

Supplementary Figure S1: Change in gene expression of heparan sulfate proteoglycan (HSPG) core proteins in undifferentiated hMSC cultures following SDC-1 knockdown (KD). Gene expression analysis was performed with RNA collected from hMSC incubated with SDC-1 specific siRNA for 72 h. (A) Syndecans-1-4 (SDC-1-4) core protein expression following SDC-1 KD resulted in a 20% reduction in SDC-1 expression. (B) Glypicans-1-4, and -6 (GPC-1-4, -6) core protein expression following SDC-1 KD. Significant differences in expression levels were detected by Student's T test, significance was set at *p<0.05. Error bars represent SEM.



YU et al. - Supplementary Figure S2

Supplementary Figure S2: Gene expression level changes in heparan sulfate biosynthesis and modification enzymes of CTRL and SDC-1 KD undifferentiated hMSC cultures. Gene expression analysis was performed with RNA collected from hMSC incubated with SDC-1 specific siRNA for 72 h. Gene expression levels of (A) exostosins 1-2 (EXT1-2) and C-5 epimerase (C5-EP), (B) *N*-deacetylase/*N*-sulfotransferases 1-2 (NDST1-2), (C) heparan sulfate 2-*O*-sulfotransferase 1 (HS2ST1) and heparan sulfate 6-*O*-

sulfotransferase 1 (HS6ST1), and **(D)** sulfatases 1-2 (SULF1-2) and heparanase (HPSE). Significant differences in expression levels were detected by Student's T test, significance was set at *P<0.05 and ***P<0.001. Error bars represent SEM.



YU et al. - Supplementary Figure S3

Supplementary Figure S3: Examination of osteogenic and adipogenic lineage marker expression changes in undifferentiated hMSC cultures following SDC-1 KD. Expression analysis was performed with RNA and protein collected from hMSC incubated with SDC-1 specific siRNA for 72 h and 96 h, respectively. (A) Gene expression changes of osteogenic markers runt-related transcription factor 2 (RUNX2), collagen, type I, alpha 1 (COL1A1), osteocalcin (OCN) and alkaline phosphatase (AP). (B) Western analysis of the osteogenic marker COL1A1 in undifferentiated CTRL and SDC-1 KD cultures, with GAPDH

as the loading control. Signal intensity was quantified using ImageJ software (NIH), error bar = SD. (C) Gene expression changes in adipogenic markers adiponectin (ADIPO-Q), CCAAT/enhancer binding protein alpha (C/EBP α), CCAAT/enhancer binding protein delta (C/EBP δ) and peroxisome proliferator-activated receptor gamma 1 (PPAR γ 1). Significant changes in gene expression was detected by Student's T test, significance level was set at *p<0.05. Error bars = SEM.



YU et al. - Supplementary Figure S4

Supplementary Figure S4: Gene expression changes in common signalling pathways implicated in osteogenic and adipogenic lineages. Gene expression analysis was performed with RNA collected from hMSC incubated with SDC-1 specific siRNA for 72 h. Changes in gene expression in undifferentiated hMSC CTRL and SDC-1 KD cultures

detected by Q-PCR of (A) Bone morphogenetic protein (BMP) signalling including the ligands BMP2 and BMP4, and BMP receptors IA (BMPR-IA) and IB (BMPR-IB). (B) Fibroblast growth factor (FGF) signalling with ligand FGF2 and the receptors FGFR1 and FGFR2. (C) Wnt signalling including Wnt3a ligand and the receptor frizzled (FZD1). (D) Sonic hedgehog signalling (SHH) including the ligand SHH, the canonical receptor Patched (PTCH1) and the G protein-coupled receptor-like receptor Smoothened (SMO). Error bars represent SEM.





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Supplementary Figure S5: Replicates of hMSC P+7 SDC-1 KD_{AD/OS} **osteogenic and adipogenic stained cultures.** Terminal differentiated (22 days) hMSC SDC-1 KD_{AD} adipogenic cultures stained with Oil Red O, and the terminal differentiated (21 days) hMSC SDC-1 KD_{OS} osteogenic cultures stained with Alizarin Red and von Kossa. Two hMSC populations were used for SDC-1 KD differentiation experiments (hMSC-1 and hMSC-2). Differentiation of each hMSC population was performed in duplicate (#1 and #2) for each condition: Untreated (UT_{AD/OS}), Scrambled (SCR_{AD/OS}) and SDC-1 knockdown (SDC-1 KD_{AD/OS}).

YU et al. - Supplementary Figure S6



C.



von Kossa



Supplementary Figure S6: Comparison of calcification and mineralisation between Untreated (UT), Scrambled (SCR) and SDC-1 knockdown (SDC-1 KD) hMSC post-KD osteogenic differentiated cultures by von Kossa and Alizarin Red staining, respectively. SDC-1 KD was performed in basal hMSC cultures, then these cultures were differentiated towards the osteogenic lineage, where terminally differentiated cultures (21 days) were examined by staining. Staining of hMSC SDC-1 KD osteogenic cultures by (**A**) Alizarin Red (red) and von Kossa stain (brown-black) with nuclear fast red-aluminium sulfate counter-stain (pink). Quantitation of (**B**) Alizarin Red (+49%) staining, and (**C**) von Kossa (+57%) in hMSC osteogenic SDC-1 KD cultures. Stained culture images were converted to 8-bit greyscale images and quantitated using ImageJ software (NIH). Optical density of CTRL cultures (UT and SCR averaged) and SDC-1 KD cultures was normalised to culture cell number recorded following SDC-1 KD, CTRL = 7.0 x 10^4 cells and SDC-1 KD = 5.0 x 10^4 cells (Fig. 1A).