



Review

# Plausible Roles for RAGE in Conditions Exacerbated by Direct and Indirect (Secondhand) Smoke Exposure

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**Abstract:** Approximately 1 billion people smoke worldwide, and the burden placed on society by primary and secondhand smokers is expected to increase. Smoking is the leading risk factor for myriad health complications stemming from diverse pathogenic programs. First- and second-hand cigarette smoke contains thousands of constituents, including several carcinogens and cytotoxic chemicals that orchestrate chronic inflammatory responses and destructive remodeling events. In the current review, we outline details related to compromised pulmonary and systemic conditions related to smoke exposure. Specifically, data are discussed relative to impaired lung physiology, cancer mechanisms, maternal-fetal complications, cardiometabolic, and joint disorders in the context of smoke exposure exacerbations. As a general unifying mechanism, the receptor for advanced glycation end-products (RAGE) and its signaling axis is increasingly considered central to smoke-related pathogenesis. RAGE is a multi-ligand cell surface receptor whose expression increases following cigarette smoke exposure. RAGE signaling participates in the underpinning of inflammatory mechanisms mediated by requisite cytokines, chemokines, and remodeling enzymes. Understanding the biological contributions of RAGE during cigarette smoke-induced inflammation may provide critically important insight into the pathology of lung disease and systemic complications that combine during the demise of those exposed.

**Keywords:** receptor for advanced glycation end-products (RAGE); secondhand smoke; disease; exposure

## 1. Introduction

### 1.1. Global Burden

Currently, it is estimated that there are nearly 1 billion smokers worldwide (WHO Fact Sheet No 339). Of this enormous number, approximately 80% live in either low- or middle-income countries where the effects or burdens of tobacco-related illness and death are the most substantial. Furthermore, while the current worldwide population of smokers is estimated at 1 billion, current projections predict that this number will rise to 1.6 billion in the next twenty-five years [1]. With such an inordinate number of smokers world-wide, roughly 6 million people are expected to die each year because of tobacco exposure (WHO Fact Sheet No 339). Of this number, over 600,000 people will die prematurely

as a result of exposure to secondhand smoke (SHS). Unfortunately, while these numbers themselves are galling, rampant tobacco use throughout the world has also had societal ramifications as exposure is believed to contribute to over \$500 billion in damages annually [2].

Because it appears that smoking prevalence will continue to rise despite its inherent dangers and costs, research is expanding in order to better understand the consequences of these trends. The intent of this review is to highlight significant health outcomes that result from SHS exposure and suggest a generally unifying mechanistic theme underlying the biological consequences of exposure. Mounting evidence suggests that the signaling effects of receptors for advanced glycation end-products (RAGE) during exposure to primary and SHS may contribute to inflammatory disease establishment and progression. RAGE is expressed in a variety of cell types including endothelial and vascular smooth muscle cells, fibroblasts, macrophages/monocytes, osteoprogenitor cells, endothelium, and epithelium [3] (personal communication). Of note, RAGE is most abundantly expressed in the lung, the tissue in which it was initially discovered. Although RAGE is predominantly expressed in the lung, it is detectable in a variety of tissues including the heart, brain, placenta, liver, kidney, pancreas, small intestine, and colon [4,5]. This review will highlight the current understanding related to a subset of smoke-related pathologies and conclude with evidence that supports a role for RAGE in disease manifestation. The biochemical assessments performed to date have linked many tobacco-related substances with negative health consequences [6]; however, much remains to be discovered.

### 1.2. Tobacco Smoke

Tobacco smoke contains over 4000 chemical substances [6], and a large portion of these entities have been correlated with damaging health outcomes. The combustion of tobacco smoke produces numerous compounds observed in both gaseous and particulate fractions. Many of these compounds are toxic components that have been demonstrated to induce inflammation, cause irritation, asphyxiation, and even carcinogenesis. Recent studies have suggested that at least 45 of these substances are known carcinogens [7]. Some of the key toxins produced by tobacco smoke include benzene (leukemogen) [8], formaldehyde (an irritant and carcinogen) [9], benzo[*a*]pyrene (carcinogen) [10], carbon monoxide and cyanide (asphyxiants) [11], acrolein (an irritant) [12], and polonium (a radioactive carcinogen) [13,14]. Additionally, combustion of tobacco products creates a non-enzymatic reaction of reducing sugars and amino groups to create compounds known as advanced glycation end-products (AGEs) [15]. Such tobacco-derived AGEs are formed by Maillard chemical pathways and are the key ligand that perpetuates pro-inflammatory RAGE signaling [16]. AGEs that bind RAGE have been implicated in a large and diverse group of diseases including respiratory inflammatory diseases [17], cardiovascular disease [18], cancer [19], diabetes [20], neurodegenerative disorders [21], placental dysfunction [22,23], osteoarthritis [24], and general inflammation [25]. In mechanistic terms, AGE-RAGE interaction initiates a cascade of events that results in the induction of chronic inflammation and impaired cell survival [26,27].

While significant damage is induced by active smoking, research has demonstrated that individuals exposed to passive smoking (or secondhand smoke, SHS) are at risk for developing significant health problems [6,28]. Indeed, literature suggests that SHS may even expose individuals to higher levels of certain deleterious compounds than those observed in mainstream smoke. SHS for example is shown to have higher levels of PAHs [29,30], tobacco-specific nitrosamines (TSNA) [31–33], aromatic amines [34], aza-arenes [29,35], carbon monoxide [36–38], nicotine [39,40], ammonia [41], pyridine [42,43], and gas phase components of acrolein, benzene, toluene, and isoprene 1,3-butadiene [44]. Recently, thirdhand smoke has also been implicated as a potent source of exposure to the toxins found in cigarettes [45]. Thirdhand smoke is obtained when tobacco smoke constituents become deposited on surfaces and such deposits may undergo oxidation and other diverse chemical processes that result in the synthesis of carcinogenic species including TSNA [46]. In fact, it has been speculated that the dangers associated with thirdhand smoke may be even more profound than active smoking [47] due to the process in which thirdhand smoke is generated. As these deposited substances

are a product of time and isolation, thirdhand smoke poses real dangers for both active smokers and nonsmokers alike.

## 2. Health Outcomes and Comorbidities

### 2.1. Chronic Obstructive Pulmonary Disease

Chronic obstructive pulmonary disease (COPD) is one of the leading causes of mortality and morbidity and currently estimated to affect roughly 5% of the world's population or about 329 million individuals (WHO, The top 10 Causes of Death Fact Sheet 2012). The data overwhelmingly implicate primary or active smoking as the greatest risk factor for developing COPD [48]; however, exposure to environmental tobacco smoke is also highly associated with increased risk for COPD in individuals who have never smoked [49]. Globally, COPD is projected to be the third leading cause of death by 2020. Economists have estimated that the economic burden (including both direct and indirect costs) resulting from COPD was \$2.1 trillion in 2010, but believe that this will rise to \$4.3 trillion by 2030 [50]. Direct costs alone have been estimated to be in the \$49.9 billion range, suggesting a greater need for preventative measures, as well as improvements in earlier diagnosis and more cost effective treatment.

Without question, pharmacological and nonpharmacological interventions have contributed to improved outcomes as they relate to the management of COPD [51,52]. However, as the worldwide prevalence of COPD is predicted to increase, so will the urgency of improved comprehensive therapy. Inflammation intensifies as COPD progresses [53] and does not “burn out” as do many other chronic inflammatory diseases [54]. Therefore, there is a pressing need for the development of new molecular targets and associated therapies, particularly as no existing treatment has been shown to reduce disease progression. New therapies for COPD may arise from improvements in existing drugs (for example, longer acting  $\beta_2$ -agonists and anticholinergics) or from the development of novel therapies when underlying disease processes are better understood. Despite recent advances in the understanding of COPD molecular pathogenesis [55], there is clearly a need for more research into its basic mechanisms. Even so, there are still several reasons why drug development in COPD has been difficult. Animal models of COPD for early drug testing are not very satisfactory [56,57]. Animal models have contributed important details related to understanding immune mechanisms; however, drugs aimed at ameliorating symptoms observed in animal models often fail in Phase II trials. One plausible explanation may be that mucus is not produced in the bronchial tree of mice, so mucous exacerbations in COPD cannot be modeled well [58]. Furthermore, human COPD is coincident with variable pathologies at different stages of COPD severity, whereas the three main current animal models merely imitate a subset of COPD characteristics following exposure to noxious stimuli, tracheal instillation of elastases, or genetic modifications.

The characterization of innovative drugs with promise is periodically met with uncertainty because of the culminating requirement of multi-year, long-term trials. Confounding trial design is the notion that COPD patients are often disqualified from participating due to notable comorbidities including diabetes and heart disease. Finally, new treatments are also slow to develop because there remains disagreement in the field as to optimal biomarkers in a patient's sputum or blood necessary to evaluate acute amelioration and long-term therapeutic capacity. Despite these limitations, a lucid understanding of disease progression during primary and SHS exposure is essential in refining patient care.

In terms of the pathophysiology, COPD is typically characterized by airflow obstruction that is minimally reversible. This airflow obstruction is due to chronic inflammation and permanent pulmonary airspace enlargement as well as the loss of elastic recoil caused by apoptosis that destroys alveolar walls observed in emphysema. Persistent inflammation in COPD patients is characteristic not only in the airways, but in the respiratory parenchyma and pulmonary vasculature as well, and results in disruption of normal lung function specifically through remodeling of the distal pulmonary airspaces. Because chronic inflammation is a major defining characteristic of the disease, extensive

research surrounding pro-inflammatory molecular mechanisms have been conducted. The key aim of such research focuses on the attenuation or removal of chronic inflammation that overcomes natural protective mechanisms and the resulting tissue damage seen with COPD. Contributors to this inflammation-related process include imbalances between proteases/antiproteases, oxidative stress, elevated apoptotic indexes, and enhanced neutrophil, macrophage, and T lymphocyte extravasation.

Recent reports have corroborated previous findings that neutrophils are increased in sputum of patients with COPD along with increased interleukin-6 (IL-6) signaling [59]. Substantial evidence has implicated primary or active smoking as a major contributor to the recruitment of these neutrophils; however, mounting evidence now suggests that secondhand smoke may have a similar effect on neutrophils [60] that is likely mediated through similar interleukin signaling [61,62]. Under normal physiological conditions, neutrophils employ proteases and small cationic peptides to attack invading bacteria, viruses, and harmful exogenous material such as particulates found in tobacco smoke. Yet, in chronic inflammatory conditions, neutrophils become major destructors of the alveolar elastic matrix. These neutrophils also release enzymes and other mediators that cleave collagen into fragments that may further activate inflammatory cells [63]. One potent signaling factor that has been demonstrated to drive neutrophilic infiltration is the chemoattractant interleukin-8 (IL-8) which is produced by exposed and damaged epithelium and endothelium [64,65]. Additionally, other chemoattractants that have been shown to induce neutrophil migration include chemokine CXC motif ligands 1, 2, 5, 8 (CXCL-1, 2, 5, 8) [66–68], leukotriene B4 (LTB4) [68], IFN- $\gamma$  [69], IL-1 $\beta$  [70,71], and TNF- $\alpha$  [72]. Current data increasingly suggest that these potent inflammatory chemoattractants are elevated with exposure to SHS [73,74]. While neutrophils are predominant mediators of chronic inflammation, they are not the only important pro-inflammatory mediator. Macrophages have also been shown to participate in the propagation of inflammation through the release of chemoattractants, and are elevated in the airways, parenchymal bronchoalveolar lavage fluid (BALF), and sputum [75–77] from affected patients. Studies involving the exposure of mice to SHS have demonstrated increased macrophages in response to SHS [78]. Furthermore, as these adaptive immunity cells play such a vital role in chemoattraction, it is unsurprising that research suggests macrophage recruitment closely corresponds with the severity of the disease [79]. Like neutrophils, macrophages migrate to injured lung tissue and enhance the release of TNF- $\alpha$ , IL-8, CXC chemokines, monocyte chemoattractant peptide-1 (MCP-1), LTB4, and other pro-inflammatory molecules [55]. Finally, it should be noted that research indicates that T-cells may act as important intermediaries in the development of emphysema [80]. In a comparison of normal patients and those with smoke-induced COPD, diseased patients demonstrated elevated levels of CD3 and CD8 [80], two cytotoxic t-cell subgroups that organize apoptotic pathways used to kill infected or damaged cells. CD8 in particular was shown to be highly correlated with increasing severity in emphysema patients [80]. A recent analysis of mice subjected to SHS resulted in increased levels of CD4 and CD8; conversely, inhibition of these cells prevented airspace enlargement, inhibited cytokine release, and reduced apoptotic signaling [81]. Mechanistically, it is likely that CD8 subtly interacts in conjunction with CD4, a T-helper cell whose activation releases cytokines and helps orchestrate the migration and activity of other inflammatory cells. These T-cell mediated processes seemingly disrupt autoimmune regulation, thus enhancing perpetual inflammation.

