

Supplementary Materials

A new *Micromonospora* strain with potent antibiotic activity isolated from the microbiome of a mid-Atlantic deep-sea sponge

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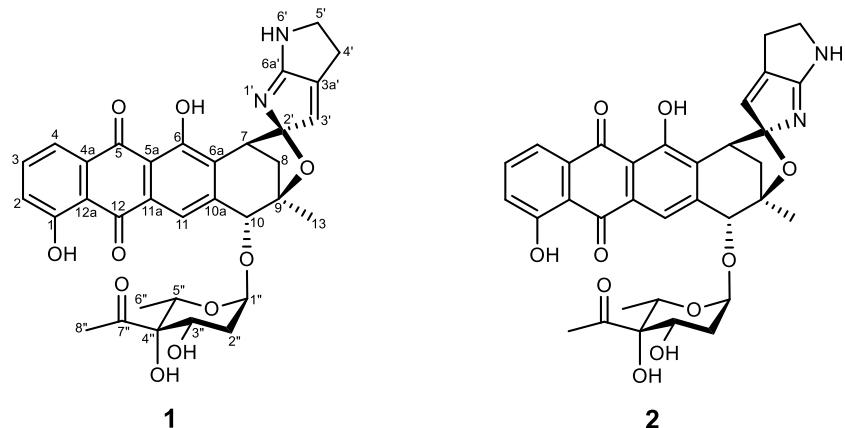
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Supplementary Section S1: LCMS analysis of the extract from 28ISP2-28^T

LCMS data were obtained on a Waters LCMS system comprising Waters 2767 autosampler, Waters 515 HPLC pump, Waters 2998 Diode Array detector, Waters 2424 ELS detector and Waters Quatro Micro mass spectrometer. HPLC grade H₂O and MeCN were added with 0.05% formic acid as solvent system. Analytical LC-MS data were obtained using a Phenomenex Kinetex column (C18, 250 x 4.60 mm, 5 µm) at a flow rate of 1 mL/min, with a gradient of 5% MeCN to 95% MeCN in 20 mins. Preparative HPLC purification were carried out using a Phenomenex Kinetex column (C18, 250 x 21.20 mm, 5 µm) at a flow rate of 16 mL/min, with a gradient of 30 to 40% of MeCN in water over 20 min. HR-ESI-MS data were obtained on a Bruker Daltonics micrOTOF II instrument. NMR data were collected on a Bruker Cryo500 NMR spectrometer.

Supplementary Section S2: NMR chemical shifts of (**2**) (Isoquinocycline B).



HR-ESI-MS: $m/z = 617.2143$ $[M+H]^+$ (617.2130 calcd for $C_{33}H_{33}N_2O_{10}$); 1H NMR (500 MHz, methanol-d₄): δ 7.94 (s, 1H, H-11), 7.87 (dd, $J = 7.6, 1.1$ Hz, 1H, H-4), 7.79 (t, $J = 8.0$ Hz, 1H, H-3), 7.37 (dd, $J = 8.4, 1.1$ Hz, 1H, H-2), 5.57 – 5.51 (m, 1H, H-3'), 5.44 (dd, $J = 5.2, 3.4$ Hz, 1H, H-1''), 4.70 – 4.63 (m, 1H, H-5'), 4.58 (d, $J = 3.2$ Hz, 1H, H-10), 4.23 (d, $J = 3.8$ Hz, 1H, H-7), 4.16 (m, 2H, H-5'), 3.85 (dd, $J = 6.0, 3.8$ Hz, 1H, H-3''), 2.89 – 2.78 (m, 2H, H-4'), 2.51 – 2.36 (m, 2H, H-8), 2.26 (s, 3H, H-8''), 2.23 (m, 1H, H-2''), 1.86 (ddd, $J = 14.5, 6.0, 3.4$ Hz, 1H, H-2''), 1.73 (s, 3H, H-13), 1.15 (d, $J = 6.5$ Hz, 3H, H-6''); ^{13}C NMR (125 MHz, methanol-d₄): δ 212.9 (C-7''), 189.3 (C-5), 188.7 (C-12), 169.2 (C-6a'), 163.9 (C-1), 159.8 (C-6), 145.5 (C-10a), 141.8 (C-3a'), 138.2 (C-3), 134.7 (C-3'), 134.6 (C-4a), 134.2 (C-6a), 133.8 (C-11a), 126.0 (C-2), 121.4 (C-11), 120.4 (C-4), 117.2 (C-12a), 116.4 (C-5a), 116.0 (C-2''), 97.6 (C-1''), 87.0 (C-9), 82.0 (C-4''), 78.3 (C-10), 71.3 (C-3''), 67.0 (C-5''), 54.7 (C-5'), 45.0 (C-7), 35.9 (C-8), 33.7 (C-2''), 27.0 (C-8''), 23.4 (C-13), 21.8 (C-4'), 14.9 (C-6'').

