

Supplementary material: Sugar-pucker force-induced transition in single-stranded DNA

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1 DNA hairpin synthesis

The short hairpin (SH) was synthesized following the protocol in Ref. [1] (named as CD4L20). The oligonucleotide forming the 3' end of the hairpin was labelled with a digoxigenin tailing. After a purification step (*QIA Nucleotide removal kit*), all the oligonucleotides forming the hairpin (Table 1) were annealed by starting at a high temperature (70 °C) and 1 °C was decreased every minute until room temperature was reached. The hairpin was next ligated using the T4 DNA ligase (New England Biolabs) in an overnight reaction (16 °C).

The long hairpins (LHs) were synthesized following a procedure based either on a PCR amplification of a dsDNA segment or digesting a segment of the linearized λ -DNA. H700, H964, H4452 and H13680 were prepared as described in Refs. [2] and [3]. Finally, H1904 and H7138 were synthesized following the protocol in Ref. [3], changing the restriction enzyme for the digestion step: *EcoRI* (New England Biolabs) for H7138, and *BspHI* (New England Biolabs) for H1904. The sequences of the oligonucleotides used for preparing the DNA hairpins are given in Sec. 2. Fig. 1(a)-(b) shows an scheme of the molecular construct for the SH and LH hairpins, respectively.

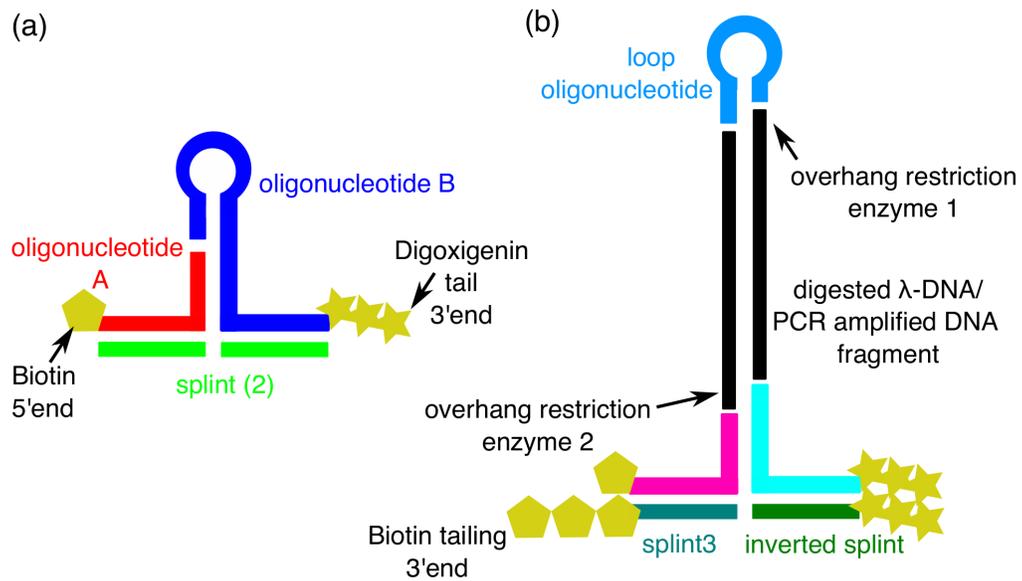


Figure 1: **Hairpin synthesis.** (a) The SH is assembled by ligating two oligonucleotides (blue and red). The oligonucleotide A is purchased biotinylated and the oligonucleotide B is end-labelled with digoxigenins using the T4 terminal transferase. The splint oligonucleotides are annealed to create dsDNA handles. (b) The LHs are assembled by ligating a set of oligonucleotides (magenta, cyan, blue) to the PCR-amplified and digested λ -phage fragment (black). Note that the complementary strands of the handles are also tailed. Color code as in Tables 1 and 2.

2 Oligonucleotides for hairpin synthesis

Name	Sequence
SH-A	5'-Biotin-AGT TAG TGG TGG AAA CAC AGT GCC AGC GCG AAC CCA CAA ACC GTG ATG GCT GTC CTT GGA GTC ATA CGC AA -3'
SH-B	5'-GAA GGA TGG AAA AAA AAA AAA AAA AAA ACA TCC TTC TTG CGT ATG ACT CCA AGG ACA GCC ATC ACG GTT TGT GGG TTC AGT TAG TGG TGG AAA CAC AGT GCC AGC GC-3'
splint	5'-GCG CTG GCA CTG TGT TTC CAC CAC TAA CT-3'

Table 1: Oligonucleotides used for the synthesis of the SH. The loop region is shown in bold.

