

MDPI

Article

Genome-Wide Screening for SNPs Associated with Stature in Diverse Cattle Breeds [†]

Alexandra S. Abdelmanova ^{1,*}, Alexander A. Sermyagin ¹, Arsen V. Dotsev ¹, Nikolay V. Bardukov ¹, Margaret S. Fornara ¹, Gottfried Brem ^{1,2} and Natalia A. Zinovieva ¹

- L.K. Ernst Federal Research Center for Animal Husbandry, Dubrovitsy, Podolsk Municipal District, 142132 Podolsk, Moscow Region, Russia
- Institute of Animal Breeding and Genetics, University of Veterinary Medicine (VMU), Veterinärplatz, A-1210 Vienna, Austria
- * Correspondence: preevetic@mail.ru
- † Presented the 2nd International Electronic Conference on Diversity—Animals, Plants and Microbes, Online, 15–31 March 2022.

Abstract: Cattle breeds used in industrial production tend to be larger than local cattle, as increased dairy and beef productivity is closely related to stature. This study aimed to identify single nucleotide polymorphisms (SNPs) significantly associated with stature in diverse cattle breeds. Thirteen local and transboundary cattle breeds (n = 670) were divided into two groups according to their stature. The high-stature group included the Angus, Ayrshire, Black-and-White, Holstein, Kholmogor, Yaroslavl, Tagil, and Istoben breeds. The low-stature group included Jersey, Kalmyk, Kyrgyz, Mongolian, and Yakut. The average height at withers was 136.3 ± 1.6 and 121.6 ± 2.8 cm in the high- and low-stature groups, respectively. The samples of the 11 breeds were genotyped using high-density DNA arrays. Genotypes of the two remaining breeds were downloaded from the publicly available WIDDE database. Genome-wide association studies revealed seven SNPs strongly associated with stature, including three at 77.3-77.8 cM on BTA4 and four at 24.5-25.2 cM on BTA14. Functional annotation showed the localization within identified regions of genes responsible for growth, exterior characteristics, protein and lipid metabolism, and feed intake. The identified SNPs can be considered useful DNA markers for marker-assisted cattle breeding aimed at increasing stature.

Keywords: local cattle; GWAS; SNP; QTL; stature; allele frequency



Citation: Abdelmanova, A.S.; Sermyagin, A.A.; Dotsev, A.V.; Bardukov, N.V.; Fornara, M.S.; Brem, G.; Zinovieva, N.A. Genome-Wide Screening for SNPs Associated with Stature in Diverse Cattle Breeds. *Diversity* 2022, 14, 692. https://doi.org/10.3390/d14080692

Academic Editor: Simone Ceccobelli

Received: 19 July 2022 Accepted: 20 August 2022 Published: 22 August 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/licenses/by/4.0/).

1. Introduction

Body size, an archetypical quantitative or complex trait that exhibits continuous variation [1], has been extensively studied because it is ecologically important, relevant in farming, and an important indicator of growth and health [2]. Stature and body weight in beef cattle breeding are important for beef production. In dairy farming, smaller or larger animals may be desirable, depending on management considerations [3,4].

Many studies have revealed a large number of quantitative trait loci (QTLs) associated with body size in cattle [5,6]. QTLs that affect the growth components of cattle have been mapped, confirming their polygenic determinism [7,8]. For example, 38 and 52 QTLs have been reported to account for more than 60% of the variation in adult height and weight, respectively, in the American Aberdeen Angus cow population [9].

A QTL on chromosome 14 with a major effect on stature and body weight was reported by Karim et al. [10] in Holstein Friesian and Jersey animals, as well as by others in many other independent populations [11–14]. Randhawa et al. [15] compared SNP data across the genome in a number of European and African breeds of *Bos taurus* with high (145–155 cM) and low (105–133 cM) withers and found the most significant selection signal associated with growth to be mapped on chromosome 14 (BTA14) in the 24.79–28.25 Mb region [16]. A detailed analysis of the fine mapping of this chromosome 14 locus [10]

Diversity 2022, 14, 692 2 of 9

yielded 13 candidate polymorphisms potentially underlying the QTL. Further functional and genetic analyses reduced the number of candidate variants to two polymorphisms in the bidirectional promoters of the *PLAG1* and *CHCHD7* genes. In particular, *PLAG1* is an obvious candidate, given *PLAG1* knockout mice suffer from slow growth rates and dwarfism [17,18]. This gene encodes a transcription factor that regulates several growth factors including *IGF2* [19,20], a key modulator of growth in both dogs and humans [21,22]. However, most studies were conducted on commercial cattle breeds subjected to strong artificial selection pressures [15,23,24].

Although comprehensive genome-wide studies of local Russian cattle breeds have been carried out [25–27], the genetic basis of stature in these breeds has not been investigated. Despite local breeds having lower productivity compared to that of commercial breeds, their adaptability to climate conditions makes them useful as reserves of genetic diversity in breeding programs.