## 2.2. Cancer

It is estimated that cigarette smoking contributes to 30% of all cancer deaths in developed countries [82]. Tobacco smoke is believed to be responsible for 70% of lung cancers deaths [83] (approximately 1.3 million deaths each year [84]) and 42% of esophageal and oral cavity cancer deaths [85]. Furthermore, tobacco smoke is believed to contribute significantly to the development of cancers of the larynx, urinary bladder, and pancreas, and to a lesser extent to cancers of the kidney, stomach, cervix, and myeloid leukemia [86]. Current evidence largely implicates active smoking as a major risk factor in cancer development; however, mounting evidence now suggests that SHS may equally participate. SHS has been shown to increase the risk of developing lung [87],

oropharyngeal [88], colorectal [89], breast [88], cervical [90], bladder [90], and pancreatic cancer [91]. Moreover, studies investigating nitrosamines, some of the most potent carcinogens in tobacco smoke, have demonstrated that high levels are present in both mainstream and SHS [92]. As nitrosamines are readily absorbed through the alveoli and then rapidly distributed through the blood, it is unsurprising that they are found to play a major role in the induction of many cancers [93,94].

Compounds such as PAHs, aromatic amines, aza-arenes, carbon monoxide, TSNA, nicotine, ammonia, pyridine, and the gas compounds of acrolein, toluene, isopentene-1,3-butadiene, and benzene are common in SHS. Overwhelmingly, the data implicate these substances in a host of cancers, although the mechanisms by which this takes place are broad and diverse. For example, recent data have demonstrated that benzene [95], toluene [95], and nicotine [96] have the ability induce the upregulation of CYP2E1, an enzyme that activates many foreign chemical compounds to become ultimate toxicants [97]. Aside from the ability to produce toxicants, the induction of CYP2E1 has been suggested to be the first step in leading to chemically induced carcinogenesis [98]. Alternatively, PAHs and TSNA has been shown to increase epithelial to mesenchymal transition (EMT), which is closely associated with an invasive or metastatic phenotype. Increased EMT is characterized by a downregulation of genes encoding for epithelial junction (claudins, occludins, e-cadherin) as well as an activation of protein products that promote mesenchymal adhesion. As these epithelial junctions are crucial in regulating cell differentiation, proliferation, and polarity, it is unsurprising that the loss of these proteins is often associated with an invasive phenotype [99]. As these tissues transition from an epithelial to mesenchymal state, the epithelial barrier is disrupted and thus a key initial line of defense in the innate immune system is compromised.

In general, tobacco smoke seems to broadly influence carcinogenesis in four ways. First, the gas and particulate phase of tobacco smoke includes at least 20 substances that can induce lung tumors in rodents [100,101]. These compounds directly contribute to carcinogenesis. Second, tobacco smoke includes substances that are not directly carcinogenic alone, but enhance the activity of carcinogens when co-administered. These substances include tumor promoters, co-carcinogens, and toxicants such as catechol, methyl catechols, and PAHs [102]. One potent example of such a compound is acrolein, which is not strongly carcinogenic when in isolation. However, acrolein expressed by ciliated epithelium can be highly toxic due to the hindrance of clearing tobacco smoke compounds from the lung, resulting in profound exposure to other carcinogens. Furthermore, acrolein reacts directly with DNA and protein, thus triggering genomic silencing of gene targets [103] that may enhance the likelihood of carcinogenesis. Third, tobacco smoke substantially influences the chronic inflammatory microenvironment. Tobacco smoke causes the recruitment of inflammatory cells, cytokine and chemokines that can act as drivers for cancer development and progression [104]. It is well documented that the infiltration of tumor-associated macrophages in tumor lesions is common to a host of cancer types, and is associated with tumor angiogenesis, invasion, and metastasis [105–108]. Finally, matrix metalloproteinases (MMP)-1, MMP-8, MMP-9, and MMP-13 are collagenases implicated in the development of COPD in response to cigarette smoke [109]. Interestingly, these same MMPs with notable importance in emphysema also function in tumor invasion and metastasis [110]. In fact, recent data supports the notion that elevated collagenases is associated with an increase in the severity of cancer. Because MMPs degrade the ECM in a fashion that allows tumor cells to be released from binding factors in their environment, greater MMP abundance increases the motility of cancer cells [111].

### 2.3. Developmental Complications

As cigarettes are known to be one of the most common teratogens [112], a number of serious obstetric complications arise with cigarette smoke exposure during pregnancy [113]. Approximately 10% of pregnant women in the US smoke, thereby exposing nearly 400,000 fetuses yearly to tobacco specific toxins [114]. Exposure to smoke during pregnancy has been demonstrated to increase the likelihood of congenital limb deficiencies [115], congenital heart defects [116], orofacial clefting [112],



and many other developmental abnormalities. Active smoking has long been considered a teratogenic agent that increases the risk of premature birth, but recent data shows that 22%–30% of nonsmoking pregnant women exposed to SHS are also at risk [117]. Developmental defects in the fetus represent substantial pregnancy complications; however, perinatally, smoke exposure further enhances mortality via increased risk of sudden infant death syndrome (SIDS) and preterm birth [118]. Altogether, it is not surprising that many researchers have suggested that cigarette exposure may be the single most important avoidable cause of adverse pregnancy outcomes [119–121].

Nicotine, one of the primary addictive compounds in tobacco smoke, is a key substance that contributes significantly to many of these health problems as even minute levels induce detectable transcriptomic modifications in small-airway epithelium [122]. Nicotine readily crosses the placenta [123] and binds to nicotinic acetylcholine receptors (nAChRs) which regulate fetal brain development [124]. Interestingly, research has demonstrated that nicotine levels are higher in the amniotic fluid, fetal serum, and placenta than in the corresponding maternal serum [125]. Studies demonstrating the adverse effects of tobacco smoke on neurodevelopment have provided compelling evidence that nicotine increases cellular damage, reduces overall cell number, impairs synaptic activity, and influences processes such as cell replication to differentiation and apoptosis [126–128]. Furthermore, nicotine has been associated with adverse neurocognitive outcomes such as behavioral disorders [129], cognitive dysfunction [130], and attention deficit hyperactivity disorder [131,132].

While a significant portion of the literature implicates tobacco smoke in neurodevelopmental pathologies, such effects are not limited to the nervous system. Prenatal tobacco smoke exposure has been demonstrated to have striking effects on respiratory development in that it reduces respiratory compliance in infants and impairs lung function in school-aged children [114,133]. Possibly contributing to impaired lung function are data that suggest that maternal smoke exposure may alter Clara cell secretory protein (CCSP) expression in fetal lungs [134]. Indeed, evidence currently suggests that maternal smoke exposure (including SHS) during pregnancy leads to the deregulation of gene expression [135]. Confirmatory primate studies have shown that in utero nicotine exposure adversely affects overall lung development by decreasing lung size and volume, elastin, while increasing Type I and Type III collagen, alveolar volume, and airway wall areas [136–139]. While the immediate ramifications are apparent, researchers have shown that nicotine exposure not only predisposes the fetus to lung dysfunction, but also has the ability to influence asthma in second and third generation offspring, likely through epigenetic modulation of the fetal program [140–142]. Aside from respiratory disorders, nicotine has further been shown to affect endocrine function [143,144], increase the likelihood of the fetus to develop chronic kidney disease (CKD) through increased mitochondrial dysfunction [145], and decrease auditory response and auditory development [146–148]. Overwhelming, the data suggests a particularly insidious role for SHS and its ability to influence development.

One further factor that may have a causal role in many developmental deficiencies is the impact of premature delivery, a risk factor that tobacco smoke has been shown to significantly increase [149]. As tobacco smoke has been shown to increase preterm birth (PTB), it should be noted that it additionally exacerbates intrauterine growth restriction (IUGR) and preeclampsia (PE), two placental diseases closely associated with PTB [113,150,151]. IUGR is a complication that stems primarily from uteroplacental vascular insufficiency, which ultimately creates an environment of chronic oxygen and nutrient deficiency, resulting in restricted fetal growth [152]. PE is another disease that impacts placentation wherein maternal hypertension and proteinuria accounts for around 20% of induced PTB [153]. Because complications such as perinatal hypoxia and asphyxia, cerebral palsy, and persistent pulmonary hypertension of the newborn have been associated with both IUGR and SHS exposure [154,155], it is likely that SHS modulates IUGR and PE symptoms that may culminate in diverse developmental pathologies.

#### 2.4. Cardiometabolic Disorders

The intimate connection, both in etiology and outcome, of cardiovascular and metabolic processes has resulted in the term, “cardiometabolic diseases”. The relevance of this is highlighted in the numbers: heart disease is the leading cause of death [156] and insulin resistance is the most common disorder in the US, affecting half of all adults [157]. Because of these startling statistics, considerable effort has been devoted over recent decades to elucidate effective strategies to reverse the trends. Overwhelmingly, these efforts have focused on the role of lifestyle variables, particularly diet. However, while diet is clearly relevant [156,158], it is also clearly not the entire solution, as cardiometabolic diseases continue unabated. Indeed, such a paradigm has left relatively unexplored that what we inhale may matter as much as what we ingest.

Insulin resistance is the “metabolic” in cardiometabolic disorders. Due to the obvious challenges of determining causality of a cigarette smoke–insulin resistance interaction, most of the findings in humans are correlational in nature [159,160], though limited data exist to establish [161,162] that cigarette smoke exposure increases insulin resistance. Typified by a reduced ability of insulin to elicit action at cells throughout the body, as well as general hyperinsulinemia, insulin resistance is at the heart of most cardiometabolic disorders, such as hypertension [163,164], atherosclerosis [163], dyslipidemia [165], cardiomyopathy [166], and more [167,168].

Unsurprisingly, cigarette smoke exposure similarly increases the risk of myriad cardiovascular complications through diverse mechanisms, though insulin resistance is clearly a dominant factor [169]. For example, dyslipidemia (i.e., increased triglycerides, reduced high density lipoprotein cholesterol), which is a key predictor in cardiovascular mortality with cigarette smoking [170], is significantly worse in smokers with insulin resistance compared with more insulin-sensitive smokers [171,172].

A second instance of the role of insulin resistance in smoke-induced cardiometabolic disorders is abnormal endothelial physiology. Blood vessels from smoke-exposed humans are less dynamic, having a reduced dilatory capacity [173], and have increased leukocyte adherence [174], increasing the risk of clot formation. In regards to endothelium-dependent vasodilation, current evidence shockingly revealed that after only 15 to 30 min of breathing SHS, vasodilation of coronary arteries in non-smokers was impaired almost to the extent of habitual smokers [175]. Intriguingly, both of these pathological processes are associated with endothelial dysfunction and are exacerbated by insulin resistance [176,177].

Among the multiple mechanisms that mediate smoke-induced cardiometabolic disorders, the effects of smoke on lipid metabolism are noteworthy insofar as they may reveal a strategy to partially mitigate the cardiometabolic consequences of smoke exposure. In particular, cigarette smoke pathologically alters sphingolipid metabolism, resulting in the accrual of ceramides, the backbone of higher-order sphingolipids, in heart [162] and skeletal muscle [178]—two key insulin-responsive tissues. Ceramide accrual in these tissues resulted in substantial disruption of mitochondrial function, including alterations in morphology and electron transport. Moreover, smoke exposure altered insulin signaling in skeletal muscle. An increase in the action of serine palmitoyltransferase, the rate-limiting step in sphingolipid biosynthesis, was necessary for the ceramide accrual, as inhibition via myriocin, was protective against the deleterious effects of smoke exposure.

Data collected over the past few decades suggest that SHS increases the incidence of coronary heart disease approximately 25%–30% [179–181]. Furthermore, although active smokers receive up to 100 times the dose of smoke than individuals exposed to SHS, an active smoker’s relative risk of coronary heart disease is 1.78 followed closely by a passive smoker at 1.31 [182]. SHS contributes to cardiovascular disease by activating blood platelets [183] likely through the combined elevation of both fibrinogen [182,184] and thromboxane [185], thus leading to the development of atherosclerosis. SHS has also been demonstrated to decrease levels of NO, the primary substrate that is implicit in the hemodynamic changes in the vascular system [186]. Research has even demonstrated that, after only 20 min of SHS exposure, direct endothelial cell injury is observed. Mechanistically, SHS exposure has been shown to increase free radicals [187] while decreasing antioxidants [188], decrease mitochondrial

function [178], decrease protective HDL levels [189,190], and increase arterial stiffness [191]. Such staggering data adds new meaning to the current warnings from the Surgeon General that state “there is no safe level of exposure to tobacco smoke” [114].

