



Review

Zfra Overrides WWOX in Suppressing the Progression of Neurodegeneration

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Abstract: We reported that a 31-amino-acid Zfra protein (zinc finger-like protein that regulates apoptosis) blocks neurodegeneration and cancer growth. Zfra binds WW domain-containing oxidoreductase (WWOX) to both N- and C-termini, which leads to accelerated WWOX degradation. WWOX limits the progression of neurodegeneration such as Alzheimer's disease (AD) by binding tau and tau-hyperphosphorylating enzymes. Similarly, Zfra binds many protein targets and accelerates their degradation independently of ubiquitination. Furthermore, Zfra4-10 peptide strongly prevents the progression of AD-like symptoms in triple-transgenic (3xTg) mice during aging. Zfra4-10 peptide restores memory loss in 9-month-old 3xTg mice by blocking the aggregation of a protein cascade, including TPC6AA, TIAF1, and SH3GLB2, by causing aggregation of tau and amyloid β . Zfra4-10 also suppresses inflammatory NF- κ B activation. Zfra-activated Hyal-2+ CD3- CD19- Z cells in the spleen, via Hyal-2/WWOX/Smad4 signaling, are potent in cancer suppression. In this perspective review, we provide mechanistic insights regarding how Zfra overrides WWOX to induce cancer suppression and retard AD progression via Z cells.

Keywords: tumor suppressor; WWOX; Zfra; Hyal-2+ Z cell; cancer; Alzheimer's disease



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1. Introduction to Molecular Signaling and Cell Fates

How a single cell maintains its highly complicated signaling network is essentially unknown. Signaling molecules are well connected in the interaction network that is essential in maintaining normal physiology. Alterations in signaling molecules, either in concentrations or structures, lead to disruption of the signaling pathway and the potential development of multiple diseases such as cancer, neurodegeneration, and others [1–3]. For example, an imbalance in L-tryptophan metabolism via the kynurenine pathway may affect and induce disorders ranging from cancer to neurodegenerative disease [1]. Dysregulation in the participating proteins in autophagy is linked to cancer development, neurodegenerative diseases, and immune defects [2]. NAD⁺ plays an indispensable role in signaling and metabolism. Controlled and balanced levels of NAD⁺ are needed to prevent metabolic diseases, cancer, neurodegeneration, and immune disorders [3].

Interestingly, inappropriate phase separation can cause aberrant formation of protein condensates or aggregates implicated in the pathogenesis of cancer and neurodegeneration [4,5]. Dysregulation of programmed cell death causes developmental disorders and may lead to cancer and neurodegeneration [6]. Nonetheless, there is still a long list showing

molecule-regulated cancer and neurodegeneration. A straightforward question to ask is how those aberrant signaling pathways can travel down a similar path for the development of both cancer and neurodegeneration.

In this perspective review, we will go through the fundamental concept of chronic inflammation in neurodegenerative diseases and cancer, update the WWOX functional properties in suppressing Alzheimer's disease (AD) and cancer, and discuss how Zfra peptide regulates AD and cancer via Hyal-2/WWOX/Smad4 signaling and how Z cell activation by Zfra works in limiting disease progression.

2. Chronic Inflammation Is Constantly Associated with Neurodegeneration and Cancer

Chronic inflammation is a common occurrence for the progression of neurodegeneration, as well as cancer growth [7–11]. Thus, suppression of inflammation may mitigate both diseases, but does not effectively provide a cure. Low-grade inflammation can be regarded as a process of aging. The aging event involves the production of inflammatory cytokines and activated immune cells. For example, the gut–brain axis is needed in the development of rotenone-induced Parkinson's disease (PD) in rats, as revealed by changes in substantial intestinal histological alterations, such as shortened villi, crypt architecture loss, and inflammation, along with upregulation of inflammatory microglial cells [7]. Using techniques for determining microglial transcriptomes and epigenomes, one can estimate the altered microglial status affecting neuroinflammation, neurodegeneration, and disease progression [8]. In this case, specific alterations in AD and related diseases partly correlate with the status of inflammatory microglia [8]. Database analysis provides information regarding the effects of metformin on inhibiting oxidative stress, gluconeogenesis, and inflammation, linking to the mechanism for improving symptoms in AD and type 2 diabetes [9]. Most recently, the cGAS-STING pathway has been identified as a driving force for aging-related inflammation in peripheral organs and the brain [10]. Suppression of cGAS-STING signaling is likely to halt neurodegeneration in old age [10]. Again, suppression of inflammation in aging may not be able to stop the aging process, nor cancer growth, completely.

Curcumin is good for preventing and suppressing neurodegeneration and cancer, as well as modulating immune functions [12,13]. Curcumin has tautomeric forms (Figure 1). The keto form of curcumin possesses antioxidant activity, whereas the enol form is not stable and tends to degrade. Curcumin induces apoptosis in cancer cells and reduces inflammation, angiogenesis, and tumor metastasis by targeting cancer signaling pathways such as p53, Ras, PI3K, AKT, Wnt- β catenin, and mTOR. The family of NADPH oxidase enzymes (Nox1-5, Duox1-2) has seven members, which are known to be involved in various diseases, such as inflammatory lung diseases, neurodegenerative diseases, and cancer. A novel Nox2 inhibitor, TG15-132, supports neuroprotection by blocking inflammatory responses, including the generation of reactive oxygen species and inflammatory cytokines [14]. Calebin-A is a natural polyphenol and a curcumin analog [15–18] (Figure 1). Calebin-A has a ferulic acid ester bond. Calebin-A supports neuronal survival from β -amyloid insult, inhibits cancer growth, and prevents obesity, partly due to its anti-inflammatory activities such as downregulating nuclear factor (NF)- κ B activation [15–18]. Whether Calebin-A reduces peritumor coats in metastatic brain cancer or causes the degradation of amyloid plaques in the brain is unknown.

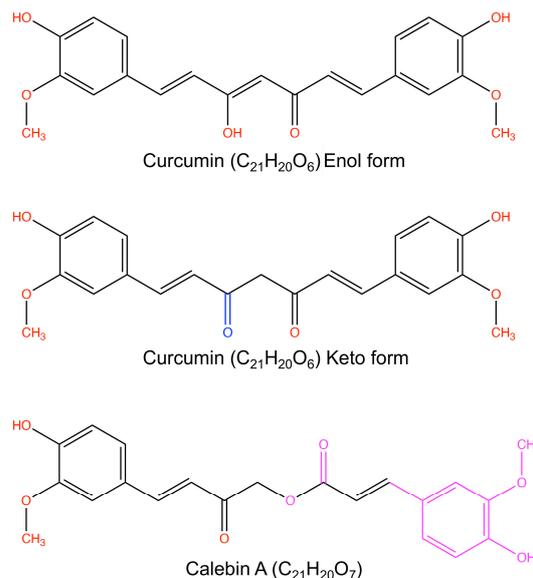


Figure 1. Structures of curcumin and calebin-A. Curcumin has a stable keto form and an unstable enol form. Calebin-A is an analog of curcumin and has a ferulic acid ester bond. The structure was drawn using a ChemDraw v22.0 software.

3. Patients Who Survive Cancer Do Not Develop AD

Epidemiological data show that AD and cancer could run in opposite directions. In most cases, patients who survive cancer do not develop AD. Conversely, patients with neurodegenerative diseases cannot develop cancer [19–21]. It has been estimated that AD patients have a 61% decreased risk of cancer incidents compared to reference subjects. It has been proposed that downregulation of Pin1 and Wnt signaling and upregulation of p53 contribute to neuronal death during cancer growth. By contrast, both Pin1 and Wnt are upregulated in cancer. This scenario fails to support the inverse relationship for AD and cancer development.

Protein aggregation in the brain occurs frequently due to the downregulation of WWOX in the brain hippocampus in middle-aged individuals [22,23]. If the protein aggregation worsens with age, the individuals will likely have AD 30 to 40 years later due to the building up of β amyloid plaques and tau aggregates in the brain. If cancer develops simultaneously in middle age, cancer cells should have a faster formation of peritumor coats containing aggregated β amyloid and amyloid fibrils [24–26]. Hypothetically, the presence of amyloid fibrils in the peritumor coats in the newly developed tumors signals the brain to slow down the protein aggregation event. Metastatic cancer cells may relocate to the brain and cause neuronal death [25]. Under this condition, both cancer growth and neural death happen simultaneously.

Drugs that are highly effective in curing AD and PD are not available. The rationale for repurposing anti-cancer drugs in treating AD, PD, and other neurodegenerative diseases is to speed up drug discovery for curing the diseases [27]. Many known kinase inhibitors have been proposed to be good for suppressing multiple protein kinases in neurodegenerative disorders. Nonetheless, whether these drugs can cure AD and PD remains questionable.

4. WWOX Exhibits Multiple Functional Properties

WW domain-containing oxidoreductase (WWOX) was first isolated in 2000 [28–30], and has been considered a candidate tumor suppressor protein [28–38]. Since then, there have been many outstanding review articles related to this gene. The WWOX/*Wwox* gene and its encoded WWOX protein participate in cancer suppression [28–32] and inhibition of AD progression [22,23]. Whether WWOX protein controls the transition between cancer susceptibility and AD resistance, or cancer resistance and AD progression has not been entirely delineated.

4.1. WWOX Primary Structure and Binding Partners

The wild-type WWOX protein possesses two N-terminal WW domains and a C-terminal short-chain alcohol dehydrogenase/reductase (SDR) domain [28–30]. A nuclear localization signal is located between the N-terminal first and second WW domains (WW1 and WW2). The WW domain is mainly responsible for protein/protein interactions. For example, WW1, possessing two tryptophan residues, binds proline-rich motifs PPXY or PPPY-containing target proteins for numerous biological functions during signaling [31–41]. Such events include cell death, differentiation, signaling, cell migration and recognition, and disease development [10–22]. WW2 has one tryptophan, whose function has not been elucidated. However, WW2 may team up with WW1 to maintain an appropriate tertiary conformation that affects the function of WW1 in protein/protein binding [42]. Nuclear localization of WWOX may occur, especially when cells are stimulated with growth factors or are under apoptotic stress [35]. The crystal structure of WWOX has not been established.

4.2. WWOX Is Not a Typical Tumor Suppressor

The human WWOX gene, encoding the WWOX protein, is located on a common fragile site, i.e., *FRA16D*—the second most frequent site of its kind [28–32]. Most cancer cells derived from breast, lung, prostate, and other organs have alterations in the WWOX gene [32–39]. Restoration of the WWOX gene and the resulting protein in cancer cells blocks their growth in vivo and in vitro [40,41]. WWOX protein is not a typical tumor suppressor, as it participates in numerous biological events, including (i) cell survival, proliferation, differentiation, cell cycle regulation, and senescence via complicated signaling pathways [38–45], (ii) aging and neurodegeneration [22,23,46,47], (iii) apoptotic cell death [30,48–51], (iv) chromosomal DNA stability [52,53], (v) bubbling cell death [54–56], and (vi) cell-to-cell recognition and migration [57]. Human newborns lacking the WWOX gene and functional WWOX protein suffer severe neural diseases but do not have spontaneous tumor formation, suggesting WWOX does not fit Knudson's two-hit hypothesis of tumorigenesis [31,35,45,46].

4.3. pY33-WWOX Maintains Normal Mitochondrial Physiology

Under physiological conditions, a portion of activated Tyr33-phosphorylated WWOX (pY33-WWOX) localizes on the outer membrane of the mitochondria to support normal physiology [28,48,58–61]. Without WWOX, cells undergo uncontrollable proliferation and die readily [43]. Under stress conditions such as exposure to UV irradiation and chemotherapeutic drugs, significantly upregulated pY33-WWOX, together with p53, induces mitochondrial apoptosis [58–61] or co-relocates to the nucleus to exert cell death [30,35,47,48]. The prosurvival TNF receptor-associated factor 2 (TRAF2) blocks the apoptotic function of WWOX and p53 [54]. The trafficking protein particle complex 6A (TRAPPC6A or TPC6A) carries WWOX to undergo nuclear translocation [22]. The protein complex dissociates in the nuclei. WWOX remains in the nuclei, and TPC6A continues to relocate to the nucleoli. Indeed, during trafficking, TPC6A travels from the mitochondria to the nucleoli and then from the nucleoli back to the mitochondria [22]. Carrying of TPC6A with proteins and associated materials between mitochondria and nucleoli is unusual but is essential for normal cell physiology [22]. The nucleolus is a center of ribosome synthesis and is abundant in small nucleolar RNA for pre-rRNA processing.

