



Review

Defining the Role of Monocytes in Sjögren's Syndrome

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Abstract: Sjögren's syndrome is one of the most prevalent autoimmune diseases after rheumatoid arthritis, with a preference for middle age, and is characterised by exocrine glandular involvement leading to xerostomia and xerophthalmia. It can have systemic implications with vascular, neurological, renal, and pulmonary involvement, and in some cases, it may evolve to non-Hodgkin's lymphoma. For a long time, B- and T-lymphocytes have been the focus of research and have been considered key players in Sjögren's syndrome pathogenesis and evolution. With the development of new technologies, including omics, more insights have been found on the different signalling pathways that lead to inflammation and activation of the immune system. New evidence indicates that a third actor linking innate and adaptive immunity plays a leading role in the Sjögren's syndrome play: the monocyte. This review summarises the recent insights from transcriptomic, proteomic, and epigenetic studies that help us to understand more about the Sjögren's syndrome pathophysiology and redefine the involvement of monocytes in this disease.

Keywords: Sjögren's syndrome; monocytes; inflammation; epigenetics; proteomics; RNA sequencing



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1. Introduction

Sjögren's syndrome (SS) is a systemic autoimmune disease of unknown origin mostly observed in middle-aged or elderly female adults [1] with a peak incidence between 50 and 60 years [2]. SS [3] can occur independently of other pathologic conditions (primary SS, pSS) or in the context of other autoimmune diseases, mostly rheumatoid arthritis, systemic lupus erythematosus, or systemic sclerosis (secondary SS) [4].

SS is mainly characterised by involvement of exocrine glands, leading to decreased tear production and xerophthalmia (ocular dryness) or decreased salivary flow production and xerostomia (oral dryness) [5]. Fatigue and joint pain are common complaints referred by SS patients and, in most severe cases, internal organs or apparatus can be affected as well. SS can indeed involve the kidneys (tubulointerstitial nephropathy and glomerulonephritis), the lungs (interstitial pneumopathy), the neurological system (central or peripheral), and the vascular (vasculitis phenomena) or the haematological system, also increasing the risk of lymphoma [6].

Similarly to other systemic autoimmune diseases, a serum autoantibody positivity and signs of immune system activation and deregulation can be found in most SS patients [7]. An antinuclear antibody (ANA) positivity can be observed in >80% of cases and extractable nuclear antigen (ENA) antibodies in SS patients directed against Ro/SSA and La/SSB antigens can be observed in 33–74% and 23–52% of cases, respectively [8]. Additionally, a serum positivity for the rheumatoid factor (RF) can be observed in 36–74% of patients who also may have hypergammaglobulinemia and indirect signs of B-cell activation, especially in conjunction with anti-Ro/SSA and/or anti-La/SSB positivity [8].

SS is an orphan disease, which is a disease that affects only a very small number of individuals or is neglected by physicians and pharmaceutical companies [9,10], and

currently, there is no targeted therapy available to globally tackle this condition. Almost all existing therapies aim at resolving or relieving specific symptoms or specific organic complications [4].

The main cell involved in this disease is the B-lymphocyte, which is supposed to have an altered function, being hyper-stimulated by factors/proteins such as the B-cell activating factor (BAFF/BLyS) [11]. The prolonged B-cell activation and proliferation is the key driver of exocrine gland involvement and may result in the appearance of lymphoma, which is one of the most prevalent causes of mortality in SS. As the immune system is composed of several subtly interlaced compartments, more and more studies are shedding light on the role of T-lymphocytes as mediators in the perpetuation of this immune response or as key activators of B-cells [12].

Monocytes are cells that play a crucial role in innate immunity, serve as a link to the adaptive immune system, and have been implicated in the development of autoimmune diseases, infiltrating target organs, being altered in number and in function, or releasing cytokines and chemokines with pro-inflammatory, proliferatory, and regulatory functions [13]. Monocytes in SS are, thus, likely part of a puzzle that has yet to fit in. Thanks to the evolution of omics (genomics, proteomics, and metabolomics) [14] and epigenetics (DNA methylation, micro-RNA, and histone modification) [15,16], concepts about the role of different immune actors—including monocytes—in SS are actively updated and revised. The purpose of this review is to investigate the role of monocytes and their different subpopulations in the onset and development of SS, as well as to highlight future therapeutic targets.

2. Monocytes

Monocytes are cells that are central both to the innate and to the adaptive immune system and participate in homeostasis and inflammatory responses [17]. Monocytes develop in the bone marrow and, from there, move to the circulatory system to exert their function [18]. They can originate both from monocyte-granulocytic and dendritic-monocytic progenitor cells and, thus, a variant plasticity is attributed to this cell. In relation to the different markers expressed on their surface, we can distinguish different subfamilies of monocytes, each characterised by peculiar functions [19]. In detail, the main subfamilies of monocytes are:

2.1. Classical ($CD14^{++}$, $CD16^{-}$)

Classical monocytes are by far the most represented population of monocytes in blood (about 80–90%) and are considered inflammatory [20]. In humans, classical monocytes highly express CD14 and lack CD16; they also express the C-C chemokine receptor type 2 (CCR2 or CD192), a molecule necessary to exit from bone marrow to the circulation. Other surface markers have been described to foster a more precise definition and isolation of classical monocytes [21], including the increased expression of the scavenger receptor CD36 [22] and the reduced expression of the CD11c [23]. Within this subpopulation, different subtypes can be further distinguished in relation to other surface markers and function. Classical monocytes expressing the CD103 have tumour-antigen-presenting functions [24]. MHCII+, SCA-1+, and CXCR3R1- monocytes can produce prostaglandin E2 (PGE₂) and interleukin 10 (IL-10) in the case of infection [25]. Dendritic cells (DCs) from MHCII+ and CD209a+ monocytes [26] also represent a subpopulation of monocytes found in small numbers (5%) and described in inflamed peripheral tissues, but not in lymphoid tissue, and is partially CCR2-dependent. These monocytes are differentiated via interferon gamma (IFN- γ), which, in this case, is secreted by natural-killer (NK) lymphocytes in response to inflammation. They play a co-stimulatory role with CD8+ T-lymphocytes. Their function is superimposable to that of type 2 DCs, which explains why, in the absence of these, the latter, monocytic-DC determines its DC function and can even present the antigen to lymphocytes, although with less efficiency. Segregated-nucleus-containing atypical monocytes (SatMS) CD115+ monocytes have been shown to appear de novo

in experimental models of fibrosis after exposure to bleomycin [27], being critical for fibrosis development.

2.2. Nonclassical ($CD14^{-}$, $CD16^{++}$)

Nonclassical monocytes express high levels of the adhesion-related-receptor C-CX-C motif chemokine receptor 1 (CX3CR1) and actively patrol the vasculature both at steady state and during inflammation to remove debris [28] and produce high levels of anti-inflammatory cytokines and pro-wound-healing factors [29]. They exert patrolling functions under either NOTCH2 and/or toll-like receptor 7 (TLR-7) stimulation in inflammatory conditions [30]; patrolling is, in part, dependent on the expression of the lymphocyte-function-associated antigen 1 (LFA1) on monocytes [31].

Within this population of monocytes, several subsets have also been described [32], including a CD61+ and 6-Sulfo LaNAc (Slan)-positive subset associated with coronary artery disease severity or a CD9+ subset involved in platelet adhesion. A slan- subtype has also been described, although it does not seem to be functionally different from the slan+ subset at the transcriptional level [32]. Additionally, a subset of monocytes expressing the angiogenic tyrosine-protein kinase receptor Tie-2 has been described to promote angiogenesis in tumours, possibly playing a role in human cancer progression [33,34].

2.3. Intermediate ($CD14^{+}$, $CD16^{+}$)

A third major subtype that is halfway between classical and nonclassical monocytes has consistently been described. This subset is part of a continuum as classical monocytes can re-convert into nonclassical monocytes, with intermediate monocytes representing a transition point: a classical monocyte in 1–2 days can become an intermediate after a stimulus and subsequently transform into a nonclassical monocyte [35]. Intermediate monocytes highly express antigen-presentation molecules [32] and CCR5 to a higher extent as compared to classical monocytes [36] and are involved in antigen processing and presentation and transendothelial migration.

3. Monocyte Activation

After an inflammatory stimulus, a complex system with many factors starts to operate to initiate emergency myelopoiesis [37]. The signals that trigger this process include cytokines and TLRs that directly differentiate macrophages. These signals influence haematopoietic stem cells and multipotent progenitor cells; as a consequence, the number of myelocytes increases, which are released from the bone marrow into the peripheral blood via CCR2 signalling. From this moment on, inflammatory markers increase (nitric oxide, IFN pathway, tumour necrosis factor (TNF), and Janus kinase-signal transducer and activation of transcription (JAK-STAT) pathway), leading to monocyte differentiation under the drive of CD8-lymphocyte stimuli (IFN- γ) [37].

During inflammation, classical monocytes can differentiate into CD11a+ MHCII+ monocytes (macrophages) or monocyte-derived dendritic cells (MoDCs), which can acquire protective or pathological functions [18]. These monocytes depend, among other molecules, on the interferon receptor factor 5 (IRF-5) signalling to differentiate into Podophyllotoxin and Rutin Modulate M1-positive (iNOS+)-producing cells; if this does not happen, they can further differentiate into M2-type CD206+ macrophages with anti-inflammatory properties [38]. Therefore, IRF5 promotes the emergence of M1 (inflammatory) and represses the M2 (anti-inflammatory) lineage, as a result of which pathogenic monocytes or macrophages may develop. In the presence of chronic or sustained inflammation, monocytes may differentiate into macrophages with haemophagocytic activity, leading to the development of a macrophage activation syndrome (MAS) [39].

4. Interferon Signature in Sjögren's Disease

The increased expression of type-1 IFN-regulated genes is commonly referred to as the "interferon signature" [40]. The type-1 IFN pathway mainly stimulates the IFN α/β

Receptor (IFNAR) that, downstream, activates the JAK-STAT pathway (Jak1 and Tyk 2), and phosphorylates STAT 1 and 2 together with the mitogen-activated protein (MAP) kinase pathway (Erk 1 and Erk 2) [34].

IFN-1 inhibits viral replication, activates NK lymphocytes and dendritic cells, and maintains the antibody response by B cells [41].

Each type of monocyte reacts differently to IFN stimulation; indeed, after IFN stimulation, the expression of STAT1, as well as the surface expression of CD169 and CD64, is higher in classical monocytes as compared to the other subsets [42]. While nonclassical monocytes that express CD169 on their surface show less affinity/sensitivity for the interferon signature [34], they also show a decrease in STAT1, which results in a decrease in IFNAR1 and its related proteomics [43]. Part of its anti-inflammatory function is due to its role as a cell debris scavenger, thus preventing a continued TLR-mediated IFN-1 stimulation [44].