### 2.5. Joint and Movement Disorders

Osteoarthritis (OA), characterized by joint pain, effusion, loss of mobility, and deformity that progresses to functional joint failure, is one of the most common chronic diseases. It is reported to be the most common disease associated with the temporomandibular joint (TMJ) [192]. There is not currently any treatment to slow or stop the progression of OA. It has become the most common cause of long-term disability, in large part because of its association with the knee and spine. The incidence of OA in the population is comparable to other major disorders such as end-stage kidney disease and heart failure. For instance, there are nearly half a million joint replacements performed annually in the United States alone [193]. Many studies, including mouse knee destabilization and TMJ misalignment models, have demonstrated a pattern of biomarker expression associated with the progression of OA [194–197]. The disease appears to be associated with an initial rise in Tgf- $\beta$ 1 expression, followed by the upregulation of HtrA1, Ddr2 and Mmp13 expression, resulting in OA as assessed by standardized joint scoring methods such as the Mankin and the Osteoarthritis Research Society International (OARSI) scoring systems [198–201]. Curiously, the expression of HtrA1 and the other factors associated with OA are attenuated in a receptor for advanced glycation end-products (RAGE) knockout (KO) mice following surgically induced OA models [195]. This suggests that inflammation may be the trigger for the initiation and onset of OA. It follows that OA is associated with cigarette smoke. It is noted that, early on, the interaction of smoking and OA was controversial [202–206]. However, it has been reported that the discrepancies between smoking and OA interaction are likely do to study design and metrics [203,207]. A correlation between smoking and OA and/or cartilage defects is now apparent [207,208]. It is interesting to note that one study showed that the harmful effects of smoking associated with OA were due to both cartilage loss as well as the development of cartilage defects in people with a family history of joint disease [207]. Suggesting that a pre-disposition may be exacerbated by smoking through some bone/cartilage development association. Finally, it is noteworthy that one group who reported no association between direct smoking and OA did report a correlation between the joint disease and indirect smoking [206]. It is unknown if constituents of tobacco smoke have direct deleterious effects on chondrocyte function or if direct and/or indirect cigarette smoke induces cartilage damage through more global means such as inflammation.

## 3. RAGE: A Plausible Unifying Mechanism

Although many interrelated mechanistic processes potentially contribute to the diversity of diseases stemming from exposure to primary smoking, SHS, and thirdhand smoke, RAGE signaling is a program that commonly emerges. An underlying mechanistic theme of the smoke-related disease states outlined in this review is chronic inflammation, in which RAGE is a key modulator. Essential to understanding the clear link between RAGE and disease progression is the key concept that RAGE expression is increased by exposure to tobacco smoke [5,209–212] and the induction of RAGE causes inflammatory disease symptoms similar or identical to the ones described herein [23,195,213–217].

RAGE is expressed in a variety of cell types including endothelial and vascular smooth muscle cells, fibroblasts, macrophages/monocytes, osteoprogenitor cells, endothelium, and epithelium [3] (personal communication). Of note, RAGE is most abundantly expressed in the lung, the tissue in which it was initially discovered. Although RAGE is predominantly expressed in the lung, it is detectable in a variety of tissues including the heart, brain, placenta, liver, kidney, pancreas, small intestine, and colon [4,5]. RAGE is a pattern recognition cell surface receptor that binds many endogenous and exogenous entities such as S100/calgranulins [218], amyloid- $\beta$ -peptide [219], HMGB-1 [220], and AGEs [221]. Following RAGE-ligand interaction, a cascade of signaling events elicits gene expression modulation via divergent signal transduction pathways [222–224]. Because RAGE expression can also



increase when ligands accumulate [225], RAGE-ligand interactions may not only induce the defects described in this review, but also contribute to the chronicity of inflammatory tobacco smoke exposure observed in these pathological states as well. RAGE activation exacerbates a host of pro-inflammatory responses via MAP kinases (ERK, JNK, and P38), NF- $\kappa$ B, reactive oxidative species (ROS), and other chemokine mediators including TNF- $\alpha$ , IL1- $\beta$ , and others [226]. While redundancies exist within the pathway, RAGE signaling generally culminates in the activation of NF- $\kappa$ B, a transcriptional regulator that not only promotes pro-inflammatory mediator elaboration, but also de novo RAGE expression. Thus, RAGE signaling via NF- $\kappa$ B represents a vicious positive feedback loop that orchestrates chronic inflammation. In contrast to short-lived cellular activation mediated by lipopolysaccharide (LPS), engagement of RAGE by its ligands results in prolonged inflammation [227] that, if left unchecked, causes severe tissue injury.

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## References

1. Mackay, J.E.M.; Shafey, O. *The Tobacco Atlas*, 2nd ed.; American Cancer Society: Atlanta, GA, USA, 2006.
2. Ekpui, V.U.; Brown, A.K. The economic impact of smoking and of reducing smoking prevalence: Review of evidence. *Tob. Use Insights* **2015**, *8*, 1–35. [[CrossRef](#)] [[PubMed](#)]
3. Brett, J.; Schmidt, A.M.; Yan, S.D.; Zou, Y.S.; Weidman, E.; Pinsky, D.; Nowygrod, R.; Neeper, M.; Przysiecki, C.; Shaw, A.; et al. Survey of the distribution of a newly characterized receptor for advanced glycation end products in tissues. *Am. J. Pathol.* **1993**, *143*, 1699–1712. [[PubMed](#)]
4. Buckley, S.T.; Ehrhardt, C. The receptor for advanced glycation end products (RAGE) and the lung. *J. Biomed. Biotechnol.* **2010**, *2010*, 917108. [[CrossRef](#)] [[PubMed](#)]
5. Nelson, M.B.; Swensen, A.C.; Winden, D.R.; Bodine, J.S.; Bikman, B.T.; Reynolds, P.R. Cardiomyocyte mitochondrial respiration is reduced by receptor for advanced glycation end-product signaling in a ceramide-dependent manner. *Am. J. Physiol. Heart Circ. Physiol.* **2015**, *309*, H63–H69. [[CrossRef](#)] [[PubMed](#)]
6. Moritsugu, K.P. The 2006 report of the surgeon general: The health consequences of involuntary exposure to tobacco smoke. *Am. J. Prev. Med.* **2007**, *32*, 542–543. [[CrossRef](#)] [[PubMed](#)]
7. Fowles, J.; Dybing, E. Application of toxicological risk assessment principles to the chemical constituents of cigarette smoke. *Tob. Control* **2003**, *12*, 424–430. [[CrossRef](#)] [[PubMed](#)]
8. Protano, C.; Andreoli, R.; Manini, P.; Guidotti, M.; Vitali, M. A tobacco-related carcinogen: Assessing the impact of smoking behaviours of cohabitants on benzene exposure in children. *Tob. Control* **2012**, *21*, 325–329. [[CrossRef](#)] [[PubMed](#)]
9. Szumska, M.; Damasiewicz-Bodzek, A.; Tyrpien-Golder, K. Environmental tobacco smoke—Assessment of formaldehyde concentration in urine samples of exposed medicine students. *Przegl. Lek.* **2015**, *72*, 140–143. [[PubMed](#)]
10. Abedin, Z.; Louis-Juste, M.; Stangl, M.; Field, J. The role of base excision repair genes OGG1, APN1 and APN2 in benzo[a]pyrene-7,8-dione induced p53 mutagenesis. *Mutat. Res.* **2013**, *750*, 121–128. [[CrossRef](#)] [[PubMed](#)]
11. Leone, A. Toxics of tobacco smoke and cardiovascular system: From functional to cellular damage. *Curr. Pharm. Des.* **2015**, *21*, 4370–4379. [[CrossRef](#)] [[PubMed](#)]
12. Noya, Y.; Seki, K.; Asano, H.; Mai, Y.; Horinouchi, T.; Higashi, T.; Terada, K.; Hatate, C.; Hoshi, A.; Nepal, P.; et al. Identification of stable cytotoxic factors in the gas phase extract of cigarette smoke and pharmacological characterization of their cytotoxicity. *Toxicology* **2013**, *314*, 1–10. [[CrossRef](#)] [[PubMed](#)]
13. Kilthau, G.F. Cancer risk in relation to radioactivity in tobacco. *Radiol. Technol.* **1996**, *67*, 217–222. [[PubMed](#)]

14. Yuan, J.M.; Murphy, S.E.; Stepanov, I.; Wang, R.; Carmella, S.G.; Nelson, H.H.; Hatsukami, D.K.; Hecht, S.S. 2-Phenethyl isothiocyanate, glutathione S-transferase M1 and T1 polymorphisms, and detoxification of volatile organic carcinogens and toxicants in tobacco smoke. *Cancer Prev. Res.* **2016**, *9*, 598–606. [[CrossRef](#)] [[PubMed](#)]
15. Nicholl, I.D.; Bucala, R. Advanced glycation endproducts and cigarette smoking. *Cell. Mol. Biol.* **1998**, *44*, 1025–1033. [[PubMed](#)]
16. Cerami, C.; Founds, H.; Nicholl, I.; Mitsuhashi, T.; Giordano, D.; Vanpatten, S.; Lee, A.; Al-Abed, Y.; Vlassara, H.; Bucala, R.; et al. Tobacco smoke is a source of toxic reactive glycation products. *Proc. Natl. Acad. Sci. USA* **1997**, *94*, 13915–13920. [[CrossRef](#)] [[PubMed](#)]
17. Robinson, A.B.; Stogsdill, J.A.; Lewis, J.B.; Wood, T.T.; Reynolds, P.R. Rage and tobacco smoke: Insights into modeling chronic obstructive pulmonary disease. *Front. Physiol.* **2012**, *3*, 301. [[CrossRef](#)] [[PubMed](#)]
18. Prasad, K.; Dhar, I.; Caspar-Bell, G. Role of advanced glycation end products and its receptors in the pathogenesis of cigarette smoke-induced cardiovascular disease. *Int. J. Angiol.* **2015**, *24*, 75–80. [[PubMed](#)]
19. Malik, P.; Chaudhry, N.; Mittal, R.; Mukherjee, T.K. Role of receptor for advanced glycation end products in the complication and progression of various types of cancers. *Biochim. Biophys. Acta* **2015**, *1850*, 1898–1904. [[CrossRef](#)] [[PubMed](#)]
20. Nowotny, K.; Jung, T.; Hohn, A.; Weber, D.; Grune, T. Advanced glycation end products and oxidative stress in type 2 diabetes mellitus. *Biomolecules* **2015**, *5*, 194–222. [[CrossRef](#)] [[PubMed](#)]
21. Espinet, C.; Gonzalo, H.; Fleitas, C.; Menal, M.J.; Egea, J. Oxidative stress and neurodegenerative diseases: A neurotrophic approach. *Curr. Drug Targets* **2015**, *16*, 20–30. [[CrossRef](#)] [[PubMed](#)]
22. Guedes-Martins, L.; Matos, L.; Soares, A.; Silva, E.; Almeida, H. Ages, contributors to placental bed vascular changes leading to preeclampsia. *Free Radic. Res.* **2013**, *47* (Suppl. S1), 70–80. [[CrossRef](#)] [[PubMed](#)]
23. Alexander, K.L.; Mejia, C.A.; Jordan, C.; Nelson, M.B.; Howell, B.M.; Jones, C.M.; Reynolds, P.R.; Arroyo, J.A. Differential receptor for advanced glycation end products expression in preeclamptic, intrauterine growth restricted, and gestational diabetic placentas. *Am. J. Reprod. Immunol.* **2016**, *75*, 172–180. [[CrossRef](#)] [[PubMed](#)]
24. Chen, Y.J.; Chan, D.C.; Chiang, C.K.; Wang, C.C.; Yang, T.H.; Lan, K.C.; Chao, S.C.; Tsai, K.S.; Yang, R.S.; Liu, S.H. Advanced glycation end-products induced VEGF production and inflammatory responses in human synoviocytes via RAGE-NF- $\kappa$ B pathway activation. *J. Orthop. Res.* **2015**, *34*, 791–800. [[CrossRef](#)] [[PubMed](#)]
25. Van Puyvelde, K.; Mets, T.; Njemini, R.; Beyer, I.; Bautmans, I. Effect of advanced glycation end product intake on inflammation and aging: A systematic review. *Nutr. Rev.* **2014**, *72*, 638–650. [[CrossRef](#)] [[PubMed](#)]
26. Ray, R.; Juranek, J.K.; Rai, V. RAGE axis in neuroinflammation, neurodegeneration and its emerging role in the pathogenesis of amyotrophic lateral sclerosis. *Neurosci. Biobehav. Rev.* **2016**, *62*, 48–55. [[CrossRef](#)] [[PubMed](#)]
27. Ibrahim, Z.A.; Armour, C.L.; Phipps, S.; Sukkar, M.B. RAGE and TLRs: Relatives, friends or neighbours? *Mol. Immunol.* **2013**, *56*, 739–744. [[CrossRef](#)] [[PubMed](#)]
28. Centers for Disease Control and Prevention (US). *The Health Consequences of Involuntary Exposure to Tobacco Smoke: A Report of the Surgeon General*; Centers for Disease Control and Prevention (US): Atlanta, GA, USA, 2006.
29. Grimmer, G.; Naujack, K.W.; Dettbarn, G. Gaschromatographic determination of polycyclic aromatic hydrocarbons, aza-arenes, aromatic amines in the particle and vapor phase of mainstream and sidestream smoke of cigarettes. *Toxicol. Lett.* **1987**, *35*, 117–124. [[CrossRef](#)]
30. Evans, W.H.; Thomas, N.C.; Boardman, M.C.; Nash, S.J. Relationships of polycyclic aromatic hydrocarbon yields with particulate matter (water and nicotine free) yields in mainstream and sidestream cigarette smoke. *Sci. Total Environ.* **1993**, *136*, 101–109. [[CrossRef](#)]
31. Brunnemann, K.D.; Yu, L.; Hoffmann, D. Assessment of carcinogenic volatile N-nitrosamines in tobacco and in mainstream and sidestream smoke from cigarettes. *Cancer Res.* **1977**, *37*, 3218–3222. [[PubMed](#)]
32. Ruhl, C.; Adams, J.D.; Hoffmann, D. Chemical studies on tobacco-specific N-nitrosamines in the smoke of selected cigarettes from the USA, west germany, and france. *J. Anal. Toxicol.* **1980**, *4*, 255–259. [[CrossRef](#)] [[PubMed](#)]
33. Hoffmann, D.; Adams, J.D.; Brunnemann, K.D.; Hecht, S.S. Assessment of tobacco-specific N-nitrosamines in tobacco products. *Cancer Res.* **1979**, *39 Pt 1*, 2505–2509. [[PubMed](#)]

