



## Supplementary information

# Human Serum Albumin in the presence of AGuIX nanoagents: structure stabilisation without direct interaction

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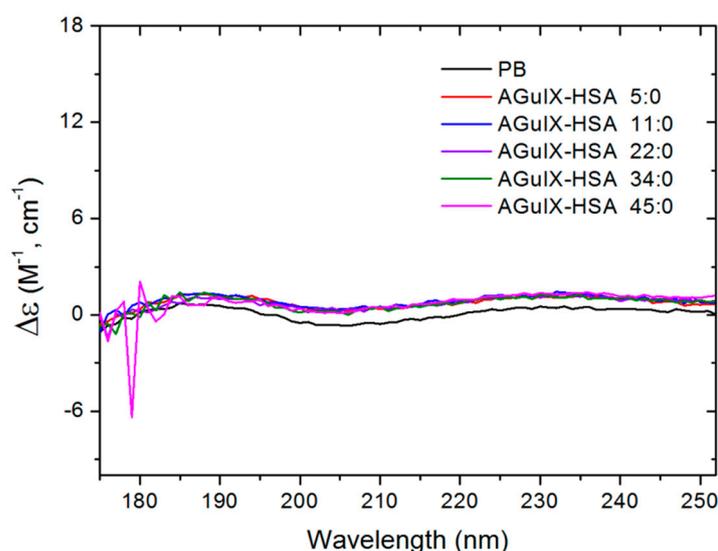
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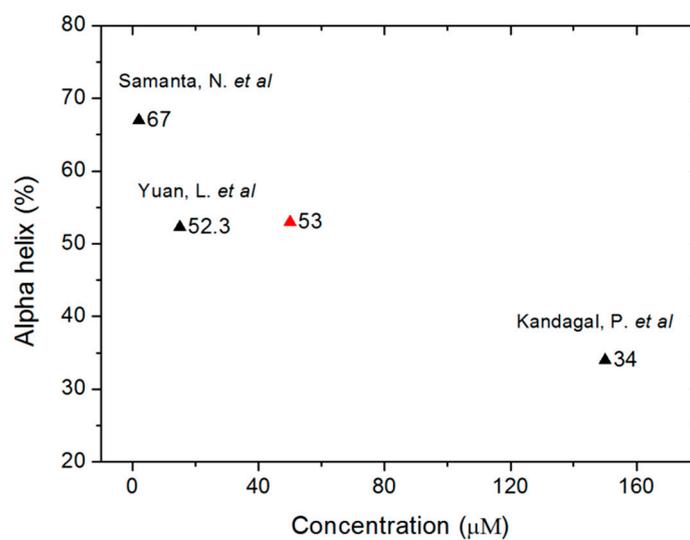
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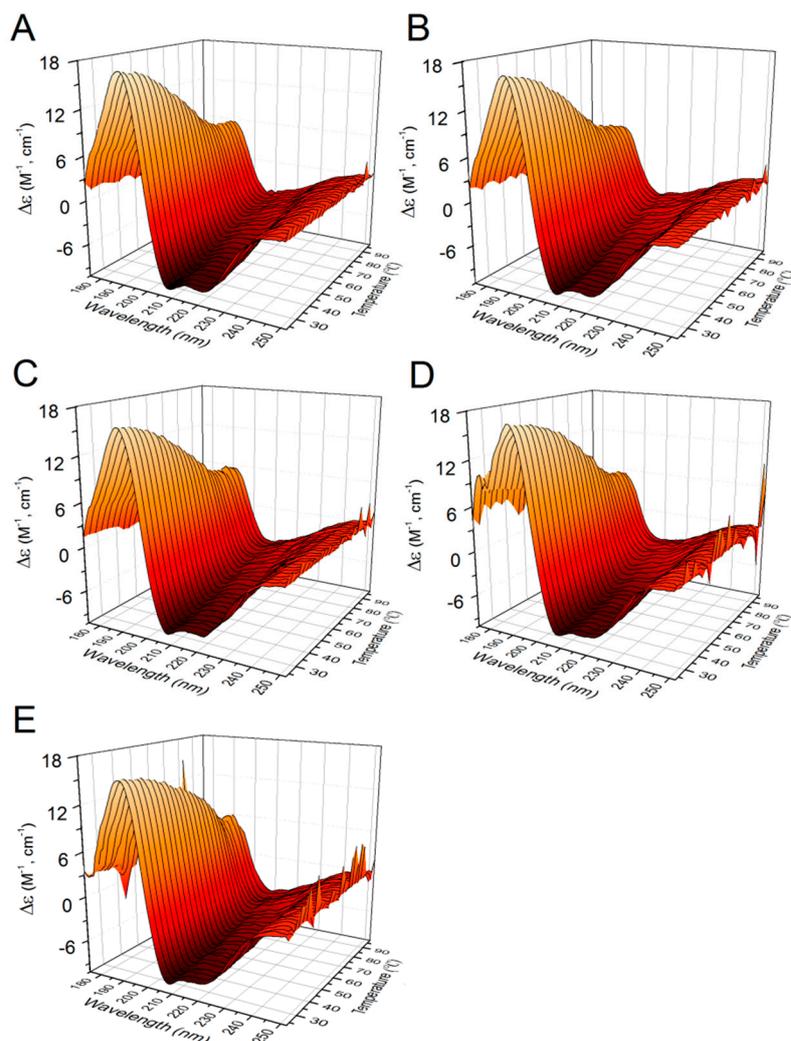
**Figure S1.** SRCD spectra of control and AGuIX® with increasing concentrations that correspond to ratios from 5:0 to 45:0 in 10 mM phosphate buffer at pH 7.4 and 37°C.

