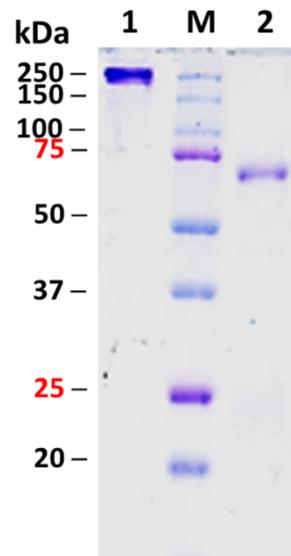
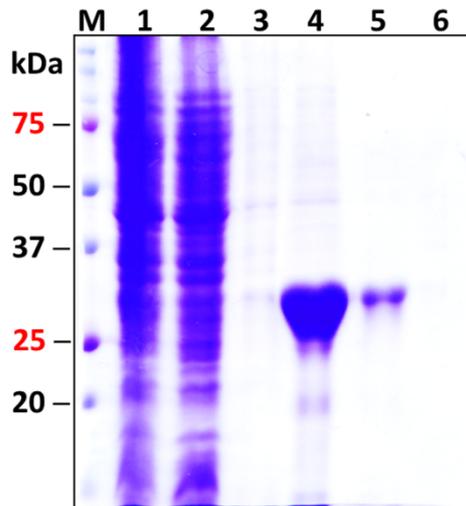


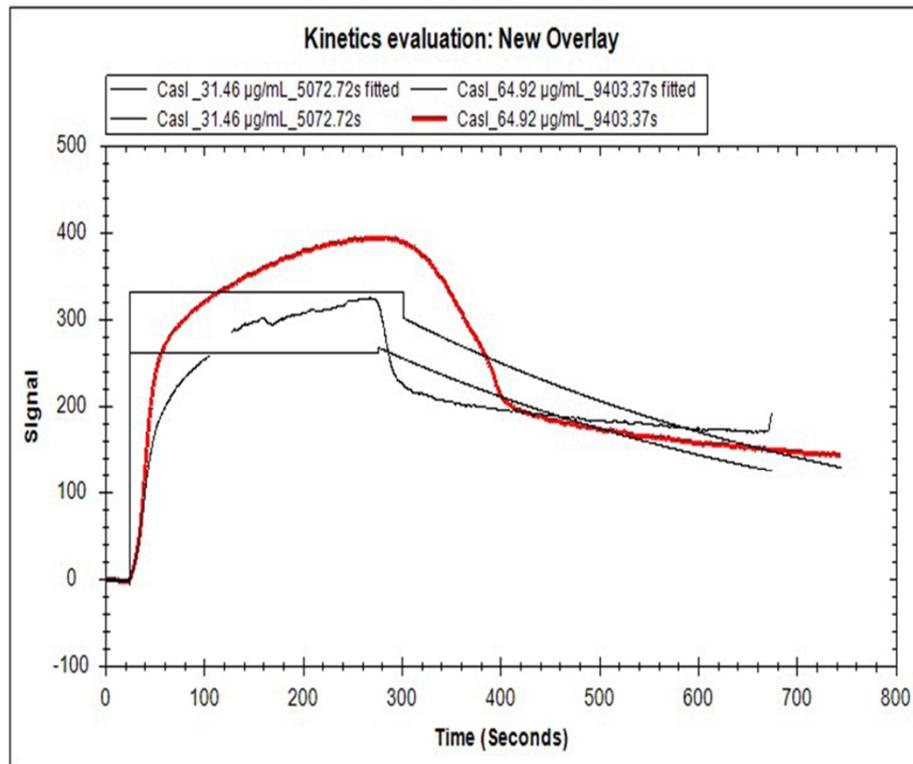
## Supplementary Data



**Figure S1.** Analysis of purified polyclonal IgY antibodies. Adult female Leghorn chickens were immunized with purified CaEno1 protein for seven times at 7-day intervals. IgY antibodies purified from the eggs laid by immunized chickens were analyzed on Coomassie blue-stained SDS-PAGE under either non-reducing (lane 1) or reducing condition (lane 2). Approximately 50–75 mg of IgY antibodies could be obtained from each egg yolk.