34. Patrianakos, C.; Hoffmann, D. Chemical studies on tobacco smoke LXIV. On the analysis of aromatic amines in cigarette smoke. *J. Anal. Toxicol.* **1979**, *3*, 150–154. [[CrossRef](#)]
35. Dong, M.; Schmeltz, I.; Jacobs, E.; Hoffmann, D. Aza-arenes in tobacco smoke. *J. Anal. Toxicol.* **1978**, *2*, 21–25. [[CrossRef](#)]
36. Hoffmann, D.; Adams, J.D.; Wynder, E.L. Formation and analysis of carbon monoxide in cigarette mainstream and sidestream smoke. *Prev. Med.* **1979**, *8*, 344–350. [[CrossRef](#)]
37. Krzych-Falta, E.; Modzelewska, D.; Samolinski, B. Levels of exhaled carbon monoxide in healthy active and passive smokers. *Przegl. Lek.* **2015**, *72*, 99–102. [[PubMed](#)]
38. Daher, N.; Saleh, R.; Jaroudi, E.; Sheheitli, H.; Badr, T.; Sepetdjian, E.; Al Rashidi, M.; Saliba, N.; Shihadeh, A. Comparison of carcinogen, carbon monoxide, and ultrafine particle emissions from narghile waterpipe and cigarette smoking: Sidestream smoke measurements and assessment of second-hand smoke emission factors. *Atmos. Environ.* **2010**, *44*, 8–14. [[CrossRef](#)] [[PubMed](#)]
39. Rickert, W.S.; Robinson, J.C.; Collishaw, N. Yields of tar, nicotine, and carbon monoxide in the sidestream smoke from 15 brands of Canadian cigarettes. *Am. J. Public Health* **1984**, *74*, 228–231. [[CrossRef](#)] [[PubMed](#)]
40. Pakhale, S.; Maru, G. Distribution of major and minor alkaloids in tobacco, mainstream and sidestream smoke of popular Indian smoking products. *Food Chem. Toxicol.* **1998**, *36*, 1131–1138. [[CrossRef](#)]
41. Brunnemann, K.D.; Hoffmann, D. Chemical studies on tobacco smoke XXXIV. Gas chromatographic determination of ammonia in cigarette and cigar smoke. *J. Chromatogr. Sci.* **1975**, *13*, 159–163. [[CrossRef](#)] [[PubMed](#)]
42. Johnson, W.; Hale, R.; Clough, S.; Chen, P. Chemistry of the conversion of nitrate nitrogen to smoke products. *Nature* **1973**, *243*, 223–225. [[CrossRef](#)] [[PubMed](#)]
43. Brunnemann, K.; Hoffmann, D. Chemical studies on tobacco smoke LIX. Analysis of volatile nitrosamines in tobacco smoke and polluted indoor environments. *IARC Sci. Publ.* **1977**, *19*, 343–356.
44. Brunnemann, K.D.; Kagan, M.R.; Cox, J.E.; Hoffmann, D. Analysis of 1,3-butadiene and other selected gas-phase components in cigarette mainstream and sidestream smoke by gas chromatography-mass selective detection. *Carcinogenesis* **1990**, *11*, 1863–1868. [[CrossRef](#)] [[PubMed](#)]
45. Ganjre, A.P.; Sarode, G.S. Third hand smoke—A hidden demon. *Oral Oncol.* **2016**, *54*, e3–e4. [[CrossRef](#)] [[PubMed](#)]
46. Sleiman, M.; Gundel, L.A.; Pankow, J.F.; Jacob, P., 3rd; Singer, B.C.; Destailats, H. Formation of carcinogens indoors by surface-mediated reactions of nicotine with nitrous acid, leading to potential thirdhand smoke hazards. *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 6576–6581. [[CrossRef](#)] [[PubMed](#)]
47. Ferrante, G.; Simoni, M.; Cibella, F.; Ferrara, F.; Liotta, G.; Malizia, V.; Corsello, G.; Viegi, G.; La Grutta, S. Third-hand smoke exposure and health hazards in children. *Monaldi Arch. Chest. Dis.* **2013**, *79*, 38–43. [[CrossRef](#)] [[PubMed](#)]
48. Casey, G. Copd: Obstructed lungs. *Nurs. N. Z.* **2016**, *22*, 20–24. [[PubMed](#)]
49. Hagstad, S.; Bjerg, A.; Ekerljung, L.; Backman, H.; Lindberg, A.; Ronmark, E.; Lundback, B. Passive smoking exposure is associated with increased risk of COPD in never smokers. *Chest* **2014**, *145*, 1298–1304. [[CrossRef](#)] [[PubMed](#)]
50. Lomborg, B. *Global Problems, Smart Solutions: Costs and Benefits*; Cambridge University Press: Cambridge, UK, 2013.
51. Rennard, S.I. Treatment of stable chronic obstructive pulmonary disease. *Lancet* **2004**, *364*, 791–802. [[CrossRef](#)]
52. Sutherland, E.R.; Cherniack, R.M. Management of chronic obstructive pulmonary disease. *N. Engl. J. Med.* **2004**, *350*, 2689–2697. [[CrossRef](#)] [[PubMed](#)]
53. Hogg, J.C.; Chu, F.; Utokaparch, S.; Woods, R.; Elliott, W.M.; Buzatu, L.; Cherniack, R.M.; Rogers, R.M.; Sciurba, F.C.; Coxson, H.O.; et al. The nature of small-airway obstruction in chronic obstructive pulmonary disease. *N. Engl. J. Med.* **2004**, *350*, 2645–2653. [[CrossRef](#)] [[PubMed](#)]
54. Barnes, P.J. Small airways in COPD. *N. Engl. J. Med.* **2004**, *350*, 2635–2637. [[CrossRef](#)] [[PubMed](#)]
55. Barnes, P.J.; Shapiro, S.D.; Pauwels, R.A. Chronic obstructive pulmonary disease: Molecular and cellular mechanisms. *Eur. Respir. J.* **2003**, *22*, 672–688. [[CrossRef](#)] [[PubMed](#)]
56. Dawkins, P.A.; Stockley, R.A. Animal models of chronic obstructive pulmonary disease. *Thorax* **2001**, *56*, 972–977. [[CrossRef](#)] [[PubMed](#)]
57. Shapiro, S.D. Animal models for COPD. *Chest* **2000**, *117* (Suppl. S1), 223S–227S. [[CrossRef](#)] [[PubMed](#)]
58. Mortaz, E.; Adcock, I.A. Limitation of COPD studies in animal modeling. *Tanaffos* **2012**, *11*, 7–8. [[PubMed](#)]

59. Ravi, A.K.; Khurana, S.; Lemon, J.; Plumb, J.; Booth, G.; Healy, L.; Catley, M.; Vestbo, J.; Singh, D. Increased levels of soluble interleukin-6 receptor and CCL3 in COPD sputum. *Respir. Res.* **2014**, *15*, 103. [[CrossRef](#)] [[PubMed](#)]
60. Menzies, D.; Nair, A.; Williamson, P.A.; Schembri, S.; Al-Khairalla, M.Z.; Barnes, M.; Fardon, T.C.; McFarlane, L.; Magee, G.J.; Lipworth, B.J. Respiratory symptoms, pulmonary function, and markers of inflammation among bar workers before and after a legislative ban on smoking in public places. *JAMA* **2006**, *296*, 1742–1748. [[CrossRef](#)] [[PubMed](#)]
61. Wu, H.; Yang, S.; Wu, X.; Zhao, J.; Zhao, J.; Ning, Q.; Xu, Y.; Xie, J. Interleukin-33/ST2 signaling promotes production of interleukin-6 and interleukin-8 in systemic inflammation in cigarette smoke-induced chronic obstructive pulmonary disease mice. *Biochem. Biophys. Res. Commun.* **2014**, *450*, 110–116. [[CrossRef](#)] [[PubMed](#)]
62. Hubeau, C.; Kubera, J.E.; Masek-Hammerman, K.; Williams, C.M. Interleukin-6 neutralization alleviates pulmonary inflammation in mice exposed to cigarette smoke and poly(I:C). *Clin. Sci.* **2013**, *125*, 483–493. [[CrossRef](#)] [[PubMed](#)]
63. Overbeek, S.A.; Braber, S.; Koelink, P.J.; Henricks, P.A.; Mortaz, E.; LoTam Loi, A.T.; Jackson, P.L.; Garssen, J.; Wagenaar, G.T.; Timens, W.; et al. Cigarette smoke-induced collagen destruction; key to chronic neutrophilic airway inflammation? *PLoS ONE* **2013**, *8*, e55612. [[CrossRef](#)]
64. Kobayashi, S.D.; DeLeo, F.R. Role of neutrophils in innate immunity: A systems biology-level approach. *Wiley Interdiscip. Rev. Syst. Biol. Med.* **2009**, *1*, 309–333. [[CrossRef](#)] [[PubMed](#)]
65. Kobayashi, S.D.; Voyich, J.M.; Burlak, C.; DeLeo, F.R. Neutrophils in the innate immune response. *Arch. Immunol. Ther. Exp. (Warsz.)* **2005**, *53*, 505–517.
66. Keatings, V.M.; Collins, P.D.; Scott, D.M.; Barnes, P.J. Differences in interleukin-8 and tumor necrosis factor- $\alpha$  in induced sputum from patients with chronic obstructive pulmonary disease or asthma. *Am. J. Respir. Crit. Care Med.* **1996**, *153*, 530–534. [[CrossRef](#)] [[PubMed](#)]
67. Tanino, M.; Betsuyaku, T.; Takeyabu, K.; Tanino, Y.; Yamaguchi, E.; Miyamoto, K.; Nishimura, M. Increased levels of interleukin-8 in BAL fluid from smokers susceptible to pulmonary emphysema. *Thorax* **2002**, *57*, 405–411. [[CrossRef](#)] [[PubMed](#)]
68. Beeh, K.M.; Kornmann, O.; Buhl, R.; Culpitt, S.V.; Giembycz, M.A.; Barnes, P.J. Neutrophil chemotactic activity of sputum from patients with COPD: Role of interleukin 8 and leukotriene b<sub>4</sub>. *Chest* **2003**, *123*, 1240–1247. [[CrossRef](#)] [[PubMed](#)]
69. Hodge, S.; Hodge, G.; Holmes, M.; Reynolds, P.N. Increased airway epithelial and t-cell apoptosis in COPD remains despite smoking cessation. *Eur. Respir. J.* **2005**, *25*, 447–454. [[CrossRef](#)] [[PubMed](#)]
70. Thacker, E.L. Lung inflammatory responses. *Vet. Res.* **2006**, *37*, 469–486. [[CrossRef](#)] [[PubMed](#)]
71. Churg, A.; Zhou, S.; Wang, X.; Wang, R.; Wright, J.L. The role of interleukin-1 $\beta$  in murine cigarette smoke-induced emphysema and small airway remodeling. *Am. J. Respir. Cell Mol. Biol.* **2009**, *40*, 482–490. [[CrossRef](#)] [[PubMed](#)]
72. Barnes, P.J.; Karin, M. Nuclear factor- $\kappa$ B: A pivotal transcription factor in chronic inflammatory diseases. *N. Engl. J. Med.* **1997**, *336*, 1066–1071. [[CrossRef](#)]
73. Merghani, T.H.; Saeed, A.; Alawad, A. Changes in plasma IL4, TNFA and CRP in response to regular passive smoking at home among healthy school children in khartoum, Sudan. *Afr. Health Sci.* **2012**, *12*, 41–47. [[PubMed](#)]
74. Flouris, A.D.; Metsios, G.S.; Carrillo, A.E.; Jamurtas, A.Z.; Stivaktakis, P.D.; Tzatzarakis, M.N.; Tsatsakis, A.M.; Koutedakis, Y. Respiratory and immune response to maximal physical exertion following exposure to secondhand smoke in healthy adults. *PLoS ONE* **2012**, *7*, e31880. [[CrossRef](#)]
75. Aldonyte, R.; Jansson, L.; Piitulainen, E.; Janciauskiene, S. Circulating monocytes from healthy individuals and COPD patients. *Respir. Res.* **2003**, *4*, 11. [[CrossRef](#)] [[PubMed](#)]
76. Finkelstein, R.; Fraser, R.S.; Ghezzi, H.; Cosio, M.G. Alveolar inflammation and its relation to emphysema in smokers. *Am. J. Respir. Crit. Care Med.* **1995**, *152 Pt 1*, 1666–1672. [[CrossRef](#)] [[PubMed](#)]
77. Shapiro, S.D. The macrophage in chronic obstructive pulmonary disease. *Am. J. Respir. Crit. Care Med.* **1999**, *160*, S29–S32. [[CrossRef](#)] [[PubMed](#)]
78. Tsuji, H.; Fujimoto, H.; Lee, K.M.; Renne, R.; Iwanaga, A.; Okubo, C.; Onami, S.; Nomura, A.K.; Nishino, T.; Yoshimura, H. Characterization of biochemical, functional and structural changes in mice respiratory organs chronically exposed to cigarette smoke. *Inhal. Toxicol.* **2015**, *27*, 342–353. [[CrossRef](#)] [[PubMed](#)]