4.4. Overexpressed pY33-WWOX Induces Apoptosis Which Is Unfavorable for Neurons But Beneficial for Eliminating Cancer Cells In Vivo

The binding of pY33-WWOX with target proteins does not require the presence of PPXY or PPPY motif in the target proteins. Indeed, pY33-WWOX expands its scope in protein binding interactions [32]. In vivo data confirms that pY33-WWOX binds and functions together with many nuclear transcription factors such as p53 [30,48,49], CREB 1 (CAMP responsive element binding protein 1) [50], and c-JUN [50], and JNK1 (c-JUN N-terminal kinase 1) [48,50]. These connections may be linked to neuronal death. For

example, during the acute phase of sciatic nerve dissection, the activation of JNK1 and WWOX occurs quickly, which leads to rapid apoptosis of large neurons in the ipsilateral side of the injury in rat brains [50]. In the chronic phase of injury, small neurons in both the ipsilateral and contralateral sides of dorsal root ganglia (DRG) have co-activation of WWOX, CREB, and NF- κ B, which leads to apoptosis [50] (Figure 2).

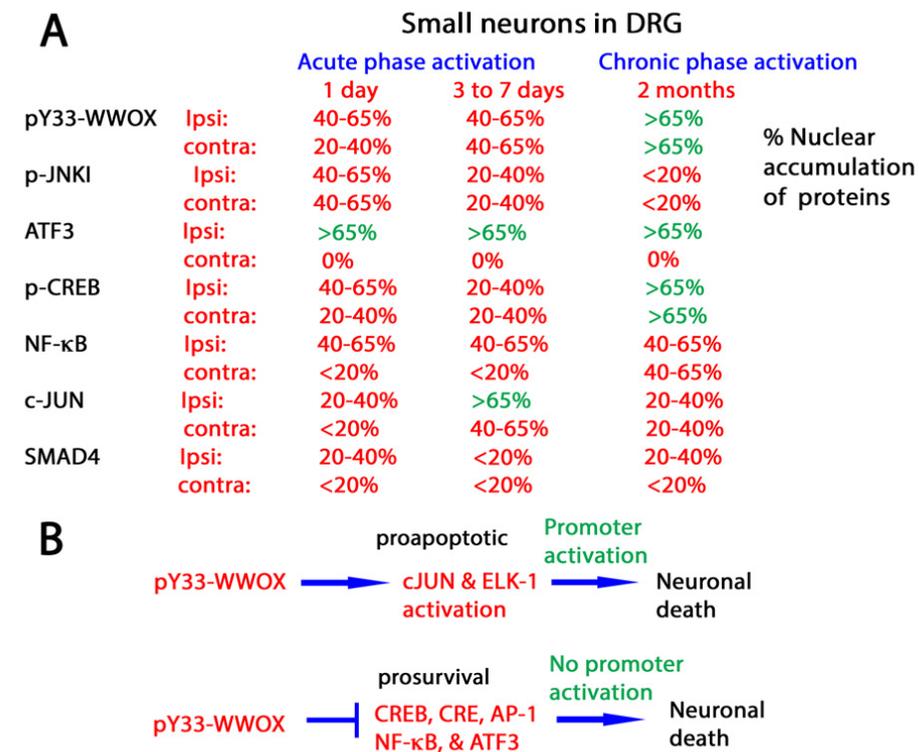


Figure 2. Nuclear accumulation of pY33-WWOX and transcription factors in small neurons in DRG during sciatic nerve dissection. (A) The extent of nuclear accumulation of an indicated protein is shown. Ipsi: ipsilateral; Contra: contralateral. (B) pY33-WWOX binds and blocks the function of JNK1, NF- κ B, CREB, and AP-1 to cause neuronal death. pY33-WWOX also enhances the promoter activation driven by c-JUN and ELK-1 to induce neuronal apoptosis. Binding of pY33-WWOX with ATF3 is unknown. Our data were tabulated and summarized from reference [50].

4.5. WWOX Modulates the Activities of Transcription Factors to Determine Neuronal Survival or Death

WWOX controls chromosomal DNA stability [52,53]. During sciatic nerve dissection, activated pY33-WWOX is co-localized in the nuclei with several transcription factors: CREB, c-JUN, ELK-1, CRE, and AP-1 [50,62]. It is very likely that these proteins co-relocate to the nuclei simultaneously. For example, at 6 hr post-sciatic nerve dissection, accumulation of pY33-WWOX goes up 40 to 65% in the nuclei of medium/large neurons in the ipsilateral DRG for at least two months [50]. p-JNK1 takes 24 h to reach a similar extent of nuclear accumulation in the medium/large neurons, followed by a reduction on day 7. c-JUN accumulation in the nuclei of medium/large neurons also takes 6 h and lasts one week only. Interestingly, ATF3 accumulation in the nuclei of medium/large neurons by more than 65% starts in 24 h and lasts two months. Transcription factors p-CREB, NF- κ B, and SMAD4 have little or no accumulation in the medium/large neurons in the ipsilateral dorsal root ganglia. Functional suppression of p-JNK1 by pY33-WWOX rapidly causes apoptosis of the medium/large neurons during the acute phase of nerve injury [48,50,63].

In vitro promoter assay revealed that transiently overexpressed WWOX activates the promoter activity driven by transcription factors c-JUN and Elk-1 [50] (Figure 2). Similarly, WW1 activates the promoter of NF- κ B [50]. Again, these interactions cause neuronal

apoptosis. In contrast, WWOX inhibits the activation of promoter elements induced by pro-survival transcription factors CREB, CRE, and AP-1, leading to neuronal death [50].

Retarded death of small neurons in two months at the chronic phase of sciatic nerve dissection is probably due to accumulation and functional balance of pY33-WWOX, ATF3, and p-CREB in the nuclei [50] (Figure 2). ATF3 is either protective or apoptosis-inducing for neurons. A nuclear calcium-CREB signaling to an ATF3-mediated neuroprotective gene repression program counteracts age- and disease-related neuronal loss [64]. Nonetheless, ATF3 modulates inflammatory responses during injury, leading to cell death [65–67].

Overall, pY33-WWOX orchestrates an array of transcription factors during sciatic nerve dissection. Binding of activated pY33-WWOX with pro-survival JNK1 [48,63], NF- κ B [50], CREB [50], and AP-1 results in inhibition of their protective function, thereby leading to neuronal death (Figure 1). Meanwhile, pY33-WWOX enhances the promoter activation driven by c-JUN and ELK-1 to induce neuronal apoptosis. pY33-WWOX physically binds c-JUN [50], whereas its binding with ELK-1 is unknown. We do not exclude the possibility of the team effort of pY33-WWOX and pS46-p53 in manipulating the function of transcription factors. pY33-WWOX binds pS46-p53, and both proteins induce apoptosis in a synergistic manner [49].

5. Zfra4-10 Activates Hyal-2/WWOX/Smad4 Signaling Pathway for Retardation of Neurodegeneration

We reported that when endogenous WWOX strongly binds intracellular protein partners in cancer cells, the cells cannot grow effectively *in vivo* [68]. When mice or cells are stimulated with the Zfra4-10 peptide, this peptide binds cell membrane hyaluronidase type II (Hyal-2) to activate the Hyal-2/WWOX/Smad4 signaling pathway [56,69–74]. The signaling activation leads to Smad4-mediated cell death or survival [69–74], cancer suppression [70,71,73,74], and restoration of memory loss in AD [71], depending upon the strength of signal activation.

5.1. Zfra Induces Activation of Spleen Hyal-2+ CD3- CD19- Z Cell

The Hyal-2/WWOX/Smad4 signaling induces spleen Hyal-2+ CD3- CD19- Z cell activation for eradicating cancer cells [70]. While Z cell function *in vivo* is largely unknown, our laboratory has been focusing on characterizing the role of Z cells in suppressing neurodegeneration in AD, PD, and seizures. Z cells are so named because Zfra activates this cell type in the spleen [70,71]. Z cells were originally isolated from T/B cell-deficient NOD-SCID mice. Z cells are not T, B, monocyte, or NK cells, but are similar in size to T and B cells. We have determined the gene expression profile of naïve Z cells (see GEO database Accession: GSE98409, ID: 200098409). Z cells are a new type of lymphoid cells. Activated Z cells enable immune-deficient NOD-SCID mice to suppress cancer growth [70,71].

Zfra-activated Z cells cause cancer cell death with or without physical contact [70]. When cancer cells are co-cultured with activated Z cells *in vitro*, Z cells undergo clonal expansion and rapidly eradicate cancer cells [68]. Without prior exposure to cancer cells or cancer cell antigens, Zfra-activated Z cells effectively induce cancer cell death [70]. Non-activated Z cells cannot kill cancer cells even though they relocate, together with T cells, to the cancer lesions [68]. Both T and Z cells group side-by-side in cancer lesions [68].

Unlike nature killer (NK) cells, naïve Z cells require pre-activation by Zfra1-31 or 4-10 or WWOX7-21. This activation allows Z cells to recognize more than ten cancer cell lines for effective eradication [70]. Thus far, Zfra1-31 or 4-10, and WWOX7-21 peptides, and antibodies against Hyal-2 or pY216-Hyal-2 are known ligands for activating Z cells [68]. Inflammatory NK cells kill cancer cells and cause damage to neurons [75–78]. In contrast, activated Z cells protect neurons from being damaged [71]. The activation markers for Z cells are being determined.

5.2. Zfra Restores Memory Loss in Triple Transgenic Mice for AD

Zfra4-10-mediated restoration of memory loss in triple transgenic mice for AD (3xTg) occurs via its inhibition of the aggregation of amyloid beta 42 (A β 42), accelerating the degradation of aggregated proteins and blocking the activation of inflammatory NF- κ B [71] (Figure 3). Zfra4-10 prevents the age-dependent progression of AD-like symptoms in 3xTg mice [72]. Whether Zfra4-10 suppresses the expansion of inflammatory glial cells in the brain is unknown. In addition to amyloid beta, there are numerous cytosolic proteins subject to degradation upon binding with Zfra—an action designated as zfraction [68,70–74].

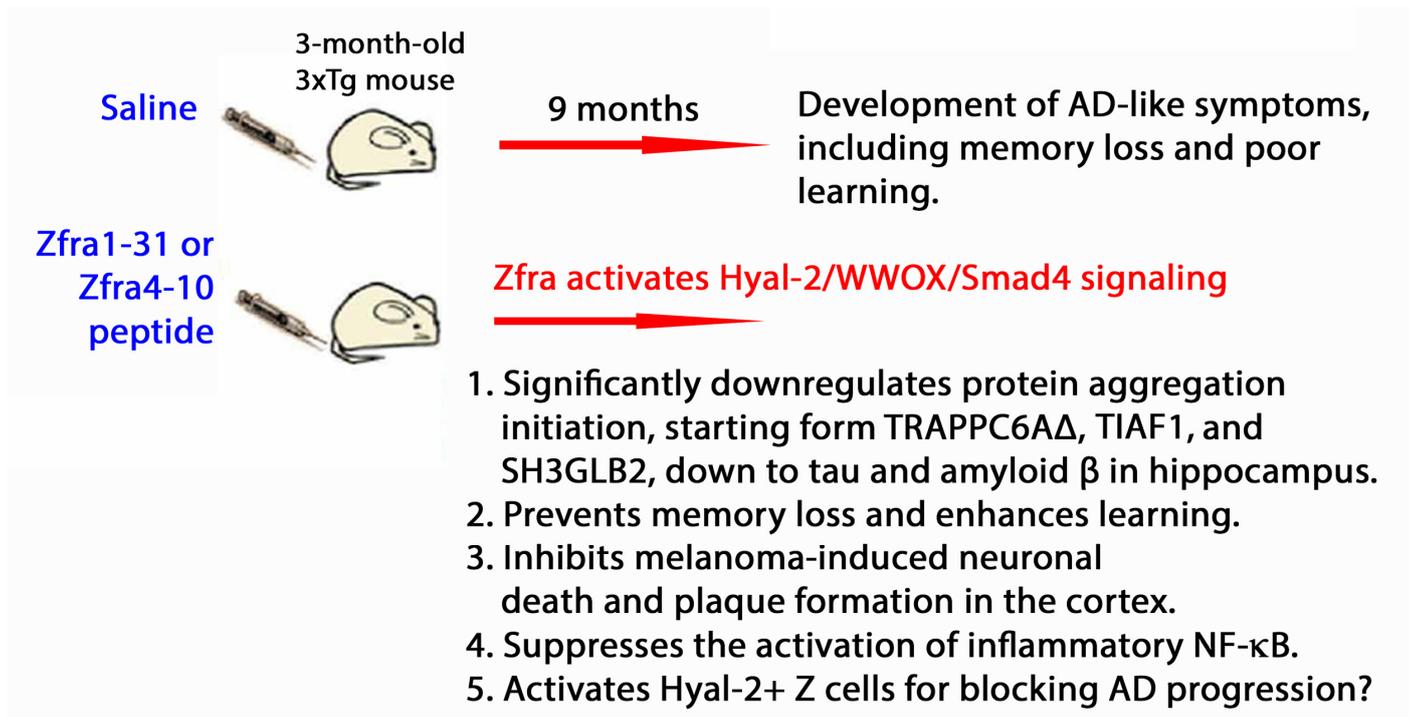


Figure 3. Zfra peptide prevents AD progression and mitigates AD-like symptoms in 3xTg mice. Zfra peptide is potent in mitigating AD-like symptoms in 9-month-old 3xTg mice [71] and prevents the age-dependent memory loss and reduced capability in learning [72].