This study aimed to identify SNPs significantly associated with stature in commercial and local cattle of different productivity types bred in Russia and nearby countries.

2. Materials and Methods

Totally 670 samples of thirteen local and transboundary cattle breeds, including Aberdeen-Angus (ANG, n = 39), Ayrshire (AYR, n = 144), Black-and-White (BLW, n = 50), Holstein (HLS, n = 184), Istoben (IST, n = 22), Jersey (JRS, n = 32), Kalmyk (KLM, n = 27), Kholmogor (KHL, n = 26), Kyrgyz (KRG, n = 24), Mongolian (MNG, n = 26), Tagil (TAG, n = 26), Yakut (YKT, n = 29) and Yaroslavl (YRS, n = 41), were subjected to our study.

The samples of 11 breeds stored in the bioresource collection "Bank of genetic materials of domestic and wild animals and birds" of the L.K. Ernst Federal Research Center for Animal Husbandry were genotyped using high-density DNA arrays (Illumina Inc., San Diego, CA, USA). The genotypes of the two remaining breeds, Aberdeen–Angus and Jersey, were downloaded from the WIDDE database [http://widde.toulouse.inra.fr/wide].

Since genotyping of the samples was carried out on SNP chips of different densities, Bovine GGP HD Bead Chip and BovineHD BeadChip (Illumina, Inc., San Diego, CA, USA), which included of ~150 K and 777 K SNPs, respectively, during data processing. SNPs common to the two chips were determined and used for further analysis. Quality control of genotyping was carried out in PLINK 1.9 [28] using the following filters: at least 90% of loci (—geno 0.1) were successfully genotyped in at least 80% of animals (—mind 0.2) and the frequency of minor alleles was at least 5% (–maf 0.05). After combining the data sets and quality control of genotyping, 115,237 SNPs were used for the subsequent genome-wide association studies (GWAS).

We compared the genome-wide genotypes of 532 animals representing eight high-stature breeds and 138 animals of five low-stature breeds. To identify SNPs that were significantly associated with withers height and body weight we carried out across-breed GWAS using PLINK 1.9. A linear regression using a linear-mixed model was applied, and defined as follows:

$$y = Xb + Wg + e$$
,

where y—a vector of phenotype traits; b—a vector of fixed effects including breed, and linear discriminant functions; g—a vector for the SNP effects; e—a vector of random residual effects with e~N(0,Io2e); and X, W—incidence matrices for b and g, respectively. To confirm the significant impact of SNPs and identify significant regions in the genome of cattle, several tests were used to check for null hypotheses by Bonferroni (threshold $p < 4.34 \times 10^{-7}$, 0.05/115,237).

The positions of the SNPs were assigned according to the *Bos taurus* genome assembly UMD 3.1.1 (https://www.ncbi.nlm.nih.gov/assembly/GCF_000003055.6, accessed on 1 October 2021).

The Domestic Animal Diversity Information System (DAD-IS) (http://www.fao.org/dad-is/en, accessed on 1 October 2021) maintained and developed by the Food and Agriculture Organization (FAO) was used to obtain the information concerning withers height and

Diversity 2022, 14, 692 3 of 9

weight for the studied breeds. Since the values of the indicators varied in different countries, the average values were calculated for each breed without considering the country of breeding and the sex of the animal.

3. Results

The cattle breeds were divided into two groups according to their stature [29]. The breeds that have their average stature above the overall average stature were categorized into the high-stature group. Other breeds were assigned to the low-stature group. The high-stature group included the Angus, Ayrshire, Black-and-White, Holstein, Kholmogor, Yaroslavl, Tagil, and Istoben breeds. The low-stature group comprised of Jersey, Kalmyk, Kyrgyz, Mongolian, and Yakut cattle. The average values for height at withers and weight in the high-stature and low-stature groups were 136.3 ± 1.6 and 121.6 ± 2.8 cm and 697.8 ± 22.8 and 457.3 ± 41.5 kg, respectively (Figure 1).



Figure 1. Distribution of height at withers and body weight in the studied breeds. Height at withers shown by colored bars in centimeters (left axis) and body weight shown by an orange line in kilograms (right axis). Breeds of different productivity types shown by blue (dairy type) and red (beef type) bars. Green dashed line indicates the overall average value of height at withers. The breeds' abbreviations are explained in the Materials and Methods section.

Genome-wide association studies (Figure 2) revealed seven SNPs strongly associated with stature: three at 24.5–25.0 cm on BTA14 were associated with body weight only (BovineHD1400007132, $p < 4.264 \times 10^{-81}$; BovineHD1400007161, $p < 1.529 \times 10^{-88}$; BovineHD1400007247, $p < 6.145 \times 10^{-82}$); one at BTA4:77613816 with height at withers only (BovineHD0400021479; $p < 1.173 \times 10^{-91}$); and three at 77.3–77.8 cm on BTA4 (Hapmap53144-ss46525999, $p < 7.747 \times 10^{-101}$ for height and $p < 4.865 \times 10^{-83}$ for weight; ARS-BFGL-NGS-116590, $p < 7.665 \times 10^{-105}$ for height and $p < 6.098 \times 10^{-87}$ for weight) and at 25.2 cM on BTA14 (BovineHD1400007259, $p < 1.324 \times 10^{-109}$ for height and $p < 4.3 \times 10^{-126}$) were associated with both.