79. Di Stefano, A.; Capelli, A.; Lusuardi, M.; Balbo, P.; Vecchio, C.; Maestrelli, P.; Mapp, C.E.; Fabbri, L.M.; Donner, C.F.; Saetta, M. Severity of airflow limitation is associated with severity of airway inflammation in smokers. *Am. J. Respir. Crit. Care Med.* **1998**, *158*, 1277–1285. [[CrossRef](#)] [[PubMed](#)]
80. Majo, J.; Ghezzi, H.; Cosio, M.G. Lymphocyte population and apoptosis in the lungs of smokers and their relation to emphysema. *Eur. Respir. J.* **2001**, *17*, 946–953. [[CrossRef](#)] [[PubMed](#)]
81. Podolin, P.L.; Foley, J.P.; Carpenter, D.C.; Bolognese, B.J.; Logan, G.A.; Long, E., 3rd; Harrison, O.J.; Walsh, P.T. T-cell depletion protects against alveolar destruction due to chronic cigarette smoke exposure in mice. *Am. J. Physiol. Lung Cell. Mol. Physiol.* **2013**, *304*, L312–L323. [[CrossRef](#)] [[PubMed](#)]
82. Pfeifer, G.P.; Denissenko, M.F.; Olivier, M.; Tretyakova, N.; Hecht, S.S.; Hainaut, P. Tobacco smoke carcinogens, DNA damage and p53 mutations in smoking-associated cancers. *Oncogene* **2002**, *21*, 7435–7451. [[CrossRef](#)] [[PubMed](#)]
83. Danaei, G.; Vander Hoorn, S.; Lopez, A.D.; Murray, C.J.; Ezzati, M. Causes of cancer in the world: Comparative risk assessment of nine behavioural and environmental risk factors. *Lancet* **2005**, *366*, 1784–1793. [[CrossRef](#)]
84. Siegel, R.; Ma, J.; Zou, Z.; Jemal, A. Cancer statistics, 2014. *CA Cancer J. Clin.* **2014**, *64*, 9–29. [[CrossRef](#)] [[PubMed](#)]
85. Lim, S.S.; Vos, T.; Flaxman, A.D.; Danaei, G.; Shibuya, K.; Adair-Rohani, H.; Amann, M.; Anderson, H.R.; Andrews, K.G.; Aryee, M.; et al. A comparative risk assessment of burden of disease and injury attributable to 67 risk factors and risk factor clusters in 21 regions, 1990–2010: A systematic analysis for the global burden of disease study 2010. *Lancet* **2012**, *380*, 2224–2260. [[CrossRef](#)]
86. Agudo, A.; Bonet, C.; Travier, N.; Gonzalez, C.A.; Vineis, P.; Bueno-de-Mesquita, H.B.; Trichopoulos, D.; Boffetta, P.; Clavel-Chapelon, F.; Boutron-Ruault, M.C.; et al. Impact of cigarette smoking on cancer risk in the European prospective investigation into cancer and nutrition study. *J. Clin. Oncol.* **2012**, *30*, 4550–4557. [[CrossRef](#)] [[PubMed](#)]
87. Hori, M.; Tanaka, H.; Wakai, K.; Sasazuki, S.; Katanoda, K. Secondhand smoke exposure and risk of lung cancer in Japan: A systematic review and meta-analysis of epidemiologic studies. *Jpn. J. Clin. Oncol.* **2016**, *46*, 942–951. [[CrossRef](#)] [[PubMed](#)]
88. Malik, A.; Jeyaraj, P.A.; Shankar, A.; Rath, G.K.; Mukhopadhyay, S.; Kamal, V.K. Passive smoking and breast cancer—A suspicious link. *Asian Pac. J. Cancer Prev.* **2015**, *16*, 5715–5719. [[CrossRef](#)] [[PubMed](#)]
89. Lee, S.H.; Hong, J.Y.; Lee, J.U.; Lee, D.R. Association between exposure to environmental tobacco smoke at the workplace and risk for developing a colorectal adenoma: A cross-sectional study. *Ann. Coloproctol.* **2016**, *32*, 51–57. [[CrossRef](#)] [[PubMed](#)]
90. Shekari, M.; Kordi-Tamandani, D.M.; MalekZadeh, K.; Solti, R.C.; Karimi, S.; Suri, V. Effect of anti-inflammatory (IL-4, IL-10) cytokine genes in relation to risk of cervical carcinoma. *Am. J. Clin. Oncol.* **2012**, *35*, 514–519. [[CrossRef](#)] [[PubMed](#)]
91. Vrieling, A.; Bueno-de-Mesquita, H.B.; Boshuizen, H.C.; Michaud, D.S.; Severinsen, M.T.; Overvad, K.; Olsen, A.; Tjonneland, A.; Clavel-Chapelon, F.; Boutron-Ruault, M.C.; et al. Cigarette smoking, environmental tobacco smoke exposure and pancreatic cancer risk in the European prospective investigation into cancer and nutrition. *Int. J. Cancer* **2010**, *126*, 2394–2403. [[CrossRef](#)] [[PubMed](#)]
92. Sleiman, M.; Maddalena, R.L.; Gundel, L.A.; Destailats, H. Rapid and sensitive gas chromatography-ion-trap tandem mass spectrometry method for the determination of tobacco-specific N-nitrosamines in secondhand smoke. *J. Chromatogr. A* **2009**, *1216*, 7899–7905. [[CrossRef](#)] [[PubMed](#)]
93. Church, T.R.; Anderson, K.E.; Caporaso, N.E.; Geisser, M.S.; Le, C.T.; Zhang, Y.; Benoit, A.R.; Carmella, S.G.; Hecht, S.S. A prospectively measured serum biomarker for a tobacco-specific carcinogen and lung cancer in smokers. *Cancer Epidemiol. Biomark. Prev.* **2009**, *18*, 260–266. [[CrossRef](#)] [[PubMed](#)]
94. Balbo, S.; James-Yi, S.; Johnson, C.S.; O'Sullivan, M.G.; Stepanov, I.; Wang, M.; Bandyopadhyay, D.; Kassie, F.; Carmella, S.; Upadhyaya, P.; et al. (S)-N'-nitrosonornicotine, a constituent of smokeless tobacco, is a powerful oral cavity carcinogen in rats. *Carcinogenesis* **2013**, *34*, 2178–2183. [[CrossRef](#)] [[PubMed](#)]
95. Gonzalez-Jasso, E.; Lopez, T.; Lucas, D.; Berthou, F.; Manno, M.; Ortega, A.; Albores, A. CYP2E1 regulation by benzene and other small organic chemicals in rat liver and peripheral lymphocytes. *Toxicol. Lett.* **2003**, *144*, 55–67. [[CrossRef](#)]
96. Joshi, M.; Tyndale, R.F. Induction and recovery time course of rat brain cyp2e1 after nicotine treatment. *Drug Metab. Dispos.* **2006**, *34*, 647–652. [[CrossRef](#)] [[PubMed](#)]



97. Guengerich, F.P.; Shimada, T. Oxidation of toxic and carcinogenic chemicals by human cytochrome p-450 enzymes. *Chem. Res. Toxicol.* **1991**, *4*, 391–407. [[CrossRef](#)] [[PubMed](#)]
98. Lieber, C.S. Cytochrome P-450E1: Its physiological and pathological role. *Physiol. Rev.* **1997**, *77*, 517–544. [[PubMed](#)]
99. Kase, S.; Sugio, K.; Yamazaki, K.; Okamoto, T.; Yano, T.; Sugimachi, K. Expression of E-cadherin and  $\beta$ -catenin in human non-small cell lung cancer and the clinical significance. *Clin. Cancer Res.* **2000**, *6*, 4789–4796. [[CrossRef](#)]
100. Hecht, S.S. Tobacco smoke carcinogens and lung cancer. *J. Natl. Cancer Inst.* **1999**, *91*, 1194–1210. [[CrossRef](#)] [[PubMed](#)]
101. Hecht, S.S. Tobacco carcinogens, their biomarkers and tobacco-induced cancer. *Nat. Rev. Cancer* **2003**, *3*, 733–744. [[CrossRef](#)] [[PubMed](#)]
102. Hecht, S.S. Lung carcinogenesis by tobacco smoke. *Int. J. Cancer* **2012**, *131*, 2724–2732. [[CrossRef](#)] [[PubMed](#)]
103. Thompson, C.A.; Burcham, P.C. Genome-wide transcriptional responses to acrolein. *Chem. Res. Toxicol.* **2008**, *21*, 2245–2256. [[CrossRef](#)] [[PubMed](#)]
104. Cho, W.C.; Kwan, C.K.; Yau, S.; So, P.P.; Poon, P.C.; Au, J.S. The role of inflammation in the pathogenesis of lung cancer. *Expert Opin. Ther. Targets* **2011**, *15*, 1127–1137. [[CrossRef](#)] [[PubMed](#)]
105. Erreni, M.; Mantovani, A.; Allavena, P. Tumor-associated macrophages (TAM) and inflammation in colorectal cancer. *Cancer Microenviron.* **2011**, *4*, 141–154. [[CrossRef](#)] [[PubMed](#)]
106. Jang, J.Y.; Lee, J.K.; Jeon, Y.K.; Kim, C.W. Exosome derived from epigallocatechin gallate treated breast cancer cells suppresses tumor growth by inhibiting tumor-associated macrophage infiltration and M2 polarization. *BMC Cancer* **2013**, *13*, 421. [[CrossRef](#)] [[PubMed](#)]
107. Mano, Y.; Aishima, S.; Fujita, N.; Tanaka, Y.; Kubo, Y.; Motomura, T.; Taketomi, A.; Shirabe, K.; Maehara, Y.; Oda, Y. Tumor-associated macrophage promotes tumor progression via STAT3 signaling in hepatocellular carcinoma. *Pathobiology* **2013**, *80*, 146–154. [[CrossRef](#)] [[PubMed](#)]
108. Ho, C.C.; Liao, W.Y.; Wang, C.Y.; Lu, Y.H.; Huang, H.Y.; Chen, H.Y.; Chan, W.K.; Chen, H.W.; Yang, P.C. TREM-1 expression in tumor-associated macrophages and clinical outcome in lung cancer. *Am. J. Respir. Crit. Care Med.* **2008**, *177*, 763–770. [[CrossRef](#)] [[PubMed](#)]
109. Ala-aho, R.; Kahari, V.M. Collagenases in cancer. *Biochimie* **2005**, *87*, 273–286. [[CrossRef](#)] [[PubMed](#)]
110. Stamenkovic, I. Matrix metalloproteinases in tumor invasion and metastasis. *Semin. Cancer Biol.* **2000**, *10*, 415–433. [[CrossRef](#)] [[PubMed](#)]
111. Stamenkovic, I. Extracellular matrix remodelling: The role of matrix metalloproteinases. *J. Pathol.* **2003**, *200*, 448–464. [[CrossRef](#)] [[PubMed](#)]
112. Ozturk, F.; Sheldon, E.; Sharma, J.; Canturk, K.M.; Otu, H.H.; Nawshad, A. Nicotine exposure during pregnancy results in persistent midline epithelial seam with improper palatal fusion. *Nicotine Tob. Res.* **2016**, *18*, 604–612. [[CrossRef](#)] [[PubMed](#)]
113. Arffin, F.; Al-Bayaty, F.H.; Hassan, J. Environmental tobacco smoke and stress as risk factors for miscarriage and preterm births. *Arch. Gynecol. Obstet.* **2012**, *286*, 1187–1191. [[CrossRef](#)] [[PubMed](#)]
114. National Center for Chronic Disease Prevention and Health Promotion (US) Office on Smoking and Health. *The Health Consequences of Smoking—50 Years of Progress: A Report of the Surgeon General*; US Department of Health and Human Services, Centers for Disease Control and Prevention, National Center for Chronic Disease Prevention and Health Promotion, Office on Smoking and Health: Atlanta, GA, USA, 2014; Volume 17.
115. Caspers, K.M.; Romitti, P.A.; Lin, S.; Olney, R.S.; Holmes, L.B.; Werler, M.M. Maternal periconceptional exposure to cigarette smoking and congenital limb deficiencies. *Paediatr. Perinat. Epidemiol.* **2013**, *27*, 509–520. [[CrossRef](#)] [[PubMed](#)]
116. Gianicolo, E.A.; Cresci, M.; Ait-Ali, L.; Foffa, I.; Andreassi, M.G. Smoking and congenital heart disease: The epidemiological and biological link. *Curr. Pharm. Des.* **2010**, *16*, 2572–2577. [[CrossRef](#)] [[PubMed](#)]
117. Salmasi, G.; Grady, R.; Jones, J.; McDonald, S.D. Environmental tobacco smoke exposure and perinatal outcomes: A systematic review and meta-analyses. *Acta Obstet. Gynecol. Scand.* **2010**, *89*, 423–441. [[CrossRef](#)] [[PubMed](#)]
118. Dietz, P.M.; England, L.J.; Shapiro-Mendoza, C.K.; Tong, V.T.; Farr, S.L.; Callaghan, W.M. Infant morbidity and mortality attributable to prenatal smoking in the U.S. *Am. J. Prev. Med.* **2010**, *39*, 45–52. [[CrossRef](#)] [[PubMed](#)]

