

Table S1. The primer sequences used in qRT-PCR.

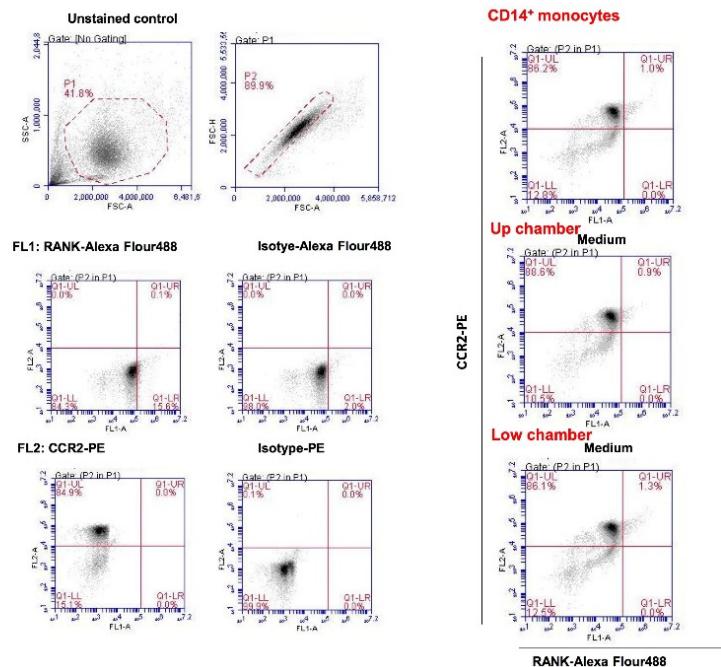
Oligo Name	Oligo Seq
hu-WNT1(F)	TCTTCGGCAAGATCGTCAAC
hu-WNT1(R)	AGTCACACGTGCAGGATT
hu-WNT2(F)	AACAGAGCTGGCAGGAAG
hu-WNT2(R)	AGAGATAATGCCCGTTTCC
hu-WNT2B(F)	CGAGAGGCAGCTTTGTATATG
hu-WNT2B(R)	CCAGTCAAAGTCCCCACG
hu-WNT3(F)	GTGTTAGTGTCCAGGGAGTTC
hu-WNT3(R)	CATTTGAGGTGCATGTGGTC
hu-WNT3A(F)	ATCAAGATTGGCATCCAGGAG
hu-WNT3A(R)	CAATGGCGTGGACAAAGG
hu-WNT4(F)	GTGCCAGTACCAAGTTCCG
hu-WNT4(R)	CACACCTGCCGAAGAGATG
hu WNT5A(F)	CCGGTACTAGCTAACTCCAA
hu WNT5A(R)	CACCATTCCACAGAGAGAGA
hu-WNT5B(F)	AAGGAGTTGTGGATGCC
hu-WNT5B(R)	GCTACGTCTGCCATCTTATACAC
hu-WNT6(F)	GAGAGTGCAGTCCAGTTC

hu-WNT6(R)	TGATGGCGAACACGAAGG
hu-WNT7A(F)	AAGGTCTTGATGCC
hu-WNT7A(R)	GCACTTACATTCCAGCTTCATG
hu-WNT7B(F)	GGATCATGCACAGAAACTTCG
hu-WNT7B(R)	GCTAGGCCAGGAATCTTGT
hu-WNT8A(F)	CGAAAATGTGGCTGTGATG
hu-WNT8A(R)	CTTCCCCTCTCCAAACTGTC
hu-WNT8B(F)	GTACACCCTGACTAGAAACTGC
hu-WNT8B(R)	CAAAC TGCTGGAAATCGCC
hu-WNT9A(F)	CAGCAAGTTCGTCAAGGAATT
hu-WNT9A(R)	TGCATGAGCCTGACACG
hu-WNT9B(F)	AGTGCCAGTTTCAGTTCCG
hu-WNT9B(R)	GGAAAGCTGTCTTTGAAGC
hu-WNT10A(F)	CTCCTGTTCTCCTACTGCTG
hu-WNT10A(R)	CACTGTGTTGGCATTGAGC
hu-WNT10B(F)	TCTCGGGATTCTTGATTCC
hu-WNT10B(R)	CATTCCGCTTCAGGTTTCAG
hu-WNT11(F)	CCAAGCCAATAAACTGATGCG
hu-WNT11(R)	GCACTTACACTTCATTCCAGAG

hu-WNT6(F)	CAGAAAGATGGAAAGGCACC
hu-WNT16(R)	ATCATGCAGTCCATCTCTC
hu-RANK(F)	CCATCATCTTGGCGTTG
hu-RANK(R)	AGCTGTGAGTGCTTCCT

Figure S1. The percentage of osteoclast precursor and the non-viable cell in this migration study was analyzed by multi-color flow cytometry. CD14⁺ magnetic microbeads were used to identify peripheral monocytes. 1X10⁵ CD14⁺ monocytes were added to the upper chamber. The CD14⁺ monocytes migrated into the lower chamber for one hour. Osteoclast precursor was characterized using RANK and CCR2. The secondary antibodies were conjugated with different fluorescence (RANK-Alexa Flour488 and CCR2-PE-A). The isotypes of individual antibodies were used as negative controls. The gating line is set according the corresponding isotype antibody. (B) To determine the percentage of non-viable cell in the migration study. The peripheral CD14⁺ monocytes, CD14⁺ monocytes in upper chamber and CD14⁺ monocytes in lower chamber after migration study, 7-Aminoactinomycin D (Cayman, Ann Arbor, Michigan, US) was used to detect non-viable cells.

(A)



(B)