The allele frequencies of the significantly associated SNPs separately for each breed (Supplementary Table S1) and average values in the two groups were calculated (Table 1).

Diversity **2022**, 14, 692 4 of 9

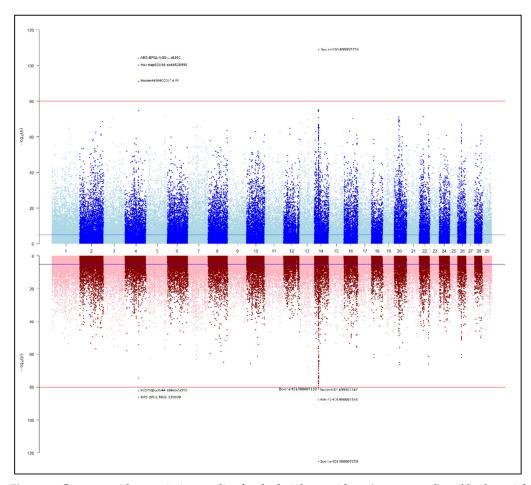


Figure 2. Genome-wide association studies for the height at withers (upper panel) and body weight (lower panel) for the studied breeds. The lines represent the significant thresholds by Bonferroni correction (blue line, $-\log_{10}(p) > 7$) and the most significant SNPs (red line, $-\log_{10}(p) > 80$).

Table 1. Frequencies of alternative alleles at the loci associated with the height at withers and body weight in different groups of cattle breeds.

Localization	Trait	SNP	Allele	Allele Frequencies		
		SINT		High-Stature Breeds	Low-Stature Breeds	
	Height	BovineHD0400021479	T	0.28	0.84	
			C a	0.72	0.16	
Cl. 4	Height and Weight	A DO DECL NICO 11/500	T	0.29	0.92	
Chr 4		ARS-BFGL-NGS-116590	G a,b	0.71	0.08	
		II	A a,b	0.71	0.09	
		Hapmap53144-ss46525999	G	0.29	0.91	
		BovineHD1400007259	G a,b	0.91	0.04	
			T	0.09	0.96	
	Weight		A b	0.95	0.45	
Cl. 14		BovineHD1400007247	G	0.05	0.55	
Chr 14		BovineHD1400007132	A ^b	0.93	0.36	
			G	0.07	0.64	
			T ^b	0.84	0.14	
		BovineHD1400007161	G	0.16	0.86	

 $^{^{\}mathrm{a}}$ —alleles associated with greater height at withers. $^{\mathrm{b}}$ —alleles associated with greater body weight.

Diversity 2022, 14, 692 5 of 9

The differences between the average frequencies of alternative alleles in the highstature group were observed for SNPs located on BTA14: 24.5–25.2. The frequencies varied from 0.05/0.95 for G/A alleles in the BovineHD1400007247 to 0.16/0.84 for G/T alleles in the BovineHD1400007161. In the low-stature group, the alleles' frequencies essentially differ for SNPs located in BTA4: 77.3–77.8. The frequencies varied from 0.08/0.92 for G/T alleles in the ARS-BFGL-NGS-116590 to 0.16/0.84 for G/T alleles in the BovineHD0400021479. The most differences between the frequencies of alternative alleles were observed for SNP BovineHD1400007259 located on BTA14 for both groups of the breeds.

A significant correlation was found between the frequencies of alternative alleles and the studied traits (height at withers and body weight) for all detected SNPs (Table 2).

Localization	SNP	Alleles	High-Stature Breeds		Low-Stature Breeds	
Localization	SINF		Height	Weight	Height	Weight
Chr 4	BovineHD0400021479	T/C a	0.708 **		-0.498 ns	
	ARS-BFGL-NGS-116590	T/G a	0.722 **	0.216 ns	-0.172 ns	-0.233 ns
	Hapmap53144-ss46525999	A a,b/G	0.719 **	0.216 ns	-0.281 ns	-0.413 ns
Chr 14	BovineHD1400007259	G ^{a,b} /T	0.678 **	0.527 ^{ns}	-0.063 ns	-0.079 ns
	BovineHD1400007247	A ^b /G		0.650 *		-0.062 ns
	BovineHD1400007132	A ^b /G		0.584 *		-0.170 ns
	BovineHD1400007161	T ^b /G		0.750 **		-0.338 ns

Table 2. Correlation coefficients of alleles' frequencies with the height at withers and body weight.

Strong and moderate correlations were observed only in the high-stature breed group. In the low-stature breed group, there was no correlation between the stature and allele frequencies (Supplementary Table S2).