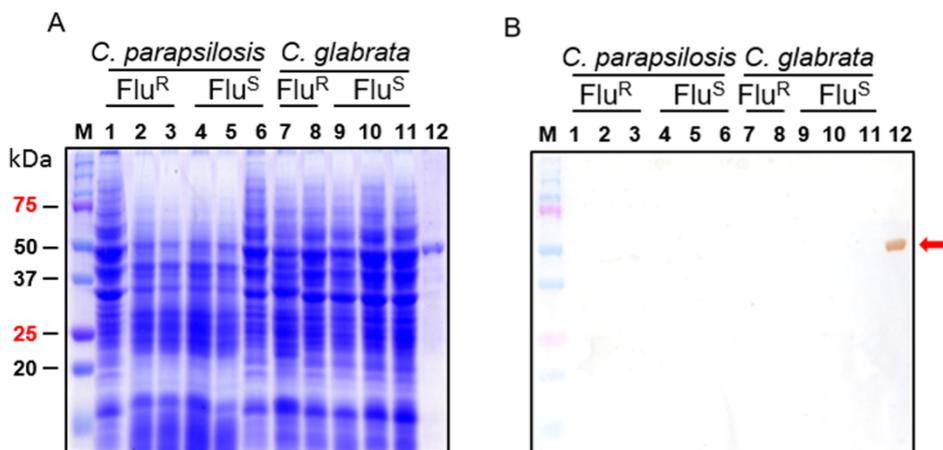
**Figure S2.** Expression and purification of CaS1 scFv antibody. After induced by IPTG, the *E. coli*-derived CaS1 scFv antibody was purified using Ni<sup>2+</sup> sepharose and analyzed on Coomassie blue-stained SDS-PAGE. The molecular weight of CaS1 scFv was around 26 kDa as expected. Approximately 1.0–1.5 mg of CaS1 could be obtained from 1 L of bacterial culture. Lane 1: total cell lysate. Lane 2: flow through. Lane 3: washing fraction. Lane 4: elution 1. Lane 5: elution 2. Lane 6: Ni<sup>2+</sup> sepharose.



Evaluation type: OneToOne

Curve name	Bmax (Signal)	ka (1/(M*s))	kd (1/s)	KD (M)	BI (Signal)	Chi2 (Signal^2)
CasI_31.46 µg/mL_5072.72s fitted	267.33	9.83e4	1.91e-3	1.95e-8	-6.04	3088.68
CasI_64.92 µg/mL_9403.37s fitted	301.64	9.83e4	1.91e-3	1.95e-8	30.07	3088.68

**Figure S3.** Binding affinity of CaS1 determined by SPR method. The rCaEno1 was first immobilized on the sensor chips. Different concentrations of CaS1 (30–65 µg/mL) were then injected over the coated surface of sensor chip. Sensorgram of the CaEno1/CaS1 interaction was recorded and analyzed using the built-in TraceDrawer software package.



**Figure S4.** Binding analysis of CaS1 scFv against Eno1 protein expressed by *C. parapsilosis* and *C. glabrata*. Total cell lysates of five *C. parapsilosis* and four *C. glabrata* were visualized by SDS-PAGE (A) and probed with CaS1 scFv (B) as described in the text. Lanes 1–6 contained the total cell lysates of 3 FLU<sup>R</sup> (CP 8-20, CP 12-27, CP 6-30) and 3 FLU<sup>S</sup> (CP 7-17, CP 8-48, BCRC 20515) *C. parapsilosis* strains, respectively. Lanes 7–11 contained total cell lysates of 2 FLU<sup>R</sup> (CG 5-8, CG 8-11) and 3 FLU<sup>S</sup> (CG 7-37, CG 5-66, BCRC 20586) *C. glabrata* strains, respectively. Lane 12 contained rCaEno1 as a positive control (red arrow).