While the action of pS14-WWOX is still being determined, it is not appropriate to consider pS14-WWOX a disabled form. pS14-WWOX is needed for T/B cell differentiation [45]. However, the pS14-WWOX-induced T/B cells cannot target cancer cells for eradication.

Full-length Zfra1-31 and the truncated Zfra4-10 undergo polymerization when the peptides are suspended in phosphate-buffered saline (PBS) but not in water [70,71]. The presence of Cys9 and Cys12 in the Zfra peptide suggests covalent binding occurs in both intra- and inter-molecular manners [70,73] (Figure 4A). Excessive polymerization of Zfra makes the peptides ineffective in blocking cancer growth [70,73]. Whether overly polymerized Zfra blocks neurodegeneration is unknown. Polymerized Zfra cannot be separated into monomers by reducing chemicals such as β -mercaptoethanol, suggesting the presence of another type of covalent binding. The likely candidate is the phosphorylation site Ser8. Compared to non-phosphorylated Zfra, pS8-Zfra is not functionally active.

Our research on Zfra peptides in mice has revealed a fascinating phenomenon. When these peptides are administered, they quickly bind with plasma proteins, forming complexes that are then sequestered in the spleen. This leads to the activation of spleen Hyal-2+ CD3- CD19- Z cells in vivo [70], a process that can last for several months due to the resistance of the Zfra/protein complex to degradation. These activated Z cells may then migrate to various organs, including the liver and lungs. However, what intrigues us is the

potential presence of Z cells in the small intestine. If they are indeed found in the gut, their activation could play a role in halting neurodegeneration via the gut–brain axis.

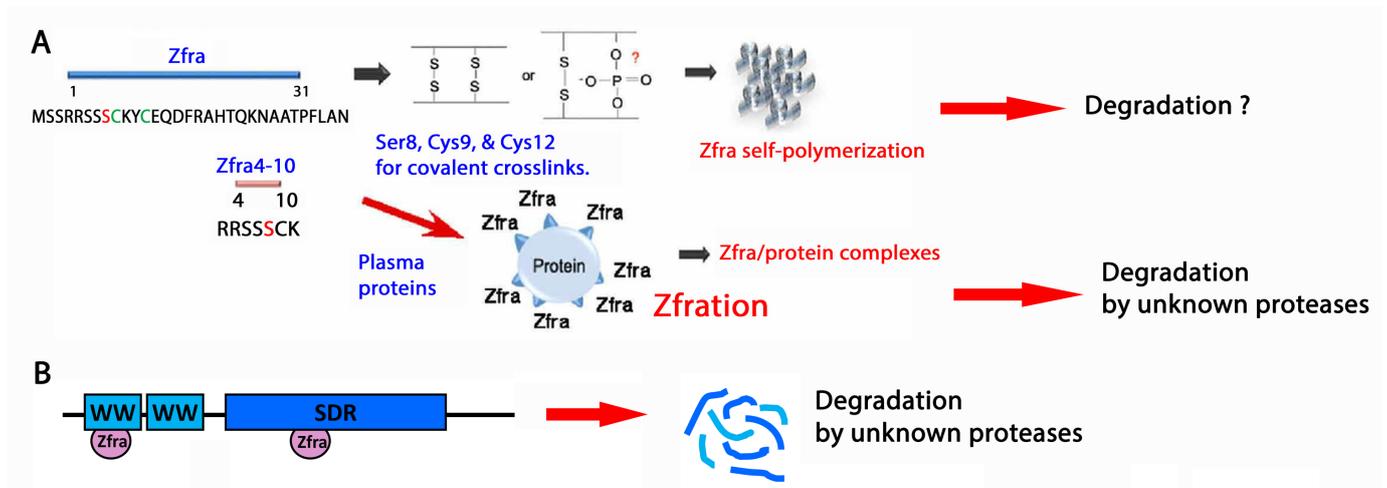


Figure 4. Zfra polymerization and protein zfractionation for degradation. (A) Zfra1-31 and 4-10 peptides can undergo self-polymerization or conjugate with cytosolic or plasma proteins (known as zfractionation). Cys9 and Cys12 are involved. Participation of Ser8 is being investigated. Degradation of the protein complexes occurs via an unknown mechanism. Known proteinase inhibitors and a proteasome inhibitor MG-132 cannot block protein complex degradation. (B) Zfra binds WW1 covalently. How Zfra binds SDR domain is still being determined. The degradation of Zfra/WWOX is not caused by ubiquitination/proteasomes or known proteases.

Additionally, activated Z cells do exist in the brain [71]. Whether these brain-activated Z cells come from the spleen is unknown. The role of brain Z cells in preventing neurodegeneration is of great interest to investigate.

5.3. Zfra Physically Interacts with Endogenous Proteins for Degradation

Zfractionation is critical for protein degradation [70–74]. Full-length Zfra1-31 proteins bind WWOX at the N-terminal WW1 domain and the C-terminal SDR domain [73], which was established by yeast two-hybrid analysis. The binding of Zfra1-31 with the recombinant WW1 domain is covalent [68] (Figure 4B). Whether the Zfra/SDR complex is covalent needs investigation. When cell lines were used to analyze Zfra-mediated protein degradation, Zfra peptides were added to the cell lysates. Covalent Zfra/WWOX or protein complexes can be seen by observing banding patterns with reducing and non-reducing gels. Compared to control cell lysates, Zfra enhances protein degradation with time. Known proteinase inhibitors and a proteasome inhibitor MG-132 cannot block the protein degradation, suggesting that a novel protease degrades the zfractionated proteins.

5.4. Zfra4-10 Covalently Interacts with WWOX7-21

A portion of cytosolic WWOX is localized on the cell membrane, and WWOX7-21 is a surface-exposed epitope [74]. Synthetic Zfra4-10 and WWOX7-21 peptides powerfully suppress the growth and metastasis of 4T1 breast cancer cells in mice [68]. However, when combined, both Zfra4-10 and WWOX7-21 peptides lose their function in blocking 4T1 growth in mice, suggesting that both peptides functionally antagonize each other via covalent binding [68].

5.5. Zfra4-10 and WWOX7-21 Strengthen the Binding of Endogenous WWOX with Target Proteins in Stem Cells, Microglia Cells, Astrocytes, and Treg Cells and in Exosomes In Vivo

We reported that the more strongly endogenous WWOX binds intracellular protein partners, the more weakly the cancer cells can grow in vivo [68]. When mice receive either Zfra4-10 or WWOX7-21 peptide via tail vein injections, endogenous WWOX strongly

increases its binding (1- to 7-fold increases) with C1qBP, CD133, p21, JNK1, COX2, p-ERK, Foxp3, and p53 in the spleen cells, as determined by co-immunoprecipitation using organ lysates (Figure 5A,B). The binding correlates with suppression of cancer growth [68] (Figure 5A–C). The stronger the pY33-WWOX binding of partner proteins, the better the suppression of cancer growth *in vivo* [68] (Figure 5D). In stark contrast, when mice receive both Zfra4-10 and WWOX7-21 peptides, a dramatically reduced binding of WWOX with the target proteins occurs, down to the baseline level or none. In the lung, Zfra4-10 or WWOX7-21 peptide alone also strongly enhances the binding of endogenous WWOX with Iba1, Oct4, ERK1/2, NF- κ B p65, GFAP, and p53 (2- to 5-fold increases).

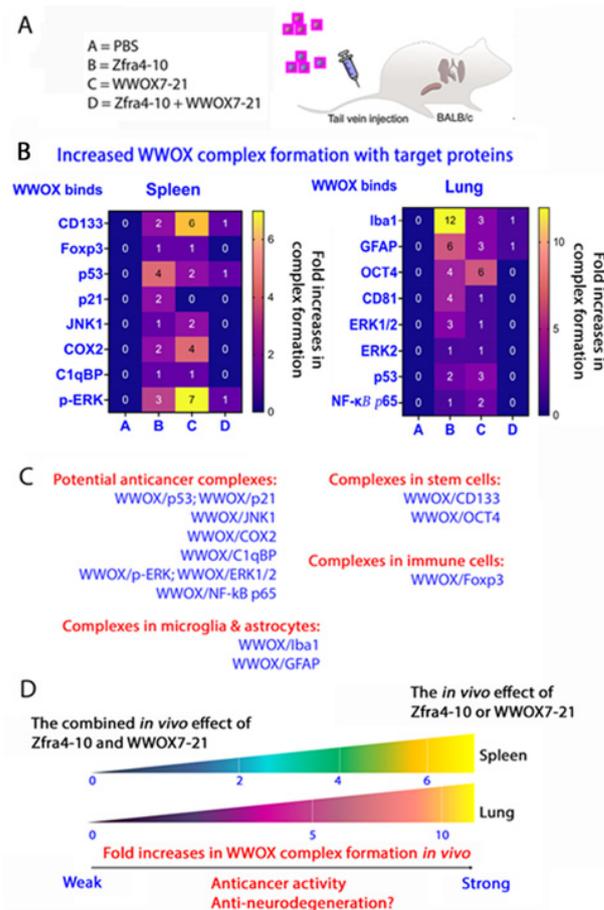


Figure 5. Zfra4-10 or WWOX7-21 upregulates the complex formation of WWOX with target proteins *in vivo*. (A) BALB/c mice received tail vein injections of Zfra4-10 and/or WWOX7-21 peptides, followed by inoculation with 4T1 breast cancer cells two weeks later and sacrifice 18 days later. By co-immunoprecipitation, increased binding of WWOX with C1qBP and other indicated proteins was shown in the spleen and lung. These complexes are present in indicated cells and exosomes. No induction of complex formation was shown when mice received the mixture of Zfra4-10 and/or WWOX7-21 peptides. Functionally, WWOX and p53 may synergistically induce apoptosis [30,48,49]. JNK1 counteracts WWOX function in inducing apoptosis [48]. The binding of WWOX with JNK1 and ERK leads to inhibition of these enzymes' activity in hyperphosphorylating tau [23]. (B) Overall, when endogenous WWOX binds strongly with interacting proteins, the extent of cancer growth is dramatically reduced *in vivo*. (C) The functional properties of WWOX and target protein complexes are shown. (D) Zfra4-10 or WWOX7-21 induces anti-cancer responses, which is due to the increased binding strength of pY33-WWOX with its partner proteins, which confers anti-cancer activity *in vivo* [68].

Upregulation of CD133 or OCT4 expression indicates expanding stem cell populations in the spleen. Foxp3-positive T regulatory (Treg) cells are also expanded in the spleen.

Interestingly, Iba1-positive microglia cells and GFAP-positive astrocytes are present in the lungs. Whether these cells can relocate to the brain is unknown.

While we used organ lysates for co-immunoprecipitation, we will repeat the experiments using tissue sections and examine the binding via antibody FRET imaging or affinity proximity assay. This will detail not only the binding affinity, but also the extent of prevalence in binding among cell types.

5.6. Inhibition of S14 Phosphorylation in WWOX by Zfra4-10 or WWOX7-21 Leads to Reduced Neurodegeneration

Overexpressed pY33-WWOX is proapoptotic, whereas an optimal level of pY33-WWOX is essential for maintaining normal physiology [48,49]. When the level of pY33-WWOX goes down, the pS14-WWOX level goes up in vivo [70–72]. pS14-WWOX supports the progression of cancer development, growth, and metastasis [70] and enhances the progression of AD [71,72]. pS14-WWOX is accumulated in the lesions of cancers and AD brains. Zfra4-10 strongly reduces the level of pS14-WWOX and thereby abolishes cancer growth and retardation of AD progression [70–72]. WWOX7-21 is also potent in blocking WWOX phosphorylation at S14. Conceivably, both Zfra4-10 and WWOX7-21 induce or activate a specific phosphatase(s) that causes pS14 dephosphorylation. The specific phosphatase(s) has yet to be identified.

While the action of pS14-WWOX is still being determined, it is not appropriate to consider pS14-WWOX as a disabled form. pS14-WWOX is needed for T/B cell differentiation [45]. However, induced T/B cannot block cancer growth.

6. Dramatic Upregulation of pY33- and pY287-WWOX in the Brain Cortex of Heterozygous *Wwox* Mice

Thus far, no reports have demonstrated that WWOX becomes aggregated in the brain. Compared to wild-type mice, heterozygous *Wwox* mice express half as much WWOX protein in tissues and organs. Most interestingly, WWOX undergoes enhanced phosphorylation at Y33 and Y287 in the brain cortex of heterozygous *Wwox* mice (~1-fold increase), but not in the brains of wild-type mice [72]. The increased phosphorylation of Y33 and Y287 in WWOX in the heterozygous mice is necessary to maintain normal physiological functions. In general, heterozygous *Wwox* mice normally behave like wild-type mice, although these mice have an accelerated neurodegeneration [71]. No increase in S14 phosphorylation of WWOX is shown [78], suggesting an inter- or intra-molecular autoregulatory mechanism for WWOX functional activation and turnover [57]. That is, binding interactions can be found for the WW or SDR domains by itself or WW/SDR self-folding or intermolecular interactions [57]. Both pY33- and pY287-WWOX do not exhibit aggregation in the brain cortex. ACK-1 phosphorylates WWOX at Y287, and pY287-WWOX can be ubiquitinated for proteasomal degradation [79]. Another report showed that when cells are under DNA single-strand break checkpoint activation, WWOX is ubiquitinated at K274 by the ubiquitin E3 ligase ITCH and interacts with ataxia telangiectasia-mutated (ATM) [53]. WWOX can undergo modification by small ubiquitin-like modifier (SUMO) proteins. As a result, the SUMOylated WWOX blocks prostate cancer growth [80].