**Table S1.** Relative content (%) of the secondary structures (alpha helix, turn, random coil) in pure HSA (3100  $\mu\text{g/mL}$ ) and in HSA mixed with AGuIX® at various concentrations, pH 7.4 and 37°C (beta sheet 0%).

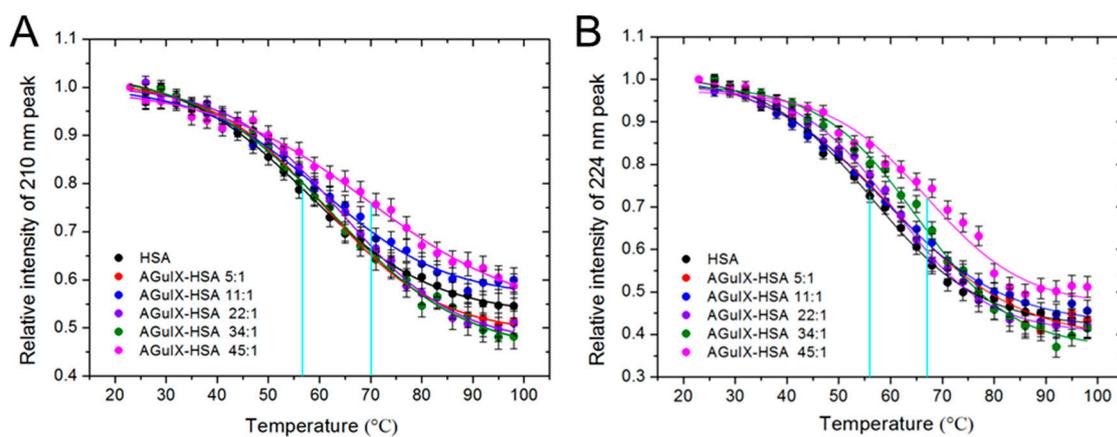
Sample	% of alpha helix	% of turn	% of random coil
HSA	53 $\pm$ 2	14 $\pm$ 1	33 $\pm$ 2
AGuIX®-HSA 5:1	53 $\pm$ 2	14 $\pm$ 1	33 $\pm$ 2
AGuIX®-HSA 11:1	52 $\pm$ 2	15 $\pm$ 1	33 $\pm$ 2
AGuIX®-HSA 22:1	53 $\pm$ 2	15 $\pm$ 2	32 $\pm$ 2
AGuIX®-HSA 34:1	64 $\pm$ 3	16 $\pm$ 2	20 $\pm$ 2
AGuIX®-HSA 45:1	71 $\pm$ 3	7 $\pm$ 1	22 $\pm$ 2



