whether it is secondary to myelin sheath destruction, or if it progresses independently [32]. The most radical concept is that MS is in fact primarily neurodegenerative, whereas autoimmune reaction is only a response to the degenerative process [33,34]. This concept was supported by the elegant neuropathological study by Barnett and Prineas in 2004, where they showed that the earliest structural change in the newly forming demyelinating lesions was oligodendrocyte apoptosis, that preceded lymphocyte infiltration and phagocyte activation [33]. Apoptotic oligodendrocytes within new MS plaques were also observed by Lucchinetti et al. [35]. The cause of oligodendrocyte destruction has so far not been established, however, its early appearance suggests that at least in some MS cases autoimmune reaction may be triggered as a secondary phenomenon. While the primary neurodegenerative origin of MS may be considered controversial, autoimmunity does involve a protective component. The examples include CD4+ Th2 cells-derived anti-inflammatory cytokines, macrophage-derived epidermal growth factor (EGF), which has myelinoand oligodendrocyte-trophic properties, and secretion of neurotrophins by peripheral blood mononuclear cells (PBMCs). Also, clinical disease activity does not correlate with the total inflammatory lesion load measured with the use of MRI. Moreover, endpoint disability in MS patients is associated with brain atrophy measures, and not with demyelinating lesions' volume [36,37].

Another interesting issue is the dual role that astrocytes may play in MS pathogenesis. Aside their structural and regulatory role within the CNS, they can be activated to exhibit immune cell functions. They are capable of acting as antigen presenting cells, providing environment for T cell activation, inhibiting myelin repair and enhancing immune reaction [38]. On the other hand, they can support oligodendrocyte and axonal regeneration, among many by inducing neurotrophin (BDNF, NT-4) synthesis [39].

Given the striking heterogeneity of MS and the limited efficacy of immunosuppressive/immunomodulatory agents, one should consider that it might be the defective neuroprotective and neuroregenerative properties that determine the disease course. So far, this aspect of MS has not been fully explored. However, more and more studies on this subject emerge.

5. Neurotrophins and MS Pathology

In 2002 Standelmann *et al.* showed that in MS-diseased brains BDNF was expressed not only in neurons, but also in T lymphocytes and macrophages, especially in the perivascular spaces, while this was not observed in healthy controls [40]. Immune cell BDNF expression was positively correlated with the inflammatory activity of demyelinating lesions. In the same study BDNF receptor, trkB, was found on neurons in close proximity of MS plaques and on reactive astrocytes within lesions. Interestingly, the highest concentrations of neurotrophic factors and their receptors in the CNS of MS patients are found within the active edge of the demyelinating plaques, where axons are at risk of bystander destruction adjacent to the neuroinflammatory core. These axons, which could be perceived as the ischaemic stroke "penumbra" equivalent, can still be salvaged with the adequate neuroprotective processes being activated [27,28]. If neuroprotective strategies fail, the area of damage becomes vaster, hence the increase of the CNS atrophy, which remains the main determinant of MS patients long-term disability. In this context it is highly probable that immune cells releasing neurotrophins into the periplaque area may in fact play a neuroprotective role. The fact that neurotrophin reactivity is higher

in MS plaques can be interpreted as the protective aspect of the inflammatory process. One could expect that the additional supply of neurotrophins, by immune cells or pharmacologically, will exert a neuroprotective effect, reflected by a better clinical outcome in patients.

The first observation that raised hopes for neurotrophic factors as potential therapeutic agents in MS was finding that leukaemia inhibitory factor (LIF), belonging to the neurotrophic factor family, reduced clinical severity and promoted oligodendrocyte survival in mice suffering from an animal equivalent of MS, namely experimental autoimmune encephalomyelitis [41]. LIF was successful no matter if administered systemically at immunization, before disease onset, or after the disease has already developed. This observation confirmed that neurotrophic factors act by facilitating regeneration, and not by suppressing the inflammatory reaction which is not present before the disease becomes clinically active.