6.1. pT12-WWOX Is an Aggregated form in the AD Brain Lesions

We have determined for the first time that pT12-WWOX aggregates or plaques are found in the neocortices and cortices of 11-month-old heterozygous *Wwox* mice but not in those of wild-type mice [72] (Figure 6A). The pT12-WWOX aggregates are about 30 to 60 μm in diameter, which are formed most likely at an earlier age in mice or at middle age in humans. Presumably, pT12-WWOX aggregates cause the activation of the protein aggregation cascade starting from the polymerization of TRAPPC6A Δ (or TPC6A Δ), followed by TIAF1 (TGF β -induced antiapoptotic protein), and SH3GLB2 (SH3 domain-containing GRB2 Like, endophilin B). The resulting complexes lead to caspase activation, degradation of amyloid precursor protein (APP), generation of amyloid beta, and neuronal apopto-

sis [22,81,82] (Figure 6B). Compared to the wild-type, TPC6AΔ isoform has an internal deletion of 14 amino acids in the N-terminus [45].

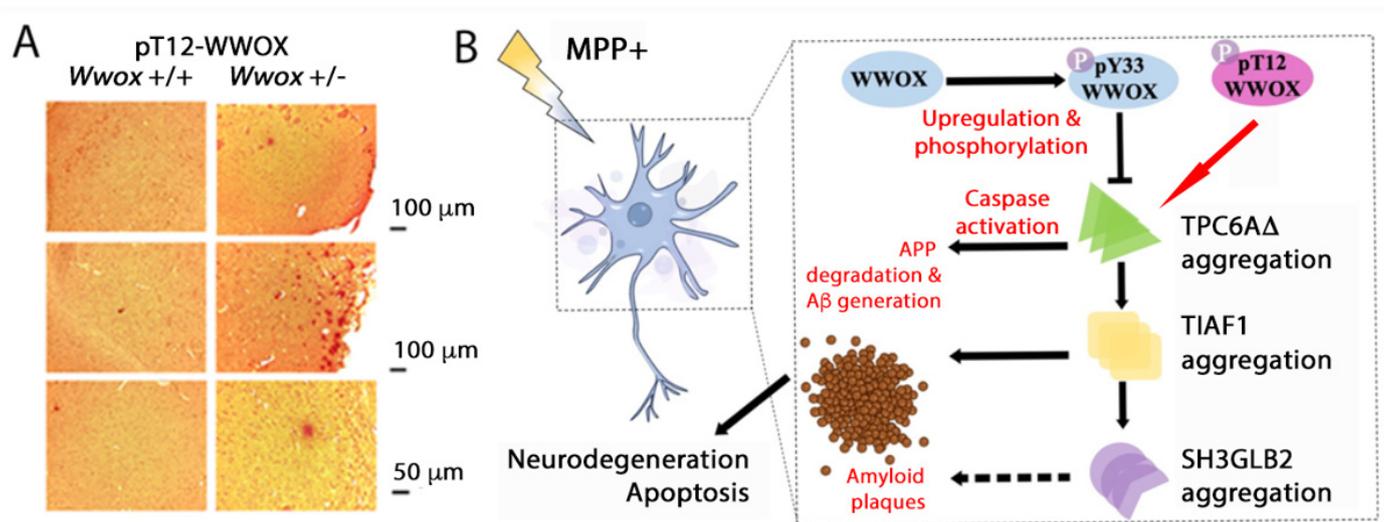


Figure 6. pT12-WWOX as aggregates may initiate the cascade of protein aggregation, leading to neurodegeneration. Identification of pT12-WWOX as aggregates in the brain cortices of 11-month-old heterozygous *Wwox* mice. (A) Presence of pT12-WWOX aggregates is shown in the cortices of heterozygous *Wwox* mice (see the red punctate). No pY287-, pY33- and pT12-WWOX aggregates were found in either the wild-type or the heterozygous *Wwox* mice. (B) A schematic graph for the action of pY33- and pT12-WWOX, which modulate the protein aggregation cascade of TPC6AΔ, TIAF1, and SH3GLB2. Stimulation of cells with a neurotoxin MPP⁺ causes neuronal apoptosis. Aggregation of TPC6AΔ, TIAF1, and SH3GLB2 occurs during MPP⁺-mediated cell death, suggesting that TPC6AΔ is a common initiator for AD and PD [72].

6.2. pT12- and pS14-WWOX in Controlling Protein Aggregation?

The protein aggregation cascade may start in middle age and keep going for 20 to 30 years before the appearance of Alzheimer's disease symptoms [22,81,82]. pS14-WWOX is involved in disease progression [31,61]. While T12 and S14 are close to each other, their relationship regarding phosphorylation and conformational changes of WWOX is unclear. However, both amino acid residues are involved in AD progression. In principle, pY33-WWOX blocks the aggregation of TPC6AΔ and related downstream proteins [22,81,82]. pT12-WWOX is likely to stimulate the aggregation cascade of TPC6AΔ, TIAF1, and SH3GLB2. When neuroblastoma SK-N-SH cells are treated with neurotoxin 1-methyl-4-phenylpyridinium (MPP⁺) for inducing PD-like symptoms, TPC6AΔ expression is increased and becomes polymerized, which causes aggregation of TIAF1, SH3GLB2, Aβ, and tau [72]. Similarly, transforming growth factor beta (TGF-β) stimulates the polymerization of TPC6AΔ, TIAF1, and SH3GLB2 in the aggregation cascade. Notably, Zfra4-10 peptide blocks the MPP⁺ effects and thereby sustains the survival of neurons.

Taken together, when WWOX protein is downregulated in the brain cortex, TPC6AΔ is expected to have an accelerated aggregation as shown in the *Wwox* knockout mice. The aggregated TPC6AΔ is at the crossroad for leading to the progression of AD or PD [72]. How TPC6AΔ directs the signaling for going to AD or PD remains to be determined. Alternatively, upregulation of pT12- or pS14-WWOX and downregulation of pY33-WWOX may compete for turning on or off to the road for AD or PD.

7. WWOX in Embryonic Development and AD Progression

WWOX plays a crucial role in affecting embryonic development. Functional deficiency or defects in the *WWOX* gene and protein lead to severe neural disorders, metabolic disorders, mental retardation, neurodegeneration, immune defects, stunted growth, and early

death [46,47,72,83,84]. WWOX gene deficiency from bi-allelic alterations in chromosome 16q leads to autosomal recessive spinocerebellar ataxia 12 (SCAR12) and WWOX-related epileptic encephalopathy (WOREE) syndromes [46,47,72,83]. Children suffering from SCAR12 have early childhood onset of cerebellar ataxia and difficulty in coordinating the movement of their muscles. WOREE is caused by bi-allelic gene mutations. Newborn patients suffer intractable epileptic seizures, severe defective development in the neural system, and movement disorders.

The WWOX gene is also regarded as a risk factor for AD [85] and is associated with the progression of PD [86]. In AD, downregulation of WWOX occurs in the hippocampus in middle age, leading to very slow and gradual aggregation of a protein cascade group over 20 to 30 years [22,78,81,82]. Balanced phosphorylation among T12, S14, Y33, and probably Y34 in the WW1 is critical to limiting disease manifestation [70,71]. For example, when Y33 phosphorylation in WWOX is downregulated, S14 phosphorylation is then upregulated. Aggregation of pT12-WWOX is likely blocked by pS14- or pY33-WWOX. Conceivably, pY34 inhibits the action of pY33. pS14-WWOX promotes the progression of cancer and AD [70,71]. Nonetheless, these presumed functions remain to be established.

Furthermore, tau hyperphosphorylating enzymes JNK and ERK can be bound and functionally blocked by Y33-phosphorylated WW1. WWOX prevents tau hyperphosphorylation by these enzymes [23]. The SDR domain binds GSK3 β and the C-terminal region of tau [23,87]. This binding leads to inhibition of tau hyperphosphorylation [23,87].

8. WWOX Antagonizes p53 for Inducing Neurodegeneration In Vivo

Protein functional antagonism may lead to the development of neurodegeneration. For example, both p53 and WWOX may functionally antagonize in regulating cancer growth, and the antagonism can lead to neurodegeneration in vivo [88]. To achieve the effect of neurodegeneration, p53 and/or WWOX cDNA expression constructs are overexpressed in p53-deficient lung cancer cells and are then inoculated in mice. Ectopic WWOX strongly inhibits lung cancer growth and inflammatory reaction in vivo [88]. In contrast, p53 does not have an inhibitory effect. In combination, the inhibitory function of WWOX is nullified by p53. Most strikingly, when mice receive p53/WWOX-expressing lung cancer cells, these mice are shown to have BACE (β -secretase 1) upregulation, APP degradation, tau tangle formation, and amyloid β generation in the brain and lung [88]. Hence, the functional opposition between p53 and WWOX leads to enhanced cancer growth and accelerated neurodegeneration in vivo [31,46,88]. Under stress conditions, pS46-p53 physically interacts with pY33-WWOX [49]. This strongly associated complex appears to be mainly responsible for inducing apoptosis [49].

However, when pY33-WWOX is de-phosphorylated and S14 phosphorylated, the pS46-p53/pS14-WWOX complex could have an opposite effect. Thus, upregulation of pS14-WWOX facilitates the growth of cancer and the progression of AD [70,71]. The specific kinase and phosphatase involved in pY33-WWOX dephosphorylation and pS14-WWOX phosphorylation is unknown. Finally, Zfra4-10 or WWOX7-21 downregulates pS14-WWOX [68,70,71], suggesting that either peptide may covalently bind pS46-p53/pS14-WWOX complex for proteolytic degradation.

9. Membrane Epitopes WWOX7-21 and WWOX286-299 and Functional Implications

WWOX is located ubiquitously in intracellular locations. Computational prediction does not show that WWOX protein has a membrane localization signal sequence. Membrane Hyal-2 binds the WW1 domain and anchor WWOX to the membrane area [56,69]. In addition, WWOX is anchored by Ezrin to the cytoskeletal-membrane area [89]. WWOX can undergo self-polymerization in the cell membrane, as confirmed by immunoelectron microscopy [56,69]. The self-polymerization is due to binding of WW1 with WW1, SDR with SDR, or WW1 with SDR in an intermolecular or intramolecular manner [57].

9.1. WWOX7-21 Epitope Confers Cancer Suppression and Probably Blocks AD Progression

Structurally, there are two epitopes in the membrane-localized WWOX, which are at amino acid 7 to 21 and 286 to 299. Synthetic peptides WWOX7-21 and WWOX286-299 and corresponding antibodies were used in functional characterization [57,90]. Treatment of 4T1 breast cancer cells with antibody against the WWOX7-21 epitope makes the cells susceptible to ceritinib, UV irradiation, and many chemotherapeutic drugs [73]. In contrast, WWOX7-21 peptide significantly increases the death of cultured breast 4T1 cells and cell sphere explosion mediated by ceritinib [73]. This peptide, which is as short as WWOX7-11, has even stronger potency in suppression and prevention of the growth and metastasis of melanoma and skin cancer cells in mice [73]. Both peptides probably mediate the signaling from membrane Hyal-2/WWOX and then recruit Smad4 for blocking cancer growth. WWOX7-21 peptide can self-polymerize [57,90], suggesting that the exogenous WWOX7-21 peptide binds the membrane WWOX7-21 epitope and probably Hyal-2 for initiating the signaling. Hyal-2 physically binds the N-terminal first WW domain in a Y33 phosphorylation dependent manner [56]. Alteration of Y33 in WWOX probably abolishes its binding with Hyal-2.

Additionally, WWOX7-21 peptide reduces ERK phosphorylation, upregulates proapoptotic pY33-WWOX, induces calcium ion influx, and abolishes I κ B α /WWOX/ERK pro-survival signaling [90]. Consequently, these events lead to cancer suppression. While pS14-WWOX supports cancer growth and enhances AD progression, pS14-WWOX7-21 peptide significantly enhances cancer growth in vivo and blocks ceritinib-mediated apoptosis in vitro [73]. In parallel, antibody against pS14-WWOX7-21 peptide reduces cancer growth [73]. Zfra peptide is potent in inducing neuroprotection [71]. Like Zfra peptide, WWOX7-21 peptide activates the Hyal-2/WWOX/Smad4 signaling, suggesting that WWOX7-21 can exert neuroprotection.