The CCR2-CCL2 molecule mediates the recruitment of classical monocytes following inflammatory stimuli [45], which, together, with the production of the IFN signature, creates the ideal environment for the development of autoimmune diseases. For instance, in systemic lupus erythematosus (SLE), it was found that monocytes were hypersensitive to the IFN receptor signal, thus increasing the number of classical monocytes and the classical vs. nonclassical monocytes ratio [34]. These alterations create an inflammatory environment that is uncontrolled by nonclassical patrolling functions and that further promotes autoimmunity.

In pSS, there is evidence of an upregulation of type-1 IFN genes in circulating monocytes; this finding is consistent with the description of IFN activation in salivary gland monocytes and peripheral blood mononucleated cells (PBMCs) [41]. Additionally, type-1 IFN was found to induce the expression of BAFF expression in monocytes and salivary gland epithelial cells [46]. In this same study, an attempt was made to find a link between BAFF and IFN-1 expression. It was found that after IFN-1 stimulation, monocytes (CD14+) increased BAFF mRNA levels and, once the stimulus was blocked, BAFF levels decreased, although BAFF expression could not be correlated with IFN-1 levels nor serum BAFF be correlated with BAFF mRNA in monocytes or PBMC.

Plasmacytoid dendritic cells (pDCs) are the main producers of type-1 IFN [41]; in pSS, their blood levels were found to be low, most likely as a consequence of margination and migration to exocrine glands [47]. pDCs react to exogenous (viral DNA/RNA) or endogenous stimuli, such as immunocomplexes of nuclear antigens and antibodies (a pSS hallmark) that bind to FcγRIIa, causing its internalisation and intracellular binding to TLR7 and TLR9 [48]. This pathway of IFN-1 production due to immunocomplex binding may provide an explanation for the association between type-1 IFN values with high autoantibodies, IgG production, and low complement in pSS.

5. Monocytes in Sjögren's Syndrome

Monocytes in SS may be studied in different compartments and by different methods; we can either study their presence by histological (salivary gland biopsy and eye biopsy) or blood (peripheral blood, PBMC) samples [49].

In salivary gland biopsies (Figure 1), an inflammatory infiltrate initially composed of CD4+ T-lymphocytes has been described, which, as the disease progresses, gives way to B-cells infiltrates that eventually lead to the formation of germinal pseudonuclei [50]. Within these nuclei, macrophages/monocytes as well as dendritic cells, in addition to lymphocytes, can be found. Their infiltration into the tissue is variable depending on the stage of the disease or the type of cell labelling. Nonetheless, the degree of infiltration was found to correlate positively with the degree of infiltration and involvement of the gland [49]. Macrophages arguably constitute a nexus between innate and adaptive immunity, connecting the different players in glandular involvement. Macrophages secrete a variety of inflammatory cytokines and mechanisms (including IL-18, IL-1, and MCP-1) and proteases in the process of monocyte-macrophage differentiation, activation of lymphocytes, or

TLRs of innate immunity [49]. On the other hand, TCD4+ lymphocytes are stimulated by local dendritic cells via MHCII and activate tissue-resident macrophages through the production of IFN- γ and other cytokines [51]. This process increases the inflammatory infiltrate and eventually increases the size of salivary glands and leads to trophic changes into the affected tissues.

In the case of keratoconjunctivitis sicca (ocular involvement), elevated IL-1 α and IL-1 β can be seen in the conjunctival tissue of pSS, causing pathological keratinisation of the eye [52]. This interleukin activates the MAPKinase pathway (p38 MAPK), which leads to the transcription of cyclic AMP (inflammatory pattern), maintaining the activation of resident cells based on CD4+ infiltration.

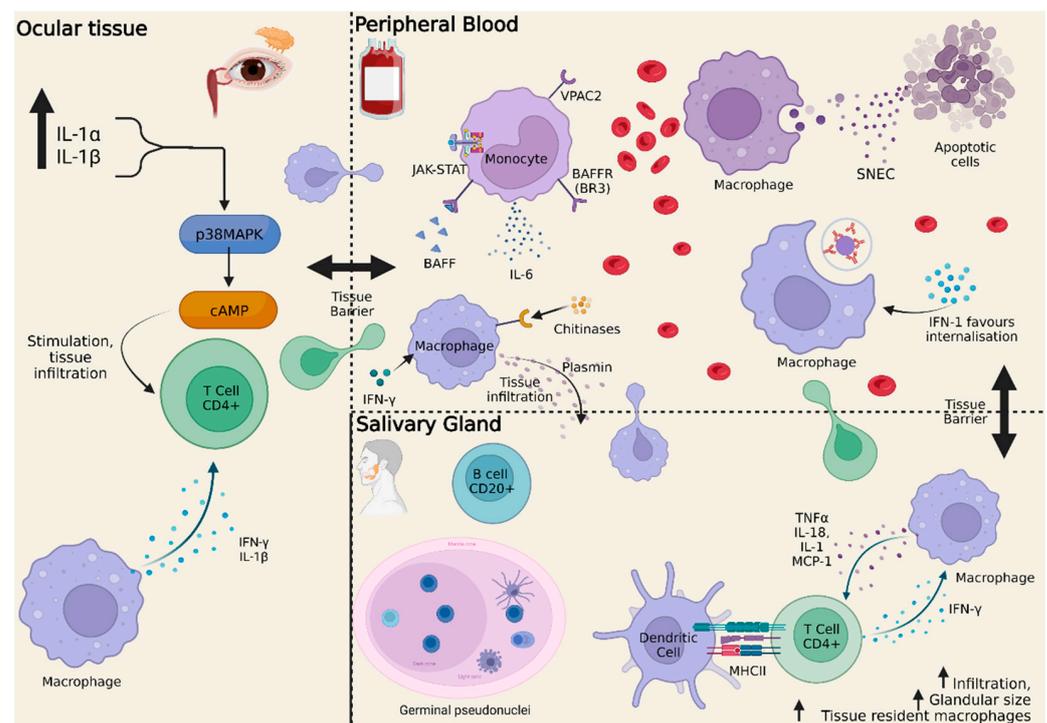


Figure 1. Monocyte and macrophage implication in Sjögren's Syndrome. This figure shows how the monocyte/macrophage infiltrates peripheral tissues, namely ocular and salivary, interacting with B and T cells, and sustaining inflammation, disease progression, and organ disease damage. IL-1 α , Interleukin-1 α ; IL-1 β ; IL-18; IL-6; p38 MAPK, p38 MAP-Kinase; cAMP, cyclic Adenosine monophosphate; IFN- γ , Interferon- γ ; IFN-1; TNF α , Tumour necrosis factor α ; MCP-1, Monocyte chemotactic protein 1; SNEC, secondary necrotic material; MHCII, Major histocompatibility complex type II; JAK, Janus-kinase; BAFF, B cell activating factor belonging to TNF family; VPAC2, Vasoactive intestinal peptide receptor. Created with www.BioRender.com. Accessed on 17 October 2022.

The balance between classical and nonclassical monocytes is fundamental to regulate inflammation. As described above, nonclassical monocytes have the function of removing apoptotic cell debris, thus preventing antigen presentation and immune system activation [28,53]. In extreme conditions, a prolonged presentation of (auto) antigens may sustain autoimmunity. This is the case, for instance, of SLE where the clearance of apoptotic debris, including alarmins, via DNase 1 is defective [54]. The accumulation of these molecules, called secondary necrotic material (SNEC), is phagocytosed by macrophages and neutrophils, which, upon activation, secrete cytokines and stimulate intracellular DNA sensors and RNA-associated nucleoproteins [55]. Both in pSS and in SLE, macrophages are overstimulated by a favourable serum environment that leads to a higher phagocytosis than under normal conditions [54]. In pSS, a significant correlation has indeed been found between defective DNase and SNEC degradation, and disease activity was found to inversely correlate to DNase levels [54]. As part of a complex system, autoantibod-

ies also play a crucial role in preventing the degradation of SNEC compounds against endonucleases, creating a kind of protective film that keeps the antigen intact until phagocytosis [56]. Additionally, type-1 interferon-related pathways favour the internalisation of immunocomplexes, facilitating the appearance of nuclear self-antigens [57].

Other factors produced by macrophages/monocytes in local tissues may also play an essential role in the pathogenesis of SS, including chitinases, plasmin, and cathepsins. Aberrant expression of human chitinase genes, including the chitinase-3-type-1 (CHI3L1/YKL-40) and the chitinase 1 (chitotyrosidase/CHIT1) gene, has been demonstrated in histological salivary gland biopsy specimens with increased expression corresponding to a more severe disease in pSS [58]. This process is involved in monocyte adhesion as well as differentiation into macrophages, and may be a direct activator of infiltration and disease development. Plasmin, a serum protease product of plasminogen activators, is actively produced by macrophages after stimulation by IFN γ ; it can lead to fibrinolysis and is capable of activating metalloproteases, which weaken the matrix of the affected tissue and facilitate lymphocyte invasion, thereby establishing tissue damage [59]. Cathepsins, also proteolytic in nature, are endosomal/lysosomal peptidases, which are activated at low pH [60]. Increased cathepsin S, as well as cathepsin H, has been found in the lacrimal glands together with CD68+ macrophages [61].

Another molecule of interest in pSS pathogenesis is the vasoactive intestinal peptide (VIP); this molecule generates immunomodulation via its receptors VPAC1 and VPAC2 in macrophages, monocytes, and T-lymphocytes [62]. pSS monocytes express increased VPAC2, whose presence is associated with impaired phagocytosis [63]. In a study with NOD mice with Sjögren-like manifestations, VIP administration was tested, restoring salivary secretion and reducing markers of autoimmunity [64]. At the cellular level, a change in the proportion of predominantly M1 macrophages (pro-inflammatory) to M2 (noninflammatory) was also observed. Moreover, classical monocytes located in salivary glands after phagocytosis show increased TNF- α levels, that is, the opposite of what is usually observed in normal conditions where monocytes/macrophages after phagocytosis of apoptotic cells secrete anti-inflammatory cytokines, with low levels of TNF- α and high levels of IL-10.

The autoimmune regulator (AIRE) is a gene whose gene transcription factor is mainly found in the thymus and whose protein coordinates the expression and presentation of tissue-specific self-antigens, thus constituting an essential role for the depletion of autoreactive lymphocytes in the thymus [65]. It has been observed that AIRE knock-out mice (KO) can develop an autoimmune disease mediated by CD4+ lymphocytes that can affect multiple organs, including exocrine glands, which is superimposable to pSS. Keratoconjunctivitis sicca, xerophthalmia, and peripheral neuropathy, among others, have been reproduced in these mice. In the glandular infiltrates of these mice, a heterogeneous population of immune cells has been observed, comprising CD4+, CD8+ and B-lymphocytes, DCs, as well as macrophage/monocytes. Macrophages/monocytes of these infiltrates highly express pro-inflammatory cytokines such as IL-1 β and IFN- γ in the ocular tissue.

