

Supplemental Material

Single cell transcriptome analysis of Niemann-Pick disease, type C1 cerebella

Cougnoux et al.

Supplemental figures and tables

	<u>Page</u>
Figure S1 t-SNE plots 7-week data	1
Figure S2 Microglia	2
Figure S3 Vascular cells	3
Figure S4 Glial cells	4
Figure S5 Purkinje neurons	6
Figure S6 Protein validation	7
Figure S7 T-SNE plots 3-week data	8
Table S1 Differentially expressed genes	9
Table S2 Pathway Analysis	See Excel file
Table S3 Antibodies, primers and reagents	18

Figure S1

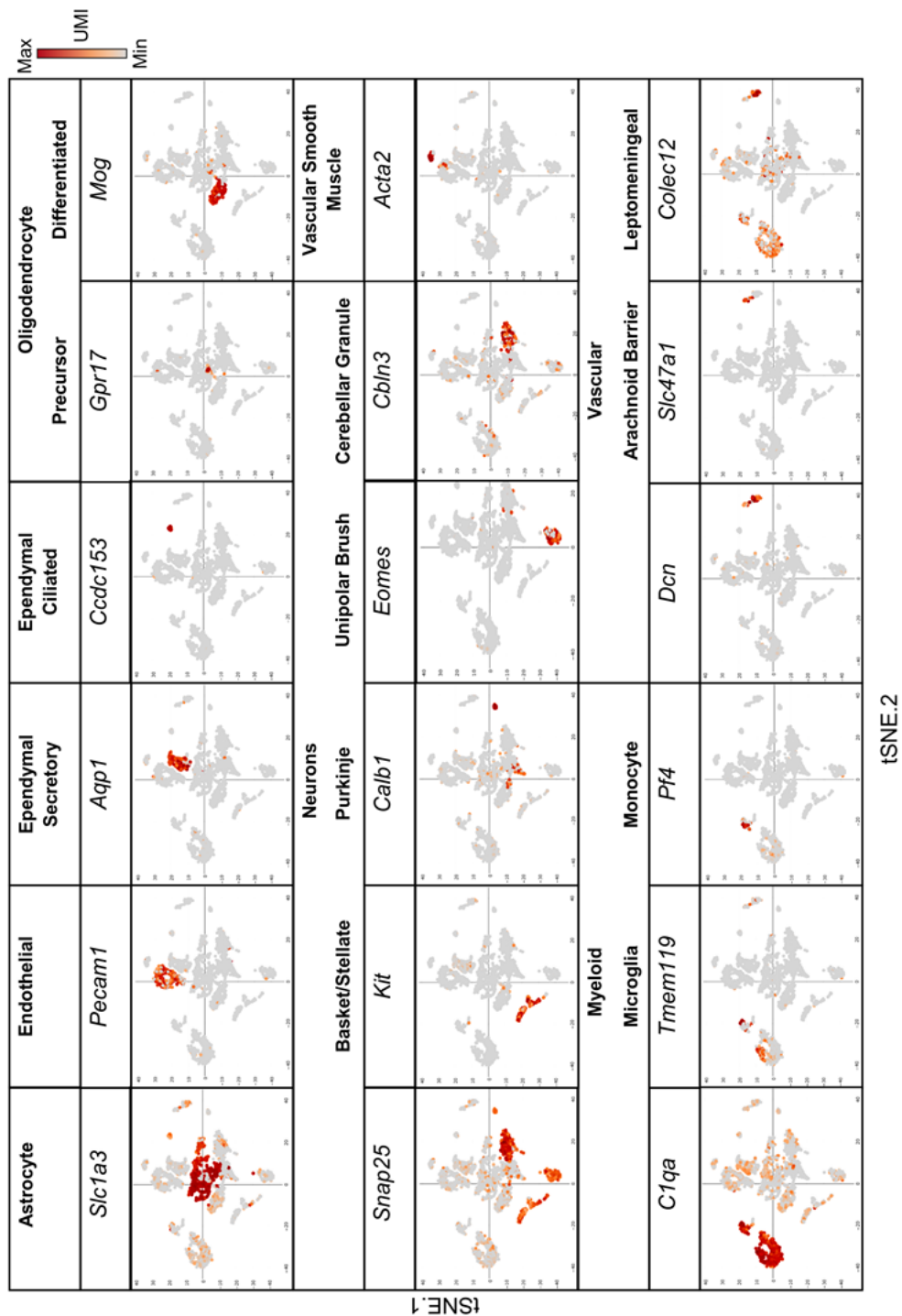


Figure S1. t-SNE plots of the different cell types identified in cerebellar tissue from 7-week-old *Npc1*^{+/+} and *Npc1*^{-/-} mice. UMI: Unique Molecular Identifier; Max: maximum; Min: minimum.