A search for genes located within ± 200 kb of the detected SNPs identified genes *PURB*, *MIR4657*, *H2AFV*, *PPIA*, *ZMIZ2*, *OGDH*, *TMED4*, *DDX56*, *NPC1L1*, *NUDCD3*, *CAMK2B*, and *YKT6*, associated with metabolism [30], composition of milk [31,32], synthesis of milk fat and protein [33], and average daily body weight gain [34] in the region BTA4: 77.3–77.8 Mb.

4. Discussion

Our results agree with the previous studies, indicating that the 24.8–25.2 Mb region on BTA 14 containing the *PLAG1* gene is strongly related to body size. Previous studies were conducted on well-selected European or American breeds of dairy (Holstein and Jersey [10], Montbeliarde [35], Swedish Holstein–Friesian and Swedish Red Cattle [36]) and beef types (Gelbvieh beef cattle [37], Simmental, Brahman [38]) or both types of breeds (multibreed cohorts [5]).

Studies on some Asian local breeds (Hanwoo and Japanese Black cattle) found significant associations of *PLAG1* gene with the carcass weight [39,40], but not in the Wagyu population [40]. At the same time, two downstream regions (BTA14: 46.5–46.9 Mb and 60.2–60.5 Mb) containing *FABP5* and *OXR1* genes, respectively, were identified and showed significant association with chuck roll weight and bicep weight for Wagyu, but not for Chinese Simmentals [41]. In contrast, Rowan et al. [42] showed associations of carcass merit with the region containing the *OXR1* gene for Red Angus and Simmental cattle.

In addition, recent studies have revealed that the region on BTA14: 26–27 Mb was associated with the traits of primal cut yields [43], indicating that a wide region on chromosome 14 was associated with the growth and development of the organism. This was common for different groups of breeds irrespective of origin, productivity type, and the intensity of selection in the population.

^a—alleles associated with greater height at withers. ^b—alleles associated with greater body weight. *p < 0.05; **p < 0.01; ns—not significant.

Diversity 2022, 14, 692 6 of 9

However, the regions described in previous studies and located on other chromosomes [14,44,45] were not identified in our sample. In addition, we identified the 77.3–77.8 Mb region on BTA 4, which was not shown to be associated with height at withers in previous studies including those using contrast breeds [15,46–48].

Structural and functional annotation revealed the presence of genes associated with metabolism, milk composition, and synthesis, as well as average daily gain.

Alternative allele frequencies were significantly different in the high- and low-stature breeds for SNPs associated with the height at withers.

No significant difference was found in the frequencies of SNPs associated with body weight on comparing the high- and low-stature cattle groups. However, beef and dairy breeds showed significant differences in allele frequencies of these SNPs (Supplementary Table S2), because the group of low-stature breeds consisted mainly of representatives of primitive breeds (Mongolian, Yakut, and Kyrgyz) in which, until recently, selection work has not been carried out. Almost all low-stature breeds (except Jersey) exhibit beef-type productivity. These have been bred for centuries by nomadic tribes in the harsh climate of the steppes and semi-deserts of Central Asia, survive under poor forage conditions in short dry summers and cold winters, and can be quickly fattened with improved feeding [49]. They were not selected to increase height, but for beef production.

The group of high-stature cattle is represented mainly by dairy breeds, which for a long time have been subjected to strong selection pressure, including increased growth.

Despite the Aberdeen Angus breed being assigned to the high-stature breed group, the same trends were observed in the distribution of allele frequencies in the SNPs located on chromosome 4 associated with height and height/weight traits in the low-stature breed group. This corresponds to the existing exterior and purpose of the breed: the animals are not high relative to specialized dairy (for example, Holstein) breeds; they have a lot of weight because of their better ability to fatten and accumulate subcutaneous fat [50]. Animals of primitive breeds (Mongolian, Yakut, and Kyrgyz) mainly formed the low-stature and beef groups.

5. Conclusions

Using across-breed genome-wide association studies, we were able to identify the SNPs, which are significantly associated with stature and body weight. The SNPs located at BTA4: 77.3–77.8 Mb were associated with the withers height (BovineHD0400021479) and withers height/body weight traits (ARS-BFGL-NGS-116590, Hapmap53144-ss46525999) for groups of high-stature and low-stature breeds and dairy-type breeds, but not for beef-type breeds. For beef-type breeds, significant differences in alleles' frequencies of above-mentioned SNPs were observed only for body weight.

The alleles' frequencies of SNPs on BTA14: 24.5–25.2 Mb associated with both the withers height/body weight (BovineHD1400007259) and body weight (BovineHD1400007247, BovineHD1400007132 and BovineHD1400007161) were significantly different in the group of intensively selected dairy breeds.

The identified SNPs can be considered useful DNA markers for marker-assisted cattle breeding to increase withers height in intensively selected commercial breeds, but not in local and primitive cattle. Additional research is required to reveal the genetic mechanisms of stature formation in primitive breeds.