There are a number of studies on neurotrophins in animal models, which show that administration of neurotrophins enhances survival of injured neurons in EAE and other neuronal injury models [42,43]. The evidence in human disease is less extensive. It is worth mentioning that one can assess the neurotrophic potential by several means and different approaches have been used by different researchers. Neurotrophin concentrations can be measured directly in PBMCs lysates with the ELISA method. One can also estimate neurotrophin mRNA expression in PBMCs, or measure the secretion of activated immune cells in cultures. In a number of studies neurotrophin levels were estimated in CSF samples. However, in our opinion this seems less useful, since CSF neurotrophin concentrations are determined mainly by the neuronal pool. Consequently, the CSF neurotrophins concentrations may in fact reflect atrophy progression, and not explain it. Importantly, there are no normative values for neurotrophins concentrations in the body fluids.

From a clinical standpoint, the role of neurotrophins in MS is undoubtedly complex. So far the most abundant literature on neurotrophins' function in MS is available for BDNF. However, the studies that were published often present obviously conflicting results. During remission BDNF levels were shown to be unchanged [44], decreased [45–47] or increased [48–50]. Data seems consistent on the fact that RRMS patients who experience relapse and are not treated with immunomodulation have higher BDNF levels [44,51]. On the contrary, these patients present lower BDNF levels at baseline, most likely because of the defective CD40-dependent stimulation, which is restored by IFN-beta [52].

So far it has been shown that PBMC-derived BDNF is related to axonoprotective potential in MS patients [45,53,54]. Our group observed that lower immune-cell NT-3 is associated with more advanced brain atrophy in RRMS patients [55]. We have also found that in RRMS patients cognitive deficit is related with immune-cell beta-NGF [56]. Similarily, Patanella *et al.* described correlations between lower PBMC-derived BDNF levels and worse performance in neuropsychological tests of MS patients [57]. A protective function of C rs2030324 BDNF allele was described in the context of visual information processing, thought to be related to thalamic volume [58].

6. Implications for Therapy

The findings that are mentioned above, although they should be interpreted cautiously, may have important implications for MS therapy. Neuroprotective properties towards neurons and

oligodendrocytes, and immunomodulatory potential of neurotrophins raise hopes for their therapeutic application.

Since neurotrophins have a relatively short (less than two minutes) plasma half-life [59,60] it is not possible to deliver them systemically. Neurotrophins cannot cross the human blood-brain barrier unless they are modified, for instance using chimeric peptide brain drug targeting technology [61]. Other options that are worth exploring would be trk receptors agonists, selective trk monoclonal antibody or small-molecule trk agonists and, finally, a cellular approach of the adoptive transfer of activated autoimmune cells, modified in vitro to secrete neurotrophins when reinjected [27]. In vivo gene therapy with viral vectors injected directly into neurons is yet another option, and such attempt already passed phase I trial for treatment of Alzheimer's disease (AD) [62]. Indeed, several attempts have been made to use neurotrophin-related pathways in therapy for other CNS disorders with a degenerative component, in which decreased neurotrophic activity was described, namely AD, Parkinson's disease (PD), amyotrophic lateral sclerosis (ALS), and Huntington's disease [63-67]. Interestingly, some of the immunomodulating agents currently used for MS, namely interferon beta, glatiramer acetate (GA) and-still under clinical trials-alemtuzumab, have been shown to increase serum and/or immune-cell levels of BDNF in MS patients, which was suggested as one of the possible modes of actions for these therapies [46,47,68]. For many years GA effects were thought to be mediated substantially by induction of a shift of GA-reactive T cells from Th1 to Th2. However, it was shown that GA-reactive T cells release BDNF locally in the CNS and thus exerting a neuroprotective effect [69]. Glatiramer acetate was also shown to augment NT-3 and NT-4 expression by T cells and resident CNS cells (neurons, astrocytes) in EAE models [70]. On the other hand, interferon beta was shown to promote NGF secretion early in the course of MS [71]. Laquinimod, which is a novel oral immunomodulatory drug developed for MS therapy-still under clinical trials, has so far demonstrated some beneficial effects in RRMS patients. Aside from its immunomodulatory mode of action, it has been shown to prevent EAE-induced BDNF loss and restore it in the chronic stage of the disease in mice [72]. Also, in MS patients, laquinimod administration resulted in the increase of serum BDNF levels [73].