Monocytes tightly interact with B-lymphocytes, a major player in pSS pathogenesis [66]. Several studies have shown that the BAFF receptor BR3 is over-expressed in peripheral monocytes, especially classical monocytes, which can actively secrete IL-6 after BAFF binding, and this mechanism is blocked by anti-BAFF antibodies [67]. Such BR3 expression may play a role in the expression of immunoglobulins as well as autoantibodies; in fact, exposure of monocytes to anti-IL-6 has been tested, inhibiting the levels of this interleukin. This finding suggests that soluble factors that are produced by BAFF-stimulated monocytes are involved in the production of immunoglobulins by IL-6-activated B-lymphocytes [68].

In addition to the IFN pathway, MAPKinases and the JAK-STAT pathway have gained prominence either as key players in the pathogenesis of pSS or as therapeutic targets [69]. The JAK-STAT pathway mediates cytokine responses, including IL6, IL-7, IL-10, IL-12, IL-17, IL-21, and TNF α , all implicated in the pathogenesis of pSS. The JAK-STAT pathway

is involved in IFN signalling as STAT tyrosine phosphorylation follows JAKs activation downstream of IFN receptors [70]. A recent study has analysed STAT1, 3, 4, 5, and 6 in peripheral blood, in T- and B-lymphocytes and monocytes in patients from pSS and healthy volunteers [71]. Phosphorylated STAT5 in monocytes as well as in B-lymphocytes strongly correlated with IgG and anti-SSB/La serum levels and, from the clinical point of view, with the presence of purpura but not with other clinical features. Regarding the other STAT molecules, STAT4 was shown to be predisposed to pSS and to affect the type-1 IFN pathway, while STAT1 and 3 can be found overexpressed at the mRNA level in PBMC.

5.1. Transcriptome Findings

In SS, recent transcriptome studies have been performed to investigate potential pathways of interest and to elucidate the pathogenesis of the disease [46]. In RNA, from CD14+ monocytes, statistically significant differences in TNFSF10 (TRAIL) expression were found in patients with pSS vs. healthy controls. The following differentially expressed genes (DEGs) were also highlighted: TMEM176B, TMEM176A, HLA-DRB5, FOS, TXNIP, ARPC1B, GRN, FGL2, SAMHD1, CEBPD, CTSZ, HLA-DQB1, SNX17, TNFSF10, WASF2, ATP5A1, ZFP36L2, and CORO1A. Among them, HLA DRB5 and TNFSF10 play a key role in the pathogenesis of many autoimmune diseases [72,73]. Enrichment/stimulation analysis of these DEGs identified neutrophil activation and IFN-associated pathways.

More studies [74], comparing the bulk transcriptome in CD14+ genes from SLE and pSS patients, showed that inflammatory as well as IFN pathways were enriched in both diseases. A propensity toward M1 macrophage differentiation appeared to be prominent in pSS.

In pSS, bulk transcriptome analysis identified a significant number of genes aberrantly expressed in monocytes: TRIM22, MX2, MS4A4A, IFI44, IFIT2, STAT2, SAMD9L, STAT1, EPST11, IFI44L, SIGLEC1, TNFSF10, CX3CR1, and ISG15 [75]. Among them, we can appreciate several associated with the IFN pathway with anti-viral and inflammatory properties (STAT, IFIT, TNFSF, among the others). Increased expression of TNFSF10 (TRAIL) in patient monocytes was identified both in scRNA-seq and bulk transcriptomic analyses. As this gene is expressed in many of the identified monocyte subsets, these data reinforce the previously described finding that TNFSF10+ may play a key role in the pathogenesis of pSS, as also suggested by other studies [76,77].

Similarly, comparing the transcriptome of CD14+ monocytes from patients with pSS, non-Sjögren's sicca (nSS), and healthy controls (HC), it was observed that the gene expression of circulating monocytes (especially intermediate and nonclassical monocytes) was highly correlated with ongoing systemic inflammation as result of local damage [78]. Overall, the expression profile of pSS was clearly distinct compared to HC monocytes, but had relatively similar profiles to nSS monocytes. Treatment with serum from pSS patients induced pSS-like transcriptome features in hallmark genes in HC-derived monocytes; these effects were mostly driven by type-1 IFNs.

The literature [41,79] reports similar results describing type-1 IFN genes (IFI44L, IFI44, IFIT3, LY6E, and MX1) overregulated in monocytes with pSS and associated with high disease activity. Other genes are related to the IFN pathway such as IFI27, IFITM1, IFIT4, and IFI44 as well. Sialic acid binding Ig-like Lectin 1 (Siglec-1), which is a biomarker of type-1 IFN activation, was highly expressed in monocytes and could also be correlated with EULAR Sjögren's syndrome disease activity index (ESSDAI) [80]. The study [75] suggested that both the IFN and virus-infection response pathways are over-regulated in pSS monocytes and play a role in its pathogenesis.

5.2. Epigenetic Findings

We refer to epigenetics when there are mitotic modifications that can influence the phenotype without the need to alter the DNA sequence. These are relatively stable changes over time that maintain the cellular identity and arise in response to different internal or

external stimuli. DNA methylation, histone modifications, and noncoding RNAs (miRNAs) are commonly studied among the epigenetic processes.

The role of monocyte miRNAs in certain pathological processes or for cell regulation [81] influencing the heterogeneity of monocytes has recently been the focus of different studies. miRNAs are molecules with a regulatory role in gene expression at the post-transcriptional level [82]. Almost half of them are clustered together and can be transcribed independently or simultaneously [83].

Many studies have found an overexpression of miR-146a/b in PBMCs from patients with pSS [84,85], and a regulatory role in the immune response via negative feedback of TLR signalling has been proposed [86]. Therefore, a dysregulation of miR-146 would promote uncontrolled inflammation, leading to the phenomenon of autoimmunity. Other studies identified overexpression of miR-181a in PBMCs in contrast to the findings in salivary tissue, in which it was found to be underexpressed together with miR-16 [87] leaving open the need for further studies to investigate a possible pathogenic factor.

In light of the involvement of miRNAs in pSS, attempts have been made to establish a sort of "miRNA signature" not dissimilar to the IFN signature found elsewhere [88]. In the study by Williams et al. [88], six miRNAs were found to be simultaneously overexpressed in pSS (miR-34b-3p, miR-300, miR-609, miR-877-3p, miR-3162-3p, and miR-4701-5p).

The involvement of these miRNAs in the transforming growth factor β (TGF β) pathway was studied, recognising that TGF β is an underappreciated pathway and could have a relevant role in the pathogenesis of pSS. Indeed, female TGF β 1-R KO mice develop inflammation in salivary glands, and TGF β 2-R2 depletion in dendritic cells resulted in multi-organ inflammation and activation of autoreactive T- and B-lymphocytes as well as in a mismatched polarisation of alternatively activated M2 macrophages. Key factors in this pathway are TGFBR3 and SMAD2, direct targets of miR-609 and miR-877-3p, respectively. Of note, expression levels in pSS monocytes of SMAD2 and 3 are high, although SMAD4 is usually reduced. Regression analyses indicated that there was a significant association between miR-300 and miR-609 on the reduction in SMAD4 expression, thus being its target and modulating its expression. The MAPKinase pathway may be regulated by pSS-related miRNAs. The direct effect of miR-34b-3p on GRB2, P38/MAPK13, and MEKK1/MAP3K1, as well as miR-877-3p on SOS1, NRAS, ERK1/MAPK1, RAC1, and HGK/MAP4K4, was studied. Likewise, with the JAK-STAT pathway: miR-877-3p regulates STAT6, which codes for the IL-4/IL-13 p transcription factor [88]. No relationship was found with the IL-12 receptor (IL12RB1 and IL12RB2), TYK2, or STAT4, related to IL-12 signalling. Regulation of the TLR/NF κ B pathway was also not found; as communication between the NF κ B and TGF β pathways is essential to coordinate cellular responses and prevent autoimmunity [89], a deficient functioning mechanism between these pathways can trigger pro-inflammatory factors in pSS.

Histones, partners in chromatin compaction in the nucleus, can undergo modifications that will involve one gene expression or another [90]. The N-terminal tails of histones protrude out of the nucleosome and are subject to a variety of post-translational covalent modifications through acetylation and methylation of the lysine residues in histones H3 and H4, the latter being the most studied modifications [91]. Acetylation leads to relaxation of the chromatin conformation in a way that allows transcription, whereas de-acetylation represses transcription by compacting the chromatin. In a review, Imgenberg-Kreuz et al. reported an association between genetic risk variants with promoter and enhancer marks in B-lymphocytes and monocytes [92]. Hypomethylated sites in pSS were observed to accumulate in enhancer regions of T- and B-lymphocytes, while hypermethylated regions predominantly overlapped with a histone mark indicating activation of gene transcription [92]. Despite this evidence, no studies are currently available to analyse histone marks in pSS cells.

Luo et al. [93] proposed that the IFN signature could be detected at the level of DNA methylation and other pathways involved in pSS pathogenesis. DNA methyltransferase 3A (DNMT3A) and methylcytosine dioxygenase translocation 10-11 (TET) play a basic role

in the incorporation and oxidation/removal of methyl groups on cytosines [94]. In the case of the monocyte, DNMT3A and TET are related to differentiation and the inflammatory response, respectively. This is an epigenetic mechanism that could reflect the influence that monocyte-mediated inflammation exerts on pSS. In this study, it was found that circulating monocytes in pSS were predominantly DNA-hypomethylated (299 out of 460 genes, 65%). Hypomethylation in MX1, PARP9, DTX3L, EPSTI1, and IFITM1, which influence the IFN pathway in pSS monocytes, was reported. These findings may support the idea that DNA methylation in IFN-signature-related genes could be a diagnostic tool for pSS in peripheral blood. This hypomethylation trend may be a trigger for dysfunctional activation of its related genes, which could elevate the IFN signature in pSS.

Differentially Methylated Positions (DMPs) corresponding to 12 genes that overlapped in pSS monocytes and salivary gland epithelial cells (SGECs) were also described [93]. These include: PTPRN2, TNK1, WDR8, TSPAN9, VIPR2, OBSCN, KCNT1, ZNF703, NEURL3, LMX1B, LOC146336, and FTSJD2. All these DMPs were related to the cell cycle, senescence and the IL-17 pathway. In the same paper, the methylation status was compared between patients who were single positive (for either anti-Ro/SSA or -La/SSB) and those who were double positive (both antibody specificities). Among single positives, only 54 DMPs related to 27 genes were found, of which 9 showed significant differences in methylation with controls. In contrast, in double positives, 1230 DMPs related to 984 genes were found, and of those, 113 showed differences in methylation in the promoter region. The double positives were related to Ras, Ribosomal, Rap1, and AMP-activated protein kinase (AMPK) signalling pathways, while the single positives only showed the NOTCH pathway. Such NOTCH genes have been described in monocytes from other autoimmune diseases (including rheumatoid arthritis) [95], and their hyperactivity may increase macrophage differentiation and promote the production of pro-inflammatory cytokines [96].

Differentially methylated genes mostly expressed between the two subtypes were in the ribosome and AMPK pathway. The AMPK-STAT3 axis plays a pivotal role in regulating monocyte-to-macrophage differentiation through increased AMPK activity [97], and thus, changes in methylation may influence IgG production by affecting monocyte differentiation. Furthermore, the NOTCH pathway (DT3XL) was highlighted as an enriched pathway in patients with elevated IgG levels.