Figure S2

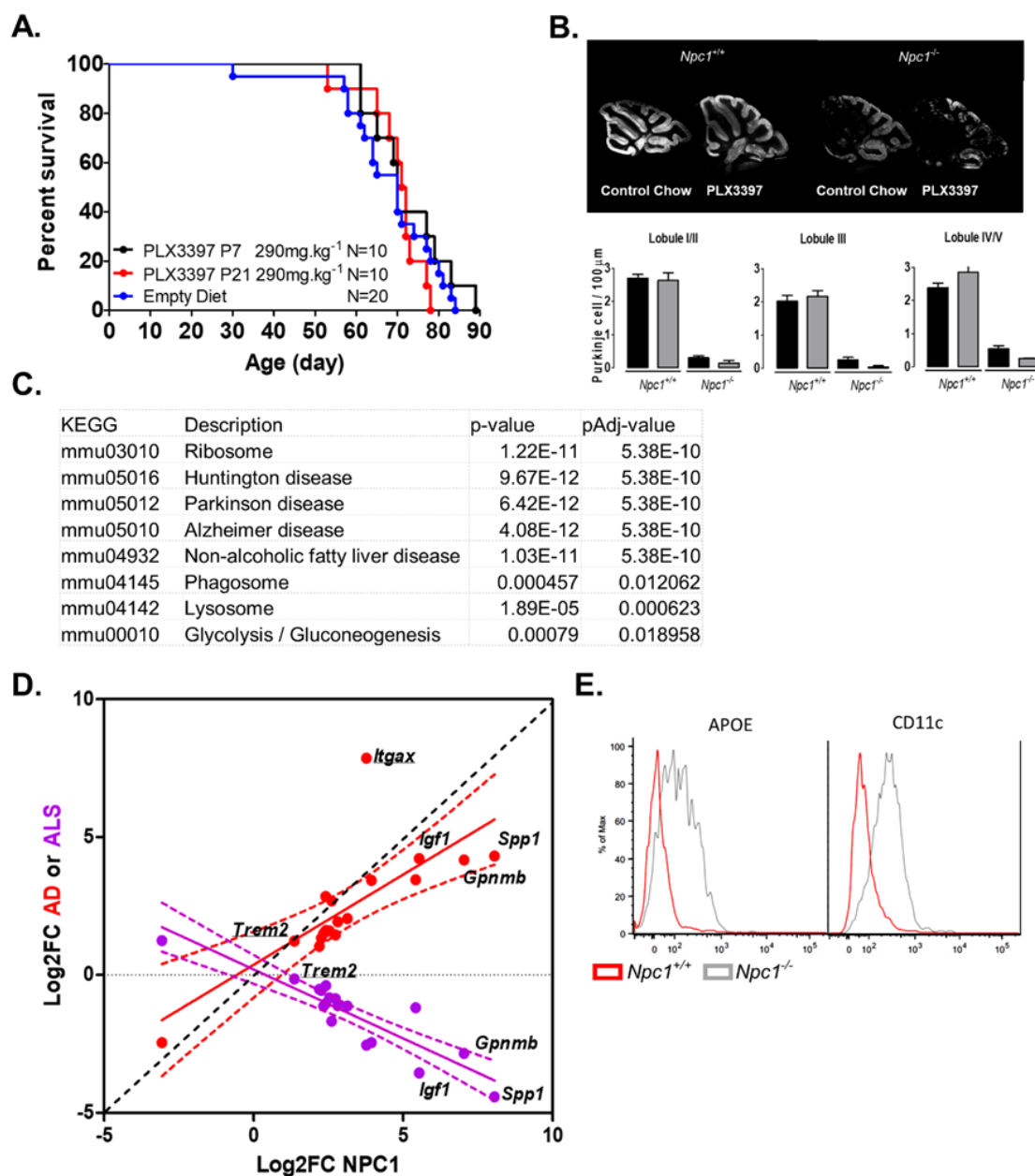


Figure S2. (A) Kaplan-Meier survival graphic of 290mg.kg⁻¹ PLX3397 treatment starting at day of life 7 (Black) 21 (Red) and control diet (Blue). (B) Purkinje neuron pathology analysis at 7 weeks of age of the mice starting at day of life 21. (C) KEGG pathway analysis demonstrated similarity between altered gene expression in *Npc1*^{-/-} microglia and microglia from mouse models of common neurodegenerative diseases. (D) Correlation of Disease Associated Microglia (DAM) profile between *Npc1*^{-/-} and either Alzheimer disease (Red) or Amyotrophic Lateral Sclerosis (Purple) mice. Data for Alzheimer and Amyotrophic Lateral Sclerosis are from Keren-Shaul et al. [36]. (E) Flow cytometry validation of increased expression of DAM microglia markers APOE and CD11C overexpression in *Npc1*^{-/-} (gray) compared to *Npc1*^{+/+} (red) 7-week-old mice cerebellar microglia.

Figure S3

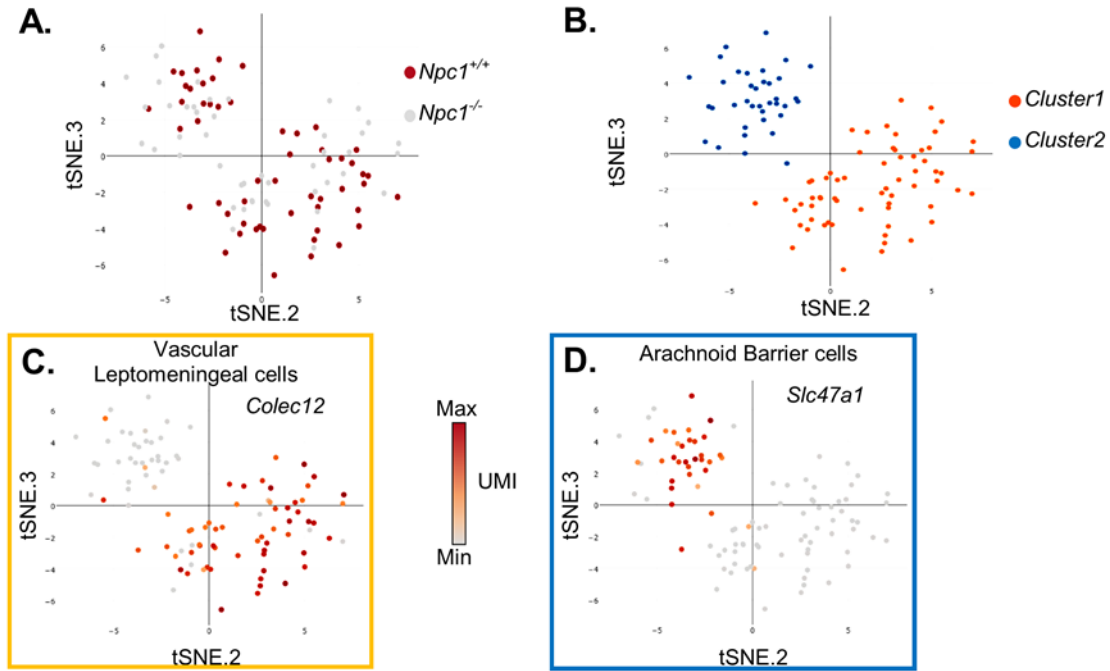


Figure S3. (A) t-SNE plot of cerebellar vascular cells that were not identified as vascular smooth muscle or endothelial cells from *Npc1*^{+/+} (red) and *Npc1*^{-/-} (gray) 7-week-old mice. (B) These cells formed two clusters which could be identified as arachnoid barrier cells (cluster 1, orange) and vascular leptomeningeal cells (cluster 2, blue) based upon expression of *Colec12* (C) in vascular leptomeningeal cell and *Slc47a1* (D) in arachnoid barrier cells.