9.2. WWOX7-21 and WWOX286-299, along with Membrane Type II TGF β Receptor (T β RII), Control Cell Migration and Cell-Cell Recognition

Cells expressing functional WWOX (WWOXf) migrate collectively and can fend off the individually migrating WWOX-deficient or -dysfunctional (WWOXd) cells to undergo retrograde migration [57,90]. Interestingly, the retrograde-migrating WWOXd cells kill a portion of WWOXf cells from a remote distance, without physical contact. Mechanistically, WWOXd cells dramatically increase the redox activity in WWOXf cells, thereby causing cell death [90]. WWOXd cells utilize the I κ B α /ERK/WWOX signaling to survive [57,90].

Specifically, when cells have surface exposure to the WWOX7-21 epitope, these cells attract any approaching WWOXf or WWOXd cells of the same or different species [57]. We have identified MIF (macrophage migration inhibitory factor) as an extracellular sensing molecule [90]. When cells are exposed to WWOX286-299 epitope on their surface, these cells strongly fend off WWOXd cells [57]. Membrane WWOX also binds membrane type II TGF β receptor (T β RII). Stimulating WWOXf cells with T β RII IgG antibodies leads treated cells to greet WWOXd cells to merge with each other. However, when WWOXd are pretreated with T β RII IgG antibody, these cells lose recognition by WWOXf cells. WWOXf cells can kill the T β RII antibody-treated WWOXd cells. The observations suggest that normal cells can be activated to attack metastatic cancer cells. Together, membrane WWOX/T β RII complex is needed for cell-to-cell recognition, maintaining the efficacy of Ca²⁺ influx and control of cell invasiveness.

9.3. WWOX in Cortical Neuron Migration and Loss of WWOX Causes Neuronal Heterotopia

Whether WWOX participates in deciding neuronal migration and positioning in the brain cortex during embryonic development is largely unknown. However, the fact is that newborn patients with WWOX deficiency develop neuronal heterotopia and severe epileptic seizures [83,84,91–93]. Without WWOX protein, neurons accelerate their migration and accumulate in the neocortex, known as neuronal heterotopia [84,94,95]. Moreover, reduced GABA-ergic inhibitory interneurons has been demonstrated in *Wwox* knockout

mice [96]. Many genes in common fragile sites are unstable and tend to undergo deletions and alterations [97]. WWOX, DAB1, and many proteins are involved in neuronal migration and lamination in the developing cerebral cortex [97–100].

Here, we propose that during cortical lamination, WWOX controls neuronal layer development via transition between epitope exposure. For example, surface exposure of the WWOX7-21 epitope in a group of cells allows the other cell group to migrate forward, and the WWOX286-299 epitope signals the other cells to stop moving. This scenario is based upon the aforementioned findings [57,90]. Nonetheless, the relationship can be complicated by phosphorylation in the epitopes such as pT12, pS14, and pY287. Given the role of Zfra in suppressing WWOX phosphorylation at T12, S14 and Y287 and inhibiting the formation of neuronal heterotopia [84], Zfra is promising in restoring neuronal functions.

10. Z Cells Have Memory Function in Eliminating Cancer Cells

10.1. Z Cell Activation by Zfra4-10, WWOX7-21, and Sonicated Hyaluronan HAson8 to Kill Cancer Cells and Retard AD Progression

Spleen Z cell activation is critical in killing cancer cells. Z cells can be activated by Zfra4-10, Zfra1-31, and WWOX7-21 peptides [68,73]. In addition, 8-h-sonicated hyaluronan (HAson8) and antibody against Hyal-2 or pY216-Hyal-2 activate Z cells to kill cancer cells [68]. Native hyaluronan has no effect [68]. These agonists bind membrane Hyal-2 and then activate the Hyal-2/WWOX/Smad4 signaling in Z cells [68]. Z cell activation requires dephosphorylation of WWOX at S14, Y33 and Y61 [45,61]. Activated Z cells exhibit memory function. How the agonists establish anti-cancer memory function in Z cells remains to be determined, because Z cells do not require pre-exposure to cancer antigens. How Z cells acquire cytotoxic capability in eradicating cancer cells in vivo and in vitro is also yet to be revealed [68]. By the same token, we strongly believe that activated Z cells have the ability to protect neurons from being damaged or degenerated.

10.2. Zfra4-10 Activates Z Cells in Immune Deficient NOD-SCID Mice

Zfra activates Z cells in normal mice and T/B cell-deficient NOD-SCID and T-deficient nude mice [68,70,71]. Zfra-activated Z cells relocates to cancer lesion to eliminate cancer cells in both immune efficient and deficient mice [68,70,71]. That is, activated Z cells restore the immune response in immunodeficient mice to limit cancer growth and probably retard age-dependent neurodegeneration.

A population of Z cells are located in the brain. This cell population can be isolated by flow cytometry. Whether these cells come from other organs or are the residential cells in the brain is unknown. Once transferred to tumor-growing mice, the purified brain-activated Z cells cause cancer cell death. Their role in mitigating the AD-like symptoms in mice is being investigated. Importantly, we are now able to activate the Z cell immune response in so-called immune-deficient mice for protection against neurodegeneration, cancer, and probably diabetes.

10.3. Zfra1-31 for Treating Hyperglycemia/Diabetes-Associated Neurodegeneration

A recent report determined that Zfra1-31 potently treats hyperglycemia/diabetes-associated neurodegeneration [101]. Diabetic rats at six months old have high levels of activated WWOX in the brain cortex and hippocampus compared to normal rats. The association of WWOX with the regulation of glucose metabolism has been reported [102]. Indeed, the WWOX/HIF1A axis is involved in altering glucose metabolism [102–107]. Substantial evidence from a genome-wide association study (GWAS) revealed the critical association of WWOX with diabetes and glucose metabolism [108–112]. For example, when the WWOX/HIF1A axis is downregulated, glucose metabolism is altered, and the changes may allow the development of metabolic disorders. Also, in early diabetic retinopathy, the knockdown of WWOX by specific siRNA in vitro suppresses superoxide production caused by the high glucose in photoreceptors from diabetic mice for two months [112]. Ablation of the WWOX gene in skeletal muscle leads to altered glucose metabolism [106].

WWOX becomes Y33-phosphorylated in response to high glucose, which leads to mitochondrial apoptosis. Indeed, overly expressed pY33-WWOX can induce apoptosis in vivo [50,51,58]. Based upon our observations [68,70,71], Zfra blocks the phosphorylation and covalently binds WWOX for degradation [46,72]. As a result, activated WWOX-mediated mitochondrial apoptosis is blocked. Overall, this finding is important and yet not surprising for Zfra. Zfra can be a universal drug for targeting many diseases via Hyal-2/WWOX/Smad4 signaling.

11. Summary and Concluding Remarks

We explored the possibility of Zfra in restoring cell function under WWOX functional deficiency or absent expression. We determined that Zfra overrides WWOX in suppressing cancer growth and the progression of neurodegeneration. First, a key mechanism is that Zfra4-10, WWOX7-21, and HAsOn8 induce the memory anti-cancer Z cells [24–28] (Route i) (Figure 7). Whether other types of immune cells are involved remains to be identified. Second, Zfra4-10 and WWOX7-21 induce the intracellular formation of the WWOX complex with target proteins [68] (Route ii). The stronger the binding of WWOX with target proteins, the better the suppression of cancer growth and retardation of neurodegeneration [68]. The turnover rate of WWOX-regulated protein complexes is unknown and remains to be elucidated. Finally, Zfra rapidly and covalently conjugates with target proteins by zfration [70,71], and the resulting complexes undergo degradation independently of ubiquitination and proteasomal degradation (Route iii). It is unclear whether the WWOX-conjugated protein complexes are stable and resist zfration for further degradation or the complexes are subjected to ubiquitination/proteasomal degradation. These scenarios remain to be determined.

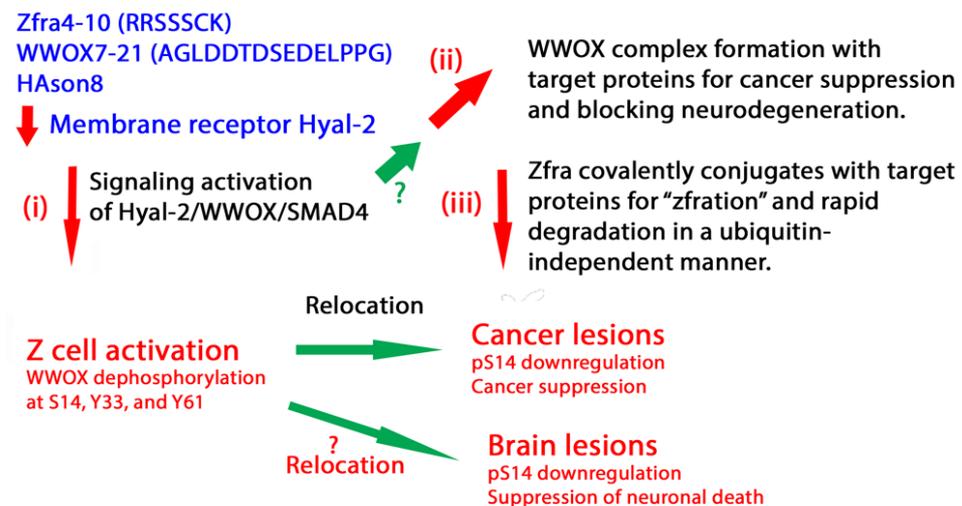


Figure 7. Agonists for activating Hyal-2/WWOX/Smad4 signaling. Suppression of neurodegeneration and cancer growth by Zfra4-10, WWOX7-21, or HAsOn8 involves (i) activation of Z cells [26], which requires dephosphorylation of pS14, pY33, and pY61 in WWOX, (ii) enhanced complex formation of intracellular WWOX with binding proteins; and (iii) rapid degradation of Zfra-conjugated proteins.

11.1. pT12-WWOX as an Initiator of Protein Aggregation Cascade?

pS14-WWOX supports the progression of many diseases such as cancer and Alzheimer's disease [70,71]. pS14-WWOX does not appear to undergo aggregation. A recent intriguing discovery is that pT12-WWOX occurs as aggregates in the cortex of heterozygous *Wwox* mice, but not in wild-type mice [78]. How pT12-WWOX becomes aggregated is unknown. pT12-WWOX aggregates could be in the middle of an aggregation cascade. Alternatively, as an initiator, phosphorylated-T12 in the first WW domain area undergoes non-stop self-polymerization.

Whether the co-presence of pS14, pY33, pY34, and pY287 with pT12 can occur in the same WWOX molecule is unknown. Perhaps co-phosphorylation among all the phosphorylation sites in WWOX is necessary to stabilize the protein and prevent self-aggregation. The WW1 domain has antiparallel β -sheets, which can undergo self-binding [57]. The crucial conditions, such as changes in intracellular pH, ion concentrations, and abnormal WWOX-binding proteins, may lead WWOX to extensively self-polymerize via the first WW domain and aggregate eventually.

While T12 and S14 are close to each other, we suspect pT12 may nullify the pathogenic function of pS14 in WWOX. Under this case, pT12 reduces the severity of pS14 in supporting disease progression such as AD development [5,53,56]. What remains to be established are the scenario and molecular mechanisms for the induction of TPC6A Δ polymerization by pT12-WWOX aggregates and the degradation of pT12-WWOX aggregates by Zfra.

Another intriguing finding is that pY33-WWOX can be released from the neurons to the extracellular matrix in the cortex [72]. pY33-WWOX is critical in maintaining cellular functions and stabilizing the normal physiology of mitochondria with p53 [9]. A portion of cytosolic pY33-WWOX is localized to the cell membrane [57]. Release of this protein is likely. We will examine whether the released pY33-WWOX binds amyloid plaques in the extracellular matrix.

11.2. Concluding Remarks

Synthetic Zfra peptides, Zfra1-31 or Zfra4-10, effectively prevent and block cancer growth in vivo. Zfra4-10 strongly retards the progression of AD-like symptoms in 3xTg mice during aging [71]. For treatment, Zfra4-10 peptide restores memory loss in 9-month-old 3xTg mice by blocking the aggregation of a group of initiating proteins, which aggregate like a cascade. Zfra activates Hyal-2+ CD3- CD19- Z cell to cause cancer cell death. Z cells are downstream effectors of Zfra-mediated inhibition of neurodegeneration and have great potential for cell therapy in AD or other neurodegenerative diseases. WWOX-deficient newborns suffer severe neurodegeneration. Zfra may override WWOX deficiency in restoring normal physiological functions.

