



SUPPLEMENT

CENTRIFUGATION PROTOCOL

The centrifugation process was performed using *Amicon*® 10K filters. The procedure is as follows: (I) concentrating the sample – (II) removing the filtrate – (III) resuspending the concentrated sample with control solution [ultrapure water pH 1.6] (i.e., buffer exchange) – (IV) re-concentrating the sample. This procedure may be repeated until the acceptable buffer threshold is reached. The manufacturer assumes a 5% sample loss at 100-fold concentration. Filter adsorption depends on the sample concentration, pH, temperature, and duration of sample-filter contact.

The removal of IL solutions from fibril samples (containing 20 μ M insulin fibrils) and concentrating fibrils to achieve higher value for ATR-FTIR measurements was performed in the following steps:

- 1) 500 μ L of the sample solution was placed into the filter, followed by centrifugation (spinning) for 15 minutes at 14 000 rcf (concentrated sample stays in the filter)
- 2) 500 μ L of the sample solution was added to the concentrate, followed by centrifugation for 15 minutes at 14 000 rcf (this step was repeated 3-times)
- 3) 300 μ L of control solution was added to the sample concentrate (resuspension), followed by centrifugation for 15 minutes at 14 000 rcf
- 4) the obtained final concentrate (~50 μ L) in the filter was invertedly centrifuged for 1 minute at 1000 rcf (i.e., transferring the sample into the test tube)

During the process, the IL solutions were removed from fibril samples (buffer exchange).