Supplementary Section S3: 16S rRNA gene sequencing of 28ISP2-46^T.

Forward 16S rRNA gene sequencing

```
TNNNNNNNGAGTCGAGCGGAAGGNCTTCGGNNANTCGAGCGGCGAACGGGTGAGTAAC  
ACGTGAGCAACCTGCCCTAGGCTTGGATAACCCCGGAAACCGGGCTAATACCGAAT  
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CTATCAGCTTGGTGGGTGATGGCCTACCAAGGCGACGGTAGCCGGCCTGAGA  
GGCGACCGGCCACACTGGGACTGAGACACGGCCCAGACTCTACGGGAGGCAGCAGTGG  
GGAATATTGCACAATGGCGGAAGCCTGATGCAGCGACGCCGCGTAGGGGATGACGGCCT  
TCGGGTTGAAACCTTTCACTGGGACGAAGCGTAAGTGACGGTACCTGCAGAAGAAG  
CGCCGGCCAACACTACGTGCCAGCAGCCGCGTAAGACGTAGGGCGCGAGCGTTGCCGAT  
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AACTCCAGGCCTGCGGTGATACGGGCAGGCTAGAGTTCGTAGGGGAGACTGGAATTCC  
TGGTGTAGCGGTGAAATGCGCAGATATCAGGAGGAACACCGGTGGCGAAGGCGGTCTCT  
GGCGCATACTGACGCTGAGGAGCGAAAGCGTGGGAGCGAACAGGATTAGATACCCTGG  
TAGTCCACGCTGTAACGTTGGCGCTAGGTGTGGGGGCTCTCCGGTCCCTGTGCCG  
CAGCTAACGCATTAAGCGCCCCGCTGGGAGTACGGCGCAAGGCTAAAACCAAAGGA  
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Reverse 16S rRNA gene sequencing

```