Figure S4

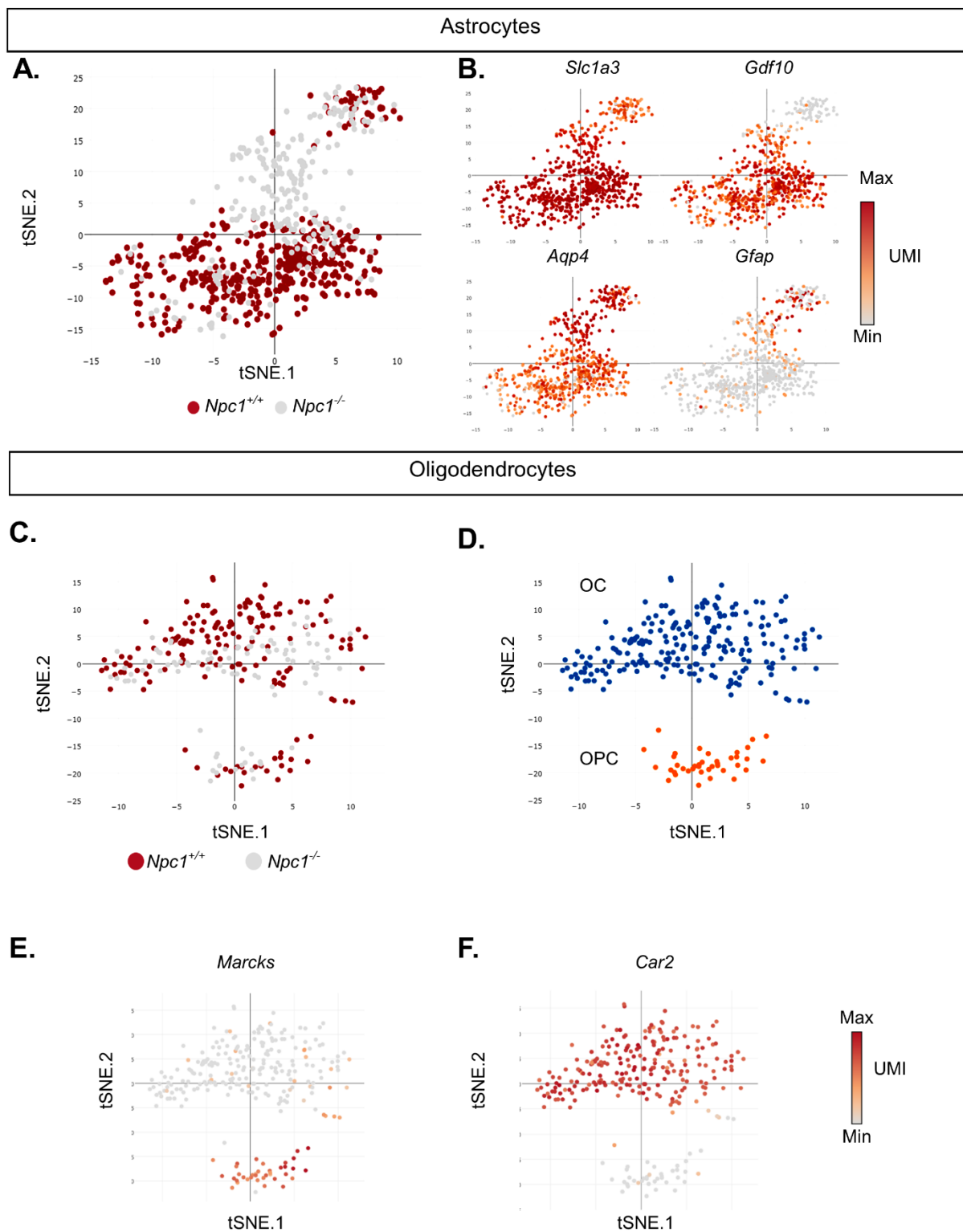


Figure S4 (continued)

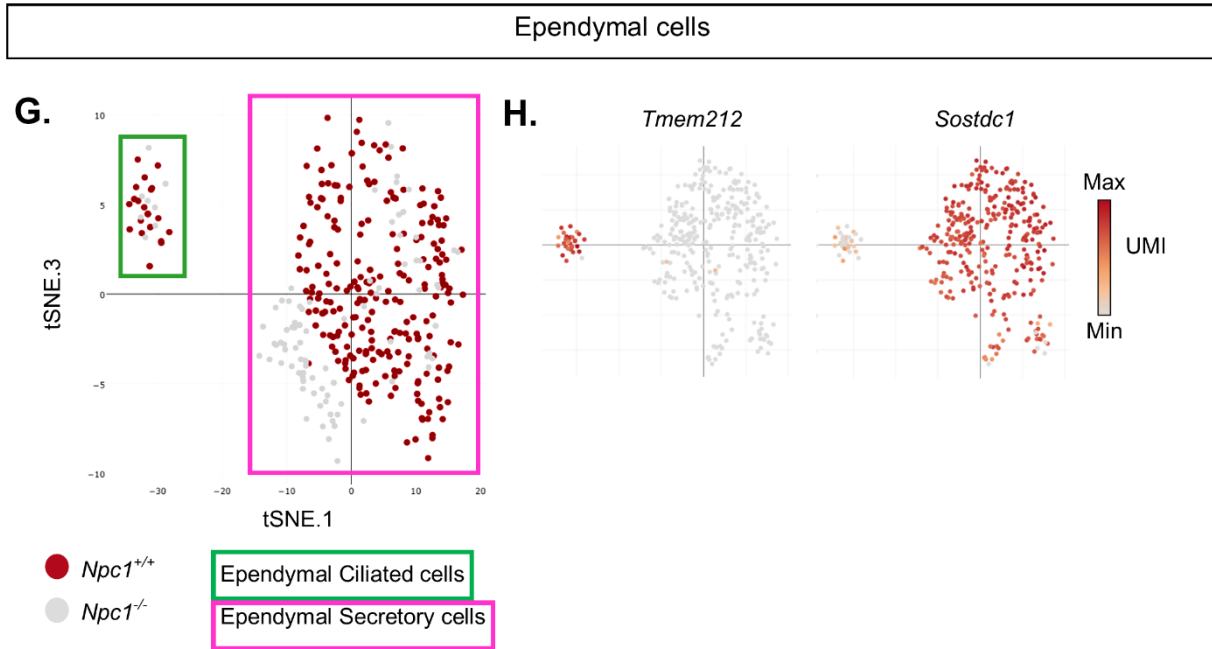


Figure S4. (A) t-SNE plot of astrocytes from *Npc1*^{+/+} (red) and *Npc1*^{-/-} (gray) 7-week-old mice. (B) t-SNE plots of *Slc1a3*, *Gfap*, *Aqp4* and *Gdf10* expression levels in astrocytes. (C) t-SNE plot of the oligodendrocytes from *Npc1*^{+/+} (red) and *Npc1*^{-/-} (gray) 7-week-old mice. (D) K-Mean clustering on t-SNE plot of the oligodendrocytes from both genotypes. OC: oligodendrocytes (blue) and OPC: oligodendrocyte precursors (orange). t-SNE plots of *Marcks* (E) and *Car2* (F) expression in oligodendrocyte precursors and oligodendrocytes, respectively. (G) t-SNE plot of ependymal cells from *Npc1*^{+/+} (red) and *Npc1*^{-/-} (gray) 7-week-old mice. Ependymal ciliated cells (green box) and ependymal secretory cells (pink box). (H) t-SNE plots of *Tmem212* and *Sostdc1* expression by ependymal cells. UMI: Unique Molecular Identifier; Max: maximum; Min: minimum.