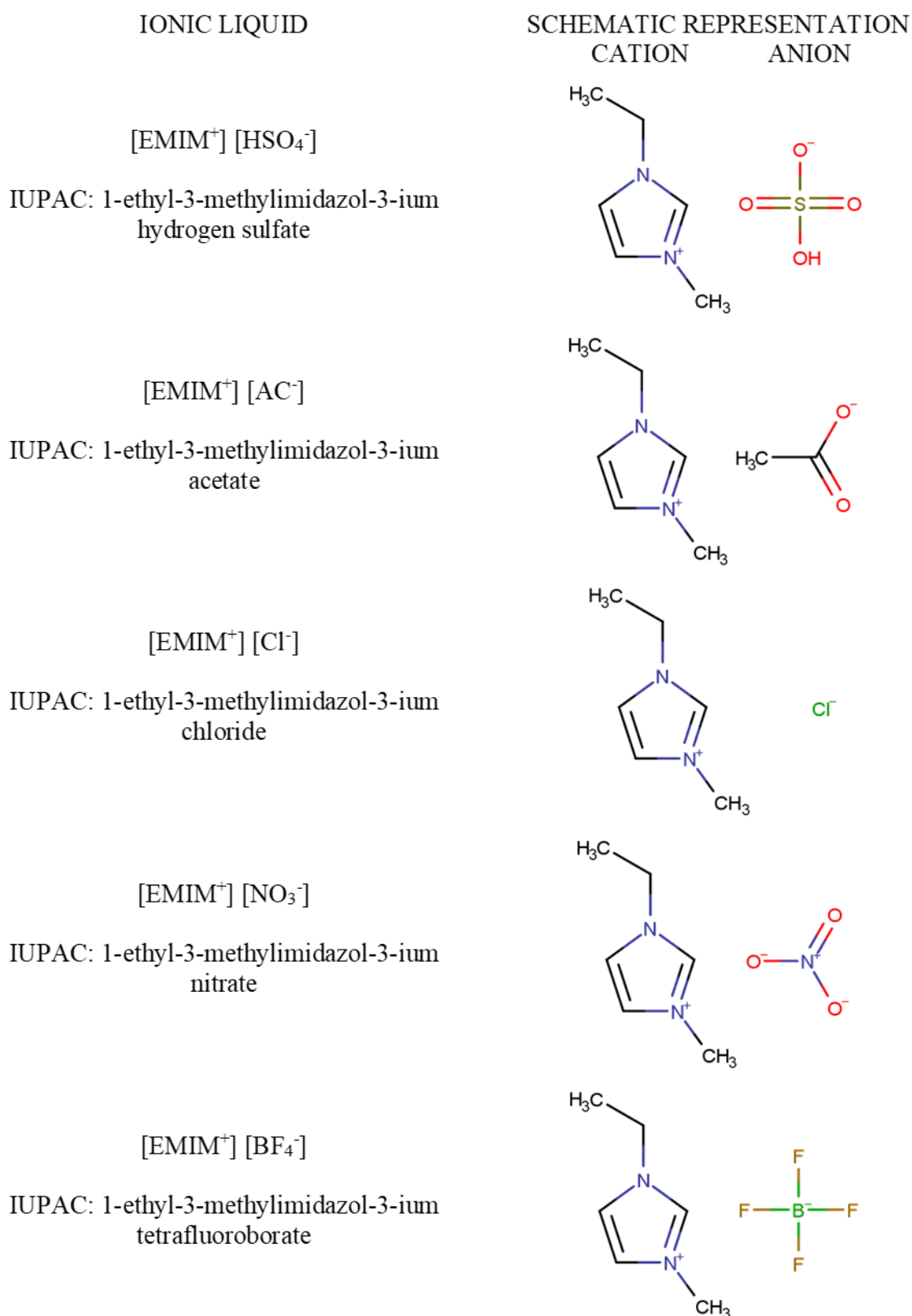


Figure S1. Schematic representation of studied ionic liquids.