119. Higgins, S.T.; Washio, Y.; Heil, S.H.; Solomon, L.J.; Gaalema, D.E.; Higgins, T.M.; Bernstein, I.M. Financial incentives for smoking cessation among pregnant and newly postpartum women. *Prev. Med.* **2012**, *55*, S33–S40. [[CrossRef](#)] [[PubMed](#)]
120. Bickerstaff, M.; Beckmann, M.; Gibbons, K.; Flenady, V. Recent cessation of smoking and its effect on pregnancy outcomes. *Aust. N. Z. J. Obstet. Gynaecol.* **2012**, *52*, 54–58. [[CrossRef](#)] [[PubMed](#)]
121. Akkar, O.B.; Yildiz, C.; Karakus, S.; Akkar, I.; Cetin, A.; Yanik, A.; Yenicesu, A.G.; Boztosun, A. Antenatal counseling against passive smoking may improve birth weight for gestational age. *Clin. Exp. Obstet. Gynecol.* **2015**, *42*, 805–809. [[PubMed](#)]
122. Strulovici-Barel, Y.; Omberg, L.; O'Mahony, M.; Gordon, C.; Hollmann, C.; Tilley, A.E.; Salit, J.; Mezey, J.; Harvey, B.G.; Crystal, R.G. Threshold of biologic responses of the small airway epithelium to low levels of tobacco smoke. *Am. J. Respir. Crit. Care Med.* **2010**, *182*, 1524–1532. [[CrossRef](#)] [[PubMed](#)]
123. Hellstrom-Lindahl, E.; Gorbounova, O.; Seiger, A.; Mousavi, M.; Nordberg, A. Regional distribution of nicotinic receptors during prenatal development of human brain and spinal cord. *Brain Res. Dev. Brain Res.* **1998**, *108*, 147–160. [[CrossRef](#)]
124. Dwyer, J.B.; McQuown, S.C.; Leslie, F.M. The dynamic effects of nicotine on the developing brain. *Pharmacol. Ther.* **2009**, *122*, 125–139. [[CrossRef](#)] [[PubMed](#)]
125. Luck, W.; Nau, H.; Hansen, R.; Steldinger, R. Extent of nicotine and cotinine transfer to the human fetus, placenta and amniotic fluid of smoking mothers. *Dev. Pharmacol. Ther.* **1985**, *8*, 384–395. [[PubMed](#)]
126. Slotkin, T.A. Fetal nicotine or cocaine exposure: Which one is worse? *J. Pharmacol. Exp. Ther.* **1998**, *285*, 931–945. [[PubMed](#)]
127. Slotkin, T.A.; Cho, H.; Whitmore, W.L. Effects of prenatal nicotine exposure on neuronal development: Selective actions on central and peripheral catecholaminergic pathways. *Brain Res. Bull.* **1987**, *18*, 601–611. [[CrossRef](#)]
128. Dwyer, J.B.; Broide, R.S.; Leslie, F.M. Nicotine and brain development. *Birth Defects Res. C Embryo Today* **2008**, *84*, 30–44. [[CrossRef](#)] [[PubMed](#)]
129. Palmer, R.H.; Bidwell, L.C.; Heath, A.C.; Brick, L.A.; Madden, P.A.; Knopik, V.S. Effects of maternal smoking during pregnancy on offspring externalizing problems: Contextual effects in a sample of female twins. *Behav. Genet.* **2016**, *46*, 403–415. [[CrossRef](#)] [[PubMed](#)]
130. Niclasen, J.; Obel, C.; Homoe, P.; Korvel-Hanquist, A.; Dammeyer, J. Associations between otitis media and child behavioural and learning difficulties: Results from a Danish cohort. *Int. J. Pediatr. Otorhinolaryngol.* **2016**, *84*, 12–20. [[CrossRef](#)] [[PubMed](#)]
131. Knopik, V.S.; Marceau, K.; Bidwell, L.C.; Palmer, R.H.; Smith, T.F.; Todorov, A.; Evans, A.S.; Heath, A.C. Smoking during pregnancy and ADHD risk: A genetically informed, multiple-rater approach. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **2016**, *171*, 971–981. [[CrossRef](#)] [[PubMed](#)]
132. De Alwis, D.; Tandon, M.; Tillman, R.; Luby, J. Nonverbal reasoning in preschool children: Investigating the putative risk of secondhand smoke exposure and attention-deficit/hyperactivity disorder as a mediator. *Scand. J. Child. Adolesc. Psychiatr. Psychol.* **2015**, *3*, 115–125. [[CrossRef](#)] [[PubMed](#)]
133. Hanrahan, J.P.; Tager, I.B.; Segal, M.R.; Tosteson, T.D.; Castile, R.G.; Van Vunakis, H.; Weiss, S.T.; Speizer, F.E. The effect of maternal smoking during pregnancy on early infant lung function. *Am. Rev. Respir. Dis.* **1992**, *145*, 1129–1135. [[CrossRef](#)] [[PubMed](#)]
134. Stillerman, K.P.; Mattison, D.R.; Giudice, L.C.; Woodruff, T.J. Environmental exposures and adverse pregnancy outcomes: A review of the science. *Reprod. Sci.* **2008**, *15*, 631–650. [[CrossRef](#)] [[PubMed](#)]
135. Votavova, H.; Dostalova Merkerova, M.; Krejcik, Z.; Fejglova, K.; Vasikova, A.; Pastorkova, A.; Tabashidze, N.; Topinka, J.; Balascak, I.; Sram, R.J.; et al. Deregulation of gene expression induced by environmental tobacco smoke exposure in pregnancy. *Nicotine Tob. Res.* **2012**, *14*, 1073–1082. [[CrossRef](#)] [[PubMed](#)]
136. Lodrup Carlsen, K.C.; Jaakkola, J.J.; Nafstad, P.; Carlsen, K.H. In utero exposure to cigarette smoking influences lung function at birth. *Eur. Respir. J.* **1997**, *10*, 1774–1779. [[CrossRef](#)] [[PubMed](#)]
137. Stocks, J.; Dezateux, C. The effect of parental smoking on lung function and development during infancy. *Respirology* **2003**, *8*, 266–285. [[CrossRef](#)] [[PubMed](#)]
138. Sekhon, H.S.; Jia, Y.; Raab, R.; Kuryatov, A.; Pankow, J.F.; Whitsett, J.A.; Lindstrom, J.; Spindel, E.R. Prenatal nicotine increases pulmonary  $\alpha 7$  nicotinic receptor expression and alters fetal lung development in monkeys. *J. Clin. Investig.* **1999**, *103*, 637–647. [[CrossRef](#)] [[PubMed](#)]

139. Sekhon, H.S.; Keller, J.A.; Proskocil, B.J.; Martin, E.L.; Spindel, E.R. Maternal nicotine exposure upregulates collagen gene expression in fetal monkey lung. Association with alpha7 nicotinic acetylcholine receptors. *Am. J. Respir. Cell Mol. Biol.* **2002**, *26*, 31–41. [[CrossRef](#)] [[PubMed](#)]
140. Rehan, V.K.; Liu, J.; Naeem, E.; Tian, J.; Sakurai, R.; Kwong, K.; Akbari, O.; Torday, J.S. Perinatal nicotine exposure induces asthma in second generation offspring. *BMC Med.* **2012**, *10*, 129. [[CrossRef](#)] [[PubMed](#)]
141. Li, Y.F.; Langholz, B.; Salam, M.T.; Gilliland, F.D. Maternal and grandmaternal smoking patterns are associated with early childhood asthma. *Chest* **2005**, *127*, 1232–1241. [[CrossRef](#)]
142. Leslie, F.M. Multigenerational epigenetic effects of nicotine on lung function. *BMC Med.* **2013**, *11*, 27. [[CrossRef](#)] [[PubMed](#)]
143. Duskova, M.; Hruskovicova, H.; Simunkova, K.; Starka, L.; Parizek, A. The effects of smoking on steroid metabolism and fetal programming. *J. Steroid Biochem. Mol. Biol.* **2014**, *139*, 138–143. [[CrossRef](#)] [[PubMed](#)]
144. Steptoe, A.; Ussher, M. Smoking, cortisol and nicotine. *Int. J. Psychophysiol.* **2006**, *59*, 228–235. [[CrossRef](#)] [[PubMed](#)]
145. Stangenberg, S.; Chen, H.; Wong, M.G.; Pollock, C.A.; Saad, S. Fetal programming of chronic kidney disease: The role of maternal smoking, mitochondrial dysfunction, and epigenetic modification. *Am. J. Physiol. Renal Physiol.* **2015**, *308*, F1189–F1196. [[CrossRef](#)] [[PubMed](#)]
146. Kable, J.A.; Coles, C.D.; Lynch, M.E.; Carroll, J. The impact of maternal smoking on fast auditory brainstem responses. *Neurotoxicol. Teratol.* **2009**, *31*, 216–224. [[CrossRef](#)] [[PubMed](#)]
147. Weitzman, M.; Govil, N.; Liu, Y.H.; Lalwani, A.K. Maternal prenatal smoking and hearing loss among adolescents. *JAMA Otolaryngol. Head Neck Surg.* **2013**, *139*, 669–677. [[PubMed](#)]
148. Katbamna, B.; Klutz, N.; Pudrith, C.; Lavery, J.P.; Ide, C.F. Prenatal smoke exposure: Effects on infant auditory system and placental gene expression. *Neurotoxicol. Teratol.* **2013**, *38*, 61–71. [[CrossRef](#)] [[PubMed](#)]
149. Wagijo, M.A.; Sheikh, A.; Duijts, L.; Been, J.V. Reducing tobacco smoking and smoke exposure to prevent preterm birth and its complications. *Paediatr. Respir. Rev.* **2015**. [[CrossRef](#)] [[PubMed](#)]
150. Suter, M.A.; Anders, A.M.; Aagaard, K.M. Maternal smoking as a model for environmental epigenetic changes affecting birthweight and fetal programming. *Mol. Hum. Reprod.* **2013**, *19*, 1–6. [[CrossRef](#)] [[PubMed](#)]
151. Horta, B.L.; Victora, C.G.; Menezes, A.M.; Halpern, R.; Barros, F.C. Low birthweight, preterm births and intrauterine growth retardation in relation to maternal smoking. *Paediatr. Perinatal Epidemiol.* **1997**, *11*, 140–151. [[CrossRef](#)]
152. Bolehovska, P.; Sehnal, B.; Driak, D.; Halaska, M.; Magner, M.; Novotny, J.; Svandova, I. Changes in placental angiogenesis and their correlation with foetal intrauterine restriction. *Ceska Gynecol.* **2015**, *80*, 144–150. [[PubMed](#)]
153. Goldenberg, R.L.; Rouse, D.J. Prevention of premature birth. *N. Engl. J. Med.* **1998**, *339*, 313–320. [[CrossRef](#)] [[PubMed](#)]
154. Brar, H.S.; Rutherford, S.E. Classification of intrauterine growth retardation. *Semin. Perinatol.* **1988**, *12*, 2–10. [[PubMed](#)]
155. Gray, P.H.; O'Callaghan, M.J.; Harvey, J.M.; Burke, C.J.; Payton, D.J. Placental pathology and neurodevelopment of the infant with intrauterine growth restriction. *Dev. Med. Child Neurol.* **1999**, *41*, 16–20. [[CrossRef](#)] [[PubMed](#)]
156. Roger, V.L.; Go, A.S.; Lloyd-Jones, D.M.; Benjamin, E.J.; Berry, J.D.; Borden, W.B.; Bravata, D.M.; Dai, S.; Ford, E.S.; Fox, C.S.; et al. Heart disease and stroke statistics—2012 update: A report from the american heart association. *Circulation* **2012**, *125*, e2–e220. [[PubMed](#)]
157. Menke, A.; Casagrande, S.; Geiss, L.; Cowie, C.C. Prevalence of and trends in diabetes among adults in the United States, 1988–2012. *JAMA* **2015**, *314*, 1021–1029. [[CrossRef](#)] [[PubMed](#)]
158. Reaven, G.M. Do high carbohydrate diets prevent the development or attenuate the manifestations (or both) of syndrome X? A viewpoint strongly against. *Curr. Opin. Lipidol.* **1997**, *8*, 23–27. [[CrossRef](#)] [[PubMed](#)]
159. Facchini, F.S.; Hollenbeck, C.B.; Jeppesen, J.; Chen, Y.D.; Reaven, G.M. Insulin resistance and cigarette smoking. *Lancet* **1992**, *339*, 1128–1130. [[CrossRef](#)]
160. Ronnemaa, T.; Ronnemaa, E.M.; Puukka, P.; Pyorala, K.; Laakso, M. Smoking is independently associated with high plasma insulin levels in nondiabetic men. *Diabetes Care* **1996**, *19*, 1229–1232. [[CrossRef](#)] [[PubMed](#)]
161. Attvall, S.; Fowelin, J.; Lager, I.; Von Schenck, H.; Smith, U. Smoking induces insulin resistance—A potential link with the insulin resistance syndrome. *J. Intern. Med.* **1993**, *233*, 327–332. [[CrossRef](#)] [[PubMed](#)]