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CGCGAAGAACCTTACCTGGTTTGACATGGCGCAAACCTGCGAGAGATGTAAGGTCTCGGGGGCGGT  
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```

16S rRNA gene sequencing (combined forward and reverse)

GCAGTCGAGCGGAAGGNCTTCGGNNANTCGAGCGCGAACGGGTGAGTAACACGTGAGCAACCTGCC
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AGGCTAGAGTTCGGTAGGGGAGACTGGAATTCTGGTGTAGCGGTGAAATGCGCAGATATCAGGAGGAAC
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TTAGATACCTGGTAGTCCACGCTGTAACGTTGGCGCTAGGTGTGGGGGCTCTCCGGTCCCTGTG
CCGCAGCTAACGCTTAAGCGCCCCGCTGGGAGTACGGCGCAAGGCTAAACTCAAAGGAATTGACG
GGGGCCCGACAAGCGCGGAGCATGCGGATTAATTGATGCAACGCGAAGAACCTTACCTGGGTTGAC
ATGGCCGAAAACCTGAGAGATGTAAGGTCTTCGGGGCGGTACAGGTGGTGCATGGCTGTCGTAG
CTCGTGTGAGATGTTGGGTTAAGTCCCGCAACGAGCGAACCGCTCGTTCGATGTTGCCAGCGCTTA
TGGCGGGGACTCATCGAAGACTGCCGGGTCAACTGGAGGAAGGTGGGATGACGTCAAGTCATCATGC
CCCTTATGTCCAGGGCTTCACGCATGCTACAATGCCGGTACAATGGCTGCGATACCGTGAGGTGGAGC
GAATCCCAAAAGCCGGTCTCAGTCGGATCGGGCTTGCAACTCGACCCGTGAAGTCGGAGTCGCTAG
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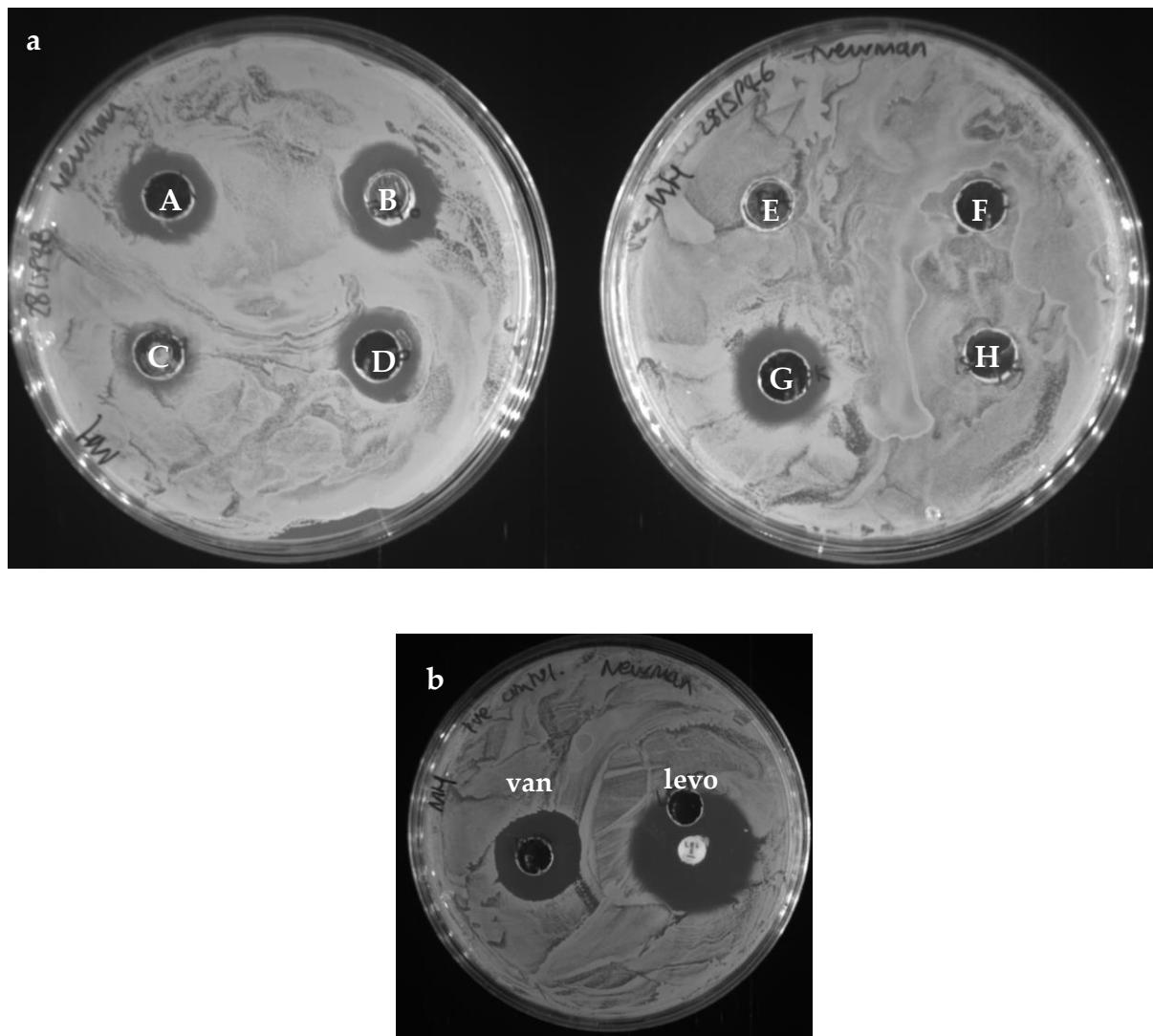