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References

1. Platten, M.; Nollen, E.A.A.; Röhrig, U.F.; Fallarino, F.; Opitz, C.A. Tryptophan metabolism as a common therapeutic target in cancer, neurodegeneration and beyond. *Nat. Rev. Drug Discov.* **2019**, *18*, 379–401. [[CrossRef](#)] [[PubMed](#)]
2. Dikic, I.; Elazar, Z. Mechanism and medical implications of mammalian autophagy. *Nat. Rev. Mol. Cell Biol.* **2018**, *19*, 349–364. [[CrossRef](#)]
3. Navas, L.E.; Carnero, A. NAD(+) metabolism, stemness, the immune response, and cancer. *Signal Transduct. Target. Ther.* **2021**, *6*, 2. [[CrossRef](#)]

4. Alberti, S.; Dormann, D. Liquid-Liquid Phase Separation in Disease. *Annu. Rev. Genet.* **2019**, *53*, 171–194. [[CrossRef](#)]
5. Taniue, K.; Akimitsu, N. Aberrant phase separation and cancer. *FEBS J.* **2022**, *289*, 17–39. [[CrossRef](#)] [[PubMed](#)]
6. Fuchs, Y.; Steller, H. Programmed cell death in animal development and disease. *Cell* **2011**, *147*, 742–758. [[CrossRef](#)] [[PubMed](#)]
7. Dos Santos, J.C.C.; Rebouças, C.; Oliveira, L.F.; Cardoso, F.D.S.; Nascimento, T.S.; Oliveira, A.V.; Lima, M.P.P.; de Andrade, G.M.; de Castro Brito, G.A.; de Barros Viana, G.S. The role of gut-brain axis in a rotenone-induced rat model of Parkinson's disease. *Neurobiol. Aging* **2023**, *132*, 185–197. [[CrossRef](#)]
8. Sun, N.; Victor, M.B.; Park, Y.P.; Xiong, X.; Scannail, A.N.; Leary, N.; Prosper, S.; Viswanathan, S.; Luna, X.; Boix, C.A.; et al. Human microglial state dynamics in Alzheimer's disease progression. *Cell* **2023**, *186*, 4386–4403. [[CrossRef](#)]
9. Shi, X.; Li, L.; Liu, Z.; Wang, F.; Huang, H. Exploring the mechanism of metformin action in Alzheimer's disease and type 2 diabetes based on network pharmacology, molecular docking, and molecular dynamic simulation. *Ther. Adv. Endocrinol. Metab.* **2023**, *14*, 20420188231187493. [[CrossRef](#)]
10. Gulen, M.F.; Samson, N.; Keller, A.; Schwabenland, M.; Liu, C.; Glück, S.; Thacker, V.V.; Favre, L.; Mangeat, B.; Kroese, L.J.; et al. cGAS-STING drives ageing-related inflammation and neurodegeneration. *Nature* **2023**, *620*, 374–380. [[CrossRef](#)]
11. Baechle, J.J.; Chen, N.; Makhijani, P.; Winer, S.; Furman, D.; Winer, D.A. Chronic inflammation and the hallmarks of aging. *Mol. Metab.* **2023**, *74*, 101755. [[CrossRef](#)] [[PubMed](#)]
12. Kasi, P.D.; Tamilselvam, R.; Skalicka-Woźniak, K.; Nabavi, S.F.; Daglia, M.; Bishayee, A.; Pazoki-Toroudi, H.; Nabavi, S.M. Molecular targets of curcumin for cancer therapy: An updated review. *Tumour Biol.* **2016**, *37*, 13017–13028. [[CrossRef](#)] [[PubMed](#)]
13. Fu, Y.S.; Chen, T.H.; Weng, L.; Huang, L.; Lai, D.; Weng, C.F. Pharmacological properties and underlying mechanisms of curcumin and prospects in medicinal potential. *Biomed. Pharmacother.* **2021**, *141*, 111888. [[CrossRef](#)] [[PubMed](#)]
14. Juric, M.; Rawat, V.; Amaradhi, R.; Zielonka, J.; Ganesh, T. Novel NADPH Oxidase-2 Inhibitors as Potential Anti-Inflammatory and Neuroprotective Agents. *Antioxidants* **2023**, *12*, 1660. [[CrossRef](#)] [[PubMed](#)]
15. Shamsnia, H.; Samanian, A.; Sabbagh Kashani, A.; Khayatan, D.; Momtaz, S.; Johnston, T.P.; Majeed, M.; Jamialahmadi, T.; Abdolghaffari, A.H.; Sahebkar, A. An Overview of the Pharmacological Properties of Calebin-A. *Curr. Med. Chem.* **2023**. [[CrossRef](#)] [[PubMed](#)]
16. Mueller, A.L.; Brockmueller, A.; Kunnumakkara, A.B.; Shakibaei, M. Calebin A, a Compound of Turmeric, Down-Regulates Inflammation in Tenocytes by NF- κ B/Scleraxis Signaling. *Int. J. Mol. Sci.* **2022**, *23*, 1695. [[CrossRef](#)]
17. Park, S.Y.; Kim, D.S. Discovery of natural products from *Curcuma longa* that protect cells from beta-amyloid insult: A drug discovery effort against Alzheimer's disease. *J. Nat. Prod.* **2002**, *65*, 1227–1231. [[CrossRef](#)]
18. Kim, D.S.; Kim, J.Y. Total synthesis of Calebin-A, preparation of its analogues, and their neuronal cell protectivity against beta-amyloid insult. *Bioorg. Med. Chem. Lett.* **2001**, *11*, 2541–2543. [[CrossRef](#)]
19. Lanni, C.; Masi, M.; Racchi, M.; Govoni, S. Cancer and Alzheimer's disease inverse relationship: An age-associated diverging derailment of shared pathways. *Mol. Psychiatry* **2021**, *26*, 280–295. [[CrossRef](#)]
20. Zabłocka, A.; Kazana, W.; Sochocka, M.; Stańczykiewicz, B.; Janusz, M.; Leszek, J.; Orzechowska, B. Inverse Correlation between Alzheimer's Disease and Cancer: Short Overview. *Mol. Neurobiol.* **2021**, *58*, 6335–6349. [[CrossRef](#)]
21. Driver, J.A. Inverse association between cancer and neurodegenerative disease: Review of the epidemiologic and biological evidence. *Biogerontology* **2014**, *15*, 547–557. [[CrossRef](#)] [[PubMed](#)]
22. Chang, J.Y.; Chang, N.S. WWOX dysfunction induces sequential aggregation of TRAPPC6A Δ , TIAF1, tau and amyloid β , and causes apoptosis. *Cell Death. Discov.* **2015**, *1*, 15003. [[CrossRef](#)]
23. Sze, C.I.; Su, M.; Pugazhenth, S.; Jambal, P.; Hsu, L.J.; Heath, J.; Schultz, L.; Chang, N.S. Down-regulation of WW domain-containing oxidoreductase induces Tau phosphorylation in vitro. A potential role in Alzheimer's disease. *J. Biol. Chem.* **2004**, *279*, 30498–30506. [[CrossRef](#)] [[PubMed](#)]
24. Pfoertner, S.; Jeron, A.; Probst-Kepper, M.; Guzman, C.A.; Hansen, W.; Westendorf, A.M.; Toepfer, T.; Schrader, A.J.; Franzke, A.; Buer, J.; et al. Signatures of human regulatory T cells: An encounter with old friends and new players. *Genome Biol.* **2006**, *7*, R54. [[CrossRef](#)]
25. Chang, J.Y.; Chiang, M.F.; Lin, S.R.; Lee, M.H.; He, H.; Chou, P.Y.; Chen, S.J.; Chen, Y.A.; Yang, L.Y.; Lai, F.J.; et al. TIAF1 self-aggregation in peritumor capsule formation, spontaneous activation of SMAD-responsive promoter in p53-deficient environment, and cell death. *Cell Death. Dis.* **2012**, *3*, e302. [[CrossRef](#)]
26. Lee, M.H.; Lin, S.R.; Chang, J.Y.; Schultz, L.; Heath, J.; Hsu, L.J.; Kuo, Y.M.; Hong, Q.; Chiang, M.F.; Gong, C.X.; et al. TGF- β induces TIAF1 self-aggregation via type II receptor-independent signaling that leads to generation of amyloid β plaques in Alzheimer's disease. *Cell Death. Dis.* **2010**, *1*, e110. [[CrossRef](#)]
27. Advani, D.; Gupta, R.; Tripathi, R.; Sharma, S.; Ambasta, R.K.; Kumar, P. Protective role of anticancer drugs in neurodegenerative disorders: A drug repurposing approach. *Neurochem. Int.* **2020**, *140*, 104841. [[CrossRef](#)] [[PubMed](#)]
28. Bednarek, A.K.; Laflin, K.J.; Daniel, R.L.; Liao, Q.; Hawkins, K.A.; Aldaz, C.M. WWOX, a novel WW domain-containing protein mapping to human chromosome 16q23.3-24.1, a region frequently affected in breast cancer. *Cancer Res.* **2000**, *60*, 2140–2145.
29. Ried, K.; Finnis, M.; Hobson, L.; Mangelsdorf, M.; Dayan, S.; Nancarrow, J.K.; Woollatt, E.; Kremmidiotis, G.; Gardner, A.; Venter, D.; et al. Common chromosomal fragile site FRA16D sequence: Identification of the FOR gene spanning FRA16D and homozygous deletions and translocation breakpoints in cancer cells. *Hum. Mol. Genet.* **2000**, *9*, 1651–1663. [[CrossRef](#)]
30. Chang, N.S.; Pratt, N.; Heath, J.; Schultz, L.; Sleeve, D.; Carey, G.B.; Zevotek, N. Hyaluronidase induction of a WW domain-containing oxidoreductase that enhances tumor necrosis factor cytotoxicity. *J. Biol. Chem.* **2001**, *276*, 3361–3370. [[CrossRef](#)]