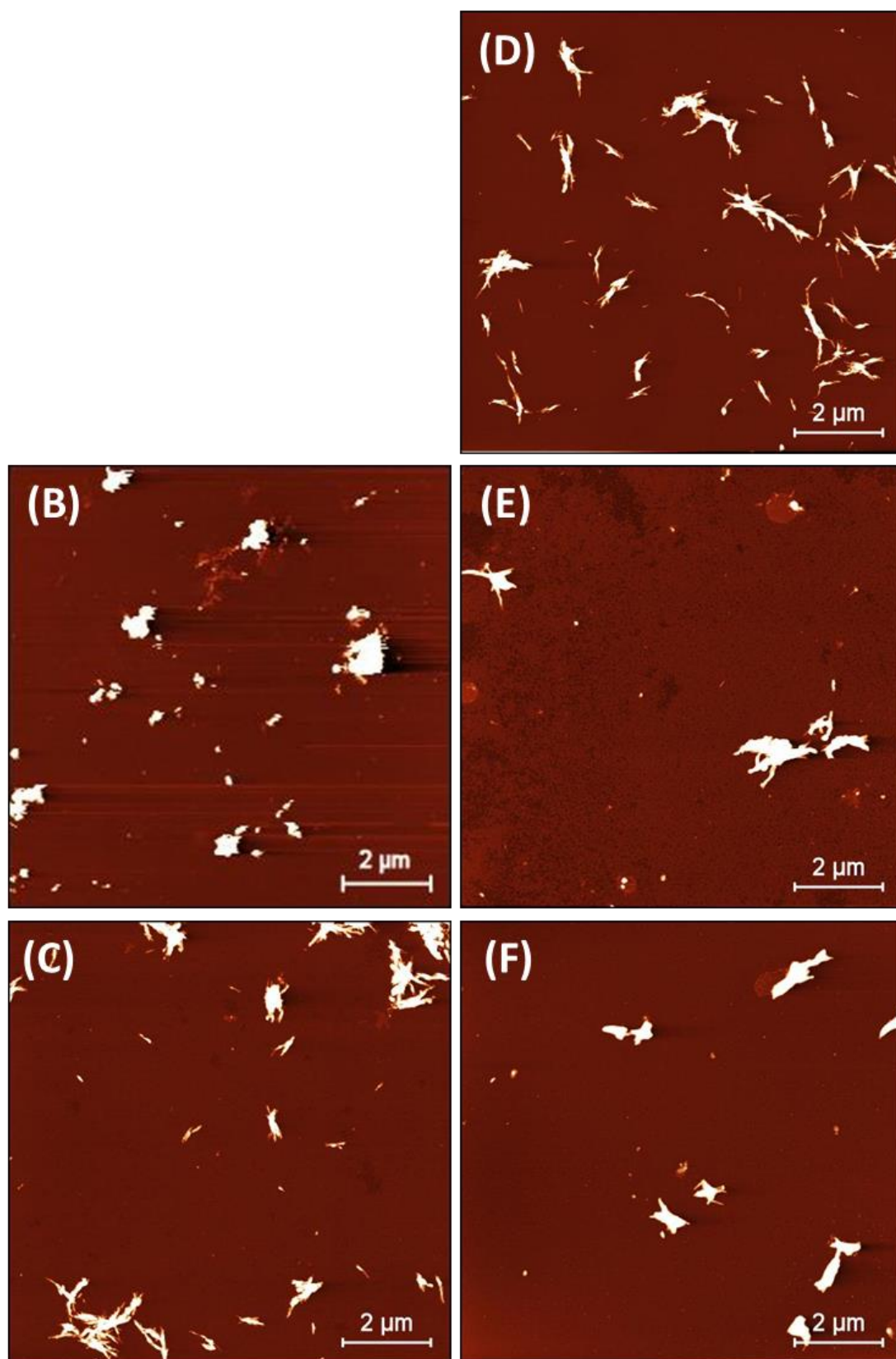


Figure S2. AFM images of insulin fibrils formed in the presence of 100 mM ILs: (B) EMIM HSO₄ 100 mM, (C) EMIM AC 100 mM, (D) EMIM Cl 100 mM, (E) EMIM NO₃ 100 mM, (F) EMIM BF₄ 100 mM.

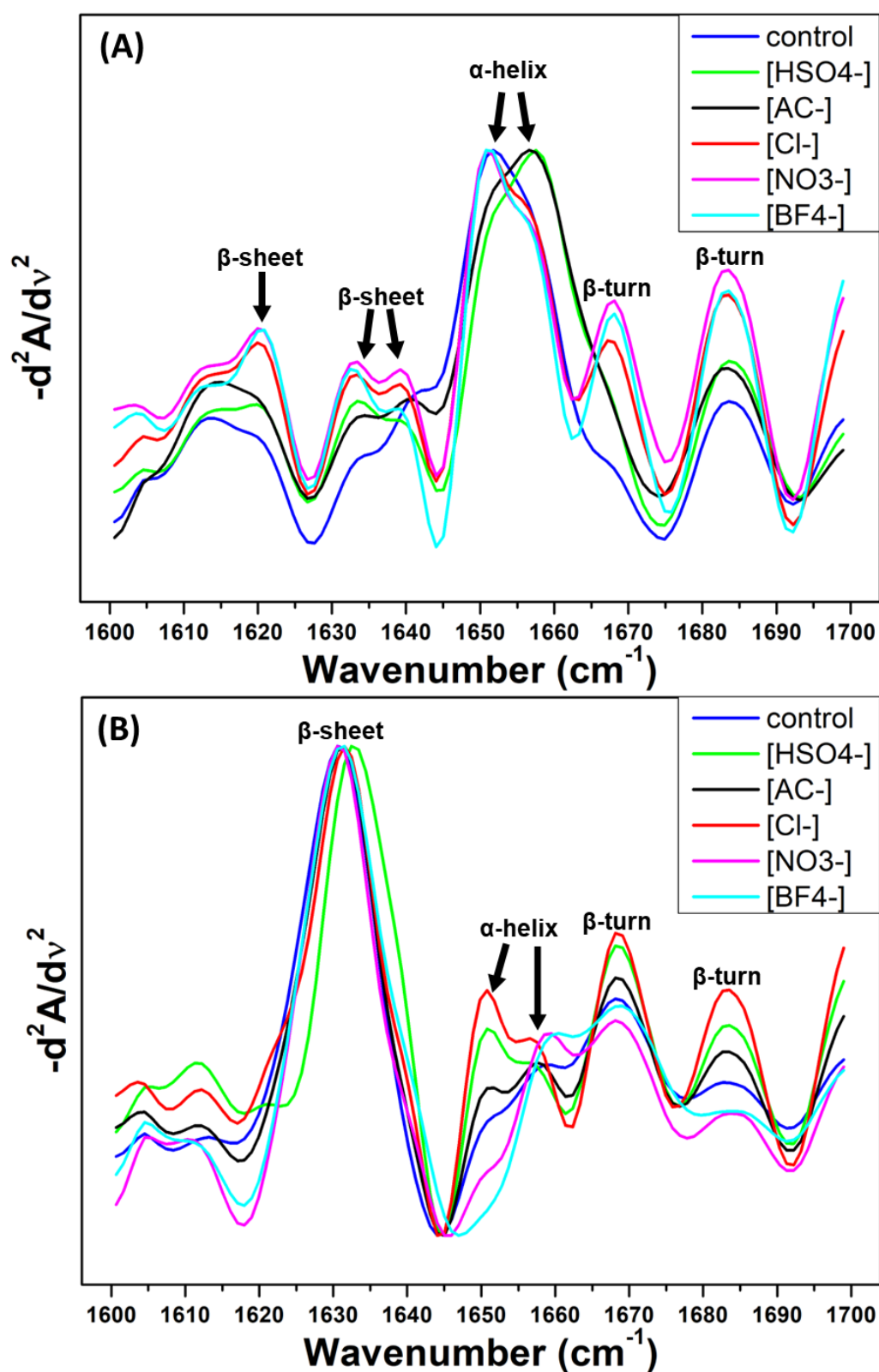


Figure S3. Second derivative of the Amide I band of (A) insulin and (B) insulin amyloid fibrils. Frequencies were assigned to particular corresponding structures according Yang¹.

