

Supplementary Data

Genome Editing of Golden SNP-Carrying Lycopene Epsilon-Cyclase (*LcyE*)/HDR Gene using the CRSPKR-Cas9 and Geminiviral Replicon System in Rice

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Figure S3. Confirmation of T-DNA insertion from calli obtained through HDR experiment in rice. M: 1kb DNA ladder; WT: wild-type; P: pGemBos::*LcyE* plasmid vector.

Table S2. The primers list used in this study.

Supplementary Table S1. Design of sgRNAs for HDR by CRISPR-Cas9 system in rice. using the Cas-designer (<http://www.rgenome.net/cas-designer/>). Under line is PAM sequence.

sgRNA	RGEN Target (5' to 3')	Direction	GC content (%, w/o PAM)	Out-of frame score	Mismatches			
					0	1	2	3
sgRNA1	CAGAGAGGAGATGTCTGACA <u>AGG</u>	-	50.0	67.8	1	0	0	0
sgRNA2	TTGAGCCGGTCGGATCAGAG <u>AGG</u>	-	60.0	77.9	1	0	0	0

LcyE (Os01g0581300) gene

→ donor and primers region, 1116 bp

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atCCCGGGCTGGAACCTCTCAAAGTCCAGTCATCAATGCTGGTAAGCATTCTCTGAT
-> : Fw1(J169)
ATTATTCAATTATTGACAAGCACACAGTATTACAAATTGGAGTACATTGCTTCAGA
TGAGGGCAGTATTAGTATAATTGGAAATGTCAGAACATTCTGGTACTAATTGC
TATGCGGTGTTGCAGCATGGAAACATTGGCCCAAGAACGGAACGTCAACGATCAT
TCTTCCTTTGGCTGGTTGATAATCCAACGTAAACGAAGGCATTGACAGACATTCT
TTGAAACCTTTCCGGTGCCTAACATGTAATTCTACTCTTGTATTGCTCTG
TTTCAGTCTATTACAAATACCATTATGTATGACCTGAAGATTGCCACCAAAAGGT
TCATCACTGTTCTTGTACTATCAGGATGTGGCAGGATTCTGGTGCACGCTTCT
TCAGTGGATCTCATCTTGCATTCTACATGTTCACAATTGGCGGAACCAAATGCGA
ATGAACCTTGTCAAGACTCTGCTCGGATCCGACCGGCTAACGATGATCAAGACCTAC
<- : Rv1(J171)          -> : Fw2(J173)
CTGACCTTGAAACCATTCAGCAGTCTACAAGAATTAGGAAATGTACAGTTGTA
<- : Rv2(J172)
GTTTGACATAACATAGTGAGAGCCAGAGGATATGGGGTTGGGGGTTACTTATCATGC
TAGAACACACAAACTGCAAGAATTATGCATGAATTGGCAAATGGAATAGATTATG
CAGAATGGAACACGTAAACGTGATGGTGTGCTGGAAACACAAGTAGGAGGAATAAA
ACCCATGGATTATGGATCTGTAGTGAGTTGCATTAGCCATTCTAGGTCTTATCATC
TCTCTCCCCCTTCCATTTCACCTCTATCCGGTTTCGACATGCCGATTACACCCCTA
TTTAGATAGCTGGCCAAATGTTCCACAAACATTAGAAAAGGAACAAATGGAAAA
GAGGAGAGAAAAAAAGAGATGGTCAAGAATGTCAGGCTATCCTGCTCCATTGCC
ATGGCATTAGGTCCAGAAGAAGGGTAGAga
<- : Rv3(J178)

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○ primer SET 1 : 555bp Section①

OLIGO start length tm gc% any 3' seq
 LEFT PRIMER 5 20 60.63 55.00 4.00 1.00 GGCCTGGAACCTCTCAAAGT
 RIGHT PRIMER 553 20 59.13 40.00 4.00 2.00 TGACAAGGTTATTGCATT

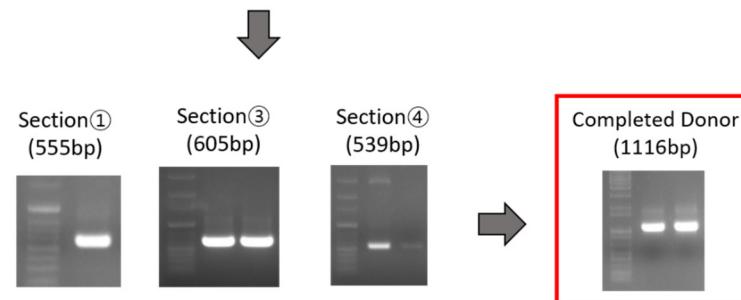
○ primer SET 2 : 84bp Section②

OLIGO start length tm gc% any 3' seq
 LEFT PRIMER 2 18 62.92 50.00 4.00 2.00 ATTGCGCCGAACCAAATG
 RIGHT PRIMER 85 22 58.20 45.45 6.00 2.00 CAGGTAGGTCTTGATCATCGTT