Figure S1. Small volume liquid culture antibiotic activity assay of 28ISP2-46^T. **(a)** Activity assay of extracts from 28ISP2-46^T: A, culture supernatant before extraction (50 µl); B, ethyl acetate fraction (50 µl); C, aqueous fraction (50 µl); D, pellet extraction with methanol (50 µl); E, ethyl acetate only control (50 µl); F, methanol only control (50 µl); G, culture supernatant before extraction treated with Proteinase K (50 µl); H, ISP2 media only treated with Proteinase K (50 µl). **(b)** Control plate with a well on the left containing 10 µg vancomycin (van) and a disk on the right containing 5 µg levofloxacin (levo).

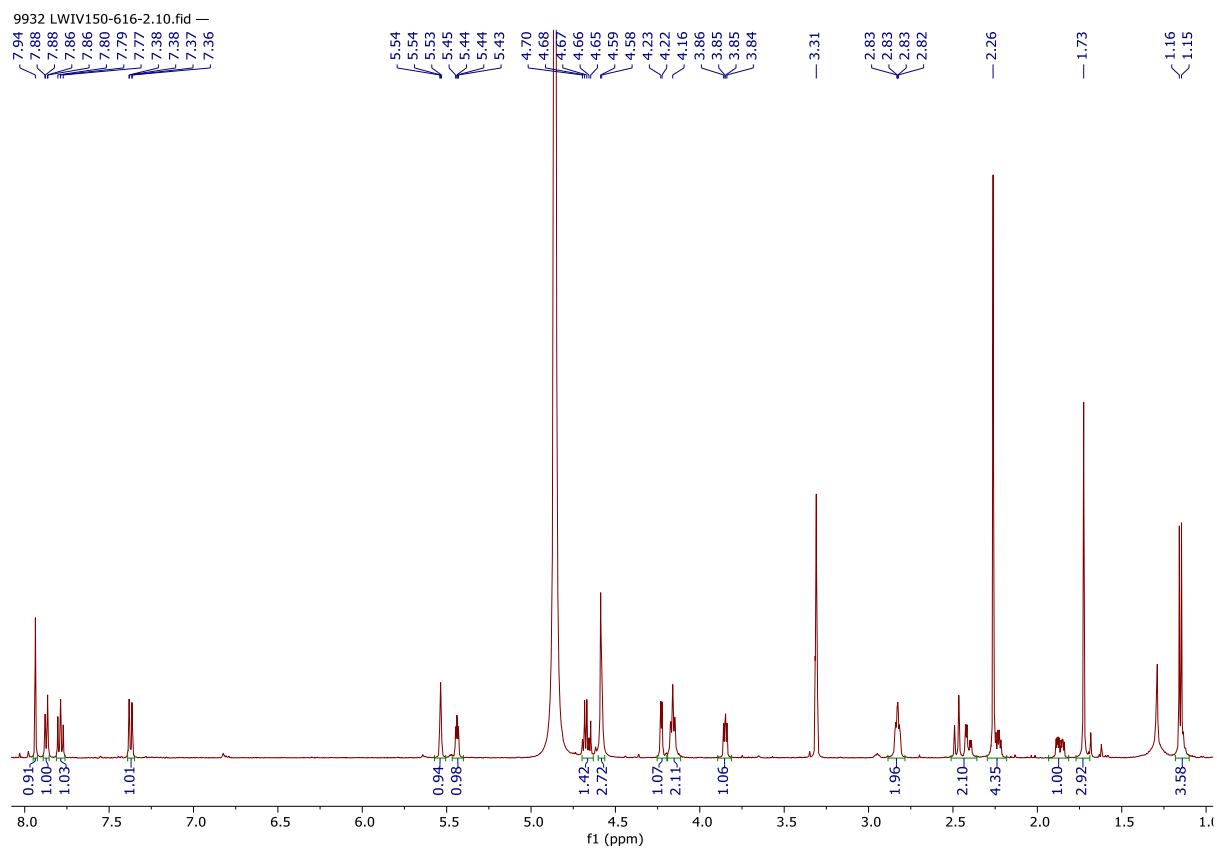


Figure S2. ^1H NMR spectrum of isoquinocycline B (**2**, CD_3OD , 500 MHz)