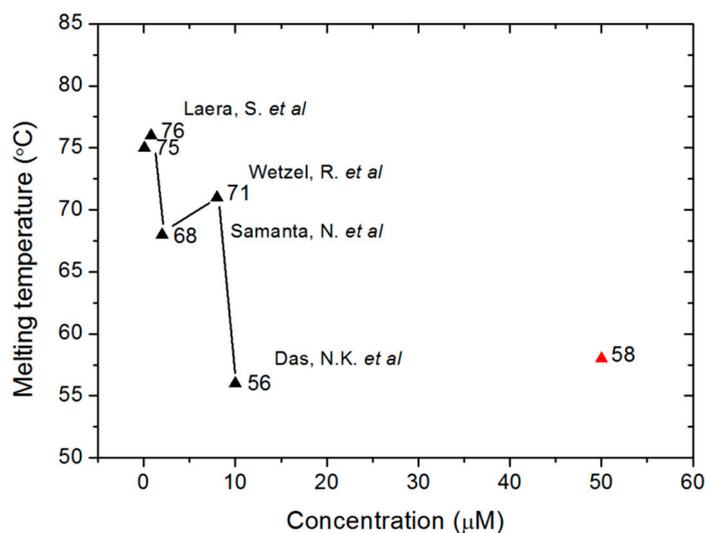
**Figure 2.** Comparison of the content of alpha helix (%) obtained by our group (red) with the data reported by Yuan, L. et al, Samanta, N. et al and Kandagal, P. et al (black).



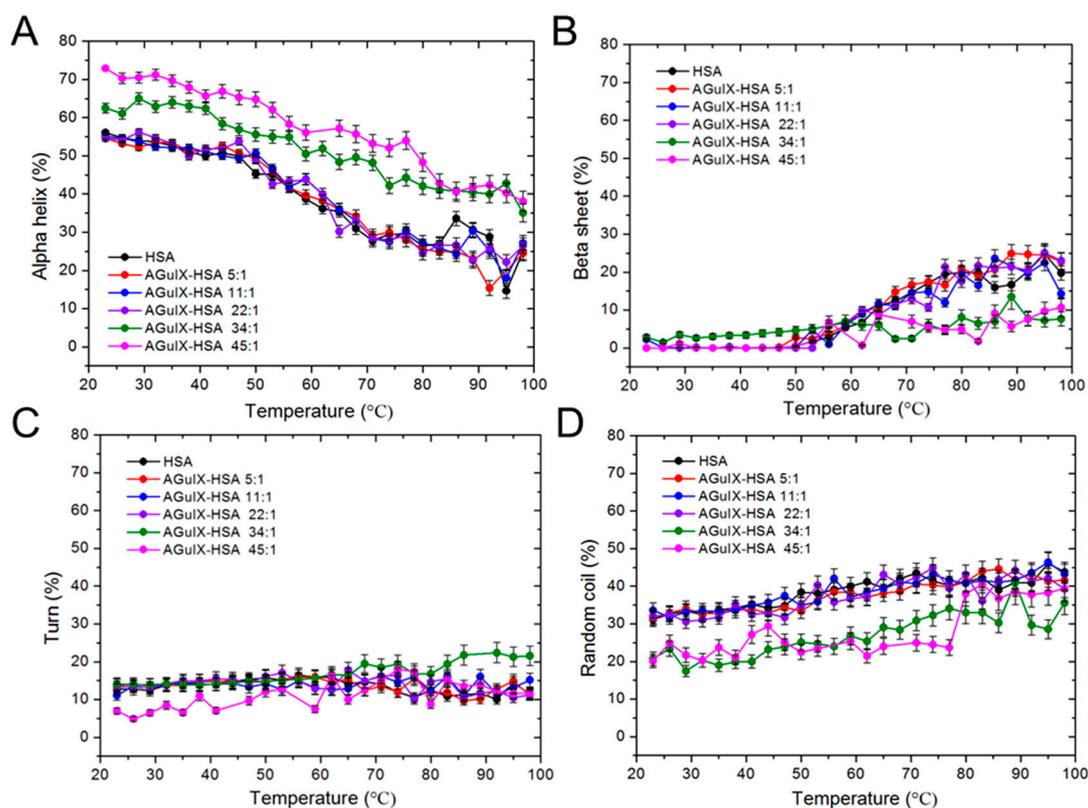
**Figure S3.** A-E. Temperature dependent SRCD spectra of HSA in 10 mM phosphate buffer at pH 7.4 recorded without (A) and in the presence of AGuIX<sup>®</sup> at the following AGuIX<sup>®</sup>:HSA ratios: 5:1 (B), 11:1 (C), 34:1 (D) and 45:1 (E).



**Figure S1.** A,B. Relative intensity of the peak at 210 nm and 224 nm for the pure HSA and HSA in the presence of AGuIX<sup>®</sup> at different ratios. The melting temperatures are determined at half maximum.



**Figure S5.** Comparison of melting temperature of pure HSA measured by our group (red) with data reported by different groups (black).



**Figure S6.** Relative content of alpha-helix (A), beta sheet (B), turn (C), and random coil (D) as function of the temperature for pure HSA and HSA mixed with AGuIX® at different concentrations.