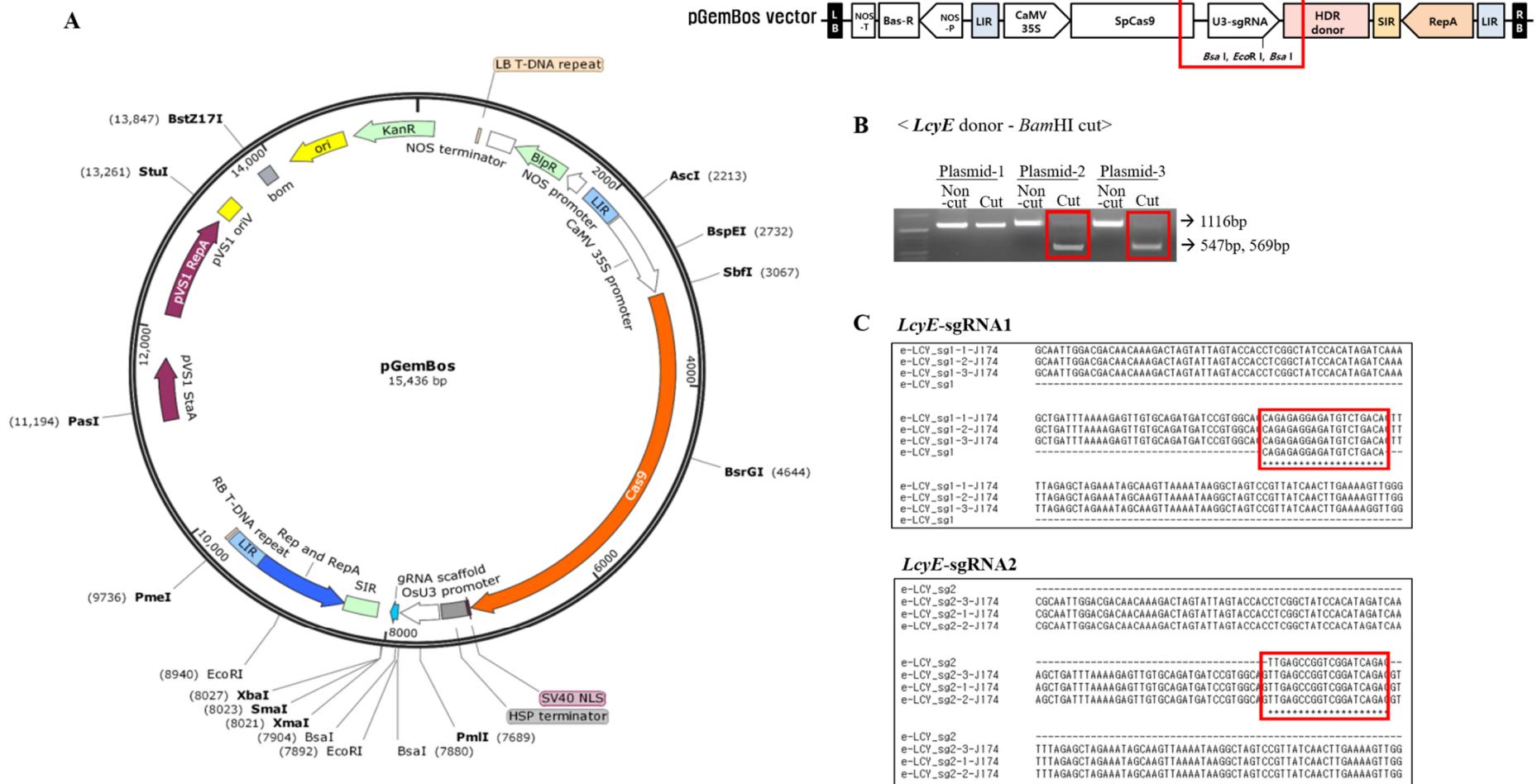
○ Section① + Section② = Section③

○ primer SET 3 : 539bp Section④

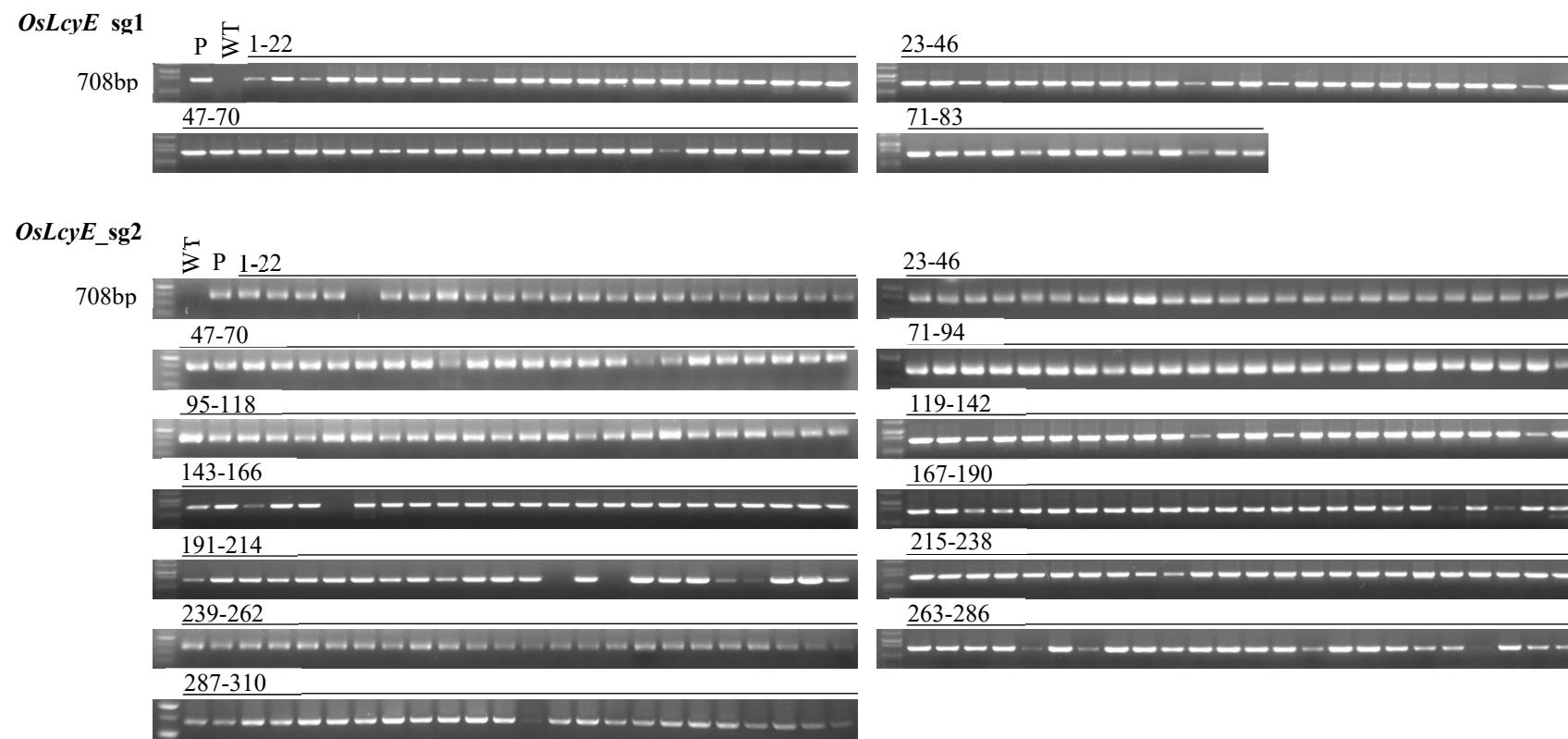
OLIGO start length tm gc% any 3' seq
 LEFT PRIMER 7 19 60.36 52.63 6.00 2.00 CGGCTCAACGATGATCAAG
 RIGHT PRIMER 543 23 61.79 52.17 6.00 1.00 TCTAGACCCCTTCTGGGACCT



Supplementary Figure S1. Design of donor template using the CRSPR-Cas9 and geminiviral replicon system.



Supplementary Figure S2. Ti-plasmid vector construction using the CRSPR-Cas9 and geminiviral replicon system (A) pGemBos vector construction (B) Confirmation of donor template into pGemBos::*LcyE* vector (C) Confirmation of sgRNA by sequencing analysis.



Supplementary Figure S3. Confirmation of T-DNA insertion from calli obtained through HDR experiment. M: 1kb DNA ladder; WT: wild-type; P: pGemBos::*LcyE* plasmid vector.

Supplementary Table S2. The primers list used in this study.