6. Current and Future Treatments

Current existing drugs for Sjögren's syndrome are not targeted to the treatment of this disease and several compounds developed with other indications have often been used on the basis of personal or anecdotal experience, yet with limited clinical efficacy [98].

Immunosuppressants are widely used in pSS, according to the notion that immunological disturbances can be addressed or, hopefully, reverted by this class of drugs even if their effect is unspecific. For instance, hydroxychloroquine has shown limited utility and no substantial effect in controlled clinical trials [99] despite its potential to interfere with the TLR system and to inhibit type-1 IFN responses. Methotrexate, a potent immune-suppressive drug with multiple effects on both acquired and innate immunity pathways, has been poorly studied in pSS despite its broad use in the rheumatology field. In the few studies conducted in pSS, methotrexate showed disappointing and conflicting results, with beneficial effects on haematological and indirect parameters of immune system activation but not on clinical symptoms [100]. Azathioprine has a long history in the treatment of autoimmune diseases, directly enhancing apoptosis of both memory and naive T cells and secondarily inhibiting B cell activation; in pSS, it is widely used as a steroid-sparing agent, yet in small double-blind randomised clinical trials, it did not show any substantial clinical effect [101]. Cyclosporine, a calcineurin inhibitor, specifically targets T cells and related responses, and has been tested in a small trial in pSS showing some limited benefit only on xerostomia, but not on other clinical parameters [102]. Leflunomide possesses the ability to inhibit the proliferation of B cells and both naive and memory CD4+ T cells; in a small trial in pSS, it ameliorated several biohumoral parameters, including cytokine

and histological alterations, and proved effective in about 50% of patients when clinical domains were globally considered [103,104]. Mycophenolate mofetil due to its tolerance and low side-effects, as well as efficacy in some clinical manifestations of pSS, has been suggested as a potential first-line therapy, yet large studies to confirm and well assess its efficacy are lacking [105].

B cell over-activation is one of the hallmarks of pSS and drugs targeted toward this cell subset have long been considered a key target for intervention. Rituximab, a depletive anti-CD20 monoclonal antibody, was first described as beneficial in some series of pSS patients [106,107], but a substantial effect was not confirmed in a larger controlled study [108]. Belimumab, an anti-BAFF/BLyS monoclonal, seems effective to reduce systemic activity, parotid enlargement, lymphadenopathies, articular manifestation, and B cell biomarkers in pSS even if concluding evidence is still lacking [109]. Sequential therapy with rituximab and belimumab has been described in selected pSS complications, including cryoglobulinemia [110] and lymphoma [111]. Other B cell-targeted therapies are currently being studied, such as Ianalumab, a BAFF receptor fully human monoclonal antibody, engineered for direct antibody-dependent cellular cytotoxicity-mediated B-cell depletion that was found to be beneficial on clinical domains in a phase II trial [112].

As far as other monoclonal antibodies are concerned (Figure 2), abatacept (CTLA4-Ig), a T cell co-stimulatory signal modulator, showed promising results in an open-label study [113], yet these findings need further confirmation in larger trials. Anti-TNF α biologicals were not found to not improve symptoms and/or disease activity [114,115]. Similarly, disappointing results were observed with anti-IL-1 or anti-IL-6 monoclonals [116].

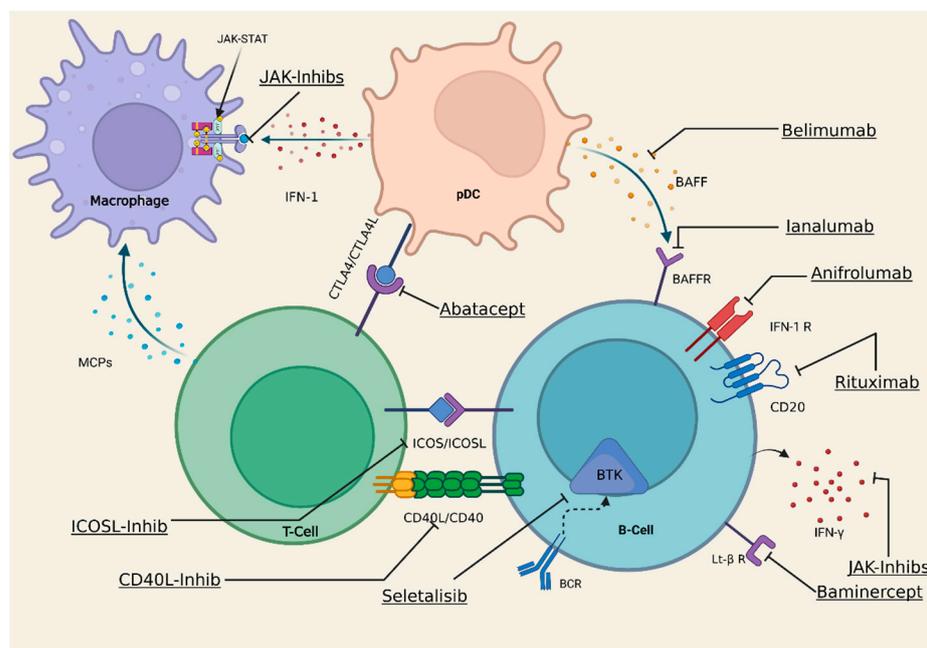


Figure 2. Sjögren's syndrome targets and future treatments. This figure illustrates some current or potential therapeutic target in relation to immune cells involved in SS pathogenesis. IFN-1, Interferon-1; IFN-1 R, Interferon-1 Receptor; IFN- γ , Interferon- γ ; pDC, plasmacytoid dendritic cell; BTK, Bruton-Kinase; JAK, Janus-Kinase; BAFF, B cell activating factor belonging to TNF family; BCR, B-cell receptor; Lt- β R, Lymphotoxin-beta-receptor; CD40L, CD40 Ligand; ICOSL, Inducible co-stimulator ligand; CTLA4L, Cytotoxic T-Lymphocyte antigen 4 ligand. Created with www.BioRender.com, accessed on 13 October 2022.

Targeting T-lymphocytes is another potential area of interest in pSS, as their inhibition/modulation would contribute to block macrophage activation and, thus, the chronicity of adaptive immune activation. In fact, drugs that block ICOS-ligand, a stimulator of T-lymphocyte pathways, are being studied with similar results to Baminercept (Lymphotoxin

β receptor IgG fusion protein) but without favourable results [117]. CD40 ligand (CD40-L) inhibitors are also being studied to prevent T cell differentiation and activation [118].

The kinases and their signalling pathways could theoretically be targeted in pSS. These include the Bruton kinase Pi3K that is capable of reducing B cells overactivation, although, for the moment, existing drugs in clinical trials have not shown favourable results (Seletalisib, anti-Pi3K) [119].

Macrophages play an important role in signalling pathways. Although a safe depletion of macrophages has not been achieved, another approach would be to inhibit macrophage chemotactic proteins (MCPs) in order to prevent their destructive involvement in target tissues. These chemokines are: CCL2 (MCP-1)/CCR2, CX3CL1 (fractalkine)/CX3CR1, and CCL5 (RANTES)/CCR5.

In light of much evidence defining IFN as one of the main effectors in pSS pathogenesis, many classes of drugs interfering with its function and related pathways are now under investigation. Suppression of IFN-stimulated JAK-STAT signalling by oral JAK-inhibitors is currently considered in ongoing clinical trials (such as with tofacitinib, a dual JAK 1/2 inhibitor) [120]. In a pilot phase I/II proof-of-concept study, baricitinib proved effective in providing benefit in the majority of patients with high disease activity [121]. Lastly, Anifrolumab, a monoclonal antibody, which directly targets IFN-1 [122] and was recently approved for the treatment of SLE [123,124], is under investigation, and the results of current trials in pSS are eagerly awaited [125].

7. Conclusions

Sjögren's syndrome is a very heterogeneous disease mostly led by dysfunction in lymphocyte function and activity, yet much evidence indicates that monocytes/macrophages are relevant in promoting inflammation, lymphocyte over-activation, and eventually structural damage. Monocytes and related signalling pathways (interferon signature) are emerging as new research targets whose relevance and precise characterisation have only recently started to be elucidated by transcriptome and epigenetic studies. The precise definition and role of monocyte-derived molecules involved in activation, inhibition, or regulation of the immune system within this intricate puzzle that is Sjögren's disease are still far from completely understood. In this complex scenario, SS emerges as a pathological entity halfway between SLE and rheumatoid arthritis, with its own identity and few globally effective therapies so far.

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References

1. Chatzis, L.G.; Goules, A.V.; Tzioufas, A.G. Searching for the “X Factor” in Sjögren's Syndrome Female Predilection. *Clin. Exp. Rheumatol.* **2021**, *39*, S206–S214. [[CrossRef](#)] [[PubMed](#)]
2. Ramos-Casals, M.; Tzioufas, A.G.; Font, J. Primary Sjögren's Syndrome: New Clinical and Therapeutic Concepts. *Ann. Rheum. Dis.* **2005**, *64*, 347–354. [[CrossRef](#)] [[PubMed](#)]
3. Negrini, S.; Emmi, G.; Greco, M.; Borro, M.; Sardanelli, F.; Murdaca, G.; Indiveri, F.; Puppo, F. Sjögren's Syndrome: A Systemic Autoimmune Disease. *Clin. Exp. Med.* **2022**, *22*, 9–25. [[CrossRef](#)] [[PubMed](#)]