Name	Sequence
13680b-loop	5'-Pho-GAT CGC CAG TTC GCG TTC GCC AGC ATC CGA CTA CGG ATG CTG GCG AAC GCG AAC TGG C-3'
7138b-loop	5'-Pho-AAT TGC CAG TTC GCG TTC GCC AGC ATC CGA CTA CGG ATG CTG GCG AAC GCG AAC TGG C-3'
4452b-loop	5'-Pho-TGA TAG CCT ACT AAG GCT ATC ACA TG-3'
1904b-loop	5'-Pho-CAT GAC AGT CGT TAG TAA CTA ACA TGA TAG TTA CTT TTG TAA CTA TCA TGT TAG TTA CTA ACG ACT GT-3'
964b-loop	5'-Pho-GTC ACT TAG TAA CTA ACA TGA TAG TTA CTT TTG TAA CTA TCA TGT TAG TTA CTA A-3'
700b-loop	5'-Pho-GTC ACT TAG TAA CTA ACA TGA TAG TTA CTT TTG TAA CTA TCA TGT TAG TTA CTA A-3'
Bio-cosRshort	5'-Bio-GAC TTC ACT AAT ACG ACT CAC TAT AGG GAA ATA GAG ACA CAT ATA TAA TAG ATC TT-3'
cosRlong	5'-Pho-GGG CGG CGA CCT AAG ATC TAT TAT ATA TGT GTC TCT ATT AGT TAG TGG TGG AAA CAC AGT GCC AGC GC-3'
Bio-cosLshort	5'-Bio-GAC TTC ACT AAT ACG ACT CAC TAT AGG GAA ATA GAG ACA CAT ATA TAA TAG ATC TT-3'
cosLlong	5'-Pho-AGG TCG CCG CCC AAG ATC TAT TAT ATA TGA GTC TCT ATT AGT TAG TGG TGG AAA CAC AGT GCC AGC GC 3'
HandBio-SMFP	5'-Bio-GAC TTC ACT AAT ACG ACT CAC TAT AGG GAA ATA GAG ACA CAT ATA TAA TAG ATC TTC GCA CTG AC -3'
HandDig-SMFP	5'-Pho-AAG ATC TAT TAT ATA TGT GTC TCT ATT AGT TAG TGG TGG AAA CAC AGT GCC AGC GC -3'
splint3	5'-TCC CTA TAG TGA GTC GTA TTA GTG AAG TC-3'
inverted-splint	3'-AAA AA-5'-5'-GCG CTG GCA CTG TGT TTC CAC CAC TAA C(SpC3)-3'

Table 2: Oligonucleotides used for the synthesis of long DNA hairpins. The loop region is shown in bold.

Name	Sequence
13680b – block – loop	5'-TAG TCG GAT GCT GGC GAA CGC GAA CTG GCG-3'
7138b – block – loop	5'-TAG TCG GAT GCT GGC GAA CGC GAA CTG GCG-3'
4452b – block – loop	5'-TAG TAG GCT ATC ACA TGC TGG CCA CCG GCT-3'
1904b – block – loop	5'-TTA CAA AAG TAA CTA TCA TGT TAG T-3'
964b – block – loop	5'-TTA CAA AAG TAA CTA TCA TGT TAG T-3'
700b – block – loop	5'-TTA CAA AAG TAA CTA TCA TGT TAG T-3'

Table 3: Blocking loop oligonucleotides used to generate ssDNA FECs for the different LHs.

References

- [1] Alemany, A.; Ritort, F. Determination of the elastic properties of short ssDNA molecules by mechanically folding and unfolding DNA hairpins, Biopolymers **2014**, 101, 1193–1199.
- [2] Camunas-Soler, J.; Manosas, M.; Frutos, S.; Tulla-Puche, J.; Albericio, F.; Ritort, F. Single-molecule kinetics and footprinting of DNA bis-intercalation: the paradigmatic case of Thiocoraline. Nucl. Acids Res. **2015**, 43, 2767–2779.
- [3] Camunas-Soler, J.; Frutos, S.; Bizarro, C.V.; de Lorenzo, S; Fuentes-Perez, M.E.; Ramsch, R.; Vilchez, S.; Solans, C.; Moreno-Herrero, F.; Albericio, F., et al. Electrostatic binding and hydrophobic collapse of peptide–nucleic acid aggregates quantified using force spectroscopy, ACS Nano **2013**, 7, 5102-5113.