Primer name	Sequence (primer direction 5'-3')
<i>LcyE</i> donor 1st Fw	TCCTGCAACCGGTACTAACAA
<i>LcyE</i> donor 1st Rv	GTGGTGCAAGGAAGGAGAAG
<i>LcyE</i> donor 2nd Fw	ATCCCAGGCCTGGAACCTCTCAAAGTTCC
<i>LcyE</i> donor 2nd Rv	TCTCTAGACCCTTCTGGACCTAAA
<i>LcyE</i> donor OE-PCR Fw1	CGGCTCAACGATGATCAAG
<i>LcyE</i> donor OE-PCR Rv1	TGACAAGGTTCATTCGCATT
<i>LcyE</i> donor OE-PCR Rv2	CAGGTAGGTCTTGATCATCGTT
T-DNA confirm-Nos ter Fw	TTGCGCGCTATATTTGTTTT
T-DNA confirm-Bar R Rv	CGTCAACCACATACATCGAGA
Replicon confirm-Rep Fw	TTCTCCCAGAGAAACTGGAA
Replicon confirm-35S Rv	CCATCTGTGGGTTAGCATT
NGS (Next Generation Sequencing) primers	
<i>LcyE</i> 1st Fw	TTCAATTCTGCAAATTCTTCA
<i>LcyE</i> 1st Rv	TCATTTTGGTCTAGTTACACATCA
<i>LcyE</i> 2nd Fw	ACACTCTTCCCTACACGACGCTTCCGATCTCTTTGGCGTATGAGCAG
<i>LcyE</i> 2nd Rv	GTGACTGGAGTTCAGACGTGTGCTCTCCGATCTGGTCATGTCCTCGGTGAAC
<i>LcyE</i> 3rd-1	AATGATA CGGCG ACCACCGAGATCTACACtatacgctACACTCTTCCCTACACGAC
<i>LcyE</i> 3rd-2	AATGATA CGGCG ACCACCGAGATCTACACataggcACACTCTTCCCTACACGAC

<i>LcyE</i> 3rd-3	AATGATA CGGC GACC ACCGAG ATCTACAC Cctatc ctAC ACTCTT CCCTAC ACGAC
<i>LcyE</i> 3rd-4	AATGATA CGGC GACC ACCGAG ATCTACAC Cggctctga AC ACTCTT CCCTAC ACGAC
<i>LcyE</i> 3rd-5	AATGATA CGGC GACC ACCGAG ATCTACAC Caggcga ag AC ACTCTT CCCTAC ACGAC
<i>LcyE</i> 3rd-6	AATGATA CGGC GACC ACCGAG ATCTACAC taat ctt a AC ACTCTT CCCTAC ACGAC
<i>LcyE</i> 3rd-7	AATGATA CGGC GACC ACCGAG ATCTACAC Caggacgt AC ACTCTT CCCTAC ACGAC
<i>LcyE</i> 3rd-8	AATGATA CGGC GACC ACCGAG ATCTACAC gtactg ac AC ACTCTT CCCTAC ACGAC
<i>LcyE</i> 3rd-9	CAAGCAGA AGAC GGGCATAC GAGA Tcgagta at GTGACT GGAG TT CAGAC GTGT
<i>LcyE</i> 3rd-10	CAAGCAGA AGAC GGGCATAC GAGA Ttctcc gga GTGACT GGAG TT CAGAC GTGT
<i>LcyE</i> 3rd-11	CAAGCAGA AGAC GGGCATAC GAGA Taat gaggc GTGACT GGAG TT CAGAC GTGT
<i>LcyE</i> 3rd-12	CAAGCAGA AGAC GGGCATAC GAGA Tggaat ct GTGACT GGAG TT CAGAC GTGT
<i>LcyE</i> 3rd-13	CAAGCAGA AGAC GGGCATAC GAGA Tttctgaa GTGACT GGAG TT CAGAC GTGT
<i>LcyE</i> 3rd-14	CAAGCAGA AGAC GGGCATAC GAGA Tacgaa tt GTGACT GGAG TT CAGAC GTGT
<i>LcyE</i> 3rd-15	CAAGCAGA AGAC GGGCATAC GAGA Tagttc ag GTGACT GGAG TT CAGAC GTGT
<i>LcyE</i> 3rd-16	CAAGCAGA AGAC GGGCATAC GAGA Tgcgattta GTGACT GGAG TT CAGAC GTGT
<i>LcyE</i> 3rd-17	CAAGCAGA AGAC GGGCATAC GAGA Tcatggc g GTGACT GGAG TT CAGAC GTGT
<i>LcyE</i> 3rd-18	CAAGCAGA AGAC GGGCATAC GAGA Ttcg cgg a GTGACT GGAG TT CAGAC GTGT
<i>LcyE</i> 3rd-19	CAAGCAGA AGAC GGGCATAC GAGA Tgcg caga GTGACT GGAG TT CAGAC GTGT
<i>LcyE</i> 3rd-20	CAAGCAGA AGAC GGGCATAC GAGA Tctatcg ct GTGACT GGAG TT CAGAC GTGT