¹ 1. Yang, H.; Yang, S.; Kong, J.; Dong, A.; Yu, S. Obtaining Information about Protein Secondary Structures in Aqueous Solution Using Fourier Transform IR Spectroscopy. *Nat. Protoc.* **2015**, *10*, 382–396, doi:10.1038/nprot.2015.024.

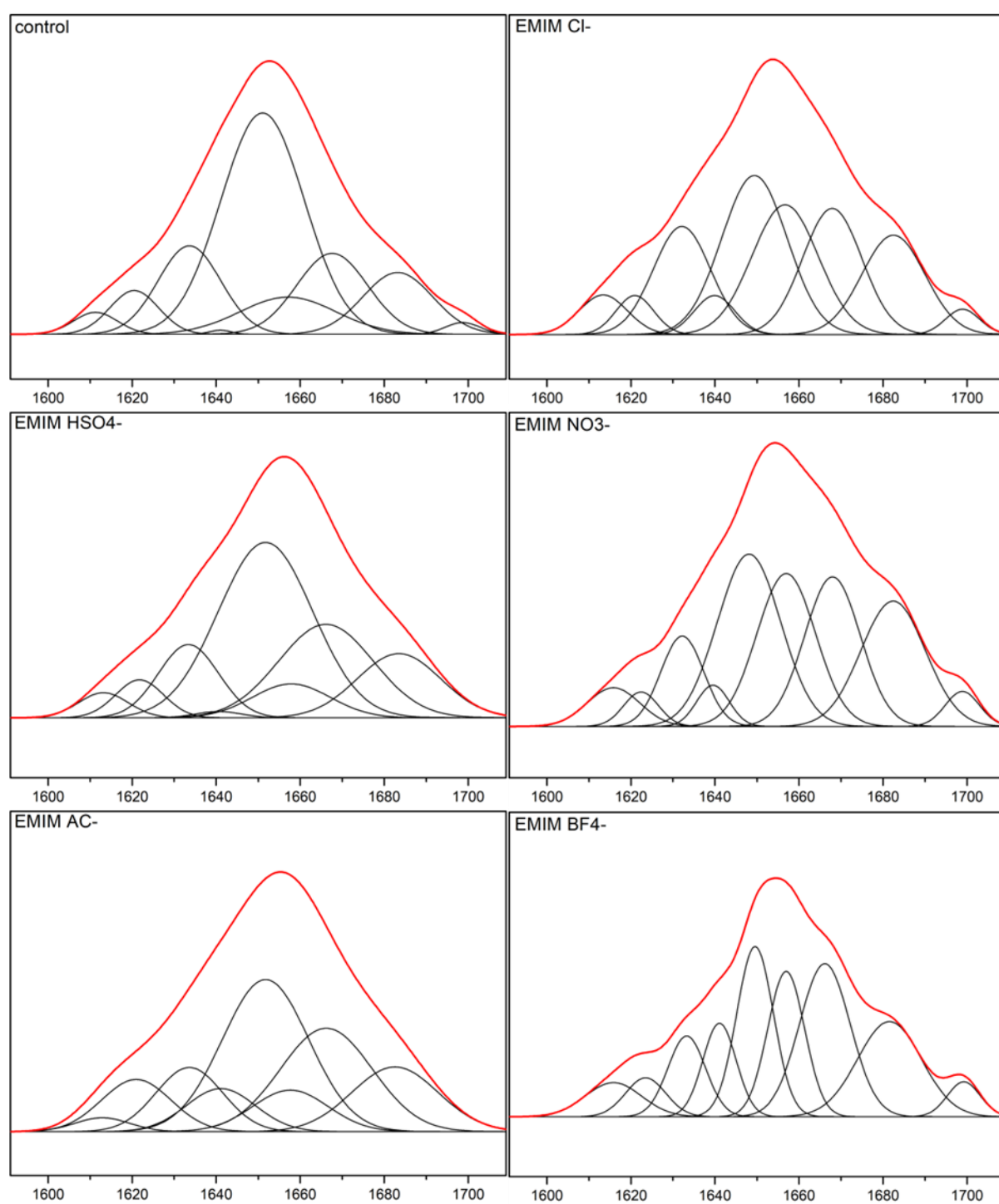


Figure S4. Deconvoluted spectra of insulin with individual peaks shown.