Supplementary Materials: The following supporting information can be downloaded at: https://www.mdpi.com/article/10.3390/d14080692/s1, Table S1: Allele' frequencies at loci that significantly associated with body weight and height at withers; Table S2: The significance of the difference in the alleles' frequencies in the SNPs associated with the withers' height and the body weight in the different groups of cattle breeds.

Author Contributions: Conceptualization, N.A.Z. and G.B.; methodology, A.S.A. and N.A.Z.; software, A.S.A. and A.V.D.; investigation, N.V.B. and M.S.F.; provision of samples, A.A.S.; writing—original draft preparation, A.S.A. and N.A.Z.; writing—review and editing, A.V.D., A.A.S., G.B. and N.A.Z.;

Diversity 2022, 14, 692 7 of 9

supervision, project administration, and funding acquisition, N.A.Z. All authors have read and agreed to the published version of the manuscript.

Funding: The genotyping of turano-mongolian cattle breeds was funded by the Ministry of Science and Higher Education of the Russian Federation within theme No. FGGN-2022-0002; the genotyping of remaining breeds and data analysis was funded by the Russian Scientific Foundation within Project No. 21-66-00007.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Genotypes of the Aberdeen–Angus and Jersey breeds available in the WIDDE database (http://widde.toulouse.inra.fr/wide, accessed on 1 October 2021). Genotypes of the Russian local cattle breeds are available on reasonable request after signing a research (MTA) agreement.

Acknowledgments: The equipment of the Center for Biological Resources and Bioengineering of Agricultural Animals (Ernst Federal Science Center for Animal Husbandry) was used in the research.

Conflicts of Interest: The authors declare no conflict of interest. The funders had no role in the design of the study: in the collection, analyses, or interpretation of data; writing of the manuscript; or decision to publish the results.

References

- Kemper, K.E.; Visscher, P.M.; Goddard, M.E. Genetic architecture of body size in mammals. Genome Biol. 2012, 13, 244. [CrossRef]
 [PubMed]
- 2. Galton, F. Regression towards mediocrity in hereditary stature. J. Anthropol. Inst. 1886, 15, 246–263. [CrossRef]
- 3. Fink, T.; Tiplady, K.; Lopdell, T.; Johnson, T.; Snell, R.G.; Spelman, R.J.; Davis, S.R.; Littlejohn, M.D. Functional confirmation of PLAG1 as the candidate causative gene underlying major pleiotropic effects on body weight and milk characteristics. *Sci. Rep.* **2017**, 21, 44793. [CrossRef] [PubMed]
- 4. Brotherstone, S. Genetic and phenotypic correlations between linear type traits and production traits in Holstein-Friesian dairy cattle. *Anim. Sci.* **1994**, *59*, 183–187. [CrossRef]
- 5. Bouwman, A.C.; Daetwyler, H.D.; Chamberlain, A.J.; Ponce, C.H.; Sargolzaei, M.; Schenkel, F.S.; Sahana, G.; Govignon-Gion, A.; Boitard, S.; Dolezal, M.; et al. Meta-analysis of genome-wide association studies for cattle stature identifies common genes that regulate body size in mammals. *Nat. Genet.* **2018**, *50*, 362–367. [CrossRef] [PubMed]
- 6. Takasuga, A. PLAG1 and NCAPG-LCORL in livestock. Anim. Sci. J. 2016, 87, 159–167. [CrossRef] [PubMed]
- 7. Purfield, D.C.; Evans, R.D.; Berry, D.P. Reaffirmation of known major genes and the identification of novel candidate genes associated with carcass-related metrics based on whole genome sequence within a large multi-breed cattle population. *BMC Genom.* **2019**, *20*, 720. [CrossRef]
- 8. Zepeda-Batista, J.L.; Núñez-Domínguez, R.; Ramírez-Valverde, R.; Jahuey-Martínez, F.J.; Herrera-Ojeda, J.B.; Parra-Bracamonte, G.M. Discovering of Genomic Variations Associated to Growth Traits by GWAS in Braunvieh Cattle. *Genes* **2021**, *12*, 1666. [CrossRef]
- 9. McClure, M.C.; Morsci, N.S.; Schnabel, R.D.; Kim, J.W.; Yao, P.; Rolf, M.M.; McKay, S.D.; Gregg, S.J.; Chapple, R.H.; Northcutt, S.L.; et al. A genome scan for quantitative trait loci influencing carcass, post-natal growth and reproductive traits in commercial Angus cattle. *Anim. Genet.* **2010**, *41*, 597–607. [CrossRef]
- 10. Karim, L.; Takeda, H.; Lin, L.; Druet, T.; Arias, J.A.; Baurain, D.; Cambisano, N.; Davis, S.R.; Farnir, F.; Grisart, B.; et al. Variants modulating the expression of a chromosome domain encompassing PLAG1 influence bovine stature. *Nat. Genet.* **2011**, 43, 405–413. [CrossRef]
- 11. Littlejohn, M.; Grala, T.; Sanders, K.; Walker, C.; Waghorn, G.; Macdonald, K.; Coppieters, W.; Georges, M.; Spelman, R.; Hillerton, E. Genetic variation in PLAG1 associates with early life body weight and peripubertal weight and growth in Bos taurus. *Anim. Genet.* **2012**, *43*, 591–594. [CrossRef] [PubMed]
- 12. Hoshiba, H.; Setoguchi, K.; Watanabe, T.; Kinoshita, A.; Mizoshita, K.; Sugimoto, Y.; Takasuga, A. Comparison of the effects explained by variations in the bovine PLAG1 and NCAPG genes on daily body weight gain, linear skeletal measurements and carcass traits in Japanese Black steers from a progeny testing program. *Anim. Sci. J.* 2013, 84, 529–534. [CrossRef] [PubMed]
- 13. Naserkheil, M.; Bahrami, A.; Lee, D.; Mehrban, H. Integrating Single-Step GWAS and Bipartite Networks Reconstruction Provides Novel Insights into Yearling Weight and Carcass Traits in Hanwoo Beef Cattle. *Animals* **2020**, *10*, 1836. [CrossRef] [PubMed]
- 14. Fortes, M.R.S.; Kemper, K.; Sasazaki, S.; Reverter, A.; Pryce, J.E.; Barendse, W.; Bunch, R.; McCulloch, R.; Harrison, B.; Bolormaa, S.; et al. Evidence for pleiotropism and recent selection in the PLAG1 region in Australian Beef cattle. *Anim. Genet.* **2013**, 44, 636–647. [CrossRef]