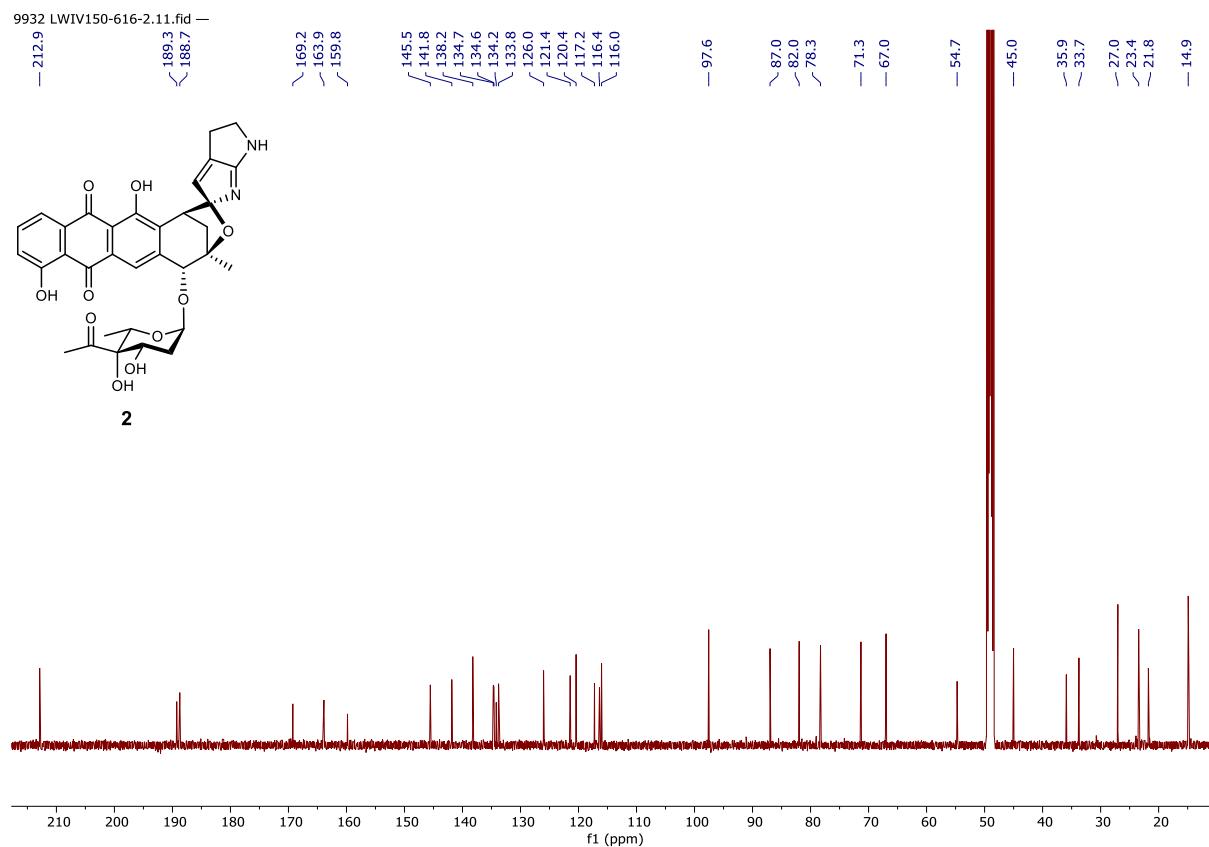


Figure S3. ^{13}C NMR spectrum of isoquinocycline B (**2**, CD_3OD , 125 MHz)

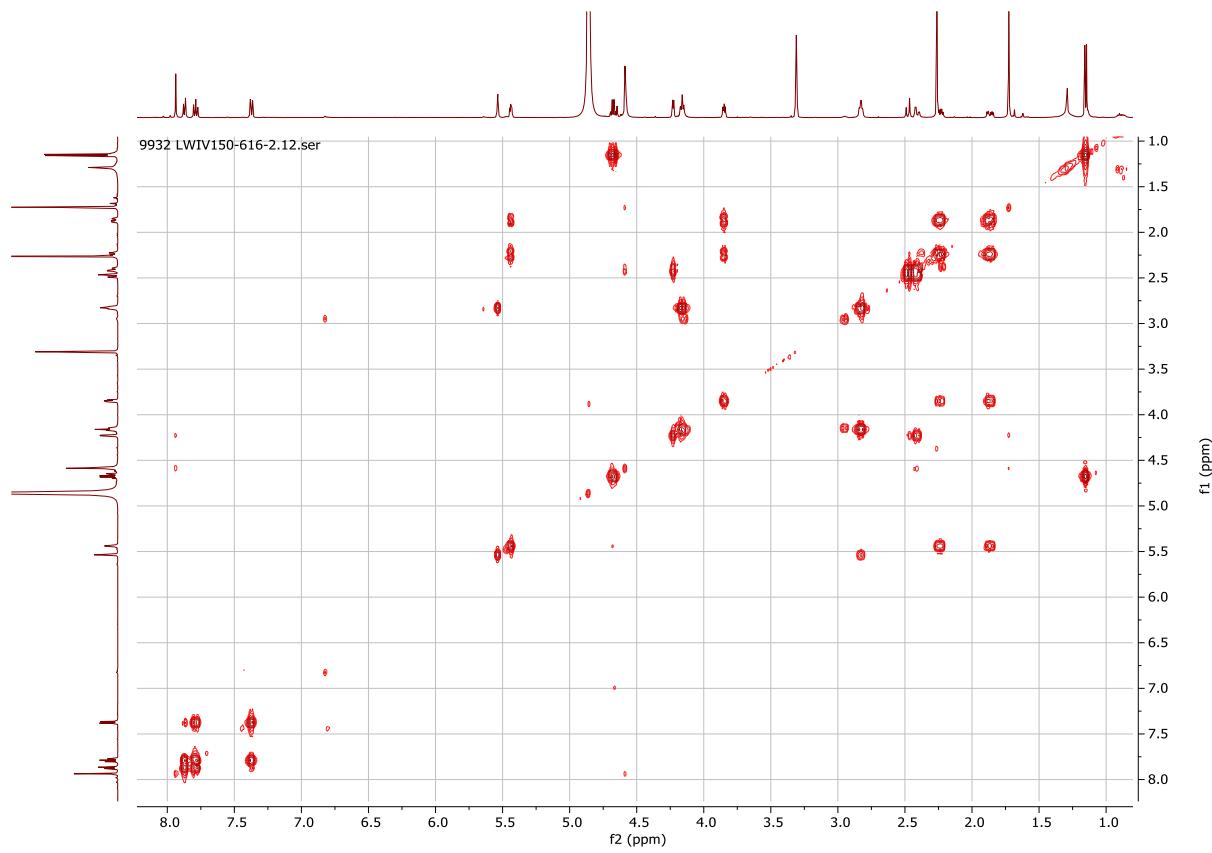


Figure S4. ¹H-¹H COSY spectrum of spectrum of isoquinocycline B (2)