The precise mechanisms in which immunomodulatory agents work in MS in not known, and their efficacy is largely established based on clinical trials. It may be so that the neuroprotective component of their mode of action is at least partially responsible for their clinical efficacy.

7. Concluding Remarks

In the healthy adult CNS neurons are the main source of neurotrophins and the role of immune cell-derived neurotrophic factors is most probably marginal, especially since it is only the activated immunological cells that can cross the blood-brain barrier and they do not get activated in the undamaged brain. On the contrary, in MS we observe dynamic interactions between the nervous and immunological systems.

It seems crucial to identify the mechanism in which PBMCs are stimulated to produce neurotrophins and release them into the plaque area. Such knowledge would be a breakthrough finding in context of therapy for MS and other CNS pathologies where neuroprotective-neuroregenerative strategies are needed. Shibata *et al.* (2003) showed that stimulation of human macrophages with

CD40L ligand increases their neurotrophic activity, resulting in enhancing protein synthesis, promoting neurite growth and facilitating synaptic activity of rat cortical neurons cultured in media conditioned with the stimulated human macrophages [74]. CD40L is expressed on activated T cells, on cerebral endothelium [75] and on astrocytes [76]. Therefore, after crossing the BBB macrophages could be activated by all these cells to produce neurotrophins. However, in many RRMS patients, regeneration processes become insufficient and the disease progresses to the secondary progressive stage. One possible explanation would be that the neuroprotective function of PBMCs is inhibited because of the proinflammatory shift of the immunological reaction. This phenomenon could account for the antigen non-specific immunological defect that leads to MS development under conditions in which subjects with no such defect would not develop the disease.

Identification of the exact role that neurotrophins play in MS pathology could serve not only for potential therapeutic approaches, but also in the diagnostic field. Flooded with a vast number of communications on detailed interactions in MS immunology, we are lacking information that could be useful in clinical practice. Currently there are no valid methods evaluating neuroprotection in patients. Standard MRI measures, with the exception of the T1-hypointense "black holes" volume, correlate poorly with axonal loss. CSF biomarkers are difficult to obtain, given the patients' reluctance to repetitive lumbar puncture procedures. At the moment there are no validated biomarkers that could be used for MS diagnosis, monitoring and prognosis of disease progression. Neurotrophins could be a plausible candidate for MS-related biomarkers, especially in light of evidence suggesting their association with brain atrophy markers and cognitive dysfunction in MS patients (see earlier). For instance, they might serve as surrogate markers of patients' neuroprotective potential that is correlated with brain atrophy.

Undoubtedly, there is more and more evidence for the role of neurotrophins as possible mediators of the neuroprotective function that inflammatory cells may exert in MS. More studies, especially observations in MS patients and not animal models, are needed in order to clarify this aspect and start considering neurotrophins as potential biomarkers and/or add-on therapies to traditional immunomodulation in multiple sclerosis patients.

References

- 1. Confavreux, C.; Aimard, G.; Devic, M. Course and prognosis of multiple sclerosis assessed by the computerized data processing of 349 patients. *Brain* **1980**, *103*, 281–300.
- Confavreux, C.; Vukusic, S.; Moreau, T.; Adeleine, P. Relapses and progression of disability in multiple sclerosis. *N. Engl. J. Med.* 2000, 343, 1430–1438.
- Amato, M.P.; Ponziani, G. A prospective study on the prognosis of multiple sclerosis. *Neurol. Sci.* 2000, 21(4 Suppl 2), S831–S838.
- Noseworthy, J.H.; Lucchinetti, C.; Rodriguez, M.; Weinshenker, B.G. Multiple sclerosis. N. Engl. J. Med. 2000, 343, 938–952.
- 5. Connor, B.; Dragunow, M. The role of neuronal growth factors in neurodegenerative disorders of the human brain. *Brain Res. Rev.* **1998**, *27*, 1–39.
- 6. Altman, L. Programmed cell death: The paths to suicide. *Trends Neurosci.* 1992, 15, 278–280.