### Thermodynamic parameter

The quantitative analysis of protein unfolding is almost exclusively based on the two-state model. It assumes that a protein in solution adopts only two conformational states, the folded (F) and unfolded (U) state. The equilibrium  $F \leftrightarrow U$  is described with a temperature-dependent equilibrium constant  $K$ :

$$K=[F]/[U]$$

[F] and [U] are the concentration of the folded and unfolded forms respectively. The fraction folded at any temperature is  $\alpha$ .

$$\alpha = [F]/([F]+[U])$$

$$\alpha = K/(1+K)$$

$$\alpha = (\theta t - \theta U)/(\theta F - \theta U)$$

$\theta t$  is the observed ellipticity at any temperature,  $\theta F$  is the ellipticity of the fully folded form and  $\theta U$  is the ellipticity of the unfolded form.

The Van't Hoff equation was created to relate equilibrium constant (K) to temperature by substituting the two free energy equations

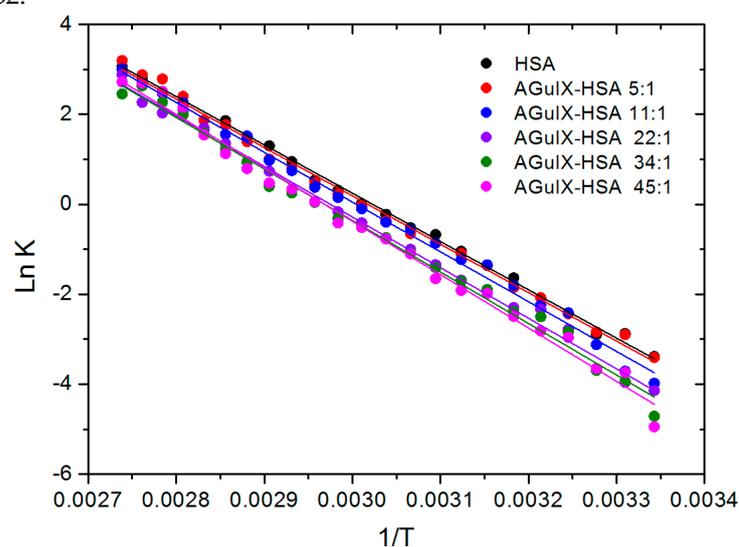
$$\Delta G^\circ = -RT \ln K \text{ (at equilibrium, } \Delta G=0)$$

$$\Delta G^\circ = \Delta H^\circ - T\Delta S^\circ$$

and rearranging to generate the Van't Hoff equation

$$\ln K = \frac{-\Delta H^\circ}{RT} + \frac{\Delta S^\circ}{R}$$

Where the natural logarithm (ln) of the folding constant, K is plotted as a function of  $1/T$ , T is the absolute temperature (Kelvin) and R is the universal gas constant =  $8.314 \text{ J}\cdot\text{mol}^{-1}\cdot\text{K}^{-1}$ . From the graph analysis, we obtain  $\Delta H^\circ$  (from the slope of the plot  $-\Delta H^\circ/R$ ) and  $\Delta S^\circ$  (from the intercept of the plot  $\Delta S^\circ/R$ ). The values are reported in the table S2.



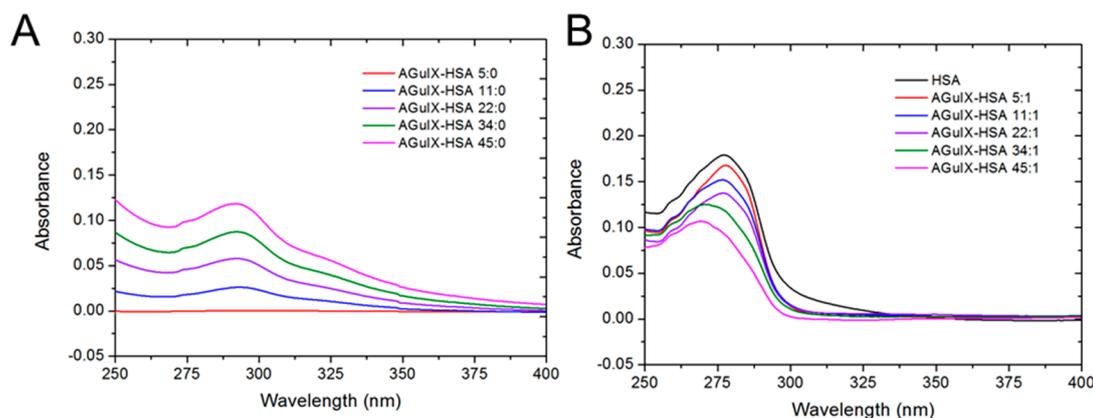
**Figure S2.** Van't Hoff plot for the calculation of thermodynamic parameters of the pure HSA and in the presence of AGuIX® at ratios from 5:1 to 45:1 (at 192 nm).