4. Cafaro, G.; Bursi, R.; Chatzis, L.G.; Fulvio, G.; Ferro, F.; Bartoloni, E.; Baldini, C. One Year in Review 2021: Sjögren's Syndrome. *Clin. Exp. Rheumatol.* **2021**, *39*, 3–13. [[CrossRef](#)] [[PubMed](#)]
5. Pertovaara, M.; Korpela, M.; Uusitalo, H.; Pukander, J.; Miettinen, A.; Helin, H.; Pasternack, A. Clinical Follow up Study of 87 Patients with Sicca Symptoms (Dryness of Eyes or Mouth, or Both). *Ann. Rheum. Dis.* **1999**, *58*, 423–427. [[CrossRef](#)] [[PubMed](#)]
6. Retamozo, S.; Acar-Denizli, N.; Rasmussen, A.; Horváth, I.F.; Baldini, C.; Priori, R.; Sandhya, P.; Hernandez-Molina, G.; Armagan, B.; Praprotnik, S.; et al. Systemic Manifestations of Primary Sjögren's Syndrome out of the ESSDAI Classification: Prevalence and Clinical Relevance in a Large International, Multi-Ethnic Cohort of Patients. *Clin. Exp. Rheumatol.* **2019**, *37*, 97–106.
7. Veenbergen, S.; Kozmar, A.; van Daele, P.L.A.; Schreurs, M.W.J. Autoantibodies in Sjögren's Syndrome and Its Classification Criteria. *J. Transl. Autoimmun.* **2022**, *5*, 100138. [[CrossRef](#)]
8. Kontny, E.; Lewandowska-Poluch, A.; Chmielińska, M.; Olesińska, M. Subgroups of Sjögren's Syndrome Patients Categorised by Serological Profiles: Clinical and Immunological Characteristics. *Reumatologia* **2018**, *56*, 346–353. [[CrossRef](#)]
9. Braconi, D.; Bernardini, G.; Spiga, O.; Santucci, A. Leveraging Proteomics in Orphan Disease Research: Pitfalls and Potential. *Expert Rev. Proteom.* **2021**, *18*, 315–327. [[CrossRef](#)]
10. Aronson, J.K. Rare Diseases and Orphan Drugs. *Br. J. Clin. Pharm.* **2006**, *61*, 243–245. [[CrossRef](#)]
11. Nocturne, G.; Mariette, X. B Cells in the Pathogenesis of Primary Sjögren Syndrome. *Nat. Rev. Rheumatol.* **2018**, *14*, 133–145. [[CrossRef](#)]
12. Verstappen, G.M.; Kroese, F.G.M.; Bootsma, H. T Cells in Primary Sjögren's Syndrome: Targets for Early Intervention. *Rheumatology* **2019**, *60*, 3088–3098. [[CrossRef](#)]
13. Ma, W.-T.; Gao, F.; Gu, K.; Chen, D.-K. The Role of Monocytes and Macrophages in Autoimmune Diseases: A Comprehensive Review. *Front. Immunol.* **2019**, *10*, 1140. [[CrossRef](#)] [[PubMed](#)]
14. Baldini, C.; Ferro, F.; Elefante, E.; Bombardieri, S. Biomarkers for Sjögren's Syndrome. *Biomark. Med.* **2018**, *12*, 275–286. [[CrossRef](#)] [[PubMed](#)]
15. Ibáñez-Cabellos, J.S.; Seco-Cervera, M.; Osca-Verdegal, R.; Pallardó, F.V.; García-Giménez, J.L. Epigenetic Regulation in the Pathogenesis of Sjögren Syndrome and Rheumatoid Arthritis. *Front. Genet.* **2019**, *10*, 1104. [[CrossRef](#)] [[PubMed](#)]
16. Kapsogeorgou, E.K.; Papageorgiou, A.; Protogerou, A.D.; Voulgarelis, M.; Tzioufas, A.G. Low MiR200b-5p Levels in Minor Salivary Glands: A Novel Molecular Marker Predicting Lymphoma Development in Patients with Sjögren's Syndrome. *Ann. Rheum. Dis.* **2018**, *77*, 1200–1207. [[CrossRef](#)]
17. Wolf, A.A.; Yáñez, A.; Barman, P.K.; Goodridge, H.S. The Ontogeny of Monocyte Subsets. *Front. Immunol.* **2019**, *10*, 1642. [[CrossRef](#)]
18. Orozco, S.L.; Canny, S.P.; Hamerman, J.A. Signals Governing Monocyte Differentiation during Inflammation. *Curr. Opin. Immunol.* **2021**, *73*, 16–24. [[CrossRef](#)]
19. Williams, M.; Mildner, A.; Yona, S. Developmental and Functional Heterogeneity of Monocytes. *Immunity* **2018**, *49*, 595–613. [[CrossRef](#)]
20. Canè, S.; Ugel, S.; Trovato, R.; Marigo, I.; de Sanctis, F.; Sartoris, S.; Bronte, V. The Endless Saga of Monocyte Diversity. *Front. Immunol.* **2019**, *10*, 1786. [[CrossRef](#)]
21. Thomas, G.D.; Hamers, A.A.J.; Nakao, C.; Marcovecchio, P.; Taylor, A.M.; McSkimming, C.; Nguyen, A.T.; McNamara, C.A.; Hedrick, C.C. Human Blood Monocyte Subsets: A New Gating Strategy Defined Using Cell Surface Markers Identified by Mass Cytometry. *Arter. Thromb. Vasc. Biol.* **2017**, *37*, 1548–1558. [[CrossRef](#)] [[PubMed](#)]
22. Martin, C.; Chevrot, M.; Poirier, H.; Passilly-Degrace, P.; Niot, I.; Besnard, P. CD36 as a Lipid Sensor. *Physiol. Behav.* **2011**, *105*, 36–42. [[CrossRef](#)] [[PubMed](#)]
23. Arnaout, M.A. Structure and Function of the Leukocyte Adhesion Molecules CD11/CD18. *Blood* **1990**, *75*, 1037–1050. [[CrossRef](#)]
24. Sharma, M.D.; Rodriguez, P.C.; Koehn, B.H.; Baban, B.; Cui, Y.; Guo, G.; Shimoda, M.; Pacholczyk, R.; Shi, H.; Lee, E.-J.; et al. Activation of P53 in Immature Myeloid Precursor Cells Controls Differentiation into Ly6c+CD103+ Monocytic Antigen-Presenting Cells in Tumors. *Immunity* **2018**, *48*, 91–106.e6. [[CrossRef](#)]
25. Askenase, M.H.; Han, S.-J.; Byrd, A.L.; Morais da Fonseca, D.; Bouladoux, N.; Wilhelm, C.; Konkel, J.E.; Hand, T.W.; Lacerda-Queiroz, N.; Su, X.; et al. Bone-Marrow-Resident NK Cells Prime Monocytes for Regulatory Function during Infection. *Immunity* **2015**, *42*, 1130–1142. [[CrossRef](#)]
26. Coillard, A.; Segura, E. Antigen Presentation by Mouse Monocyte-Derived Cells: Re-Evaluating the Concept of Monocyte-Derived Dendritic Cells. *Mol. Immunol.* **2021**, *135*, 165–169. [[CrossRef](#)] [[PubMed](#)]
27. Satoh, T.; Nakagawa, K.; Sugihara, F.; Kuwahara, R.; Ashihara, M.; Yamane, F.; Minowa, Y.; Fukushima, K.; Ebina, I.; Yoshioka, Y.; et al. Identification of an Atypical Monocyte and Committed Progenitor Involved in Fibrosis. *Nature* **2017**, *541*, 96–101. [[CrossRef](#)]
28. Auffray, C.; Fogg, D.; Garfa, M.; Elain, G.; Join-Lambert, O.; Kayal, S.; Sarnacki, S.; Cumano, A.; Lauvau, G.; Geissmann, F. Monitoring of Blood Vessels and Tissues by a Population of Monocytes with Patrolling Behavior. *Science* **2007**, *317*, 666–670. [[CrossRef](#)]
29. Thomas, G.; Tacke, R.; Hedrick, C.C.; Hanna, R.N. Nonclassical Patrolling Monocyte Function in the Vasculature. *Arterioscler. Thromb. Vasc. Biol.* **2015**, *35*, 1306–1316. [[CrossRef](#)]