Figure S5

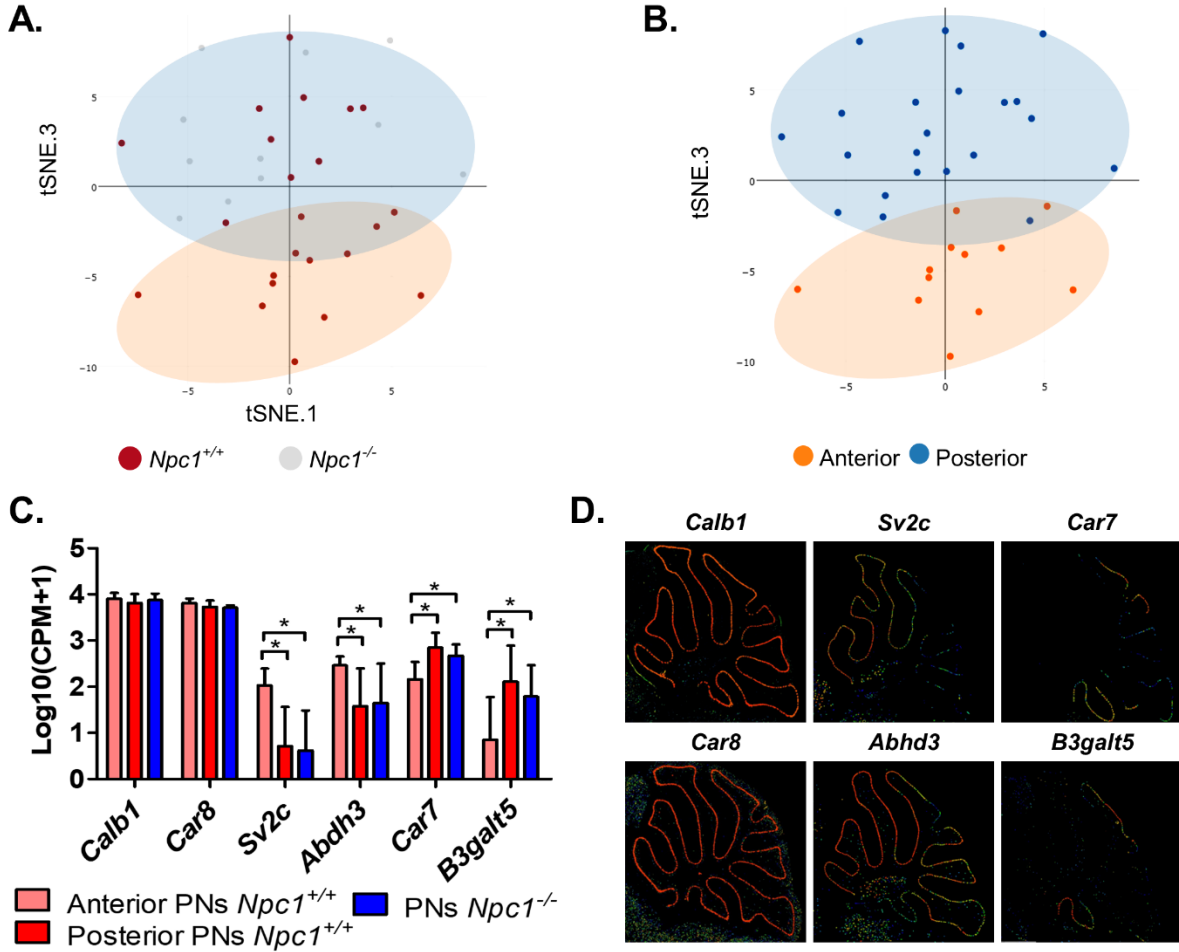


Figure S5 extended Figure 5: (A) t-SNE plot of the Purkinje cells from *Npc1*^{+/+} (red) and *Npc1*^{-/-} (gray) 7-week-old mice. The light blue ellipse surrounds the posterior Purkinje cells. The light orange ellipse surrounds the anterior Purkinje cells. (B) K-Mean clustering on t-SNE plot of the Purkinje cells from both genotypes of 7-week-old mice. The 2 clusters separate cells from the posterior (blue) and anterior (orange) cerebellum. The light blue ellipse surrounds the posterior Purkinje cells. The light orange ellipse surrounds the anterior Purkinje cells. (C) Expression level of transcripts used to differentiate anterior and posterior Purkinje neurons. (D) Corresponding *in situ* expression of genes used to identify anterior and posterior Purkinje neurons from the Allen brain atlas (<https://portal.brain-map.org/>).

Figure S6

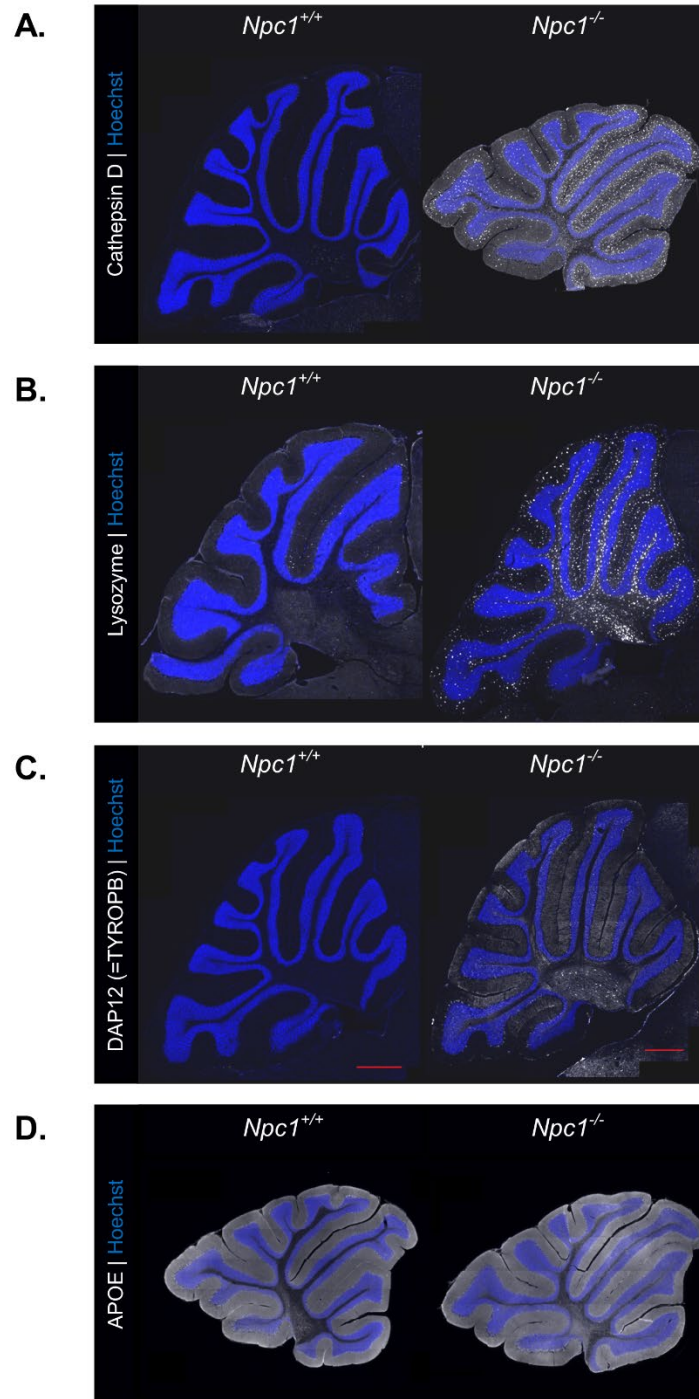


Figure S6. Representative immunostaining of 7-week-old *Npc1*^{+/+} and *Npc1*^{-/-} cerebellum stained for Cathepsin D (A), Lysozyme (B), DAP12 (C) and APOE (D). Nuclei were counterstained with Hoechst (Blue).