**Table S1.** Values of the slope, intercept and  $R^2$  of the Van't Hoff plot of HSA and AGuIX®-HSA with different AGuIX® concentrations (from the SRCD measurements at 192 nm).

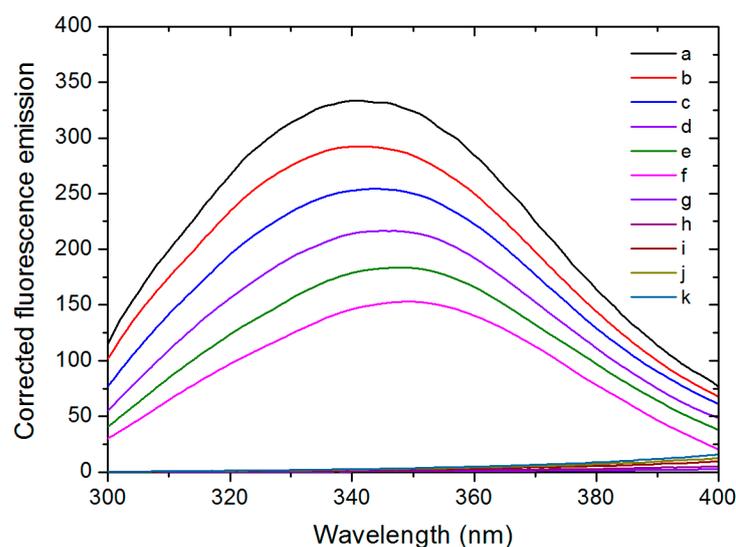
Sample	Slope	Intercept	$R^2$
HSA	-10742.1±130.8	32.475±0.399	0.997
AGuIX®-HSA 5:1	-10759.8±167.7	32.476±0.407	0.995
AGuIX®-HSA 11:1	-11071.3±202.6	33.265±0.406	0.994
AGuIX®-HSA 22:1	-11253.0±183.0	33.477±0.414	0.994
AGuIX®-HSA 34:1	-11455.6±224.0	34.008±0.418	0.993
AGuIX®-HSA 45:1	-11875.1±254.9	35.251±0.420	0.991

### UV-vis absorption spectra

A CARY 300 spectrophotometer (Int. Agilent) was used to scan the UV-visible spectra. For the inner filter effect correction of the fluorescence quenching data, the UV-visible absorption spectra of HSA (4.7  $\mu\text{M}$ ) in the absence and presence of AGuIX<sup>®</sup> at different ratios (5:1 to 45:1) in a total volume of 1 mL were recorded at the 250–400 nm wavelength range.



**Figure S3.** (A) UV-vis absorption spectra of free AGuIX<sup>®</sup> at different concentrations from 5:0 to 45:0 in 67mM phosphate buffer, pH 7.4. (B) UV-vis absorption spectra of HSA without and with increasing AGuIX<sup>®</sup> concentrations (AGuIX<sup>®</sup>:HSA ratios from 5:1 to 45:1) in 67mM phosphate buffer, pH 7.4. The spectra were obtained after subtracting the absorption spectra of AGuIX<sup>®</sup> from the respective absorption spectra of AGuIX<sup>®</sup>-HSA samples.



**Figure S4.** Corrected fluorescence emission spectra of HSA without (a) and with increasing AGuIX<sup>®</sup> concentrations from 5:1 to 45:1 (b-f), AGuIX<sup>®</sup> alone at corresponding concentration (g-k) in 67mM phosphate buffer, pH 7.4 at 37°C upon excitation at 280 nm.

**Table S3.** The Stern-Volmer constant ( $K_{sv}$ ,  $\text{M}^{-1}$ ) and the quenching rate constant ( $k_q$  in  $\text{M}^{-1}\cdot\text{s}^{-1}$ ) characteristics of the AGuIX<sup>®</sup>-HSA interactions, at 25°C and 37°C, pH 7.4.

T/°C	$K_{sv}$ , $\text{M}^{-1}$	$k_q$ , $\text{M}^{-1}\cdot\text{s}^{-1}$
25	$5.2 \times 10^3$	$8.15 \times 10^{11}$
37	$5.9 \times 10^3$	$9.25 \times 10^{11}$