162. Thatcher, M.O.; Tippetts, T.S.; Nelson, M.B.; Swensen, A.C.; Winden, D.R.; Hansen, M.E.; Anderson, M.C.; Johnson, I.E.; Porter, J.P.; Reynolds, P.R.; et al. Ceramides mediate cigarette smoke-induced metabolic disruption in mice. *Am. J. Physiol. Endocrinol. Metabol.* **2014**, *307*, E919–E927. [[CrossRef](#)] [[PubMed](#)]
163. DeFronzo, R.A.; Ferrannini, E. Insulin resistance. A multifaceted syndrome responsible for NIDDM, obesity, hypertension, dyslipidemia, and atherosclerotic cardiovascular disease. *Diabetes Care* **1991**, *14*, 173–194. [[CrossRef](#)] [[PubMed](#)]
164. Ferrannini, E.; Buzzigoli, G.; Bonadonna, R.; Giorico, M.A.; Oleggini, M.; Graziadei, L.; Pedrinelli, R.; Brandi, L.; Bevilacqua, S. Insulin resistance in essential hypertension. *N. Engl. J. Med.* **1987**, *317*, 350–357. [[CrossRef](#)] [[PubMed](#)]
165. Reaven, G.M. Insulin resistance and compensatory hyperinsulinemia: Role in hypertension, dyslipidemia, and coronary heart disease. *Am. Heart J.* **1991**, *121 Pt 2*, 1283–1288. [[CrossRef](#)]
166. Witteles, R.M.; Tang, W.H.; Jamali, A.H.; Chu, J.W.; Reaven, G.M.; Fowler, M.B. Insulin resistance in idiopathic dilated cardiomyopathy: A possible etiologic link. *J. Am. Coll. Cardiol.* **2004**, *44*, 78–81. [[CrossRef](#)] [[PubMed](#)]
167. Olefsky, J.M.; Glass, C.K. Macrophages, inflammation, and insulin resistance. *Annu. Rev. Physiol.* **2010**, *72*, 219–246. [[CrossRef](#)] [[PubMed](#)]
168. Urakawa, H.; Katsuki, A.; Sumida, Y.; Gabazza, E.C.; Murashima, S.; Morioka, K.; Maruyama, N.; Kitagawa, N.; Tanaka, T.; Hori, Y.; et al. Oxidative stress is associated with adiposity and insulin resistance in men. *J. Clin. Endocrinol. Metab.* **2003**, *88*, 4673–4676. [[CrossRef](#)] [[PubMed](#)]
169. Reaven, G.; Tsao, P.S. Insulin resistance and compensatory hyperinsulinemia: The key player between cigarette smoking and cardiovascular disease? *J. Am. Coll. Cardiol.* **2003**, *41*, 1044–1047. [[CrossRef](#)]
170. Jeppesen, J.; Hein, H.O.; Suadicani, P.; Gyntelberg, F. Low triglycerides-high high-density lipoprotein cholesterol and risk of ischemic heart disease. *Arch. Intern. Med.* **2001**, *161*, 361–366. [[CrossRef](#)] [[PubMed](#)]
171. Sijbrands, E.J.; Westendorp, R.G.; Hoffer, M.J.; Havekes, L.M.; Frants, R.R.; Meinders, A.E.; Frolich, M.; Smelt, A.H. Effect of insulin resistance, apoE2 allele, and smoking on combined hyperlipidemia. *Arterioscleros. Thrombos.* **1994**, *14*, 1576–1580. [[CrossRef](#)]
172. Tahtinen, T.M.; Vanhala, M.J.; Oikarinen, J.A.; Keinänen-Kiukaanniemi, S.M. Effect of smoking on the prevalence of insulin resistance-associated cardiovascular risk factors among Finnish men in military service. *J. Cardiovasc. Risk* **1998**, *5*, 319–323. [[CrossRef](#)] [[PubMed](#)]
173. Heitzer, T.; Yla-Herttuala, S.; Luoma, J.; Kurz, S.; Munzel, T.; Just, H.; Olschewski, M.; Drexler, H. Cigarette smoking potentiates endothelial dysfunction of forearm resistance vessels in patients with hypercholesterolemia. Role of oxidized LDL. *Circulation* **1996**, *93*, 1346–1353. [[CrossRef](#)] [[PubMed](#)]
174. Adams, M.R.; Jessup, W.; Celermajer, D.S. Cigarette smoking is associated with increased human monocyte adhesion to endothelial cells: Reversibility with oral L-arginine but not vitamin c. *J. Am. Coll. Cardiol.* **1997**, *29*, 491–497. [[CrossRef](#)]
175. Otsuka, R.; Watanabe, H.; Hirata, K.; Tokai, K.; Muro, T.; Yoshiyama, M.; Takeuchi, K.; Yoshikawa, J. Acute effects of passive smoking on the coronary circulation in healthy young adults. *JAMA* **2001**, *286*, 436–441. [[CrossRef](#)] [[PubMed](#)]
176. Chen, N.G.; Holmes, M.; Reaven, G.M. Relationship between insulin resistance, soluble adhesion molecules, and mononuclear cell binding in healthy volunteers. *J. Clin. Endocrinol. Metab.* **1999**, *84*, 3485–3489. [[CrossRef](#)] [[PubMed](#)]
177. Stuhlinger, M.C.; Abbasi, F.; Chu, J.W.; Lamendola, C.; McLaughlin, T.L.; Cooke, J.P.; Reaven, G.M.; Tsao, P.S. Relationship between insulin resistance and an endogenous nitric oxide synthase inhibitor. *JAMA* **2002**, *287*, 1420–1426. [[CrossRef](#)] [[PubMed](#)]
178. Tippetts, T.S.; Winden, D.R.; Swensen, A.C.; Nelson, M.B.; Thatcher, M.O.; Saito, R.R.; Condie, T.B.; Simmons, K.J.; Judd, A.M.; Reynolds, P.R.; et al. Cigarette smoke increases cardiomyocyte ceramide accumulation and inhibits mitochondrial respiration. *BMC Cardiovasc. Disord.* **2014**, *14*, 165. [[CrossRef](#)] [[PubMed](#)]
179. Barnoya, J.; Glantz, S.A. Cardiovascular effects of secondhand smoke: Nearly as large as smoking. *Circulation* **2005**, *111*, 2684–2698. [[CrossRef](#)] [[PubMed](#)]
180. Faught, B.E.; Flouris, A.D.; Cairney, J. Epidemiological evidence associating secondhand smoke exposure with cardiovascular disease. *Inflamm. Allergy Drug Targets* **2009**, *8*, 321–327. [[CrossRef](#)] [[PubMed](#)]



181. Institute of Medicine (US) Committee on Secondhand Smoke Exposure and Acute Coronary Events. *Secondhand Smoke Exposure and Cardiovascular Effects: Making Sense of the Evidence*; National Academy of Sciences: Washington, DC, USA, 2010.
182. Law, M.R.; Morris, J.K.; Wald, N.J. Environmental tobacco smoke exposure and ischaemic heart disease: An evaluation of the evidence. *BMJ* **1997**, *315*, 973–980. [[CrossRef](#)] [[PubMed](#)]
183. Rubenstein, D.A.; Morton, B.E.; Yin, W. The combined effects of sidestream smoke extracts and glycated serum albumin on endothelial cells and platelets. *Cardiovasc. Diabetol.* **2010**, *9*, 28. [[CrossRef](#)] [[PubMed](#)]
184. Iso, H.; Shimamoto, T.; Sato, S.; Koike, K.; Iida, M.; Komachi, Y. Passive smoking and plasma fibrinogen concentrations. *Am. J. Epidemiol.* **1996**, *144*, 1151–1154. [[CrossRef](#)] [[PubMed](#)]
185. Schmid, P.; Karanikas, G.; Kritz, H.; Pirich, C.; Stamatopoulos, Y.; Peskar, B.A.; Sinzinger, H. Passive smoking and platelet thromboxane. *Thrombos. Res.* **1996**, *81*, 451–460. [[CrossRef](#)]
186. Barua, R.S.; Ambrose, J.A.; Eales-Reynolds, L.J.; deVoe, M.C.; Zervas, J.G.; Saha, D.C. Dysfunctional endothelial nitric oxide biosynthesis in healthy smokers with impaired endothelium-dependent vasodilatation. *Circulation* **2001**, *104*, 1905–1910. [[CrossRef](#)] [[PubMed](#)]
187. Burke, A.; Fitzgerald, G.A. Oxidative stress and smoking-induced vascular injury. *Prog. Cardiovasc. Dis.* **2003**, *46*, 79–90. [[CrossRef](#)]
188. Tribble, D.L.; Giuliano, L.J.; Fortmann, S.P. Reduced plasma ascorbic acid concentrations in nonsmokers regularly exposed to environmental tobacco smoke. *Am. J. Clin. Nutr.* **1993**, *58*, 886–890. [[PubMed](#)]
189. Gotto, A.M., Jr.; Brinton, E.A. Assessing low levels of high-density lipoprotein cholesterol as a risk factor in coronary heart disease: A working group report and update. *J. Am. Coll. Cardiol.* **2004**, *43*, 717–724. [[CrossRef](#)] [[PubMed](#)]
190. DeFaria Yeh, D.; Freeman, M.W.; Meigs, J.B.; Grant, R.W. Risk factors for coronary artery disease in patients with elevated high-density lipoprotein cholesterol. *Am. J. Cardiol.* **2007**, *99*, 1–4. [[CrossRef](#)] [[PubMed](#)]
191. Mack, W.J.; Islam, T.; Lee, Z.; Selzer, R.H.; Hodis, H.N. Environmental tobacco smoke and carotid arterial stiffness. *Prev. Med.* **2003**, *37*, 148–154. [[CrossRef](#)]
192. El-Hakim, I.E.; Elyamani, A.O. Preliminary evaluation of histological changes found in a mechanical arthropatic temporomandibular joint (TMJ) exposed to an intra-articular hyaluronic acid (HA) injection, in a rat model. *J. Cranio Maxill. Surg.* **2011**, *39*, 610–614. [[CrossRef](#)] [[PubMed](#)]
193. Abramson, S.; Krasnokutsky, S. Biomarkers in osteoarthritis. *Bull. NYU Hosp. Jt. Dis.* **2006**, *64*, 77–81. [[PubMed](#)]
194. Xu, L.; Golshirazian, I.; Asbury, B.J.; Li, Y. Induction of high temperature requirement A1, a serine protease, by TGF- $\beta$ 1 in articular chondrocytes of mouse models of oa. *Histol. Histopathol.* **2014**, *29*, 609–618. [[PubMed](#)]
195. Larkin, D.J.; Kartchner, J.Z.; Doxey, A.S.; Hollis, W.R.; Rees, J.L.; Wilhelm, S.K.; Draper, C.S.; Peterson, D.M.; Jackson, G.G.; Ingersoll, C.; et al. Inflammatory markers associated with osteoarthritis after destabilization surgery in young mice with and without receptor for advanced glycation end-products (RAGE). *Front. Physiol.* **2013**, *4*, 121. [[CrossRef](#)] [[PubMed](#)]
196. Holt, D.W.; Henderson, M.L.; Stockdale, C.E.; Farrell, J.T.; Kooyman, D.L.; Bridgewater, L.C.; Seegmiller, R.E. Osteoarthritis-like changes in the heterozygous sedc mouse associated with the HtrA1-Ddr2-Mmp-13 degradative pathway: A new model of osteoarthritis. *Osteoarthr. Cartil.* **2012**, *20*, 430–439. [[CrossRef](#)] [[PubMed](#)]
197. Matias, E.M.; Mecham, D.K.; Black, C.S.; Graf, J.W.; Steel, S.D.; Wilhelm, S.K.; Andersen, K.M.; Mitchell, J.A.; Macdonald, J.R.; Hollis, W.R.; et al. Malocclusion model of temporomandibular joint osteoarthritis in mice with and without receptor for advanced glycation end products. *Arch. Oral Biol.* **2016**, *69*, 47–62. [[CrossRef](#)] [[PubMed](#)]
198. Mankin, H.J.; Dorfman, H.; Lippiello, L.; Zarins, A. Biochemical and metabolic abnormalities in articular cartilage from osteo-arthritic human hips. II. Correlation of morphology with biochemical and metabolic data. *J. Bone Jt. Surg. Am.* **1971**, *53*, 523–537. [[CrossRef](#)]
199. Mankin, H.J. The reaction of articular cartilage to injury and osteoarthritis (first of two parts). *N. Engl. J. Med.* **1974**, *291*, 1285–1292. [[CrossRef](#)] [[PubMed](#)]
200. Glasson, S.S.; Chambers, M.G.; Van Den Berg, W.B.; Little, C.B. The oars histopathology initiative—Recommendations for histological assessments of osteoarthritis in the mouse. *Osteoarthr. Cartil.* **2010**, *18* (Suppl. S3), S17–S23. [[CrossRef](#)] [[PubMed](#)]