Figure S7

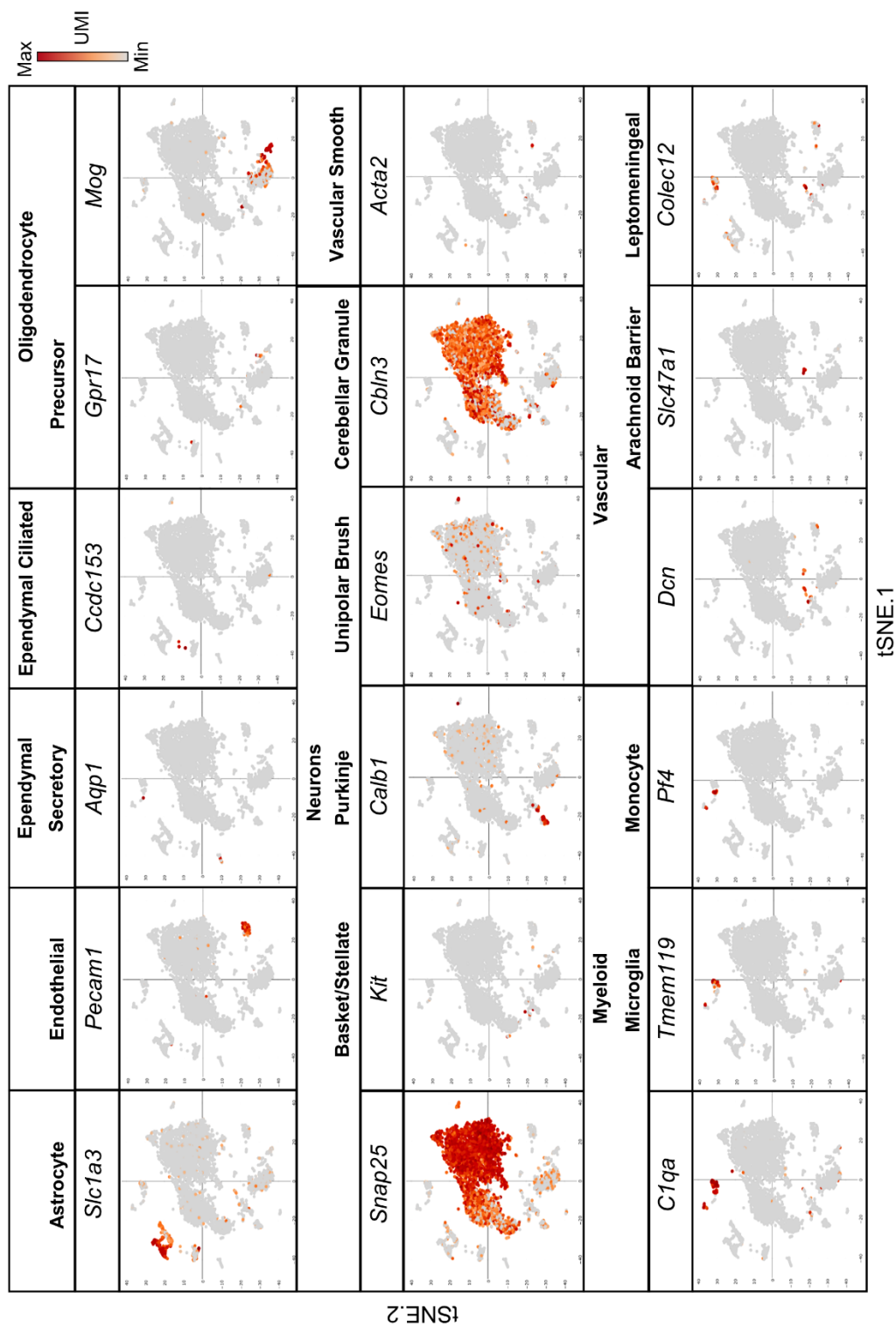


Figure S7. t-SNE plots of the different cell types identified in cerebellar tissue from 3-week-old *Npc1*^{+/+} and *Npc1*^{-/-} mice. UMI: Unique Molecular Identifier; Max: maximum; Min: minimum.

Table S1

	Glial Cells					Vascular Cells			Neurons					
Gene	Astrocytes (A)	Ependymal ciliated cells (ECC)	Ependymal Secretory cells (ESC)	Oligodendrocytes (O)	Oligodendrocytes Precursor (OP)	Endothelial cells (EC)	Vascular Leptomeningeal cells (VLC)	Vascular Smooth muscle cells (VSM)	Basket/Stellate cells (B/S)	Cerebellar granule cells (GC)	Interneurons (IN)	Purkinje neurons (PN)	Unipolar Brush Cell (UBC)	Gene description and relevance to NPC1

7-week-old NPC1 mutant expression

<i>Agt</i>	↑													<i>Agt</i> , encodes angiotensinogen precursor previously described to be a mature astrocyte marker important for blood-brain-barrier integrity [1].
<i>Apoe</i>			↑	↑	↑		↑	↑		↑	↑	↑	↑	<i>Apoe</i> encodes Apolipoprotein E, the main extracellular cholesterol carrier in the central nervous system. Variants of apolipoprotein E have been associated with Alzheimer and Niemann-Pick type C disease severity [2, 3].
<i>Aqp4</i>	↑													<i>Aqp4</i> encodes Aquaporin 4, an intrinsic plasma membrane protein that functions as water-selective channels and controls brain water homeostasis and blood-brain permeability [4].
<i>Arhgap31</i>					↑									<i>Arhgap31</i> encodes Rho GTPase-activating protein 31 which interacts with Rho GTPases RAC1 and CDC42 [5]. Endothelial cell expression of <i>Arhgap31</i> is important for vascular development and VEGF mediated angiogenesis, but its function in oligodendrocytes is not known [5]. Pathogenic variants of <i>ARHGAP31</i> are causative of Adams-Oliver syndrome [6].

Table S1 (continued)

Gene	A	ECC	ESC	O	OP	EC	VLC	VSM	B/S	GC	IN	PN	UBC	Gene description and relevance to NPC1
<i>B2m</i>			↑	↑								↑		<i>B2m</i> encodes beta 2 microglobulin, a protein essential for the expression of properly folded MHC-I protein [7].
<i>C1qa</i>		↑	↑									↑		<i>C1qa</i>, <i>C1qb</i> and <i>C1qc</i> encode subunits of complement C1. A previous study by Lopez <i>et al.</i> , demonstrated the lack of a role played by <i>C1qa</i> in <i>Npc1</i> ^{-/-} cerebellar disease progression [8].
<i>C1qb</i>			↑											
<i>C1qc</i>			↑											
<i>Cd63</i>				↑										<i>Cd63</i> /CD63 (LAMP3) is a member of the transmembrane 4 superfamily or tetraspanins expressed in late endosomes, lysosomes and at the surface of exosomes [9]. Loss of <i>Npc1</i> has been studied in oligodendrocytes and shown to enhance exosome secretion. Thus, increased expression of <i>Cd63</i> may be a cellular response to decrease unesterified cholesterol storage. Strauss <i>et al.</i> , [10] previously demonstrated in NPC1 that exosomal release of unesterified cholesterol can bypass the intracellular trafficking defect to maintain cellular cholesterol homeostasis.
<i>Ctla2a</i>						↑								<i>Ctla2a</i> encodes Cytotoxic T-Lymphocyte Antigen-2α, a protein with immunomodulatory property in addition to its Cathepsin L like cysteine protease inhibition function [11].
<i>Ctsb</i>	↑			↑		↑								<i>Ctsb</i> encodes the lysosomal cysteine protease cathepsin B. Cathepsin B appears to be neuroprotective in NPC1 given that loss of cathepsin B function in <i>Npc1</i> ^{-/-} : <i>Ctsb</i> ^{-/-} mice leads to a more severe NPC1 phenotype [12].
<i>Ctsd</i>	↑			↑	↑	↑		↑	↑	↑	↑	↑	↑	<i>Ctsd</i> encodes Cathepsin D, a lysosomal aspartyl protease mutated in type 10 neuronal ceroid lipofuscinosis [13]. It has previously been reported to be overexpressed in NPC1 cerebellum over the course of the disease in microglia and neurons [14].