Diversity 2022, 14, 692 8 of 9

15. Randhawa, I.A.S.; Khatkar, M.S.; Thomson, P.C.; Raadsma, H.W. Composite selection signals for complex traits exemplified through bovine stature using multibreed cohorts of European and African Bos taurus. *G3 Genes Genomes Genet.* **2015**, *5*, 1391–1401. [CrossRef]

- 16. Zimin, A.V.; Delcher, A.L.; Florea, L.; Kelley, D.R.; Schatz, M.C.; Puiu, D.; Hanrahan, F.; Pertea, G.; Van Tassell, C.P.; Sonstegard, T.S.; et al. A whole-genome assembly of the domestic cow, Bos taurus. *Genome Biol.* **2009**, *10*, R42. [CrossRef]
- 17. Hensen, K.; Braem, C.; Declercq, J.; Van Dyck, F.; Dewerchin, M.; Fiette, L.; Denef, C.; Van de Ven, W.J. Targeted disruption of the murine Plag1 proto-oncogene causes growth retardation and reduced fertility. *Dev. Growth Differ.* 2004, 46, 459–470. [CrossRef]
- 18. Wei, Z.; Wang, K.; Wu, H.; Wang, Z.; Pan, C.; Chen, H.; Lan, X. Detection of 15-bp Deletion Mutation within PLAG1 Gene and Its Effects on Growth Traits in Goats. *Animals* **2021**, *11*, 2064. [CrossRef]
- 19. Voz, M.L.; Agten, N.S.; Van de Ven, W.J.; Kas, K. PLAG1, the main translocation target in pleomorphic adenoma of the salivary glands, is a positive regulator of IGF-II. *Cancer Res.* **2000**, *60*, 106–113.
- 20. Van Dyck, F.; Declercq, J.; Braem, C.V.; Van de Ven, W.J.M. PLAG1, the prototype of the PLAG gene family: Versatility in tumour development (review). *Int. J. Oncol.* **2007**, *30*, 765–774. [CrossRef]
- 21. De Vos, L.; Declercq, J.; Rosas, G.G.; Van Damme, B.; Roebroek, A.; Vermorken, F.; Ceuppens, J.; van de Ven, W.; Creemers, J. MMTV-cre-mediated fur inactivation concomitant with PLAG1 proto-oncogene activation delays salivary gland tumorigenesis in mice. *Int. J. Oncol.* 2008, 32, 1073–1083. [CrossRef] [PubMed]
- DeChiara, T.M.; Efstratiadis, A.; Robertson, E.J. A growth-deficiency phenotype in heterozygous mice carrying an insulin-like growth factor II gene disrupted by targeting. *Nature* 1990, 345, 78–80. [CrossRef] [PubMed]
- 23. Boitard, S.; Boussaha, M.; Capitan, A.; Rocha, D.; Servin, B. Uncovering adaptation from sequence data: Lessons from genome resequencing of four cattle breeds. *Genetics* **2016**, 203, 433–450. [CrossRef]
- 24. Hou, J.; Qu, K.; Jia, P.; Hanif, Q.; Zhang, J.; Chen, N.; Dang, R.; Chen, H.; Huang, B.; Lei, C. A SNP in PLAG1 is associated with body height trait in Chinese cattle. *Anim. Genet.* **2020**, *51*, 87–90. [CrossRef]
- 25. Abdelmanova, A.S.; Kharzinova, V.R.; Volkova, V.V.; Mishina, A.I.; Dotsev, A.V.; Sermyagin, A.A.; Boronetskaya, O.I.; Petrikeeva, L.V.; Chinarov, R.Y.; Brem, G.; et al. Genetic diversity of historical and modern populations of Russian cattle breeds revealed by microsatellite analysis. *Genes* 2020, 11, 940. [CrossRef] [PubMed]
- 26. Zinovieva, N.A.; Sermyagin, A.A.; Dotsev, A.V.; Boronetslaya, O.I.; Petrikeeva, L.V.; Abdelmanova, A.S.; Brem, G. Animal Genetic resources: Developing the research of allele pool of Russian cattle breeds—minireview. *Sel'skokhozyaistvennaya Biol. Agric. Biol.* **2019**, *54*, 631–641. [CrossRef]