- 7. Johnson, E.M.; Chang, J.Y.; Koike, T.; Martin, D.P. Why do neurons die when deprived of trophic factors? *Neurobiol. Aging* **1989**, *10*, 549–552.
- 8. Thoenen, H. Neurotrophins and activity-dependent plasticity. *Prog. Brain Res.* 2000, *128*, 183–191.
- 9. Raff, M.C.; Whitmore, A.V.; Finn, J.T. Axonal self-destruction and neurodegeneration. *Science* **2002**, *296*, 868–871.
- 10. Levi-Montalcini, R. Effects of mouse tumor transplantation on the nervous system. *Ann. N. Y. Acad. Sci.* **1952**, *55*, 330–3440.
- 11. Huang, E.J.; Reichardt, L.F. Neurotrophins: Roles in neuronal development and function. *Annu. Rev. Neurosci.* 2001, 24, 677–736.
- 12. Lee, R.; Kermani, P.; Teng, K.K.; Hempstead, B.L. Regulation of cell survival by secreted proneurotrophins. *Science* **2001**, *294*, 1945–1948.
- 13. Kaplan, D.R.; Miller, F.D. Neurotrophin signal transduction in the nervous system. *Curr. Opin. Neurobiol.* **2000**, *10*, 381–391.
- 14. Chao, M.V. The p75 neurotrophin receptor. J. Neurobiol. 1994, 25, 1373–1385.
- 15. Chao, M.V. Neurotrophins and their receptors: A convergence point for many signalling pathways. *Nat. Rev. Neurosci.* **2003**, *4*, 299–309.
- 16. Harrington, A.W.; Kim, J.Y.; Yoon, S.O. Activation of Rac GTPase by p75 is necessary for c-jun *N*-terminal kinase-mediated apoptosis. *J. Neurosci.* **2002**, *22*, 156–166.
- 17. Khursigara, G.; Bertin, J.; Yano, H.; Moffett, H.; DiStefano, P.S.; Chao, M.V. A prosurvival function for the p75 receptor death domain mediated via the caspase recruitment domain receptor-interacting protein 2. *J. Neurosci.* **2001**, *21*, 5854–5863.
- 18. Faustman, D.; Davis, M. TNF receptor 2 pathway: Drug target for autoimmune diseases. *Nat. Rev. Drug Discov.* **2010**, *9*, 482–493.
- Beattie, M.S.; Harrington, A.W.; Lee, R.; Kim, J.Y.; Boyce, S.L.; Longo, F.M.; Bresnahan, J.C.; Hempstead, B.L.; Yoon, S.O. ProNGF induces p75-mediated death of oligodendrocytes following spinal cord injury. *Neuron* 2002, *36*, 375–386.
- Lin, S.Y.; Wu, K.; Levine, E.S.; Mount, H.T.; Suen, P.C.; Black, I.B. BDNF acutely increases tyrosine phosphorylation of the NMDA receptor subunit 2B in cortical and hippocampal postsynaptic densities. *Brain Res. Mol. Brain Res.* 1998, 55, 20–27.
- 21. Tucker, K.; Fadool, D.A. Neurotrophin modulation of voltage-gated potassium channels in rat through TrkB receptors is time and sensory experience dependent. *J. Physiol.* **2002**, *542*, 413–429.
- 22. Wong, S.T.; Henley, J.R.; Kanning, K.C.; Huang, K.H.; Bothwell, M.; Poo, M.M. A p75 (NTR) and Nogo receptor complex mediates repulsive signaling by myelin-associated glycoprotein. *Nat. Neurosci* .2002, *5*, 1302–1308.
- 23. Wang, K.C.; Kim, J.A.; Sivasankaran, R.; Segal, R.; He, Z. P75 interacts with the Nogo receptor as a co-receptor for Nogo, MAG and OMgp. *Nature* **2002**, *420*, 74–78.
- 24. Linker, R.A.; Gold, R.; Lühder, F. Function of neurotrophic factors beyond the nervous system: Inflammation and autoimmune demyelination. *Crit. Rev. Immunol.* **2009**, *29*, 43–68.
- 25. Reichardt, L.F. Neurotrophin-regulated signalling pathways. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* **2006**, *361*, 1545–1564.