201. Polur, I.; Lee, P.L.; Servais, J.M.; Xu, L.; Li, Y. Role of HTRA1, a serine protease, in the progression of articular cartilage degeneration. *Histol. Histopathol.* **2010**, *25*, 599–608. [[PubMed](#)]
202. Felson, D.T.; Anderson, J.J.; Naimark, A.; Hannan, M.T.; Kannel, W.B.; Meenan, R.F. Does smoking protect against Osteo-arthritis. *Arthritis Rheum.* **1989**, *32*, 166–172. [[CrossRef](#)] [[PubMed](#)]
203. Dube, C.E.; Liu, S.H.; Driban, J.B.; McAlindon, T.E.; Eaton, C.B.; Lapane, K.L. The relationship between smoking and knee osteoarthritis in the osteoarthritis initiative. *Osteoarthr. Cartil.* **2016**, *24*, 465–472. [[CrossRef](#)] [[PubMed](#)]
204. Mnataganian, G.; Ryan, P.; Norman, P.; Davidson, D.; Hiller, J. Smoking, body weight, physical exercise and risk of lower limb total joint replacement in a population-based cohort of men. *J. Epidemiol. Commun. Health* **2011**, *65*, 2523–2530. [[CrossRef](#)] [[PubMed](#)]
205. Harrison, B.J.; Silman, A.J.; Wiles, N.J.; Scott, D.G.; Symmons, D.P. The association of cigarette smoking with disease outcome in patients with early inflammatory polyarthritis. *Arthritis Rheum.* **2001**, *44*, 323–330. [[CrossRef](#)]
206. Kang, K.; Shin, J.S.; Lee, J.; Lee, Y.J.; Kim, M.R.; Park, K.B.; Ha, I.H. Association between direct and indirect smoking and osteoarthritis prevalence in Koreans: A cross-sectional study. *BMJ Open* **2016**, *6*, e010062. [[CrossRef](#)] [[PubMed](#)]
207. Ding, C.; Cicuttini, F.; Blizzard, L.; Jones, G. Smoking interacts with family history with regard to change in knee cartilage volume and cartilage defect development. *Arthritis Rheum.* **2007**, *56*, 1521–1528. [[CrossRef](#)] [[PubMed](#)]
208. Villaverde-Garcia, V.; Cobo-Ibanez, T.; Candelas-Rodriguez, G.; Seoane-Mato, D.; Campo-Fontecha, P.D.; Guerra, M.; Munoz-Fernandez, S.; Canete, J.D. The effect of smoking on clinical and structural damage in patients with axial spondyloarthritis: A systematic literature review. *Semin. Arthritis Rheum.* **2016**. [[CrossRef](#)] [[PubMed](#)]
209. Winden, D.R.; Barton, D.B.; Betteridge, B.C.; Bodine, J.S.; Jones, C.M.; Rogers, G.D.; Chavarria, M.; Wright, A.J.; Jergensen, Z.R.; Jimenez, F.R.; et al. Antenatal exposure of maternal secondhand smoke (SHS) increases fetal lung expression of rage and induces rage-mediated pulmonary inflammation. *Respir. Res.* **2014**, *15*, 129. [[CrossRef](#)] [[PubMed](#)]
210. Wood, T.T.; Winden, D.R.; Marlor, D.R.; Wright, A.J.; Jones, C.M.; Chavarria, M.; Rogers, G.D.; Reynolds, P.R. Acute secondhand smoke-induced pulmonary inflammation is diminished in rage knockout mice. *Am. J. Physiol. Lung Cell. Mol. Physiol.* **2014**, *307*, L758–L764. [[CrossRef](#)] [[PubMed](#)]
211. Robinson, A.B.; Johnson, K.D.; Bennion, B.G.; Reynolds, P.R. Rage signaling by alveolar macrophages influences tobacco smoke-induced inflammation. *Am. J. Physiol. Lung Cell. Mol. Physiol.* **2012**, *302*, L1192–L1199. [[CrossRef](#)] [[PubMed](#)]
212. Reynolds, P.R.; Kasteler, S.D.; Schmitt, R.E.; Hoidal, J.R. Receptor for advanced glycation end-products signals through Ras during tobacco smoke-induced pulmonary inflammation. *Am. J. Respir. Cell Mol. Biol.* **2011**, *45*, 411–418. [[CrossRef](#)] [[PubMed](#)]
213. Bodine, B.G.; Bennion, B.G.; Leatham, E.; Jimenez, F.R.; Wright, A.J.; Jergensen, Z.R.; Erickson, C.J.; Jones, C.M.; Johnson, J.P.; Knapp, S.M.; et al. Conditionally induced RAGE expression by proximal airway epithelial cells in transgenic mice causes lung inflammation. *Respir. Res.* **2014**, *15*, 133. [[CrossRef](#)] [[PubMed](#)]
214. Winden, D.R.; Ferguson, N.T.; Bukey, B.R.; Geyer, A.J.; Wright, A.J.; Jergensen, Z.R.; Robinson, A.B.; Stogsdill, J.A.; Reynolds, P.R. Conditional over-expression of rage by embryonic alveolar epithelium compromises the respiratory membrane and impairs endothelial cell differentiation. *Respir. Res.* **2013**, *14*, 108. [[CrossRef](#)] [[PubMed](#)]
215. Stogsdill, M.P.; Stogsdill, J.A.; Bodine, B.G.; Fredrickson, A.C.; Sefcik, T.L.; Wood, T.T.; Kasteler, S.D.; Reynolds, P.R. Conditional overexpression of receptors for advanced glycation end-products in the adult murine lung causes airspace enlargement and induces inflammation. *Am. J. Respir. Cell Mol. Biol.* **2013**, *49*, 128–134. [[CrossRef](#)] [[PubMed](#)]
216. Stogsdill, J.A.; Stogsdill, M.P.; Porter, J.L.; Hancock, J.M.; Robinson, A.B.; Reynolds, P.R. Embryonic overexpression of receptors for advanced glycation end-products by alveolar epithelium induces an imbalance between proliferation and apoptosis. *Am. J. Respir. Cell Mol. Biol.* **2012**, *47*, 60–66. [[CrossRef](#)] [[PubMed](#)]

217. Reynolds, P.R.; Stogsdill, J.A.; Stogsdill, M.P.; Heimann, N.B. Up-regulation of receptors for advanced glycation end-products by alveolar epithelium influences cytodifferentiation and causes severe lung hypoplasia. *Am. J. Respir. Cell Mol. Biol.* **2011**, *45*, 1195–1202. [[CrossRef](#)] [[PubMed](#)]
218. Reddy, M.A.; Li, S.L.; Sahar, S.; Kim, Y.S.; Xu, Z.G.; Lanting, L.; Natarajan, R. Key role of src kinase in S100B-induced activation of the receptor for advanced glycation end products in vascular smooth muscle cells. *J. Biol. Chem.* **2006**, *281*, 13685–13693. [[CrossRef](#)] [[PubMed](#)]
219. Cai, C.; Dai, X.; Zhu, Y.; Lian, M.; Xiao, F.; Dong, F.; Zhang, Q.; Huang, Y.; Zheng, Q. A specific rage-binding peptide biopanning from phage display random peptide library that ameliorates symptoms in amyloid  $\beta$  peptide-mediated neuronal disorder. *Appl. Microbiol. Biotechnol.* **2016**, *100*, 825–835. [[CrossRef](#)] [[PubMed](#)]
220. Huang, W.; Zhao, H.; Dong, H.; Wu, Y.; Yao, L.; Zou, F.; Cai, S. High-mobility group box 1 impairs airway epithelial barrier function through the activation of the RAGE/ERK pathway. *Int. J. Mol. Med.* **2016**, *37*, 1189–1198. [[PubMed](#)]
221. Khodeer, D.M.; Zaitone, S.A.; Farag, N.E.; Moustafa, Y.M. Cardioprotective effect of pioglitazone in diabetic and non-diabetic rats subjected to acute myocardial infarction involves suppression of AGE-RAGE axis and inhibition of apoptosis. *Can. J. Physiol. Pharmacol.* **2016**, *94*, 463–476. [[CrossRef](#)] [[PubMed](#)]
222. Kim, V.; Rogers, T.J.; Criner, G.J. New concepts in the pathobiology of chronic obstructive pulmonary disease. *Proc. Am. Thoracic Soc.* **2008**, *5*, 478–485. [[CrossRef](#)] [[PubMed](#)]
223. Toure, F.; Zahm, J.M.; Garnotel, R.; Lambert, E.; Bonnet, N.; Schmidt, A.M.; Vitry, F.; Chanard, J.; Gillery, P.; Rieu, P. Receptor for advanced glycation end-products (RAGE) modulates neutrophil adhesion and migration on glycooxidated extracellular matrix. *Biochem. J.* **2008**, *416*, 255–261. [[CrossRef](#)] [[PubMed](#)]
224. Hudson, B.I.; Kalea, A.Z.; Del Mar Arriero, M.; Harja, E.; Boulanger, E.; D'Agati, V.; Schmidt, A.M. Interaction of the rage cytoplasmic domain with diaphanous-1 is required for ligand-stimulated cellular migration through activation of Rac1 and Cdc42. *J. Biol. Chem.* **2008**, *283*, 34457–34468. [[CrossRef](#)] [[PubMed](#)]
225. Schmidt, A.M.; Yan, S.D.; Yan, S.F.; Stern, D.M. The multiligand receptor rage as a progression factor amplifying immune and inflammatory responses. *J. Clin. Investig.* **2001**, *108*, 949–955. [[CrossRef](#)] [[PubMed](#)]
226. Bianchi, R.; Giambanco, I.; Donato, R. S100B/RAGE-dependent activation of microglia via NF- $\kappa$ B and AP-1 Co-regulation of COX-2 expression by S100B, IL-1 $\beta$  and TNF- $\alpha$ . *Neurobiol. Aging* **2010**, *31*, 665–677. [[CrossRef](#)] [[PubMed](#)]
227. Lin, L.; Park, S.; Lakatta, E.G. Rage signaling in inflammation and arterial aging. *Front. Biosci.* **2009**, *14*, 1403–1413. [[CrossRef](#)]



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