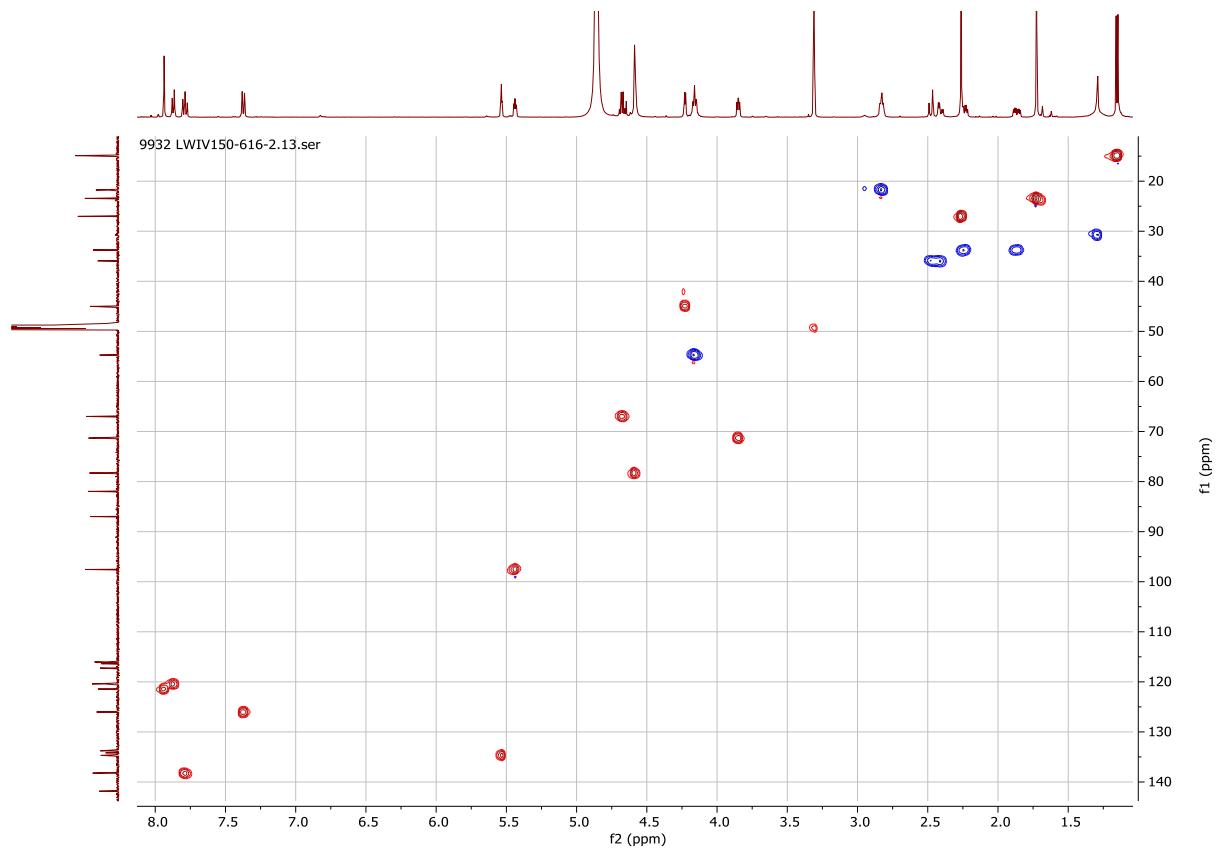


Figure S5. HSQC spectrum of isoquinocycline B (**2**)