- 26. Vega, J.A.; Garc á-Su árez, O.; Hannestad, J.; P érez-P érez, M.; German à, A. Neurotrophins and the immune system. *J. Anat.* **2003**, *203*, 1–19.
- 27. Hohlfeld, R. Neurotrophic cross-talk between the nervous and immune systems: Relevance for repair strategies in multiple sclerosis? *J. Neurol. Sci.* **2008**, *265*, 93–96.
- Kerschensteiner, M.; Stadelmann, C.; Dechant, G.; Wekerle, H.; Hohlfeld, R. Neurotrophic cross-talk between the nervous and immune systems: Implications for neurological diseases. *Ann. Neurol.* 2003, 53, 292–304.
- Tabakman, R.; Lecht, S.; Sephanova, S.; Arien-Zakay, H.; Lazarovici, P. Interactions between the cells of the immune and nervous system: Neurotrophins as neuroprotection mediators in CNS injury. *Prog. Brain Res.* 2004, *146*, 387–401.
- Linker, R.; Lee, D.-H.; Siglienti, I.; Gold, R. Is there a role for neurotrophins in the pathology of multiple sclerosis? *J. Neurol.* 2007, 254(Suppl. 1), I/33–I/40.
- Wekerle, H.; Lassman, H. The immunology of inflammatory demyelinating disease. In McAlpine's Multiple Sclerosis, 4th ed.; Compston, A., Confavreux, C., Lassmann, H., McDonald, I., Miller, D., Noseworthy, J., Smith, K., Werkerle, H., Eds.; Churchill Livingstone Elsevier, Linn, MO, USA, 2006; pp. 491–555.
- 32. Lassman, H. What drives disease in multiple sclerosis: Inflammation or neurodegeneration? *Clin. Exp. Neuroimmunol.* **2010**, *1*, 2–11.
- 33. Barnett, M.H.; Prineas, J.W. Relapsing and remitting multiple sclerosis: Pathology of the newly forming lesion. *Ann. Neurol.* **2004**, *55*, 458–468.
- Stys, P.K. Multiple sclerosis: Autoimmune disease or autoimmune reaction? *Can. J. Neurol. Sci.* 2010, *37*(*Suppl. 2*), S16–S23.
- Lucchinetti, C.; Brück, W.; Parisi, J.; Scheithauer, B.; Rodriguez, M.; Lassmann, H. Heterogeneity of multiple sclerosis lesions: Implications for the pathogenesis of demyelination. *Ann. Neurol.* 2000, 47, 707–717.
- Bjartmar, C.; Kidd, G.; Mörk, S.; Rudick, R.; Trapp, B.D. Neurological disability correlates with spinal cord axonal loss and reduced *N*-acetyl aspartate in chronic multiple sclerosis patients. *Ann. Neurol.* 2000, 48, 893–901.
- Fisher, E.; Rudick, R.A.; Simon, J.H.; Cutter, G.; Baier, M.; Lee, J.C.; Miller, D.; Weinstock-Guttman, B.; Mass, M.K.; Dougherty, D.S.; *et al.* Eight-year follow-up study of brain atrophy in patients with MS. *Neurology* 2002, *59*, 1412–1420.
- 38. Nair. A.; Frederick, T.J.; Miller, S.D. Astrocytes in multiple sclerosis: A product of their environment. *Cell Mol. Life Sci.* **2008**, *65*, 2702–2720.
- Bsibsi, M.; Persoon-Deen, C.; Verwer, R.W.; Meeuwsen, S.; Ravid, R.; van Noort, J.M. Toll-like receptor 3 on adult human astrocytes triggers production of neuroprotective mediators. *Glia* 2006, 53, 688–695.
- Standelmann, C.; Kerschensteiner, M.; Misgeld, T.; Brück, W.; Hohlfeld, R.; Lassmann, H. BDNF and gp145trkB in multiple sclerosis brain lesions: Neuroprotective interactions between immune and neuronal cells? *Brain* 2002, *125(Pt 1)*, 75–85.