Table S1 (continued)

Gene	A	ECC	ESC	O	OP	EC	VLC	VSM	B/S	GC	IN	PN	UBC	Gene description and relevance to NPC1
<i>Ctsl</i>			↑											<i>Ctsl</i> encodes the protein Cathepsin L1, a lysosomal cysteine protease involved in intracellular protein catabolism. The loss of <i>Ctsl</i> leads to a <i>NPC1</i> -like phenotype with cholesterol sequestration, suggesting its overexpression could be a compensatory mechanism to the loss of <i>Npc1</i> [15].
<i>Ctss</i>			↑									↑		<i>Ctss</i> and <i>Ctsz</i> encode Cathepsin S and Z which are lysosomal cysteine proteases involved in intracellular protein catabolism and CTSS is known to be increased at the protein level in <i>Npc1</i> ^{-/-} mouse tissues [16].
<i>Ctsz</i>			↑									↑		
<i>Fabp5</i>			↑	↑										<i>Fabp5</i> encodes Fatty Acid Binding Protein 5 which is involved in fatty acid uptake, transport, and metabolism. May be a compensation mechanism due to the defective lysosomal lipid storage.
<i>Fcer1g</i>			↑											<i>Fcer1g</i> encodes for the high affinity immunoglobulin epsilon receptor subunit gamma, a receptor involved in axon regeneration previously reported [17] to be expressed glial cells [18].
<i>Fstl4</i>												↓		<i>Fstl4</i> encodes a protein of unknown function: Follistatin Like 4
<i>Fxyd7</i>												↓		<i>Fxyd7</i> is encodes FXYD7, a brain specific member of the FXYD family. FXYD7 functions to regulate neuronal excitability by controlling Na,K-ATPase $\alpha 1$ - β isozymes K ⁺ affinity [19]. However, the significant change in <i>Fxyd7</i> expression is not associated with electrophysiological differences in Purkinje neurons following the loss of <i>Npc1</i> [20].
<i>Hexb</i>			↑											<i>Hexb</i> encodes hexosaminidase subunit beta which catalyzes the lysosomal degradation of the ganglioside GM2. GM2 is known to accumulate in NPC [21]. Carrying pathogenic variants on both alleles leads to Sandhoff disease. Its expression at the

Table S1 (continued)

														protein level was previously reported to be increased in <i>Npc1</i> ^{-/-} tissues [16].
Gene	A	ECC	ESC	O	OP	EC	VLC	VSM	B/S	GC	IN	PN	UBC	Gene description and relevance to NPC1
<i>Gfap</i>	↑													<i>Gfap</i> encodes one of the major intermediate filament proteins of mature astrocytes known to be overexpressed in reactive astrocytes [22].
<i>Gstp1</i>				↑										<i>Gstp1</i> encodes Glutathione S-transferase P1, an enzyme involved in detoxification of reactive species, potentially detrimental, by conjugation with glutathione. Increased expression of GSTP1 has been proposed to be neuroprotective [23].
<i>Lyz2</i>	↑	↑	↑	↑	↑		↑	↑	↑		↑	↑	↑	<i>Lyz2</i> gene encodes lysozyme M, an antibacterial protein homologous to human lysozyme. Neuronal expression of lysozyme may be detrimental in Sanfilippo syndrome type B [24].
<i>Manf</i>			↓											<i>Manf</i> encodes Mesencephalic Astrocyte Derived Neurotrophic Factor, a neuroprotective factor involved in the control of endoplasmic reticulum stress signaling [25]. Its downregulation is suggested to affect sleep and promote neurodegeneration by promoting neuroinflammation, a hallmark of NPC [4, 26].
<i>Map3k7cl</i>								↑						<i>Map3k7cl</i> encodes MAP3 Kinase 7 C-Terminal Like protein. The function of this protein is not known.
<i>Mt1</i>	↑													<i>Mt1</i> and <i>Mt2</i> are members of the metallothionein family of genes, coding for antioxidant proteins which bind divalent heavy metal ions. These receptors when binding to melatonin have pro-survival and anti-inflammatory functions [27].
<i>Mt2</i>	↑													
<i>Npc2</i>				↑										NPC2 functions in concert with NPC1 to transport unesterified cholesterol out of the endolysosomal compartment [28] and increased expression of <i>Npc2</i> has been reported in <i>Npc1</i> ^{-/-} oligodendrocytes [10].

Table S1 (continued)

Gene	A	ECC	ESC	O	OP	EC	VLC	VSM	B/S	GC	IN	PN	UBC	Gene description and relevance to NPC1
<i>Ptgds</i>				↑										<i>Ptgds</i> encodes prostaglandin-H2 D-isomerase (PTGDS), an enzyme that converts prostaglandin H2 to prostaglandin D2. Prostaglandin D2 is a known neuromodulator and trophic factor in the CNS [29] suggesting its overexpression is neuroprotective in an attempt to slow the progressive cerebellar degeneration.
<i>S100a1</i>				↑										<i>S100a1</i> encodes S100 calcium-binding protein A1 (S100A1). The function of S100A1 has been extensively studied in the skeletal muscle cells for its calcium dependent interaction with ryanodine 1, a well-established Purkinje neuron marker [30]. Disrupted calcium signaling appears to be involved in NPC cellular pathology and Purkinje cell loss [31-33].
<i>Spp1</i>			↑											<i>Spp1</i> encodes the Osteopontin, a protein with anti-apoptotic and immunomodulatory functions [4].
<i>Them6</i>												↑		<i>Them6</i> encodes thioesterase superfamily member 6, a protein of unknown function.
<i>Tmsb4x</i>			↑											<i>Tmsb4x</i> encodes Thymosin beta-4 a protein of unknown function expressed in the CNS.
<i>Trem2</i>			↑											TREM2 (triggering receptor expressed on myeloid cells 2) is thought to function as a receptor for TYROBP [34]. Mutations of either <i>TYROBP</i> or <i>TREM2</i> are found in Nasu-Hakola disease, an autosomal recessive disorder characterized by progressive dementia and bone defects [35]. Variants of <i>TREM2</i> are risk factors for Alzheimer disease [34]. Increased expression of <i>Trem2</i> is associated with neuroinflammation in general [36], but has specifically been observed in microglia from <i>Npc1</i> ^{-/-} mutant mice [37].

Table S1 (continued)

Gene	A	ECC	ESC	O	OP	EC	VLC	VSM	B/S	GC	IN	PN	UBC	Gene description and relevance to NPC1
<i>Ttr</i>	↑								↑					<i>Ttr</i> encodes for transthyretin, a protein present in serum and cerebrospinal fluid which transports thyroid hormone and retinol. TTR misfolding and aggregation is associated with amyloid diseases [38].
<i>Tyrobp</i>		↑	↑		↑		↑					↑	↑	<i>Tyrobp</i> encodes TYRO protein tyrosine kinase-binding protein, a transmembrane signaling protein also known as DAP12. This protein appears to have multiple functions including involvement in myelination and [39]. Loss of TYROBP function delays disease progression in Alzheimer Disease mice [40]; however, given its involvement in dementia, it is not clear whether inhibition of DAP12 function would be of potential benefit in NPC1.