31. Zhao, Y.; Wang, W.; Pan, W.; Yu, Y.; Huang, W.; Gao, J.; Zhang, Y.; Zhang, S. WWOX promotes apoptosis and inhibits autophagy in paclitaxel-treated ovarian carcinoma cells. *Mol. Med. Rep.* **2021**, *23*, 115. [[CrossRef](#)]
32. Taouis, K.; Driouch, K.; Lidereau, R.; Lallemand, F. Molecular Functions of WWOX Potentially Involved in Cancer Development. *Cells* **2021**, *10*, 1051. [[CrossRef](#)]
33. Iliopoulos, D.; Guler, G.; Han, S.Y.; Druck, T.; Ottey, M.; McCorkell, K.A.; Huebner, K. Roles of FHIT and WWOX fragile genes in cancer. *Cancer Lett.* **2006**, *232*, 27–36. [[CrossRef](#)]
34. Smith, D.I.; McAvoy, S.; Zhu, Y.; Perez, D.S. Large common fragile site genes and cancer. *Semin. Cancer Biol.* **2007**, *17*, 31–41. [[CrossRef](#)]
35. Chang, N.S.; Hsu, L.J.; Lin, Y.S.; Lai, F.J.; Sheu, H.M. WW domain-containing oxidoreductase: A candidate tumor suppressor. *Trends Mol. Med.* **2007**, *13*, 12–22. [[CrossRef](#)]
36. Gardenswartz, A.; Aqeilan, R.I. WW domain-containing oxidoreductase's role in myriad cancers: Clinical significance and future implications. *Exp. Biol. Med.* **2014**, *239*, 253–263. [[CrossRef](#)]
37. Aldaz, C.M.; Hussain, T. WWOX Loss of Function in Neurodevelopmental and Neurodegenerative Disorders. *Int. J. Mol. Sci.* **2020**, *21*, 8922. [[CrossRef](#)] [[PubMed](#)]
38. Abu-Remaileh, M.; Joy-Dodson, E.; Schueler-Furman, O.; Aqeilan, R.I. Pleiotropic Functions of Tumor Suppressor WWOX in Normal and Cancer Cells. *J. Biol. Chem.* **2015**, *290*, 30728–30735. [[CrossRef](#)] [[PubMed](#)]
39. Bonin, F.; Taouis, K.; Azorin, P.; Pétitalot, A.; Tariq, Z.; Nola, S.; Bouteille, N.; Tury, S.; Vacher, S.; Bièche, I.; et al. VOPP1 promotes breast tumorigenesis by interacting with the tumor suppressor WWOX. *BMC Biol.* **2018**, *16*, 109. [[CrossRef](#)] [[PubMed](#)]
40. Park, D.; Gharghabi, M.; Reczek, C.R.; Plow, R.; Yungvirt, C.; Aldaz, C.M.; Huebner, K. Wwox Binding to the Murine Brca1-BRCT Domain Regulates Timing of Brip1 and CtIP Phospho-Protein Interactions with This Domain at DNA Double-Strand Breaks, and Repair Pathway Choice. *Int. J. Mol. Sci.* **2022**, *23*, 3729. [[CrossRef](#)] [[PubMed](#)]
41. Li, G.; Sun, L.; Mu, Z.; Huang, Y.; Fu, C.; Hu, B. Ectopic WWOX Expression Inhibits Growth of 5637 Bladder Cancer Cell In Vitro and In Vivo. *Cell Biochem. Biophys.* **2015**, *73*, 417–425. [[CrossRef](#)]
42. Liu, C.W.; Chen, P.H.; Yu, T.J.; Lin, K.J.; Chang, L.C. WWOX Modulates ROS-Dependent Senescence in Bladder Cancer. *Molecules* **2022**, *27*, 7388. [[CrossRef](#)]
43. Cheng, H.C.; Huang, P.H.; Lai, F.J.; Jan, M.S.; Chen, Y.L.; Chen, S.Y.; Chen, W.L.; Hsu, C.K.; Huang, W.; Hsu, L.J. Loss of fragile WWOX gene leads to senescence escape and genome instability. *Cell. Mol. Life Sci.* **2023**, *80*, 338. [[CrossRef](#)]
44. Gao, K.; Yin, J.; Dong, J. Deregulated WWOX is involved in a negative feedback loop with microRNA-214-3p in osteosarcoma. *Int. J. Mol. Med.* **2016**, *38*, 1850–1856. [[CrossRef](#)]
45. Saigo, C.; Kito, Y.; Takeuchi, T. Cancerous Protein Network That Inhibits the Tumor Suppressor Function of WW Domain-Containing Oxidoreductase (WWOX) by Aberrantly Expressed Molecules. *Front. Oncol.* **2018**, *8*, 350. [[CrossRef](#)]
46. Shaukat, Q.; Hertecant, J.; El-Hattab, A.W.; Ali, B.R.; Suleiman, J. West syndrome, developmental and epileptic encephalopathy, and severe CNS disorder associated with WWOX mutations. *Epileptic Disord.* **2018**, *20*, 401–412. [[CrossRef](#)] [[PubMed](#)]
47. Liu, C.C.; Ho, P.C.; Lee, I.T.; Chen, Y.A.; Chu, C.H.; Teng, C.C.; Wu, S.N.; Sze, C.I.; Chiang, M.F.; Chang, N.S. WWOX Phosphorylation, Signaling, and Role in Neurodegeneration. *Front. Neurosci.* **2018**, *12*, 563. [[CrossRef](#)]
48. Chang, N.S.; Doherty, J.; Ensign, A. JNK1 physically interacts with WW domain-containing oxidoreductase (WOX1) and inhibits WOX1-mediated apoptosis. *J. Biol. Chem.* **2003**, *278*, 9195–9202. [[CrossRef](#)] [[PubMed](#)]
49. Chang, N.S.; Doherty, J.; Ensign, A.; Schultz, L.; Hsu, L.J.; Hong, Q. WOX1 is essential for tumor necrosis factor-, UV light-, staurosporine-, and p53-mediated cell death, and its tyrosine 33-phosphorylated form binds and stabilizes serine 46-phosphorylated p53. *J. Biol. Chem.* **2005**, *280*, 43100–43108. [[CrossRef](#)] [[PubMed](#)]
50. Li, M.Y.; Lai, F.J.; Hsu, L.J.; Lo, C.P.; Cheng, C.L.; Lin, S.R.; Lee, M.H.; Chang, J.Y.; Subhan, D.; Tsai, M.S.; et al. Dramatic co-activation of WWOX/WOX1 with CREB and NF-kappaB in delayed loss of small dorsal root ganglion neurons upon sciatic nerve transection in rats. *PLoS ONE* **2009**, *4*, e7820. [[CrossRef](#)] [[PubMed](#)]
51. Chen, S.T.; Chuang, J.I.; Cheng, C.L.; Hsu, L.J.; Chang, N.S. Light-induced retinal damage involves tyrosine 33 phosphorylation, mitochondrial and nuclear translocation of WW domain-containing oxidoreductase in vivo. *Neuroscience* **2005**, *130*, 397–407. [[CrossRef](#)] [[PubMed](#)]
52. Abu-Odeh, M.; Salah, Z.; Herbel, C.; Hofmann, T.G.; Aqeilan, R.I. WWOX, the common fragile site FRA16D gene product, regulates ATM activation and the DNA damage response. *Proc. Natl. Acad. Sci. USA* **2014**, *111*, E4716–E4725. [[CrossRef](#)] [[PubMed](#)]
53. Abu-Odeh, M.; Hereema, N.A.; Aqeilan, R.I. WWOX modulates the ATR-mediated DNA damage checkpoint response. *Oncotarget* **2016**, *7*, 4344–4355. [[CrossRef](#)]
54. Chen, S.J.; Lin, P.W.; Lin, H.P.; Huang, S.S.; Lai, F.J.; Sheu, H.M.; Hsu, L.J.; Chang, N.S. UV irradiation/cold shock-mediated apoptosis is switched to bubbling cell death at low temperatures. *Oncotarget* **2015**, *6*, 8007–8018. [[CrossRef](#)] [[PubMed](#)]
55. Chang, N.S. Bubbling cell death: A hot air balloon released from the nucleus in the cold. *Exp. Biol. Med.* **2016**, *241*, 1306–1315. [[CrossRef](#)] [[PubMed](#)]
56. Hsu, L.J.; Hong, Q.; Chen, S.T.; Kuo, H.L.; Schultz, L.; Heath, J.; Lin, S.R.; Lee, M.H.; Li, D.Z.; Li, Z.L.; et al. Hyaluronan activates Hyal-2/WWOX/Smad4 signaling and causes bubbling cell death when the signaling complex is overexpressed. *Oncotarget* **2017**, *8*, 19137–19155. [[CrossRef](#)]

57. Chen, Y.A.; Sie, Y.D.; Liu, T.Y.; Kuo, H.L.; Chou, P.Y.; Chen, Y.J.; Lee, K.T.; Chen, P.J.; Chen, S.T.; Chang, N.S. Normal cells repel WWOX-negative or -dysfunctional cancer cells via WWOX cell surface epitope 286–299. *Commun. Biol.* **2021**, *4*, 753. [[CrossRef](#)]
58. Chang, N.S.; Doherty, J.; Ensign, A.; Lewis, J.; Heath, J.; Schultz, L.; Chen, S.T.; Oppermann, U. Molecular mechanisms underlying WOX1 activation during apoptotic and stress responses. *Biochem. Pharmacol.* **2003**, *66*, 1347–1354. [[CrossRef](#)]
59. Chang, N.S.; Schultz, L.; Hsu, L.J.; Lewis, J.; Su, M.; Sze, C.I. 17beta-Estradiol upregulates and activates WOX1/WWOXv1 and WOX2/WWOXv2 in vitro: Potential role in cancerous progression of breast and prostate to a premetastatic state in vivo. *Oncogene* **2005**, *24*, 714–723. [[CrossRef](#)]
60. Lin, H.P.; Chang, J.Y.; Lin, S.R.; Lee, M.H.; Huang, S.S.; Hsu, L.J.; Chang, N.S. Identification of an In Vivo MEK/WOX1 Complex as a Master Switch for Apoptosis in T Cell Leukemia. *Genes Cancer* **2011**, *2*, 550–562. [[CrossRef](#)] [[PubMed](#)]
61. Huang, S.S.; Su, W.P.; Lin, H.P.; Kuo, H.L.; Wei, H.L.; Chang, N.S. Role of WW Domain-containing Oxidoreductase WWOX in Driving T Cell Acute Lymphoblastic Leukemia Maturation. *J. Biol. Chem.* **2016**, *291*, 17319–17331. [[CrossRef](#)]
62. Gaudio, E.; Palamarchuk, A.; Palumbo, T.; Trapasso, F.; Pekarsky, Y.; Croce, C.M.; Aqeilan, R.I. Physical association with WWOX suppresses c-Jun transcriptional activity. *Cancer Res.* **2006**, *66*, 11585–11589. [[CrossRef](#)]
63. Aderca, I.; Moser, C.D.; Veerasamy, M.; Bani-Hani, A.H.; Bonilla-Guerrero, R.; Ahmed, K.; Shire, A.; Cazanave, S.C.; Montoya, D.P.; Mettler, T.A.; et al. The JNK inhibitor SP600129 enhances apoptosis of HCC cells induced by the tumor suppressor WWOX. *J. Hepatol.* **2008**, *49*, 373–383. [[CrossRef](#)] [[PubMed](#)]
64. Zhang, S.J.; Buchthal, B.; Lau, D.; Hayer, S.; Dick, O.; Schwaninger, M.; Veltkamp, R.; Zou, M.; Weiss, U.; Bading, H. A signaling cascade of nuclear calcium-CREB-ATF3 activated by synaptic NMDA receptors defines a gene repression module that protects against extrasynaptic NMDA receptor-induced neuronal cell death and ischemic brain damage. *J. Neurosci.* **2011**, *31*, 4978–4990. [[CrossRef](#)] [[PubMed](#)]
65. Li, Y.; Fan, Q.; Li, F.; Pang, R.; Chen, C.; Li, P.; Wang, X.; Xuan, W.; Yu, W. The multifaceted roles of activating transcription factor 3 (ATF3) in inflammatory responses—Potential target to regulate neuroinflammation in acute brain injury. *J. Cereb. Blood Flow Metab.* **2023**, *43*, 8–17. [[CrossRef](#)] [[PubMed](#)]
66. Kang, L.; Piao, M.; Liu, N.; Gu, W.; Feng, C. Sevoflurane Exposure Induces Neuronal Cell Ferroptosis Initiated by Increase of Intracellular Hydrogen Peroxide in the Developing Brain via ER Stress ATF3 Activation. *Mol. Neurobiol.* **2023**. [[CrossRef](#)] [[PubMed](#)]
67. Inaba, Y.; Hashiuchi, E.; Watanabe, H.; Kimura, K.; Oshima, Y.; Tsuchiya, K.; Murai, S.; Takahashi, C.; Matsumoto, M.; Kitajima, S.; et al. The transcription factor ATF3 switches cell death from apoptosis to necroptosis in hepatic steatosis in male mice. *Nat. Commun.* **2023**, *14*, 167. [[CrossRef](#)] [[PubMed](#)]
68. Su, W.P.; Wang, W.J.; Chang, J.Y.; Ho, P.C.; Liu, T.Y.; Wen, K.Y.; Kuo, H.L.; Chen, Y.J.; Huang, S.S.; Subhan, D.; et al. Therapeutic Zfra4-10 or WWOX7-21 Peptide Induces Complex Formation of WWOX with Selective Protein Targets in Organs that Leads to Cancer Suppression and Spleen Cytotoxic Memory Z Cell Activation In Vivo. *Cancers* **2020**, *12*, 2189. [[CrossRef](#)] [[PubMed](#)]
69. Hsu, L.J.; Schultz, L.; Hong, Q.; Van Moer, K.; Heath, J.; Li, M.Y.; Lai, F.J.; Lin, S.R.; Lee, M.H.; Lo, C.P.; et al. Transforming growth factor beta1 signaling via interaction with cell surface Hyal-2 and recruitment of WWOX/WOX1. *J. Biol. Chem.* **2009**, *284*, 16049–16059. [[CrossRef](#)] [[PubMed](#)]
70. Lee, M.H.; Su, W.P.; Wang, W.J.; Lin, S.R.; Lu, C.Y.; Chen, Y.A.; Chang, J.Y.; Huang, S.S.; Chou, P.Y.; Ye, S.R.; et al. Zfra activates memory Hyal-2+ CD3- CD19- spleen cells to block cancer growth, stemness, and metastasis in vivo. *Oncotarget* **2015**, *6*, 3737–3751. [[CrossRef](#)]
71. Lee, M.H.; Shih, Y.H.; Lin, S.R.; Chang, J.Y.; Lin, Y.H.; Sze, C.I.; Kuo, Y.M.; Chang, N.S. Zfra restores memory deficits in Alzheimer's disease triple-transgenic mice by blocking aggregation of TRAPPC6AΔ, SH3GLB2, tau, and amyloid β, and inflammatory NF-κB activation. *Alzheimers. Dement.* **2017**, *3*, 189–204. [[CrossRef](#)]
72. Lin, Y.H.; Shih, Y.H.; Yap, Y.V.; Chen, Y.W.; Kuo, H.L.; Liu, T.Y.; Hsu, L.J.; Kuo, Y.M.; Chang, N.S. Zfra inhibits the TRAPPC6AΔ-initiated pathway of neurodegeneration. *Int. J. Mol. Sci.* **2022**, *23*, 14510. [[CrossRef](#)]
73. Hong, Q.; Hsu, L.J.; Schultz, L.; Pratt, N.; Mattison, J.; Chang, N.S. Zfra affects TNF-mediated cell death by interacting with death domain protein TRADD and negatively regulates the activation of NF-kappaB, JNK1, p53 and WOX1 during stress response. *BMC Mol. Biol.* **2007**, *8*, 50. [[CrossRef](#)]
74. Wang, W.J.; Ho, P.C.; Nagarajan, G.; Chen, Y.A.; Kuo, H.L.; Subhan, D.; Su, W.P.; Chang, J.Y.; Lu, C.Y.; Chang, K.T.; et al. WWOX possesses N-terminal cell surface-exposed epitopes WWOX(7-21) and WWOX(7-11) for signaling cancer growth suppression and prevention in vivo. *Cancers* **2019**, *11*, 1818. [[CrossRef](#)]
75. Lv, M.; Zhang, Z.; Cui, Y. Unconventional T cells in brain homeostasis, injury and neurodegeneration. *Front. Immunol.* **2023**, *14*, 1273459. [[CrossRef](#)] [[PubMed](#)]
76. Zhang, H.; Zhang, X.; Li, H.; Wang, B.; Chen, P.; Meng, J. The roles of macrophage migration inhibitory factor in retinal diseases. *Neural Regen. Res.* **2024**, *19*, 309–315. [[CrossRef](#)] [[PubMed](#)]
77. Melo, A.M.; Taher, N.A.; Doherty, D.G.; Molloy, E.J. The role of lymphocytes in neonatal encephalopathy. *Brain Behav. Immun. Health* **2021**, *18*, 100380. [[CrossRef](#)] [[PubMed](#)]
78. Davies, A.J.; Rinaldi, S.; Costigan, M.; Oh, S.B. Cytotoxic Immunity in Peripheral Nerve Injury and Pain. *Front. Neurosci.* **2020**, *14*, 142. [[CrossRef](#)] [[PubMed](#)]
79. Mahajan, N.P.; Whang, Y.E.; Mohler, J.L.; Earp, H.S. Activated tyrosine kinase Ack1 promotes prostate tumorigenesis: Role of Ack1 in polyubiquitination of tumor suppressor Wwox. *Cancer Res.* **2005**, *65*, 10514–10523. [[CrossRef](#)] [[PubMed](#)]