- Butzkueven, H.; Zhang, J.G.; Soilu-Hanninen, M.; Hochrein, H.; Chionh, F.; Shipham, K.A.; Emery, B.; Turnley, A.M.; Petratos, S.; Ernst, M.; *et al.* LIF receptor signaling limits immune-mediated demyelination by enhancing oligodendrocyte survival. *Nat. Med.* 2002, *8*, 613–619.
- 42. Yan, Q.; Elliot, J.; Snider, W.D. Brain-derived nerotrophic factor rescues spinal motor neurons from axotomy-induced cell death. *Nature* **1992**, *360*, 753–755.
- 43. Mo, L.; Yang, Z.; Zhang, A.; Li, X. The repair of the injured adult rat hippocampus with NT-3-chitosan carriers. *Biomaterials* **2010**, *31*, 2184–2192.
- 44. Sarchielli, P.; Greco, L.; Stipa, A.; Floridi, A.; Gallai, V. Brain-derived neurotrophic factor in patients with multiple sclerosis. *J. Neuroimmunol.* **2002**, *132*, 180–188.
- 45. Azoulay, D.; Urshansky, A.; Karni, A. Low and dysregulated BDNF secretion from immune cells of MS patients is related to reduced neuroprotection. *J. Neuroimmunol.* **2008**, *195*, 186–193.
- Lalive, P.H.; Kantengwa, S.; Benkhoucha, M.; Juillard, C.; Chofflon, M. Interferon-β induces brain-derived neurotrophic factor in peripheral blood mononuclear cells of multiple sclerosis patients. J. Neuroimmunol. 2008, 197, 147–151.
- Azoulay, D.; Vachapova, V.; Shihman, B.; Miler, A.; Karni, A. Lower brain-derived neurotrophic factor in serum of relapsing-remitting MS: Reversal by glatiramer acetate. *J. Neuroimmunol.* 2005, 167, 215–218.
- Gielen, A.; Khademi, M.; Muhallab, S.; Olsson, T.; Piehl, F. Increased brain-derived neurotrophic factor expression in white blood cells of relapsing-remitting multiple sclerosis patients. *Scand. J. Immunol.* 2003, 57,493–497.
- 49. Petereit, H.F.; Lindemann, H.; Schoppe, S. Effect of immunomodulatory drugs on *in vitro* production of brain-derived neurotrophic factor. *Mult. Scler.* **2003**, *9*, 16–20.
- Liguori, M.; Fera, F.; Patitucci, A.; Manna, I.; Condino, F.; Valentino, P.; Telarico, P.; Cerasa, A.; Gioia, M.C.; di Palma, G.; *et al.* A longitudinal observation of brain derived neurotrophic factor mRNA levels in patients with relapsing-remitting multiple sclerosis. *Brain Res.* 2009, *1256*, 123–128.
- Sarchielli, P.; Zaffaroni, M.; Floridi, A.; Greco, L.; Candeliere, A.; Mattioni, A.; Tenaglia, S.; di Filippo, M.; Calabresi, P. Production of brain derived neurotrophic factor by mononuclear cells of patients with multiple sclerosis treated with glatiramer acetate, interferon-beta 1a, and high doses of immunoglobulins. *Mult. Scler.* 2007, *13*, 313–331.
- 52. Azoulay, D.; Mausner-Fainberg, K.; Urshansky, N.; Fahoum, F.; Karni, A. Interferon-beta therapy upregulates BDNF secretion from PBMCs of MS patients through a CD40-dependent mechanism. *J. Neuroimmunol.* **2009**, *211*, 114–119.
- Weinstock-Guttman, B.; Zivadinov, R.; Tamano-Blanco, M.; Abdelrahman, N.; Badgett, D.; Durfee, J.; Hussein, S.; Feichter, J.; Patrick, K.; Benedict, R.; *et al.* Immune cell BDNF secretion is associated with white matter voulume in multiple sclerosis. *J. Neuroimmunol.* 2007, 188, 167–174.
- Linker, R.A.; Lee, D.H.; Demir, S.; Wiese, S.; Kuse, N.; Siglienti, I.; Gerhardt, E.; Neumann, H.; Sendtner, M.; Luehder, F.; *et al.* Functional role of brain-derived neurotrophic factor in neuroprotective autoimmunity: Therapeutic implications in a model of multiple sclerosis. *Brain* 2010, *133*, 2248–2263.