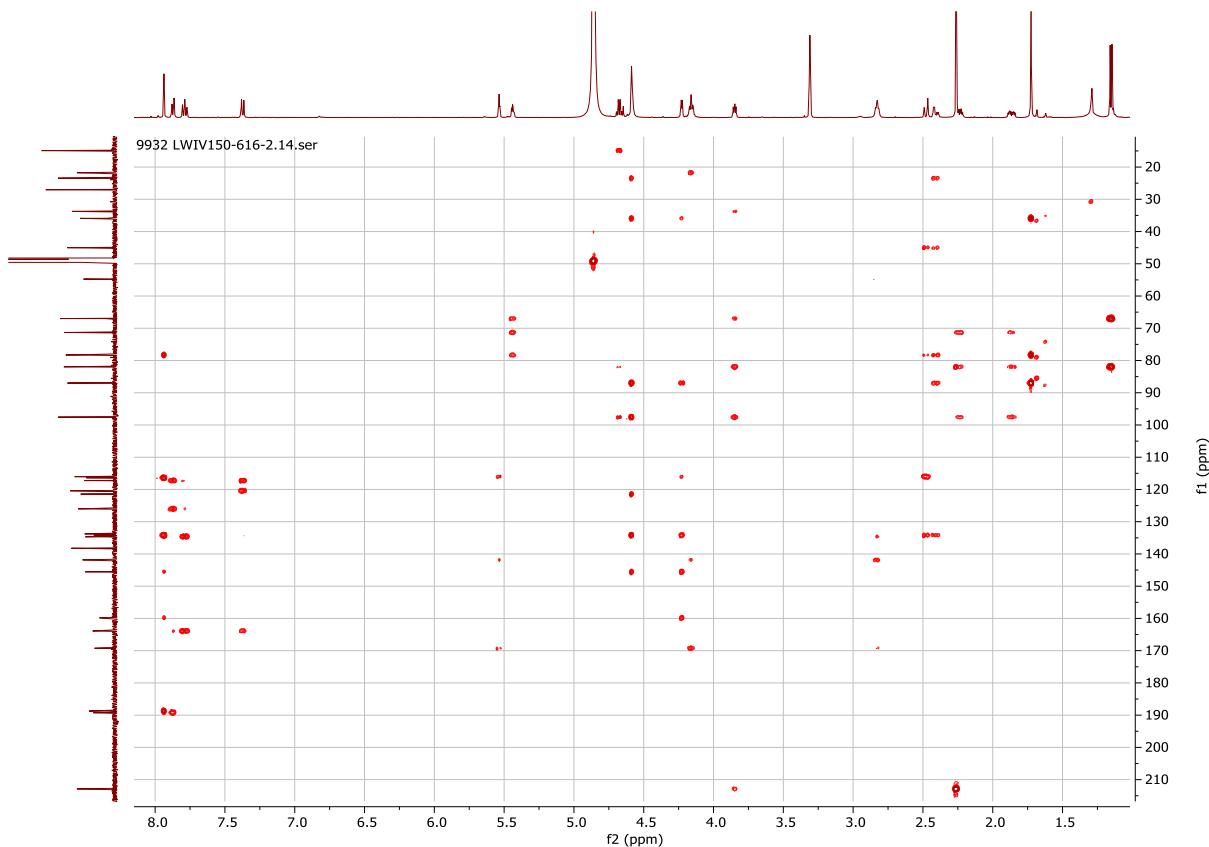


Figure S6. HMBC spectrum of isoquinocycline B (**2**)

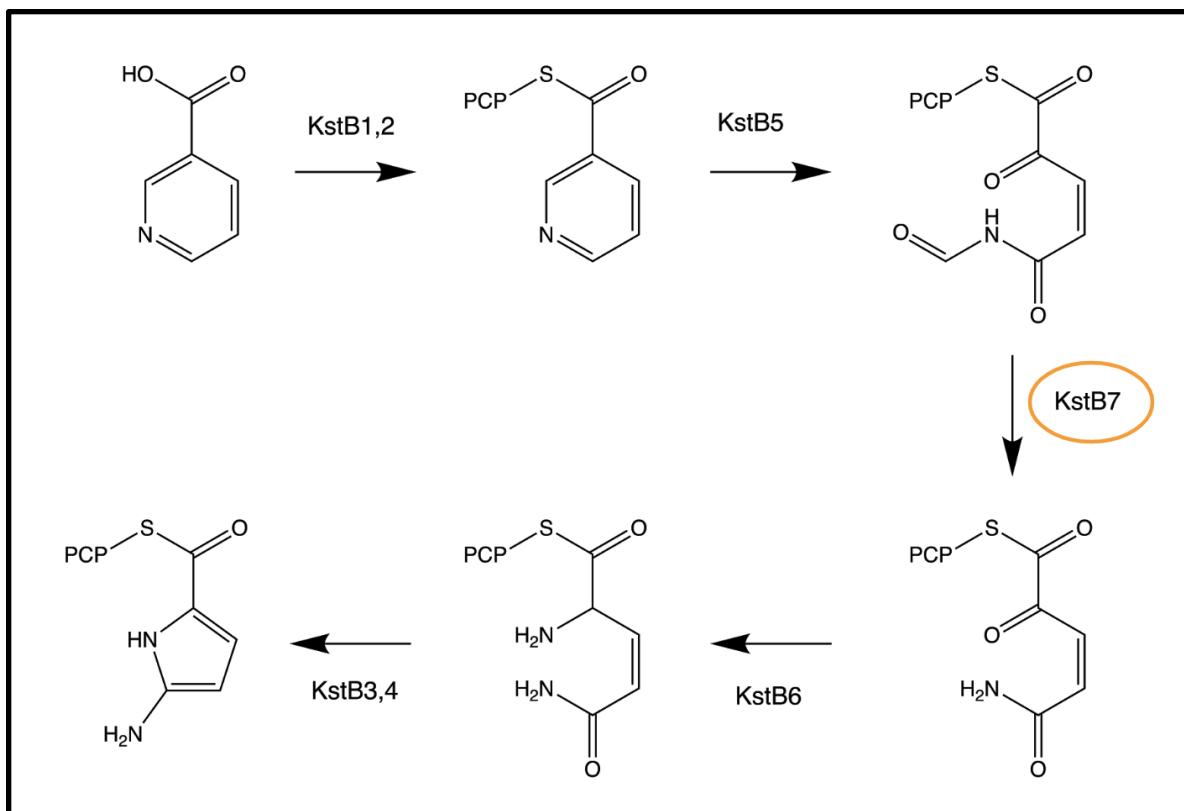


Figure S7. The predicted function of KstB7 in pyrrolopyrrole biosynthesis according to Ma *et al.* 2013 [1].