80. Choi, H.J.; Park, J.H.; Park, J.H.; Lee, K.B.; Oh, S.M. Pc2-mediated SUMOylation of WWOX is essential for its suppression of DU145 prostate tumorigenesis. *FEBS Lett.* **2015**, *589*, 3977–3988. [[CrossRef](#)]
81. Sze, C.I.; Kuo, Y.M.; Hsu, L.J.; Fu, T.F.; Chiang, M.F.; Chang, J.Y.; Chang, N.S. A cascade of protein aggregation bombards mitochondria for neurodegeneration and apoptosis under WWOX deficiency. *Cell Death Dis.* **2015**, *6*, e1881. [[CrossRef](#)]
82. Chang, J.Y.; Lee, M.H.; Lin, S.R.; Yang, L.Y.; Sun, H.S.; Sze, C.I.; Hong, Q.; Lin, Y.S.; Chou, Y.T.; Hsu, L.J.; et al. Trafficking protein particle complex 6A delta (TRAPPC6A Δ) is an extracellular plaque-forming protein in the brain. *Oncotarget* **2015**, *6*, 3578–3589. [[CrossRef](#)]
83. Piard, J.; Hawkes, L.; Milh, M.; Villard, L.; Borgatti, R.; Romaniello, R.; Fradin, M.; Capri, Y.; Héron, D.; Nougues, M.C.; et al. The phenotypic spectrum of WWOX-related disorders: 20 additional cases of WOREE syndrome and review of the literature. *Genet. Med.* **2019**, *21*, 1308–1318. [[CrossRef](#)]
84. Cheng, Y.Y.; Chou, Y.T.; Lai, F.J.; Jan, M.S.; Chang, T.H.; Jou, I.M.; Chen, P.S.; Lo, J.Y.; Huang, S.S.; Chang, N.S.; et al. Wwox deficiency leads to neurodevelopmental and degenerative neuropathies and glycogen synthase kinase 3 β -mediated epileptic seizure activity in mice. *Acta Neuropathol. Commun.* **2020**, *8*, 6. [[CrossRef](#)]
85. Kunkle, B.W.; Grenier-Boley, B.; Sims, R.; Bis, J.C.; Damotte, V.; Naj, A.C.; Boland, A.; Vronskaya, M.; van der Lee, S.J.; Amlie-Wolf, A.; et al. Genetic meta-analysis of diagnosed Alzheimer’s disease identifies new risk loci and implicates A β , tau, immunity and lipid processing. *Nat. Genet.* **2019**, *51*, 414–430. [[CrossRef](#)]
86. Liu, G.; Peng, J.; Liao, Z.; Locascio, J.J.; Corvol, J.C.; Zhu, F.; Dong, X.; Maple-Grødem, J.; Campbell, M.C.; Elbaz, A.; et al. Genome-wide survival study identifies a novel synaptic locus and polygenic score for cognitive progression in Parkinson’s disease. *Nat. Genet.* **2021**, *53*, 787–793. [[CrossRef](#)] [[PubMed](#)]
87. Wang, H.Y.; Juo, L.I.; Lin, Y.T.; Hsiao, M.; Lin, J.T.; Tsai, C.H.; Tzeng, Y.H.; Chuang, Y.C.; Chang, N.S.; Yang, C.N.; et al. WW domain-containing oxidoreductase promotes neuronal differentiation via negative regulation of glycogen synthase kinase 3 β . *Cell Death Differ.* **2012**, *19*, 1049–1059. [[CrossRef](#)] [[PubMed](#)]
88. Chou, P.Y.; Lin, S.R.; Lee, M.H.; Schultz, L.; Sze, C.I.; Chang, N.S. A p53/TIAF1/WWOX triad exerts cancer suppression but may cause brain protein aggregation due to p53/WWOX functional antagonism. *Cell Commun. Signal.* **2019**, *17*, 76. [[CrossRef](#)] [[PubMed](#)]
89. Jin, C.; Ge, L.; Ding, X.; Chen, Y.; Zhu, H.; Ward, T.; Wu, F.; Cao, X.; Wang, Q.; Yao, X. PKA-mediated protein phosphorylation regulates ezrin-WWOX interaction. *Biochem. Biophys. Res. Commun.* **2006**, *341*, 784–791. [[CrossRef](#)] [[PubMed](#)]
90. Chou, P.Y.; Lai, F.J.; Chen, Y.A.; Sie, Y.D.; Kuo, H.L.; Su, W.P.; Wu, C.Y.; Liu, T.Y.; Wen, K.Y.; Hsu, L.J.; et al. Strategies by which WWOX-deficient metastatic cancer cells utilize to survive via dodging, compromising, and causing damage to WWOX-positive normal microenvironment. *Cell Death Discov.* **2019**, *5*, 97. [[CrossRef](#)] [[PubMed](#)]
91. Oliver, K.L.; Trivisano, M.; Mandelstam, S.A.; De Dominicis, A.; Francis, D.I.; Green, T.E.; Muir, A.M.; Chowdhary, A.; Hertzberg, C.; Goldhahn, K.; et al. WWOX developmental and epileptic encephalopathy: Understanding the epileptology and the mortality risk. *Epilepsia* **2023**, *64*, 1351–1367. [[CrossRef](#)]
92. Al Baradie, R.; Mir, A.; Alsaif, A.; Ali, M.; Al Ghamdi, F.; Bashir, S.; Howsawi, Y. Epilepsy in patients with WWOX-related epileptic encephalopathy (WOREE) syndrome. *Epileptic Disord.* **2022**, *24*, 697–712. [[CrossRef](#)] [[PubMed](#)]
93. Repudi, S.; Steinberg, D.J.; Elazar, N.; Breton, V.L.; Aquilino, M.S.; Saleem, A.; Abu-Swai, S.; Vainshtein, A.; Eshed-Eisenbach, Y.; Vijayaragavan, B.; et al. Neuronal deletion of Wwox, associated with WOREE syndrome, causes epilepsy and myelin defects. *Brain* **2021**, *144*, 3061–3077. [[CrossRef](#)] [[PubMed](#)]
94. Roberts, B. Neuronal migration disorders. *Radiol. Technol.* **2018**, *89*, 279–295.
95. Francis, F.; Cappello, S. Neuronal migration and disorders—An update. *Curr. Opin. Neurobiol.* **2021**, *66*, 57–68. [[CrossRef](#)]
96. Hussain, T.; Kil, H.; Hattiangady, B.; Lee, J.; Kodali, M.; Shuai, B.; Attaluri, S.; Takata, Y.; Shen, J.; Abba, M.C.; et al. Wwox deletion leads to reduced GABA-ergic inhibitory interneuron numbers and activation of microglia and astrocytes in mouse hippocampus. *Neurobiol. Dis.* **2019**, *121*, 163–176. [[CrossRef](#)]
97. McAvoy, S.; Zhu, Y.; Perez, D.S.; James, C.D.; Smith, D.I. Disabled-1 is a large common fragile site gene, inactivated in multiple cancers. *Genes Chromosomes Cancer* **2008**, *47*, 165–174. [[CrossRef](#)] [[PubMed](#)]
98. Yang, T.; Hergenreder, T.; Ye, B. Analysis of mouse brain sections by live-cell time-lapse confocal microscopy. *Bio-Protocol* **2023**, *13*, e4648. [[CrossRef](#)]
99. Honda, T.; Hirota, Y.; Nakajima, K. Heterozygous *Dab1* null mutation disrupts neocortical and hippocampal development. *eNeuro* **2023**, *10*, ENEURO.0433-22.2023. [[CrossRef](#)]
100. Kounoupa, Z.; Tivodar, S.; Theodorakis, K.; Kyriakis, D.; Denaxa, M.; Karagogeos, D. Rac1 and Rac3 GTPases and TPC2 are required for axonal outgrowth and migration of cortical interneurons. *J. Cell Sci.* **2023**, *136*, jcs260373. [[CrossRef](#)]
101. Carvalho, C.; Correia, S.C.; Seica, R.; Moreira, P.I. WWOX inhibition by Zfra1-31 restores mitochondrial homeostasis and viability of neuronal cells exposed to high glucose. *Cell Mol Life Sci.* **2022**, *79*, 487. [[CrossRef](#)]
102. Abu-Remaileh, M.; Aqeilan, R.I. Tumor suppressor WWOX regulates glucose metabolism via HIF1 α modulation. *Cell Death Differ.* **2014**, *21*, 1805–1814. [[CrossRef](#)]
103. Abu-Remaileh, M.; Aqeilan, R.I. The tumor suppressor WW domain-containing oxidoreductase modulates cell metabolism. *Exp. Biol. Med.* **2015**, *240*, 345–350. [[CrossRef](#)] [[PubMed](#)]
104. Abu-Remaileh, M.; Seewaldt, V.L.; Aqeilan, R.I. WWOX loss activates aerobic glycolysis. *Mol. Cell Oncol.* **2014**, *2*, e965640. [[CrossRef](#)] [[PubMed](#)]

105. Janczar, S.; Nautiyal, J.; Xiao, Y.; Curry, E.; Sun, M.; Zanini, E.; Paige, A.J.; Gabra, H. WWOX sensitises ovarian cancer cells to paclitaxel via modulation of the ER stress response. *Cell Death Dis.* **2017**, *8*, e2955. [[CrossRef](#)] [[PubMed](#)]
106. Abu-Remaileh, M.; Abu-Remaileh, M.; Akkawi, R.; Knani, I.; Udi, S.; Pacold, M.E.; Tam, J.; Aqeilan, R.I. WWOX somatic ablation in skeletal muscles alters glucose metabolism. *Mol. Metab.* **2019**, *22*, 132–140. [[CrossRef](#)]
107. Baryła, I.; Styczeń-Binkowska, E.; Płuciennik, E.; Kośła, K.; Bednarek, A.K. The WWOX/HIF1A Axis Downregulation Alters Glucose Metabolism and Predispose to Metabolic Disorders. *Int. J. Mol. Sci.* **2022**, *23*, 3326. [[CrossRef](#)] [[PubMed](#)]
108. Wittekind, D.A.; Scholz, M.; Kratzsch, J.; Löffler, M.; Horn, K.; Kirsten, H.; Witte, V.; Villringer, A.; Kluge, M. Genome-wide association and transcriptome analysis suggests total serum ghrelin to be linked with GFRAL. *Eur. J. Endocrinol.* **2021**, *184*, 847–856. [[CrossRef](#)]
109. Baryła, I.; Płuciennik, E.; Kośła, K.; Wojcik, M.; Zieleniak, A.; Zurawska-Klis, M.; Cypriak, K.; Wozniak, L.A.; Bednarek, A.K. Identification of a novel association for the WWOX/HIF1A axis with gestational diabetes mellitus (GDM). *PeerJ.* **2021**, *9*, e10604. [[CrossRef](#)]
110. Permana, S.; Lukman, H.; Norahmawati, E.; Eka Puspita, O.; Faisal Moh Al Zein, D.; Kawamoto, Y.; Tri Endharti, A. East Asian Genome-wide association study derived loci in relation to type 2 diabetes in the Han Chinese population. *Acta Biochim. Pol.* **2019**, *66*, 679–686.
111. Chang, Y.C.; Chiu, Y.F.; Liu, P.H.; Shih, K.C.; Lin, M.W.; Sheu, W.H.; Quertermous, T.; Curb, J.D.; Hsiung, C.A.; Lee, W.J.; et al. Replication of genome-wide association signals of type 2 diabetes in Han Chinese in a prospective cohort. *Clin. Endocrinol.* **2012**, *76*, 365–372. [[CrossRef](#)] [[PubMed](#)]
112. Saadane, A.; Du, Y.; Thoreson, W.B.; Miyagi, M.; Lessieur, E.M.; Kiser, J.; Wen, X.; Berkowitz, B.A.; Kern, T.S. Photoreceptor Cell Calcium Dysregulation and Calpain Activation Promote Pathogenic Photoreceptor Oxidative Stress and Inflammation in Prodromal Diabetic Retinopathy. *Am. J. Pathol.* **2021**, *191*, 1805–1821. [[CrossRef](#)] [[PubMed](#)]

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