- Kalinowska-Łyszczarz, A.; Pawlak, M.A.; Michalak, S.; Paprzycki, W.; Losy, J. Immune cell NT-3 expression is associated with brain atrophy in multiple sclerosis patients. *J. Neuroimmunol.* 2011, 240–241, 109–113.
- 56. Kalinowska-Łyszczarz, A.; Pawlak, M.A.; Michalak, S.; Losy, J. Cognitive deficit is related to immune-cell beta-NGF in multiple sclerosis patients. *J. Neurol. Sci.* **2012**, *321*, 43–48.
- Patanella, A.K.; Zinno, M.; Quaranta, D.; Nociti, V.; Frisullo, G.; Gainotti, G.; Tonali, P.A.; Batocchi, A.P.; Marra, C. Correlations between peripheral blood mononuclear cell production of BDNF, TNF-alpha, IL-6, IL-10 and cognitive performances in multiple sclerosis patients. *J. Neurosci. Res.* 2010, 88, 1106–1112.
- Weinstock-Guttman, B.; Benedict, R.H.; Tamaño-Blanco, M.; Ramasamy, D.P.; Stosic, M.; Polito, J.; Zivadinov, R.; Ramanathan, M. The rs2030324 SNP of brain-derived neurotrophic factor (BDNF) is associated with visual cognitive processing in multiple sclerosis. *Pathophysiology* 2011, 18, 43–52.
- Pradat, P.F.; Kennel, P.; Naimi-Sadaoui, S.; Finiels, F.; Orsini, C.; Revah, F.; Delaere, P.; Mallet, J. Continuous delivery of neurotrophin 3 by gene therapy has a neuroprotective effect in experimental models of diabetic and acrylamide neuropathies. *Hum. Gene. Ther.* 2001, *12*, 2237–2249.
- Pradat, P.F.; Kennel, P.; Naimi-Sadaoui, S.; Finiels, F.; Scherman, D.; Orsinim, C.; Delaere, P.; Mallet, J.; Revah, F. Viral and non-viral gene therapy partially prevents experimental cisplatin-induced neuropathy. *Gene. Ther.* 2002, *19*, 1333–1337.
- 61. Pardridge, W.M. Neurotrophins, neuroprotection and the blood-brain barrier. *Curr. Opin. Investig. Drugs.* **2002**, *3*, 1753–1757.
- 62. Blesch, A. Neurotrophin gene therapy for Alzheimer's disease. Future Neurol. 2006, 1, 179–187
- Dawbarn, D.; Allen, S.J. Neurotrophins and neurodegeneration. *Neuropathol. Appl. Neurobiol.* 2003, 29, 211–230.
- 64. Tuszynski, M.H.; Thal, L.; Pay, M.; Salmon, D.P.; U, H.S.; Bakay, R.; Patel, P.; Blesch, A.; Vahlsing, H.L.; Ho, G.; *et al.* A phase 1 clinical trial of nerve growth factor gene therapy for Alzheimer disease. *Nat. Med.* **2005**, *11*, 551–555.
- 65. Drinkut, A.; Tereshchenko, Y.; Schulz, J.B.; Bähr, M.; Kügler, S. Efficient gene therapy for Parkinson's disease using astrocytes as hosts for localized neurotrophic factor delivery. *Mol. Ther.* **2012**, *20*, 534–543.
- 66. Calvo, A.C.; Moreno-Igoa, M.; Mancuso, R.; Manzano, R.; Oliván, S.; Muñoz, M.J.; Penas, C.; Zaragoza, P.; Navarro, X.; Osta, R. Lack of a synergistic effect of a non-viral ALS gene therapy based on BDNF and a TTC fusion molecule. *Orphanet. J. Rare Dis.* 2011, *6*, 10.
- 67. Giralt, A.; Carretón, O.; Lao-Peregrin, C.; Mart ń, E.D.; Alberch, J. Conditional BDNF release under pathological conditions improves Huntington's disease pathology by delaying neuronal dysfunction. *Mol. Neurodegener.* **2011**, *6*, 71.
- 68. Jones, J.L.; Anderson, J.M.; Phuah, C.L.; Fox, E.J.; Selmaj, K.; Margolin, D.; Lake, S.L.; Palmer, J.; Thompson, S.J.; Wilkins, A.; *et al.* Improvement in disability after alemtuzumab treatment of multiple sclerosis is associated with neuroprotective autoimmunity. *Brain* **2010**, *133*, 2232–2247.