- 27. Sermyagin, A.A.; Dotsev, A.V.; Gladyr, E.A.; Traspov, A.A.; Deniskova, T.E.; Kostyunina, O.V.; Reyer, H.; Wimmers, K.; Barbato, M.; Paronyan, I.A.; et al. Whole-genome SNP analysis elucidates the genetic structure of Russian cattle and its relationship with Eurasian taurine breeds. *Genet. Sel. Evol.* 2018, *50*, 37. [CrossRef] [PubMed]
- 28. Chang, C.C.; Chow, C.C.; Tellier, L.C.; Vattikuti, S.; Purcell, S.M.; Lee, J.J. Second-generation PLINK: Rising to the challenge of larger and richer datasets. *Gigascience* **2015**, *4*, s13742-015. [CrossRef]
- 29. Abdelmanova, A.S.; Sermyagin, A.A.; Dotsev, A.V.; Bardukov, N.V.; Fornara, M.S.; Brem, G.; Zinovieva, N.A. Genome-Wide Screening for SNPs Associated with Stature in Diverse Cattle Breeds. *Biol. Life Sci. Forum* **2022**, *15*, 24. [CrossRef]
- 30. Lee, S.H.; Zhu, C.; Peng, Y.; Johnson, D.T.; Lehmann, L.; Sun, Z. Identification of a novel role of ZMIZ2 protein in regulating the activity of the Wnt/β-catenin signaling pathway. *J. Biol. Chem.* **2013**, *288*, 35913–35924. [CrossRef]
- 31. Sanchez, M.-P.; Ramayo-Caldas, Y.; Wolf, V.; Laithier, C.; El Jabri, M.; Michenet, A.; Boussaha, M.; Taussat, S.; Fritz, S.; Delacroix-Buchet, A.; et al. Sequence-based GWAS, network and pathway analyses reveal genes co-associated with milk cheesemaking properties and milk composition in Montbéliarde cows. *Genet. Sel. Evol.* 2019, 51, 34. [CrossRef] [PubMed]
- 32. Sanchez, M.P.; Govignon-Gion, A.; Croiseau, P.; Fritz, S.; Hozé, C.; Miranda, G.; Martin, P.; Barbat-Leterrier, A.; Letaïef, R.; Rocha, D.; et al. Within-breed and multi-breed GWAS on imputed whole-genome sequence variants reveal candi-date mutations affecting milk protein composition in dairy cattle. *Genet. Sel. Evol.* **2017**, *49*, 68. [CrossRef] [PubMed]
- 33. Huo, N.; Yu, M.; Li, X.; Zhou, C.; Jin, X.; Gao, X. PURB is a positive regulator of amino acid-induced milk synthesis in bovine mammary epithelial cells. *J. Cell. Physiol.* **2019**, 234, 6992–7003. [CrossRef] [PubMed]
- 34. Lu, D.; Miller, S.; Sargolzaei, M.; Kelly, M.; Vander Voort, G.; Caldwell, T.; Wang, Z.; Plastow, G.; Moore, S. Genome-wide association analyses for growth and feed efficiency traits in beef cattle. *J. Anim. Sci.* **2013**, *91*, 3612–3633. [CrossRef]
- 35. Marete, A.G.; Guldbrandtsen, B.; Lund, M.S.; Fritz, S.; Sahana, G.; Boichard, D. A Meta-Analysis Including Pre-selected Sequence Variants Associated With Seven Traits in Three French Dairy Cattle Populations. *Front. Genet.* **2018**, *9*, 522. [CrossRef]
- 36. Ghoreishifar, S.M.; Eriksson, S.; Johansson, A.M.; Khansefid, M.; Moghaddaszadeh-Ahrabi, S.; Parna, N.; Davoudi, P.; Javanmard, A. Signatures of selection reveal candidate genes involved in economic traits and cold acclimation in five Swedish cattle breeds. *Genet. Sel. Evol.* **2020**, *52*, 1–15. [CrossRef]
- 37. Smith, J.L.; Wilson, M.L.; Nilson, S.M.; Rowan, T.N.; Oldeschulte, D.L.; Schnabel, R.D.; Decker, J.E.; Seabury, C.M. Genome-wide association and genotype by environment interactions for growth traits in U.S. Gelbvieh cattle. *BMC Genom.* **2019**, *20*, 926. [CrossRef]
- 38. Saatchi, M.; Schnabel, R.D.; Taylor, J.F.; Garrick, D.J. Large-effect pleiotropic or closely linked QTL segregate within and across ten US cattle breeds. *BMC Genom.* **2014**, *15*, 442. [CrossRef]