30. Gamrekelashvili, J.; Kapanadze, T.; Sablotny, S.; Ratiu, C.; Dastagir, K.; Lochner, M.; Karbach, S.; Wenzel, P.; Sitnow, A.; Fleig, S.; et al. Notch and TLR Signaling Coordinate Monocyte Cell Fate and Inflammation. *Elife* **2020**, *9*, e57007. [[CrossRef](#)]
31. Carlin, L.M.; Stamatiades, E.G.; Auffray, C.; Hanna, R.N.; Glover, L.; Vizcay-Barrena, G.; Hedrick, C.C.; Cook, H.T.; Diebold, S.; Geissmann, F. Nr4a1-Dependent Ly6C(Low) Monocytes Monitor Endothelial Cells and Orchestrate Their Disposal. *Cell* **2013**, *153*, 362–375. [[CrossRef](#)] [[PubMed](#)]
32. Hamers, A.A.J.; Dinh, H.Q.; Thomas, G.D.; Marcovecchio, P.; Blatchley, A.; Nakao, C.S.; Kim, C.; McSkimming, C.; Taylor, A.M.; Nguyen, A.T.; et al. Human Monocyte Heterogeneity as Revealed by High-Dimensional Mass Cytometry. *Arterioscler. Thromb. Vasc. Biol.* **2019**, *39*, 25–36. [[CrossRef](#)] [[PubMed](#)]
33. Venneri, M.A.; de Palma, M.; Ponzoni, M.; Pucci, F.; Scielzo, C.; Zonari, E.; Mazzieri, R.; Doglioni, C.; Naldini, L. Identification of Proangiogenic TIE2-Expressing Monocytes (TEMs) in Human Peripheral Blood and Cancer. *Blood* **2007**, *109*, 5276–5285. [[CrossRef](#)] [[PubMed](#)]
34. Han, S.; Zhuang, H.; Lee, P.Y.; Li, M.; Yang, L.; Nigrovic, P.A.; Reeves, W.H. Differential Responsiveness of Monocyte and Macrophage Subsets to Interferon. *Arthritis Rheumatol.* **2020**, *72*, 100–113. [[CrossRef](#)]
35. Coillard, A.; Segura, E. In Vivo Differentiation of Human Monocytes. *Front. Immunol.* **2019**, *10*, 1907. [[CrossRef](#)]
36. Weber, C.; Belge, K.U.; von Hundelshausen, P.; Draude, G.; Steppich, B.; Mack, M.; Frankenberger, M.; Weber, K.S.; Ziegler-Heitbrock, H.W. Differential Chemokine Receptor Expression and Function in Human Monocyte Subpopulations. *J. Leukoc. Biol.* **2000**, *67*, 699–704. [[CrossRef](#)]
37. Pinho, V.; Italiani, P.; Mitroulis, I.; Kalafati, L.; Bornhäuser, M.; Hajishengallis, G.; Chavakis, T. Regulation of the Bone Marrow Niche by Inflammation. *Front. Immunol.* **2020**, *11*, 1540. [[CrossRef](#)]
38. Seneviratne, A.N.; Edsfeldt, A.; Cole, J.E.; Kassiteridi, C.; Swart, M.; Park, I.; Green, P.; Khoiratty, T.; Saliba, D.; Goddard, M.E.; et al. Interferon Regulatory Factor 5 Controls Necrotic Core Formation in Atherosclerotic Lesions by Impairing Efferocytosis. *Circulation* **2017**, *136*, 1140–1154. [[CrossRef](#)]
39. Crayne, C.B.; Albeituni, S.; Nichols, K.E.; Cron, R.Q. The Immunology of Macrophage Activation Syndrome. *Front. Immunol.* **2019**, *10*, 119. [[CrossRef](#)] [[PubMed](#)]
40. Marketos, N.; Cinoku, I.; Rapti, A.; Mavragani, C.P. Type I Interferon Signature in Sjögren’s Syndrome: Pathophysiological and Clinical Implications. *Clin. Exp. Rheumatol.* **2019**, *37*, S185–S191.
41. Brkic, Z.; Maria, N.I.; van Helden-Meeuwssen, C.G.; van de Merwe, J.P.; van Daele, P.L.; Dalm, V.A.; Wildenberg, M.E.; Beumer, W.; Drexhage, H.A.; Versnel, M.A. Prevalence of Interferon Type I Signature in CD14 Monocytes of Patients with Sjogren’s Syndrome and Association with Disease Activity and BAFF Gene Expression. *Ann. Rheum. Dis.* **2013**, *72*, 728–735. [[CrossRef](#)] [[PubMed](#)]
42. Bourgoin, P.; Biéché, G.; Ait Belkacem, I.; Morange, P.-E.; Malergue, F. Role of the Interferons in CD64 and CD169 Expressions in Whole Blood: Relevance in the Balance between Viral- or Bacterial-Oriented Immune Responses. *Immun. Inflamm. Dis.* **2020**, *8*, 106–123. [[CrossRef](#)] [[PubMed](#)]
43. Shemesh, M.; Lochte, S.; Piehler, J.; Schreiber, G. IFNAR1 and IFNAR2 Play Distinct Roles in Initiating Type I Interferon-Induced JAK-STAT Signaling and Activating STATs. *Sci. Signal.* **2021**, *14*, eabe4627. [[CrossRef](#)] [[PubMed](#)]
44. Boyette, L.B.; Macedo, C.; Hadi, K.; Elinoff, B.D.; Walters, J.T.; Ramaswami, B.; Chalasani, G.; Taboas, J.M.; Lakkis, F.G.; Metes, D.M. Phenotype, Function, and Differentiation Potential of Human Monocyte Subsets. *PLoS ONE* **2017**, *12*, e0176460. [[CrossRef](#)]
45. Deshmane, S.L.; Kremlev, S.; Amini, S.; Sawaya, B.E. Monocyte Chemoattractant Protein-1 (MCP-1): An Overview. *J. Interferon Cytokine Res.* **2009**, *29*, 313–325. [[CrossRef](#)]
46. Nordmark, G.; Eloranta, M.-L.; Ronnblom, L. Primary Sjögren’s Syndrome and the Type I Interferon System. *Curr. Pharm. Biotechnol.* **2012**, *13*, 2054–2062. [[CrossRef](#)] [[PubMed](#)]
47. Wildenberg, M.E.; van Helden-Meeuwssen, C.G.; van de Merwe, J.P.; Drexhage, H.A.; Versnel, M.A. Systemic Increase in Type I Interferon Activity in Sjögren’s Syndrome: A Putative Role for Plasmacytoid Dendritic Cells. *Eur. J. Immunol.* **2008**, *38*, 2024–2033. [[CrossRef](#)]
48. Bao, M.; Liu, Y.J. Regulation of TLR7/9 Signaling in Plasmacytoid Dendritic Cells. *Protein Cell* **2013**, *4*, 40–52. [[CrossRef](#)]
49. Zhou, D.; McNamara, N.A. Macrophages: Important Players in Primary Sjögren’s Syndrome? *Expert Rev. Clin. Immunol.* **2014**, *10*, 513–520. [[CrossRef](#)]
50. Lee, K.E.; Kang, J.H.; Yim, Y.R.; Kim, J.E.; Lee, J.W.; Wen, L.; Park, D.J.; Kim, T.J.; Park, Y.W.; Yoon, K.C.; et al. The Significance of Ectopic Germinal Centers in the Minor Salivary Gland of Patients with Sjögren’s Syndrome. *J. Korean Med. Sci.* **2016**, *31*, 190–195. [[CrossRef](#)]
51. Florencia Quiroga, M.; Arbour, N.; Dario Motrich, R.; Ishimaru, N.; Ushio, A.; Arakaki, R.; Otsuka, K.; Yamada, A.; Tsunematsu, T.; Kudo, Y.; et al. CCL22-Producing Resident Macrophages Enhance T Cell Response in Sjögren’s Syndrome. *Front. Immunol.* **2018**, *9*, 2594. [[CrossRef](#)]
52. Solomon, A.; Dursun, D.; Liu, Z.; Xie, Y.; Macri, A.; Pflugfelder, S.C. Pro- and Anti-Inflammatory Forms of Interleukin-1 in the Tear Fluid and Conjunctiva of Patients with Dry-Eye Disease. *Investig. Ophthalmol. Vis. Sci.* **2001**, *42*, 2283–2292.
53. Ożańska, A.; Szymczak, D.; Rybka, J. Pattern of Human Monocyte Subpopulations in Health and Disease. *Scand. J. Immunol.* **2020**, *92*, e12883. [[CrossRef](#)]
54. Fragoulis, G.E.; Vakrakou, A.G.; Papadopoulou, A.; Germenis, A.; Kanavakis, E.; Moutsopoulos, H.M.; Manoussakis, M.N. Impaired Degradation and Aberrant Phagocytosis of Necrotic Cell Debris in the Peripheral Blood of Patients with Primary Sjögren’s Syndrome. *J. Autoimmun.* **2015**, *56*, 12–22. [[CrossRef](#)] [[PubMed](#)]

55. Muñoz, L.E.; Janko, C.; Grossmayer, G.E.; Frey, B.; Voll, R.E.; Kern, P.; Kalden, J.R.; Schett, G.; Fietkau, R.; Herrmann, M.; et al. Remnants of Secondarily Necrotic Cells Fuel Inflammation in Systemic Lupus Erythematosus. *Arthritis Rheum.* **2009**, *60*, 1733–1742. [[CrossRef](#)] [[PubMed](#)]
56. Grossmayer, G.E.; Munoz, L.E.; Weber, C.K.; Franz, S.; Voll, R.E.; Kern, P.M.; Kalden, J.R.; Schett, G.; Herrmann, M.; Gaip, U.S. IgG Autoantibodies Bound to Surfaces of Necrotic Cells and Complement C4 Comprise the Phagocytosis Promoting Activity for Necrotic Cells of Systemic Lupus Erythematosis Sera. *Ann. Rheum. Dis.* **2008**, *67*, 1626–1632. [[CrossRef](#)]
57. Mavragani, C.P.; Crow, M.K. Activation of the Type I Interferon Pathway in Primary Sjogren's Syndrome. *J. Autoimmun.* **2010**, *35*, 225–231. [[CrossRef](#)] [[PubMed](#)]
58. Greenwell-Wild, T.; Moutsopoulos, N.M.; Gliozzi, M.; Kapsogeorgou, E.; Rangel, Z.; Munson, P.J.; Moutsopoulos, H.M.; Wahl, S.M. Chitinases in the Salivary Glands and Circulation of Patients with Sjögren's Syndrome: Macrophage Harbingers of Disease Severity. *Arthritis Rheum.* **2011**, *63*, 3103–3115. [[CrossRef](#)]
59. Gliozzi, M.; Greenwell-Wild, T.; Jin, W.; Moutsopoulos, N.M.; Kapsogeorgou, E.; Moutsopoulos, H.M.; Wahl, S.M. A Link between Interferon and Augmented Plasmin Generation in Exocrine Gland Damage in Sjögren's Syndrome. *J. Autoimmun.* **2013**, *40*, 122–133. [[CrossRef](#)]
60. Klinngam, W.; Janga, S.R.; Lee, C.; Ju, Y.; Yarber, F.; Shah, M.; Guo, H.; Wang, D.; MacKay, J.A.; Edman, M.C.; et al. Inhibition of Cathepsin S Reduces Lacrimal Gland Inflammation and Increases Tear Flow in a Mouse Model of Sjögren's Syndrome. *Sci. Rep.* **2019**, *9*, 9559. [[CrossRef](#)]
61. Li, X.; Wu, K.; Edman, M.; Schenke-Layland, K.; MacVeigh-Aloni, M.; Janga, S.R.; Schulz, B.; Hamm-Alvarez, S.F. Increased Expression of Cathepsins and Obesity-Induced Proinflammatory Cytokines in Lacrimal Glands of Male NOD Mouse. *Investig. Ophthalmol. Vis. Sci.* **2010**, *51*, 5019–5029. [[CrossRef](#)] [[PubMed](#)]
62. Delgado, M.; Abad, C.; Martinez, C.; Leceta, J.; Gomariz, R.P. Vasoactive Intestinal Peptide Prevents Experimental Arthritis by Downregulating Both Autoimmune and Inflammatory Components of the Disease. *Nat. Med.* **2001**, *7*, 563–568. [[CrossRef](#)]
63. Hauk, V.; Fraccaroli, L.; Grasso, E.; Eimon, A.; Ramhorst, R.; Hubscher, O.; Pérez Leirós, C. Monocytes from Sjögren's Syndrome Patients Display Increased Vasoactive Intestinal Peptide Receptor 2 Expression and Impaired Apoptotic Cell Phagocytosis. *Clin. Exp. Immunol.* **2014**, *177*, 662–670. [[CrossRef](#)]
64. Lodde, B.M.; Mineshiba, F.; Wang, J.; Cotrim, A.P.; Afione, S.; Tak, P.P.; Baum, B.J. Effect of Human Vasoactive Intestinal Peptide Gene Transfer in a Murine Model of Sjogren's Syndrome. *Ann. Rheum. Dis.* **2006**, *65*, 195–200. [[CrossRef](#)]
65. Bruserud, Ø.; Oftedal, B.E.; Wolff, A.B.; Husebye, E.S. AIRE-Mutations and Autoimmune Disease. *Curr. Opin. Immunol.* **2016**, *43*, 8–15. [[CrossRef](#)]
66. Mackay, F.; Groom, J.R.; Tangye, S.G. An Important Role for B-Cell Activation Factor and B Cells in the Pathogenesis of Sjögren's Syndrome. *Curr. Opin. Rheumatol.* **2007**, *19*, 406–413. [[CrossRef](#)] [[PubMed](#)]
67. Yoshimoto, K.; Tanaka, M.; Kojima, M.; Setoyama, Y.; Kameda, H.; Suzuki, K.; Tsuzaka, K.; Ogawa, Y.; Tsubota, K.; Abe, T.; et al. Regulatory Mechanisms for the Production of BAFF and IL-6 Are Impaired in Monocytes of Patients of Primary Sjögren's Syndrome. *Arthritis Res.* **2011**, *13*, R170. [[CrossRef](#)]
68. Yoshimoto, K.; Suzuki, K.; Takei, E.; Ikeda, Y.; Takeuchi, T. Elevated Expression of BAFF Receptor, BR3, on Monocytes Correlates with B Cell Activation and Clinical Features of Patients with Primary Sjögren's Syndrome. *Arthritis Res.* **2020**, *22*, 157. [[CrossRef](#)]
69. Charras, A.; Arvaniti, P.; le Dantec, C.; Dalekos, G.N.; Zachou, K.; Bordron, A.; Renaudineau, Y. JAK Inhibitors and Oxidative Stress Control. *Front. Immunol.* **2019**, *10*, 2814. [[CrossRef](#)] [[PubMed](#)]
70. Xin, P.; Xu, X.; Deng, C.; Liu, S.; Wang, Y.; Zhou, X.; Ma, H.; Wei, D.; Sun, S. The Role of JAK/STAT Signaling Pathway and Its Inhibitors in Diseases. *Int. Immunopharmacol.* **2020**, *80*, 106210. [[CrossRef](#)]
71. Pertovaara, M.; Silvennoinen, O.; Isomäki, P. STAT-5 Is Activated Constitutively in T Cells, B Cells and Monocytes from Patients with Primary Sjögren's Syndrome. *Clin. Exp. Immunol.* **2015**, *181*, 29–38. [[CrossRef](#)] [[PubMed](#)]
72. Nguyen, V.; Cudrici, C.; Zernetkina, V.; Niculescu, F.; Rus, H.; Drachenberg, C.; Rus, V. TRAIL, DR4 and DR5 Are Upregulated in Kidneys from Patients with Lupus Nephritis and Exert Proliferative and Proinflammatory Effects. *Clin. Immunol.* **2009**, *132*, 32–42. [[CrossRef](#)] [[PubMed](#)]
73. Castellino, G.; Corallini, F.; Trotta, F.; Secchiero, P. Elevated Levels of TRAIL in Systemic Lupus Erythematosus Are Associated to the Presence of Anti-SSA/SSB Antibodies. *Lupus* **2007**, *16*, 479–482. [[CrossRef](#)] [[PubMed](#)]
74. Lee, K.E.; Mun, S.; Kim, S.-M.; Shin, W.; Jung, W.; Paek, J.; Lee, J.; Hudson, E.; Reeves, W.H.; Han, K.; et al. The Inflammatory Signature in Monocytes of Sjögren's Syndrome and Systemic Lupus Erythematosus, Revealed by the Integrated Reactome and Drug Target Analysis. *Genes Genom.* **2022**, *44*, 1215–1229. [[CrossRef](#)] [[PubMed](#)]
75. He, Y.; Chen, R.; Zhang, M.; Wang, B.; Liao, Z.; Shi, G.; Li, Y. Abnormal Changes of Monocyte Subsets in Patients With Sjögren's Syndrome. *Front. Immunol.* **2022**, *13*, 864920. [[CrossRef](#)]
76. Matsumura, R.; Umemiya, K.; Kagami, M.; Tomioka, H.; Tanabe, E.; Sugiyama, T.; Sueishi, M.; Kayagaki, N.; Yagita, H.; Okumura, K. Expression of TNF-Related Apoptosis Inducing Ligand (TRAIL) on Infiltrating Cells and of TRAIL Receptors on Salivary Glands in Patients with Sjögren's Syndrome. *Clin. Exp. Rheumatol.* **2002**, *20*, 791–798.
77. Chen, W.S.; Lin, K.C.; Chen, C.H.; Liao, H.T.; Wang, H.P.; Li, W.Y.; Lee, H.T.; Tsai, C.Y.; Chou, C.T. Autoantibody and Biopsy Grading Are Associated with Expression of ICAM-1, MMP-3, and TRAIL in Salivary Gland Mononuclear Cells of Chinese Patients with Sjogren's Syndrome. *J. Rheumatol.* **2009**, *36*, 989–996. [[CrossRef](#)]