- 69. Ziemssen, T.; Kuempfel, T.; Klinkert, W.E.F.; Neuhaus, O.; Hohlfeld, R. Glatiramer acetate-specific T-helper 1- and 2-type cell lines produce BDNF: Implications for multiple sclerosis therapy. *Brain* **2002**, *125*, 2381–2391.
- Aharoni, R.; Eilam, R.; Domev, H.; Labunskay, G.; Sela, M.; Arnon, R. The immunomodulator glatiramer acetate augments the expression of neurotrophic factors in brains of experimental autoimmune encephalomyelitis mice. *Proc. Natl. Acad. Sci. USA* 2005, *102*, 19045–19050.
- Biernacki, K.; Antel, J.P.; Blain, M.; Narayanan, S.; Arnold, D.L.; Prat, A. Interferon Beta Promotes Nerve Growth Factor Secretion Early in the Course of Multiple Sclerosis. *Arch. Neurol.* 2005, *62*, 563–568.
- Aharoni, R.; Saada, R.; Eilam, R.; Hayardeny, L.; Sela, M.; Arnon, R. Oral treatment with laquinimod augments regulatory T-cells and brain-derived neurotrophic factor expression and reduces injury in the CNS of mice with experimental autoimmune encephalomyelitis. *J. Neuroimmunol.* 2012, 251, 14–24.
- 73. Thöne, J.; Ellrichmann, G.; Seubert, S.; Peruga, I.; Lee, D.H.; Conrad, R; Hayardeny, L.; Comi, G.; Wiese, S.; Linker, R.A.; *et al.* Modulation of autoimmune demyelination by laquinimod via induction of brain-derived neurotrophic factor. *Am. J. Pathol.* 2012, *180*, 267–274.
- Shibata, A.; Zelivyanskaya, M.; Limoges, J.; Carlson, K.A.; Gorantla, S.; Branecki, C.; Bishu, S.; Xiong, H.; Gendelman H.E. Peripheral nerve induces macrophage neurotrophic activities: Regulation of neuronal process outgrowth, intracellular signaling and synaptic function. *J. Neuroimmunol.* 2003, 142, 112–129.
- 75. Mach, F.; Schönbeck, U.; Sukhova, G.K.; Bourcier, T.; Bonnefoy, J.Y.; Pober, J.S.; Libby, P. Functional CD40 ligand is expressed on human vascular endothelial cells, smooth muscle cells, and macrophages: Implications for CD40-CD40 ligand signaling in atherosclerosis. *Proc. Natl. Acad. Sci. USA* 1997, 94, 1931–1936.
- 76. Calingasan, N.Y.; Erdely, H.A.; Altar, C.A. Identification of CD40 ligand in Alzheimer's disease and in animal models of Alzheimer's disease and brain injury. *Neurobiol. Aging* **2002**, *23*, 31–39.

© 2012 by the authors; licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution license (http://creativecommons.org/licenses/by/3.0/).