Table S1: Bacterial strains used in this study.

Organism	Strain	Details	Reference
28ISP2-46 ^T	<i>Micromonospora ferruginea</i> sp. nov.		This study [2]
<i>Staphylococcus aureus</i>	Newman		[2]
<i>S. aureus</i>	Mu50	Methicillin & vancomycin resistant	[3]
<i>Pseudomonas aeruginosa</i>	PAO1		[4]
<i>Escherichia coli</i>	BW25113		[5]
<i>Klebsiella pneumoniae</i>	NCTC 5055		[6]
<i>Acinetobacter baumannii</i>	ATCC 19606		[7]
<i>A. baumannii</i>	ATCC 17978		[8]
<i>A. baumannii</i>	CCD167h1.1	ATCC 17978 $\Delta adeIJ$, $\Delta A1S_3447-3446$	
<i>E. coli</i>	CCD121a8	BW25113 $\Delta acrAB$, $\Delta acrD$, $\Delta emrAB$, $\Delta macAB$, $\Delta mdfA$, $\Delta mdtK$	
<i>E. coli</i>	CCD181-1	BW25113 <i>imp4213</i>	
<i>K. pneumoniae</i>	ATCC 10031		?
<i>P. aeruginosa</i>	ATCC 33359		[9]
<i>P. aeruginosa</i>	CCD047	NCTC 11451 $\Delta mexAB$, $\Delta mexCD$, $\Delta mexVW$, $\Delta mexXY$	
<i>S. aureus</i>	SH1000		[10]

Table S2: Sequence identity comparison of the 28ISP2-46^T 16S rRNA gene sequence with the most closely related strains. All *Micromonospora* strains.

Strain	Coverage (%)	Identity (%)
<i>M. humi</i>	99	99
<i>M. maritima</i>	99	99
<i>M. marina</i>	99	99
<i>M. coxensis</i>	99	99
<i>M. aurantiaca</i>	99	99
<i>M. sediminicola</i>	99	99
<i>M. purpureochromogenes</i>	99	99
<i>M. chaiyaphumensis</i>	99	99

Table S3. BCG comparison of *Micromonospora* group Ia strains (*M. humi*, *M. aurantiaca*, *M. auratinigra*, *M. chalcea*, *M. chaiyaphumensis*, *M. chersina*, *M. marina*, *M. sediminicola*, and *M. tulbaghiae*) with strain 28ISP2-46^T.

28ISP2-46 ^T BCG region	BCG class	Closest BGC relative (genes with homologs) ^a	Group Ia species with homologous BCGs
1	Type III PKS	Alkyl-O-dihydrogeranyl-methoxyhydroquinones (71%)	All
2	Terpene	Isorenieratene (25%)	All
3	Terpene	Phosphonoglycans (3%)	All
4	N-acetylglutaminylglutamine amide (NAGGN)	None	All
5	NRPS / type I PKS	Bleomycin (6%)	All but <i>M. aurantiaca</i> , <i>M. marina</i> , <i>M. tulbaghiae</i>
6	Lanthipeptide	None	<i>M. chalcea</i>
7	NRPS, type I PKS	Nostopeptolide A2 (25%)	All but <i>M. sediminicola</i>
8	Lanthipeptide	SapB (75%)	<i>M. chaiyaphumensis</i> , <i>M. chersina</i> , <i>M. humi</i>
9	Type I PKS, NRPS-like	Rifamycin (38%)	<i>M. aurantiaca</i> , <i>M. chalcea</i> , <i>M. sediminicola</i>
10	NRPS, arylpolyene	Kedarcidin (13%)	<i>M. chaiyaphumensis</i> , <i>M. humi</i>
11	Terpene	Nocathiacin (4%)	All but <i>M. chaiyaphumensis</i> , <i>M. chersina</i>
12	Type II PKS	Formicamycins A-M (13%)	All
13	NRPS	Azicemicin B (13%)	<i>M. aurantiaca</i> , <i>M. humi</i> , <i>M. marina</i>
14	Siderophore	Desferrioxamine E (100%)	All but <i>M. aurantiaca</i> , <i>M. chalcea</i>
15	Oligosaccharide, terpene, lanthipeptide	Lobosamide A/B/C (13%)	<i>M. aurantiaca</i> , <i>M. chalcea</i> , <i>M. humi</i> , <i>M. marina</i>

16	NRPS, type II PKS	Feglymycin (63%)	None
17	Type II PKS, ectoine, NRPS, type I PKS, other	Kosinostatin (77%)	None
18	Bacteriocin	Lymphostatin (33%)	All
19	Terpene	None	All

^a Percentage values represent sequence identity between the closest BGC relative with strain 28ISP2-46^T.

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