3-week-old NPC1 mutant expression														
Gene	A	ECC	ESC	O	OP	EC	VLC	VSM	B/S	GC	IN	PN	UBC	Gene description and relevance to NPC1
<i>Slc16a1</i>						↓								<i>Slc16a1</i> or solute carrier family 16-member 1 gene encodes for monocarboxylate transporter 1 (MCT1) that mediates the movement of monocarboxylates through the plasma membrane [41]. This overexpression might prevent the shift toward a glycolytic metabolism reported in other cell type following the loss of <i>Npc1</i> [37].
<i>Timp3</i>						↑								<i>Timp3</i> encodes tissue inhibitor of metalloproteinase 3. Metalloproteinases are a group of enzymes involved in degradation of the extracellular matrix that functions to facilitate cell movement and inflammation.

Table S1 (continued)

Table S1 References

1. Wosik, K., et al., *Angiotensin II controls occludin function and is required for blood-brain barrier maintenance: Relevance to multiple sclerosis*. Journal of Neuroscience, 2007. **27**(34): p. 9032-9042.
2. Fu, R., et al., *Apolipoprotein E genotype and neurological disease onset in Niemann-Pick disease, type C1*. Am J Med Genet A, 2012. **158A**(11): p. 2775-80.
3. Liu, C.C., et al., *Apolipoprotein E and Alzheimer disease: risk, mechanisms and therapy*. Nat Rev Neurol, 2013. **9**(2): p. 106-18.
4. Ikeshima-Kataoka, H., *Neuroimmunological Implications of AQP4 in Astrocytes*. Int J Mol Sci, 2016. **17**(8).
5. Caron, C., et al., *CdGAP/ARHGAP31, a Cdc42/Rac1 GTPase regulator, is critical for vascular development and VEGF-mediated angiogenesis*. Sci Rep, 2016. **6**: p. 27485.
6. Isrie, M., et al., *Isolated terminal limb reduction defects: extending the clinical spectrum of Adams-Oliver syndrome and ARHGAP31 mutations*. Am J Med Genet A, 2014. **164A**(6): p. 1576-9.
7. Smith, L.K., et al., *beta2-microglobulin is a systemic pro-aging factor that impairs cognitive function and neurogenesis*. Nat Med, 2015. **21**(8): p. 932-7.
8. Lopez, M.E., A.D. Klein, and M.P. Scott, *Complement is dispensable for neurodegeneration in Niemann-Pick disease type C*. J Neuroinflammation, 2012. **9**: p. 216.
9. Tancini, B., et al., *Insight into the Role of Extracellular Vesicles in Lysosomal Storage Disorders*. Genes (Basel), 2019. **10**(7).
10. Strauss, K., et al., *Exosome secretion ameliorates lysosomal storage of cholesterol in Niemann-Pick type C disease*. J Biol Chem, 2010. **285**(34): p. 26279-88.
11. Sugita, S., et al., *Induction of T regulatory cells by cytotoxic T-lymphocyte antigen-2alpha on corneal endothelial cells*. Invest Ophthalmol Vis Sci, 2011. **52**(5): p. 2598-605.
12. Chung, C., et al., *Genetic and pharmacological evidence implicates cathepsins in Niemann-Pick C cerebellar degeneration*. Human Molecular Genetics, 2016. **25**(7): p. 1434-1446.
13. Koch, S., et al., *Morphologic and functional correlates of synaptic pathology in the cathepsin D knockout mouse model of congenital neuronal ceroid lipofuscinosis*. J Neuropathol Exp Neurol, 2011. **70**(12): p. 1089-96.

Table S1 (continued)

14. Amritraj, A., et al., *Increased activity and altered subcellular distribution of lysosomal enzymes determine neuronal vulnerability in Niemann-Pick type C1-deficient mice*. Am J Pathol, 2009. **175**(6): p. 2540-56.
15. Cermak, S., et al., *Loss of Cathepsin B and L Leads to Lysosomal Dysfunction, NPC-Like Cholesterol Sequestration and Accumulation of the Key Alzheimer's Proteins*. Plos One, 2016. **11**(11).
16. Sleat, D.E., et al., *Proteomic analysis of mouse models of Niemann-Pick C disease reveals alterations in the steady-state levels of lysosomal proteins within the brain*. Proteomics, 2012. **12**(23-24): p. 3499-3509.
17. Martin, K.B., et al., *Identification of Novel Pathways Associated with Patterned Cerebellar Purkinje Neuron Degeneration in Niemann-Pick Disease, Type C1*. Int J Mol Sci, 2019. **21**(1).
18. Zhang, G., et al., *Fcgamma receptor-mediated inflammation inhibits axon regeneration*. PLoS One, 2014. **9**(2): p. e88703.
19. Beguin, P., et al., *FXVD7 is a brain-specific regulator of Na,K-ATPase alpha 1-beta isozymes*. Embo Journal, 2002. **21**(13): p. 3264-3273.
20. Elrick, M.J., et al., *Conditional Niemann-Pick C mice demonstrate cell autonomous Purkinje cell neurodegeneration*. Human Molecular Genetics, 2010. **19**(5): p. 837-847.
21. Vanier, M.T., *Lipid changes in Niemann-Pick disease type C brain: personal experience and review of the literature*. Neurochem Res, 1999. **24**(4): p. 481-9.
22. Liddelow, S.A., et al., *Neurotoxic reactive astrocytes are induced by activated microglia*. Nature, 2017. **541**(7638): p. 481-487.
23. Sun, K.H., et al., *Glutathione-S-transferase P1 is a critical regulator of Cdk5 kinase activity*. J Neurochem, 2011. **118**(5): p. 902-14.
24. Ohmi, K., et al., *Sanfilippo syndrome type B, a lysosomal storage disease, is also a tauopathy*. Proceedings of the National Academy of Sciences of the United States of America, 2009. **106**(20): p. 8332-8337.
25. Renko, J.M., et al., *Mesencephalic Astrocyte-Derived Neurotrophic Factor (MANF) Elevates Stimulus-Evoked Release of Dopamine in Freely-Moving Rats*. Molecular Neurobiology, 2018. **55**(8): p. 6755-6768.
26. Walkowicz, L., et al., *Downregulation of DmMANF in Glial Cells Results in Neurodegeneration and Affects Sleep and Lifespan in Drosophila melanogaster*. Frontiers in Neuroscience, 2017. **11**.
27. Tarocco, A., et al., *Melatonin as a master regulator of cell death and inflammation: molecular mechanisms and clinical implications for newborn care*. Cell Death Dis, 2019. **10**(4): p. 317.
28. Vanier, M.T., *Niemann-Pick disease type C*. Orphanet J Rare Dis, 2010. **5**: p. 16.