**Table S1.** Clinical fluconazole resistant and susceptible *Candida spp.* and their MIC.

Specimen	Organism	Drug	MIC ( $\mu\text{g}/\text{mL}$ )	Interpretation *
CA6-17	<i>C. albicans</i>	fluconazole	16	R
CA7-26	<i>C. albicans</i>	fluconazole	8	R
CA7-3	<i>C. albicans</i>	fluconazole	2	S
CA10-50	<i>C. albicans</i>	fluconazole	2	S
CA7-30	<i>C. albicans</i>	fluconazole	1	S
CA10-65	<i>C. albicans</i>	fluconazole	1	S
CT6-29	<i>C. tropicalis</i>	fluconazole	32	R
CT11-52	<i>C. tropicalis</i>	fluconazole	32	R
CT6-50	<i>C. tropicalis</i>	fluconazole	8	R
CT12-54	<i>C. tropicalis</i>	fluconazole	8	R
CG5-8	<i>C. glabrata</i>	fluconazole	64	R
CG8-11	<i>C. glabrata</i>	fluconazole	64	R
CG7-37	<i>C. glabrata</i>	fluconazole	32	S
CG5-66	<i>C. glabrata</i>	fluconazole	16	S
CP8-20	<i>C. parapsilosis</i>	fluconazole	256	R
CP12-37	<i>C. parapsilosis</i>	fluconazole	256	R
CP6-20	<i>C. parapsilosis</i>	fluconazole	16	R
CP7-17	<i>C. parapsilosis</i>	fluconazole	8	S
CP8-48	<i>C. parapsilosis</i>	fluconazole	4	S

\* R: resistance; S: susceptible.

**Table S2.** The threshold cycle (Ct) values of *C. albicans* in tissues by real-time PCR.

Group	<i>C. albicans</i> Ct	GAPDH Ct	$\Delta\text{Ct}$ <i>C. albicans</i> -GAPDH
A2	22.26 $\pm$ 0.16	30.13 $\pm$ 0.15	-7.87
A4	20.64 $\pm$ 0.26	28.78 $\pm$ 0.19	-8.14
B1	38.18 $\pm$ 0.35	30.14 $\pm$ 0.01	8.04
B2	37.21 $\pm$ 0.04	30.56 $\pm$ 0.06	6.65
B4	35.55 $\pm$ 0.01	29.06 $\pm$ 0.47	6.49
C2	-	29.37 $\pm$ 0.36	#
C4	-	31.67 $\pm$ 0.54	#
C5	-	28.97 $\pm$ 0.01	#
D2	-	28.65 $\pm$ 0.33	#
D4	-	28.61 $\pm$ 0.28	#
D5	-	28.52 $\pm$ 0.06	#

-: Undetectable; #: not available.

**Table S3.** Primers used in the amplification of  $V_H$  and  $V_L$  genes.

Primers	Nucleotide Sequences
CSCVH <sub>o</sub> -F	5'-GGTCAGTCCAGATCTTCCGCCGTGACGTTGGACGAG-3'
CSCVH <sub>o</sub> -FL	5'-GGTCAGTCCTCTAGATCTTCCGGCGGTGGTGGCAGCTCCGGTGGTG GCGGTTCCGCCGTGACGTTGGACGAG-3'
CSCG-B	5'-CTGGCCGGCCTGGCCACTAGTGGAGGAGACGATGACTTCCGGTCC-3'
CSCVK	5'-GTGGCCAGGCGGCCCTGACTCAGCCGTCTCGGTGTC-3'
CKJ <sub>o</sub> -B	5'-GGAAGATCTAGAGGACTGACCTAGGACGGTCAGG-3'
CSC-F	5'-GAGGAGGAGGAGGAGGAGGTGGCCAGGCGGCCCTGACTCAG-3'
CSC-B	5'-GAGGAGGAGGAGGAGGAGGAGCTGGCCGGCCTGGCCACTAGTGGAGG-3'