Diversity **2022**, 14, 692 9 of 9

39. Lee, S.H.; Choi, B.H.; Lim, D.; Gondro, C.; Cho, Y.M.; Dang, C.G.; Sharma, A.; Jang, G.W.; Lee, K.T.; Yoon, D.; et al. Genome-wide association study identifies major loci for carcass weight on BTA14 in Hanwoo (Korean Cattle). *PLoS ONE* **2013**, *8*, e74677. [CrossRef]

- 40. Nishimura, S.; Watanabe, T.; Mizoshita, K.; Tatsuda, K.; Fujita, T.; Watanabe, N.; Sugimoto, Y.; Takasuga, A. Genome-wide association study identified three major QTL for carcass weight including the PLAG1-CHCHD7 QTN for stature in Japanese Black cattle. *BMC Genet.* 2012, *13*, 40. [CrossRef]
- 41. Zhang, R.; Miao, J.; Song, Y.; Zhang, W.; Xu, L.; Chen, Y.; Zhang, L.; Gao, H.; Zhu, B.; Li, J.; et al. Genome-wide association study identifies the PLAG1-OXR1 region on BTA14 for carcass meat yield in cattle. *Physiol. Genom.* **2019**, *51*, 137–144. [CrossRef] [PubMed]
- 42. Rowan, T.N.; Schnabel, R.D.; Decker, J.E. Uncovering the architecture of selection in two Bos taurus cattle breeds. *bioRxiv* **2021**. [CrossRef]
- 43. Naserkheil, M.; Mehrban, H.; Lee, D.; Park, M.N. Genome-wide Association Study for Carcass Primal Cut Yields Using Single-step Bayesian Approach in Hanwoo Cattle. *Front. Genet.* **2021**, *12*, 752424. [CrossRef] [PubMed]
- 44. Setoguchi, K.; Furuta, M.; Hirano, T.; Nagao, T.; Watanabe, T.; Sugimoto, Y.; Takasuga, A. Cross-breed comparisons identified a critical 591-kb region for bovine carcass weight QTL (CW-2) on chromosome 6 and the Ile-442-Met substitution in NCAPG as a positional candidate. *BMC Genet.* **2009**, *10*, 43. [CrossRef] [PubMed]
- 45. Vanvanhossou, S.; Scheper, C.; Dossa, L.H.; Yin, T.; Brügemann, K.; König, S. A multi-breed GWAS for morphometric traits in four Beninese indigenous cattle breeds reveals loci associated with conformation, carcass and adaptive traits. *BMC Genom.* **2020**, 21, 783. [CrossRef]
- 46. Doyle, J.L.; Berry, D.P.; Veerkamp, R.F.; Carthy, T.R.; Walsh, S.W.; Evans, R.D.; Purfield, D.C. Genomic regions associated with skeletal type traits in beef and dairy cattle are common to regions associated with carcass traits, feed intake and calving difficulty. *Front. Genet.* **2020**, *11*, 20. [CrossRef]
- 47. Pryce, J.E.; Hayes, B.J.; Bolormaa, S.; Goddard, M.E. Polymorphic regions affecting human height also control stature in cattle. *Genetics* **2011**, *187*, 981–984. [CrossRef]
- 48. An, B.; Xu, L.; Xia, J.; Wang, X.; Miao, J.; Chang, T.; Song, M.; Ni, J.; Xu, L.; Zhang, L.; et al. Multiple association analysis of loci and candidate genes that regulate body size at three growth stages in Simmental beef cattle. *BMC Genet.* **2020**, 21, 32. [CrossRef]
- 49. Abdelmanova, A.S.; Kharzinova, V.R.; Volkova, V.V.; Dotsev, A.V.; Sermyagin, A.A.; Chinarov, R.Y.; Zinovieva, N.A.; Boronetskaya, O.I.; Lutshikhina, E.M.; Sölkner, J.; et al. Comparative study of the genetic diversity of local steppe cattle breeds from Russia, Kazakhstan and Kyrgyzstan by microsatellite analysis of museum and modern samples. *Diversity* **2021**, *13*, 351. [CrossRef]
- 50. Bown, M.D.; Muir, P.D.; Thomson, B.C. Dairy and beef breed effects on beef yield, beef quality and profitability: A review. *New Zealand J. Agric. Res.* **2016**, *59*, 74–184. [CrossRef]