Table S1 (continued)

29. Cruz Duarte, P., B. St-Jacques, and W. Ma, *Prostaglandin E2 contributes to the synthesis of brain-derived neurotrophic factor in primary sensory neuron in ganglion explant cultures and in a neuropathic pain model*. *Exp Neurol*, 2012. **234**(2): p. 466-81.
30. Wright, N.T., et al., *S100A1 and calmodulin compete for the same binding site on ryanodine receptor*. *J Biol Chem*, 2008. **283**(39): p. 26676-83.
31. Lloyd-Evans, E., et al., *Niemann-Pick disease type C1 is a sphingosine storage disease that causes deregulation of lysosomal calcium*. *Nat Med*, 2008. **14**(11): p. 1247-55.
32. Yamamoto, T., T. Tokoro, and Y. Eto, *The attenuated elevation of cytoplasmic calcium concentration following the uptake of low density lipoprotein in type C Niemann-Pick fibroblasts*. *Biochem Biophys Res Commun*, 1994. **198**(2): p. 438-44.
33. Ferrante, A., et al., *The adenosine A2A receptor agonist T1-11 ameliorates neurovisceral symptoms and extends the lifespan of a mouse model of Niemann-Pick type C disease*. *Neurobiol Dis*, 2018. **110**: p. 1-11.
34. Carmona, S., et al., *The role of TREM2 in Alzheimer's disease and other neurodegenerative disorders*. *Lancet Neurol*, 2018. **17**(8): p. 721-730.
35. Kaneko, M., et al., *Nasu-Hakola disease: The first case reported by Nasu and review: The 50th Anniversary of Japanese Society of Neuropathology*. *Neuropathology*, 2010. **30**(5): p. 463-70.
36. Keren-Shaul, H., et al., *A Unique Microglia Type Associated with Restricting Development of Alzheimer's Disease*. *Cell*, 2017. **169**(7): p. 1276-1290 e17.
37. Cougnoux, A., et al., *Microglia activation in Niemann-Pick disease, type C1 is amendable to therapeutic intervention*. *Hum Mol Genet*, 2018. **27**(12): p. 2076-2089.
38. Ueda, M. and Y. Ando, *Recent advances in transthyretin amyloidosis therapy*. *Transl Neurodegener*, 2014. **3**: p. 19.
39. Colonna, M., *DAP12 signaling: from immune cells to bone modeling and brain myelination*. *J Clin Invest*, 2003. **111**(3): p. 313-4.
40. Audrain, M., et al., *Integrative approach to sporadic Alzheimer's disease: deficiency of TYROBP in a tauopathy mouse model reduces C1q and normalizes clinical phenotype while increasing spread and state of phosphorylation of tau*. *Mol Psychiatry*, 2019. **24**(9): p. 1383-1397.
41. Al-Khawaga, S., et al., *A SLC16A1 Mutation in an Infant With Ketoacidosis and Neuroimaging Assessment: Expanding the Clinical Spectrum of MCT1 Deficiency*. *Front Pediatr*, 2019. **7**: p. 299.

Table S3

Primary Antibody	Dilution	Incubation	Application	Host	Company	Clone/Catalog No.
GFAP	1:400	Overnight, 4°C	IF	Chicken	Novus	NPP1-05198
Calbindin	1:400	Overnight, 4°C	IF	Mouse	Sigma-Aldrich Millipore	CB-955
Calbindin	1:400	Overnight, 4°C	IF	Rabbit	Cell Signaling Tech.	13176S
CD117/KIT	1:400	Overnight, 4°C	IF	Rat	R&D Systems	MAB1356
ALDH1L1	1:100	Overnight, 4°C	IF	Rabbit	Invitrogen	PA532127
AQP4	1:400	Overnight, 4°C	IF	Mouse	LS Bio	LS-C413484
IBA1	1:200	Overnight, 4°C	IF	Rabbit	Wako	019-19741
OLIG2	1:200	Overnight, 4°C	IF	Rabbit	Sigma-Aldrich Millipore	AB9610
CD68	1:800	Overnight, 4°C	IF	Rat	Sigma-Aldrich Millipore	FA-11
TBR2	1:400	Overnight, 4°C	IF	Rabbit	Sigma-Aldrich Millipore	AB2283
AQP1	1:400	Overnight, 4°C	IF	Mouse	abcam	ab9566
Parvalbumin	1:1000	Overnight, 4°C	IF	Rabbit	Abcam	ab11427
NeuroD1	1:1000	Overnight, 4°C	IF	Rabbit	Abcam	ab213725
CBLN3	1:400	30 min, 4°C	FC	Rabbit	Novusbio	NBP1-85844
TTR	1:400	Overnight, 4°C	IF	Rabbit		
NEUN	1:400	Overnight, 4°C	IF	Mouse	Sigma-Aldrich Millipore	A-60
APOE	1:200	30 min, 4°C	FC	Rabbit	Biodesign International	K23100R
CD11c-FITC	1:100	30 min, 4°C	FC	Rat	Biolegend	N418
Secondary Antibody	Dilution	Incubation	Application	Host	Company	Clone/Catalog No.
anti-Chicken conjugated to Alexa-Fluor 488	1:2000	1 hour, room temp	IF	Goat	ThermoFischer Scientific	A-11039
anti-Mouse conjugated to Alexa-Fluor 488	1:2000	1 hour, room temp	IF	Goat	ThermoFischer Scientific	A28175
anti-Mouse conjugated to DyLight-594	1:2000	1 hour, room temp	IF	Goat	ThermoFischer Scientific	35510
anti-Mouse conjugated to Alexa-Fluor-633	1:2000	1 hour, room temp	IF	Goat	ThermoFischer Scientific	A-21052
anti-Rabbit conjugated to Alexa-Fluor 488	1:2000	1 hour, room temp	IF	Goat	ThermoFischer Scientific	A27034
anti-Rabbit conjugated to AlexaFluor-594	1:2000	1 hour, room temp	IF	Goat	ThermoFischer Scientific	R37119
anti-Rabbit conjugated to Alexa-Fluor-633	1:2000	1 hour, room temp	IF	Goat	ThermoFischer Scientific	A-21071
anti-Rabbit conjugated to Alexa-Fluor 488	1:2000	1 hour, 4°C	FC	Goat	ThermoFischer Scientific	A27034

Table S3 (continued)

Reagent	Dilution	Incubation	Application	Host	Company	Clone/Catalog No.
Hoechst 3342	1:5000	15 min, room temp	IF	N/A	ThermoFischer Scientific	62249
Goat Serum	1:20	1 hour, 4°C	IF		Sigma-Aldrich Millipore	G9023
Neurotrace	1:1000	1 hour, 4°C	IF		ThermoFischer Scientific	N21483
30% Bovine Serum Albumin	1:10	10 min, 4°C	FACS	Bovine	Sigma-Aldrich Millipore	A9576-50ML
TruStain FcX	1:100	10 min, 4°C	FACS	Rat	Biolegend	101320
Mouse B2M / Beta 2 Microglobulin ELISA Kit			ELISA		LSBio	LS-F14141
Human Lysozyme ELISA Kit			ELISA		LSBio	NBP2-60511
Human CTSD / Cathepsin D ELISA Kit			ELISA		LSBio	LS-F6862-1
Bovine Serum Albumin	2% w/v		WB		Sigma-Aldrich Millipore	A7030
Paraformaldehyde	4% w/v		IF		Sigma-Aldrich Millipore	A6148
Papain dissociation system			Cerebellum dissociation		Worington	LK003150
KAPA Taq Ready Mix			Genotyping		Sigma-Aldrich Millipore	KK1006
PLX3397	290mg/kg	Mouse feed. RT	Microglia depletion		Selleckchem	S7818
Software					Company	Version
Prism 5					Graphpad	Version 5.01
R/Rstudio					CRAN foundation	Version 1.1.463
MS Office 365 and 2016					Microsoft	
ZEN2(Blue)					Zeiss	
ImageJ					NIH	Version 1.52i
Primers	Sequence				Company	
Npc1_F	GGACAGCCAAGTAGGCGAC				Integrated DNA Technologies	
Npc1_R	GACGTGTGCACCCCCTTTC				Integrated DNA Technologies	