78. Lopes, A.P.; Bekker, C.P.J.; Hillen, M.R.; Blokland, S.L.M.; Hinrichs, A.C.; Pandit, A.; Kruize, A.A.; Radstake, T.R.D.J.; van Roon, J.A.G. The Transcriptomic Profile of Monocytes from Patients with Sjögren's Syndrome Is Associated with Inflammatory Parameters and Is Mimicked by Circulating Mediators. *Front. Immunol.* **2021**, *12*, 701656. [[CrossRef](#)]
79. Maria, N.I.; Brkic, Z.; Waris, M.; van Helden-Meeuwssen, C.G.; Heezen, K.; van de Merwe, J.P.; van de Merwe, P.L.; Dalm, V.A.S.H.; Drexhage, H.A.; Versnel, M.A. MxA as a Clinically Applicable Biomarker for Identifying Systemic Interferon Type i in Primary Sjögren's Syndrome. *Ann. Rheum. Dis.* **2014**, *73*, 1052–1059. [[CrossRef](#)]
80. Rose, T.; Szelinski, F.; Lisney, A.; Reiter, K.; Fleischer, S.J.; Burmester, G.R.; Radbruch, A.; Hiepe, F.; Grützkau, A.; Biesen, R.; et al. SIGLEC1 Is a Biomarker of Disease Activity and Indicates Extraglandular Manifestation in Primary Sjögren's Syndrome. *RMD Open* **2016**, *2*, e000292. [[CrossRef](#)]
81. Duroux-Richard, I.; Robin, M.; Peilleux, C.; Apparailly, F. MicroRNAs: Fine Tuners of Monocyte Heterogeneity. *Front. Immunol.* **2019**, *10*, 2145. [[CrossRef](#)] [[PubMed](#)]
82. Pu, M.; Chen, J.; Tao, Z.; Miao, L.; Qi, X.; Wang, Y.; Ren, J. Regulatory Network of MiRNA on Its Target: Coordination between Transcriptional and Post-Transcriptional Regulation of Gene Expression. *Cell Mol. Life Sci.* **2019**, *76*, 441–451. [[CrossRef](#)] [[PubMed](#)]
83. Cullen, B.R. Transcription and Processing of Human MicroRNA Precursors. *Mol. Cell* **2004**, *16*, 861–865. [[CrossRef](#)] [[PubMed](#)]
84. Boldin, M.P.; Taganov, K.D.; Rao, D.S.; Yang, L.; Zhao, J.L.; Kalwani, M.; Garcia-Flores, Y.; Luong, M.; Devrekanli, A.; Xu, J.; et al. MiR-146a Is a Significant Brake on Autoimmunity, Myeloproliferation, and Cancer in Mice. *J. Exp. Med.* **2011**, *208*, 1189–1201. [[CrossRef](#)] [[PubMed](#)]
85. Nakasa, T.; Shibuya, H.; Nagata, Y.; Niimoto, T.; Ochi, M. The Inhibitory Effect of MicroRNA-146a Expression on Bone Destruction in Collagen-Induced Arthritis. *Arthritis Rheum.* **2011**, *63*, 1582–1590. [[CrossRef](#)]
86. Taganov, K.D.; Boldin, M.P.; Chang, K.J.; Baltimore, D. NF-KappaB-Dependent Induction of MicroRNA MiR-146, an Inhibitor Targeted to Signaling Proteins of Innate Immune Responses. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 12481–12486. [[CrossRef](#)]
87. Wang, Y.; Zhang, G.; Zhang, L.; Zhao, M.; Huang, H. Decreased MicroRNA-181a and -16 Expression Levels in the Labial Salivary Glands of Sjögren Syndrome Patients. *Exp. Med.* **2018**, *15*, 426–432. [[CrossRef](#)]
88. Williams, A.E.G.; Choi, K.; Chan, A.L.; Lee, Y.J.; Reeves, W.H.; Bubb, M.R.; Stewart, C.M.; Cha, S. Sjögren's Syndrome-Associated MicroRNAs in CD14(+) Monocytes Unveils Targeted TGFβ Signaling. *Arthritis Res.* **2016**, *18*, 95. [[CrossRef](#)]
89. Arsur, M.; Wu, M.; Sonenshein, G.E. TGF Beta 1 Inhibits NF-Kappa B/Rel Activity Inducing Apoptosis of B Cells: Transcriptional Activation of I Kappa B Alpha. *Immunity* **1996**, *5*, 31–40. [[CrossRef](#)]
90. Black, J.C.; van Rechem, C.; Whetstone, J.R. Histone Lysine Methylation Dynamics: Establishment, Regulation, and Biological Impact. *Mol. Cell* **2012**, *48*, 491–507. [[CrossRef](#)]
91. Smolle, M.; Workman, J.L. Transcription-Associated Histone Modifications and Cryptic Transcription. *Biochim. Biophys. Acta* **2013**, *1829*, 84–97. [[CrossRef](#)] [[PubMed](#)]
92. Imgenberg-Kreuz, J.; Sandling, J.K.; Almlöf, J.C.; Nordlund, J.; Signér, L.; Norheim, K.B.; Omdal, R.; Rönnblom, L.; Eloranta, M.L.; Syvänen, A.C.; et al. Genome-Wide DNA Methylation Analysis in Multiple Tissues in Primary Sjögren's Syndrome Reveals Regulatory Effects at Interferon-Induced Genes. *Ann. Rheum. Dis.* **2016**, *75*, 2029–2036. [[CrossRef](#)] [[PubMed](#)]
93. Luo, X.; Peng, Y.; Chen, Y.-Y.; Wang, A.-Q.; Deng, C.-W.; Peng, L.-Y.; Wu, Q.-J.; Zhao, Y.; Fei, Y.-Y.; Zhang, W. Genome-Wide DNA Methylation Patterns in Monocytes Derived from Patients with Primary Sjögren Syndrome. *Chin. Med. J.* **2021**, *134*, 1310–1316. [[CrossRef](#)] [[PubMed](#)]
94. Garcia-Gomez, A.; Li, T.; Kerick, M.; Català-Moll, F.; Comet, N.R.; Rodríguez-Ubreva, J.; de La Rica, L.; Branco, M.R.; Martín, J.; Ballestar, E. TET2- and TDG-Mediated Changes Are Required for the Acquisition of Distinct Histone Modifications in Divergent Terminal Differentiation of Myeloid Cells. *Nucleic Acids Res.* **2017**, *45*, 10002–10017. [[CrossRef](#)] [[PubMed](#)]
95. Gamrekelashvili, J.; Giagnorio, R.; Jussofie, J.; Soehnlein, O.; Duchene, J.; Briseño, C.G.; Ramasamy, S.K.; Krishnasamy, K.; Limbourg, A.; Kapanadze, T.; et al. Regulation of Monocyte Cell Fate by Blood Vessels Mediated by Notch Signalling. *Nat. Commun.* **2016**, *7*, 12597. [[CrossRef](#)]
96. Ohishi, K.; Varnum-Finney, B.; Serda, R.E.; Anasetti, C.; Bernstein, I.D. The Notch Ligand, Delta-1, Inhibits the Differentiation of Monocytes into Macrophages but Permits Their Differentiation into Dendritic Cells. *Blood* **2001**, *98*, 1402–1407. [[CrossRef](#)]
97. Vasamsetti, S.B.; Karnewar, S.; Kanugula, A.K.; Thatipalli, A.R.; Kumar, J.M.; Kotamraju, S. Metformin Inhibits Monocyte-to-Macrophage Differentiation via AMPK-Mediated Inhibition of STAT3 Activation: Potential Role in Atherosclerosis. *Diabetes* **2015**, *64*, 2028–2041. [[CrossRef](#)]
98. Seror, R.; Nocturne, G.; Mariette, X. Current and Future Therapies for Primary Sjögren Syndrome. *Nat. Rev. Rheumatol.* **2021**, *17*, 475–486. [[CrossRef](#)]
99. Gottenberg, J.-E.; Ravaud, P.; Puéchal, X.; le Guern, V.; Sibia, J.; Goeb, V.; Larroche, C.; Dubost, J.-J.; Rist, S.; Saraux, A.; et al. Effects of Hydroxychloroquine on Symptomatic Improvement in Primary Sjögren Syndrome: The JOQUER Randomized Clinical Trial. *JAMA* **2014**, *312*, 249–258. [[CrossRef](#)]
100. Winzer, M.; Aringer, M. Use of Methotrexate in Patients with Systemic Lupus Erythematosus and Primary Sjögren's Syndrome. *Clin. Exp. Rheumatol.* **2010**, *28*, S156–9.
101. Price, E.J.; Rigby, S.P.; Clancy, U.; Venables, P.J. A Double Blind Placebo Controlled Trial of Azathioprine in the Treatment of Primary Sjögren's Syndrome. *J. Rheumatol.* **1998**, *25*, 896–899. [[PubMed](#)]
102. Drosos, A.A.; Skopouli, F.N.; Galanopoulou, V.K.; Kitridou, R.C.; Moutsopoulos, H.M. Cyclosporin A Therapy in Patients with Primary Sjögren's Syndrome: Results at One Year. *Scand. J. Rheumatol. Suppl.* **1986**, *61*, 246–249. [[PubMed](#)]