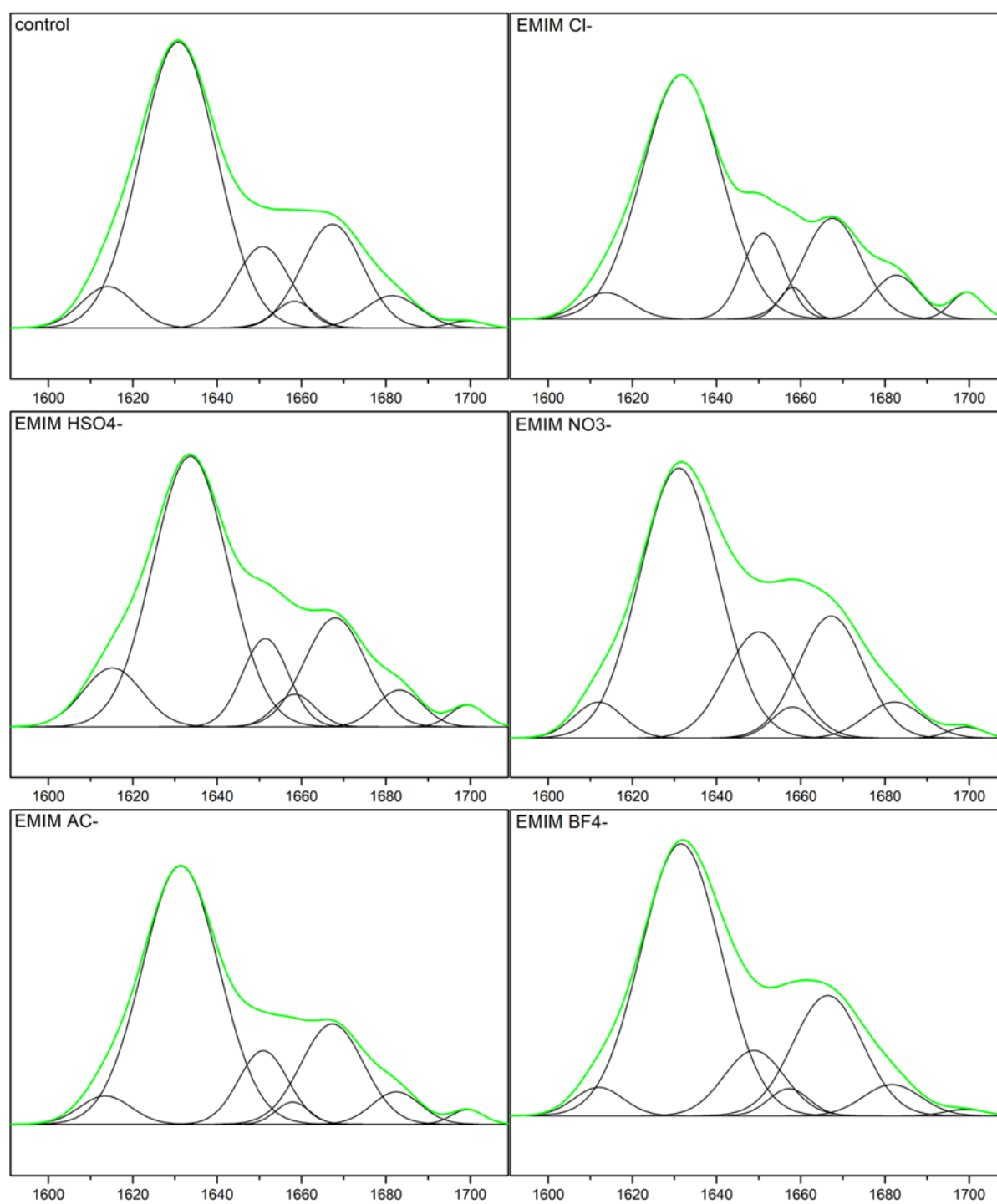


Figure S5. Deconvoluted spectra of insulin fibrils with individual peaks shown.