103. Bikker, A.; van Woerkom, J.-M.; Kruize, A.A.; van der Wurff-Jacobs, K.M.G.; Bijlsma, J.W.J.; Lafeber, F.P.J.G.; van Roon, J.A.G. Clinical Efficacy of Leflunomide in Primary Sjogren's Syndrome Is Associated with Regulation of T-Cell Activity and Upregulation of IL-7 Receptor α Expression. *Ann. Rheum. Dis.* **2012**, *71*, 1934–1941. [[CrossRef](#)] [[PubMed](#)]
104. Van Woerkom, J.M.; Kruize, A.A.; Geenen, R.; van Roon, E.N.; Goldschmeding, R.; Verstappen, S.M.M.; van Roon, J.A.G.; Bijlsma, J.W.J. Safety and Efficacy of Leflunomide in Primary Sjögren's Syndrome: A Phase II Pilot Study. *Ann. Rheum. Dis.* **2007**, *66*, 1026–1032. [[CrossRef](#)] [[PubMed](#)]
105. Chen, W.; Lin, J. Mycophenolate for the Treatment of Primary Sjögren's Syndrome. *J. Transl. Int. Med.* **2020**, *8*, 146–149. [[CrossRef](#)] [[PubMed](#)]
106. Meijer, J.M.; Meiners, P.M.; Vissink, A.; Spijkervet, F.K.L.; Abdulahad, W.; Kamminga, N.; Brouwer, E.; Kallenberg, C.G.M.; Bootsma, H. Effectiveness of Rituximab Treatment in Primary Sjögren's Syndrome: A Randomized, Double-Blind, Placebo-Controlled Trial. *Arthritis Rheum.* **2010**, *62*, 960–968. [[CrossRef](#)]
107. Devauchelle-Pensec, V.; Mariette, X.; Jousse-Joulin, S.; Berthelot, J.-M.; Perdriger, A.; Puéchal, X.; le Guern, V.; Sibilia, J.; Gottenberg, J.-E.; Chiche, L.; et al. Treatment of Primary Sjögren Syndrome with Rituximab: A Randomized Trial. *Ann. Intern. Med.* **2014**, *160*, 233–242. [[CrossRef](#)]
108. Bowman, S.J.; Everett, C.C.; O'Dwyer, J.L.; Emery, P.; Pitzalis, C.; Ng, W.-F.; Pease, C.T.; Price, E.J.; Sutcliffe, N.; Gendi, N.S.T.; et al. Randomized Controlled Trial of Rituximab and Cost-Effectiveness Analysis in Treating Fatigue and Oral Dryness in Primary Sjögren's Syndrome. *Arthritis Rheumatol.* **2017**, *69*, 1440–1450. [[CrossRef](#)]
109. Álvarez-Rivas, N.; Sang-Park, H.; Díaz Del Campo, P.; Fernández-Castro, M.; Corominas, H.; Andreu, J.L.; Navarro-Compán, V. Efficacy of Belimumab in Primary Sjögren's Syndrome: A Systematic Review. *Reum. Clin.* **2021**, *17*, 170–174. [[CrossRef](#)]
110. Chevalier, K.; Belkhir, R.; Seror, R.; Mariette, X.; Nocturne, G. Efficacy of a Sequential Treatment by Anti-CD 20 Monoclonal Antibody and Belimumab in Type II Cryoglobulinaemia Associated with Primary Sjögren Syndrome Refractory to Rituximab Alone. *Ann. Rheum. Dis.* **2020**, *79*, 1257–1259. [[CrossRef](#)]
111. De Vita, S.; Quartuccio, L.; Salvin, S.; Picco, L.; Scott, C.A.; Rupolo, M.; Fabris, M. Sequential Therapy with Belimumab Followed by Rituximab in Sjögren's Syndrome Associated with B-Cell Lymphoproliferation and Overexpression of BAFF: Evidence for Long-Term Efficacy. *Clin. Exp. Rheumatol.* **2014**, *32*, 490–494. [[PubMed](#)]
112. Bowman, S.J.; Fox, R.; Dörner, T.; Mariette, X.; Papas, A.; Grader-Beck, T.; Fisher, B.A.; Barcelos, F.; de Vita, S.; Schulze-Koops, H.; et al. Safety and Efficacy of Subcutaneous Ianalumab (VAY736) in Patients with Primary Sjögren's Syndrome: A Randomised, Double-Blind, Placebo-Controlled, Phase 2b Dose-Finding Trial. *Lancet* **2022**, *399*, 161–171. [[CrossRef](#)]
113. Meiners, P.M.; Vissink, A.; Kroese, F.G.M.; Spijkervet, F.K.L.; Smitt-Kamminga, N.S.; Abdulahad, W.H.; Bulthuis-Kuiper, J.; Brouwer, E.; Arends, S.; Bootsma, H. Abatacept Treatment Reduces Disease Activity in Early Primary Sjögren's Syndrome (Open-Label Proof of Concept ASAP Study). *Ann. Rheum. Dis.* **2014**, *73*, 1393–1396. [[CrossRef](#)] [[PubMed](#)]
114. Sankar, V.; Brennan, M.T.; Kok, M.R.; Leakan, R.A.; Smith, J.A.; Manny, J.; Baum, B.J.; Pillemer, S.R. Etanercept in Sjögren's Syndrome: A Twelve-Week Randomized, Double-Blind, Placebo-Controlled Pilot Clinical Trial. *Arthritis Rheum.* **2004**, *50*, 2240–2245. [[CrossRef](#)]
115. Mariette, X.; Ravaud, P.; Steinfeld, S.; Baron, G.; Goetz, J.; Hachulla, E.; Combe, B.; Puéchal, X.; Pennec, Y.; Sauvezie, B.; et al. Inefficacy of Infliximab in Primary Sjögren's Syndrome: Results of the Randomized, Controlled Trial of Remicade in Primary Sjögren's Syndrome (TRIPSS). *Arthritis Rheum.* **2004**, *50*, 1270–1276. [[CrossRef](#)]
116. IL-6 Receptor Inhibition in Primary Sjögren Syndrome: Results from a Randomized Multicenter Academic Double Blind Placebo-Controlled Trial of Tocilizumab in 110 Patients-ACR Meeting Abstracts. Available online: <https://acrabstracts.org/abstract/il-6-receptor-inhibition-in-primary-sjogren-syndrome-results-from-a-randomized-multicenter-academic-double-blind-placebo-controlled-trial-of-tocilizumab-in-110-patients/> (accessed on 27 September 2022).
117. St.Clair, E.W.; Baer, A.N.; Wei, C.; Noaiseh, G.; Parke, A.; Coca, A.; Utset, T.O.; Genovese, M.C.; Wallace, D.J.; McNamara, J.; et al. Clinical Efficacy and Safety of Baminercept, a Lymphotoxin β Receptor Fusion Protein, in Primary Sjögren's Syndrome: Result from a Phase II Randomized, Double-Blind, Placebo-Controlled Trial. *Arthritis Rheumatol.* **2018**, *70*, 1470–1480. [[CrossRef](#)]
118. Safety, Tolerability, Pharmacokinetics, and Therapeutic Efficacy of SAR441344 in Primary Sjögren's Syndrome (PSjS) (PhaethuSA). US National Library of Medicine. Available online: <https://clinicaltrials.gov/ct2/show/NCT04572841> (accessed on 29 September 2022).
119. Juarez, M.; Diaz, N.; Johnston, G.I.; Nayar, S.; Payne, A.; Helmer, E.; Cain, D.; Williams, P.; Devauchelle-Pensec, V.; Fisher, B.A.; et al. A Phase 2 Randomized, Double-Blind, Placebo-Controlled, Proof-of-Concept Study of Oral Seletalisib in Primary Sjögren's Syndrome. *Rheumatology* **2021**, *60*, 1364–1375. [[CrossRef](#)]
120. Safety of Tofacitinib, an Oral Janus Kinase Inhibitor, in Primary Sjogren's Syndrome-Full Text View-ClinicalTrials.Gov. Available online: <https://clinicaltrials.gov/ct2/show/NCT04496960> (accessed on 27 September 2022).
121. Bai, W.; Liu, H.; Dou, L.; Yang, Y.; Leng, X.; Li, M.; Zhang, W.; Zhao, Y.; Zeng, X. Pilot Study of Baricitinib for Active Sjogren's Syndrome. *Ann. Rheum. Dis.* **2022**, *81*, 1050–1052. [[CrossRef](#)]
122. Narain, S.; Berman, N.; Furie, R. Biologics in the Treatment of Sjogren's Syndrome, Systemic Lupus Erythematosus, and Lupus Nephritis. *Curr. Opin. Rheumatol.* **2020**, *32*, 609–616. [[CrossRef](#)]
123. Morand, E.F.; Furie, R.; Tanaka, Y.; Bruce, I.N.; Askanase, A.D.; Richez, C.; Bae, S.-C.; Brohawn, P.Z.; Pineda, L.; Berglind, A.; et al. Trial of Anifrolumab in Active Systemic Lupus Erythematosus. *N. Engl. J. Med.* **2020**, *382*, 211–221. [[CrossRef](#)]

124. Furie, R.; Khamashta, M.; Merrill, J.T.; Werth, V.P.; Kalunian, K.; Brohawn, P.; Illei, G.G.; Drappa, J.; Wang, L.; Yoo, S. Anifrolumab, an Anti-Interferon- α Receptor Monoclonal Antibody, in Moderate-to-Severe Systemic Lupus Erythematosus. *Arthritis Rheumatol.* **2017**, *69*, 376–386. [[CrossRef](#)] [[PubMed](#)]
125. Anifrolumab Treatment for 24 Weeks in Patients with Primary Sjögren’s Syndrome (ANISE-II). US National Library of Medicine. Available online: <https://clinicaltrials.gov/ct2/show/NCT05383677> (accessed on 29 September 2022).