

Supplementary Materials

Deciphering the Glycosylation Steps in the Biosynthesis of P-1894B and Grincamycin Isolated from Marine-derived *Streptomyces lusitanus* SCSIO LR32.

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Abstract: Recently, we re-isolated the glycosylated angucycline antibiotics P-1894B (**1**) and grincamycin (**1'**) from the marine-derived *Streptomyces lusitanus* SCSIO LR32 as potent antitumor agents and identified their biosynthesis gene cluster *gcn*. Both P-1894B (**1**) and grincamycin (**1'**) possess a trisaccharide and a disaccharide moiety comprised of five deoxysugars. In this work, three genes encoding glycosyltransferases (GcnG1, GcnG2, and GcnG3) responsible for the assembly of deoxysugars into angucycline aglycone were identified from the biosynthesis gene cluster *gcn*. Gene inactivations of *gcnG1*, *gcnG2*, *gcnG3*, and *gcnG1G2* by lambda-RED-mediated gene replacements led to the construction of four mutants, in which the glycosyltransferase genes were disrupted, respectively. The metabolites from the mutants were purified and identified, including two new analogues designated as grincamycin U (**3a**) and V (**3'**). The sequential glycosylation steps in the biosynthesis of P-1894B (**1**) and grincamycin (**1'**) catalyzed by GcnG3, GcnG1, and GcnG2 were elucidated.

Keywords: angucycline; glycosylation; biosynthesis; P-1894B; grincamycin; marine *Streptomyces*

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Table S1. Deduced functions of ORFs containing *gcnG1~G3* and *gcnQ* genes in the grincamycin biosynthetic gene cluster.

gene	Size(a.a)	Proposed function	Protein homologue and origin	Identity/ similarity	accession number
<i>gcnG1</i>	430	O-Glycosyltransferase	SaqGT2[<i>Micromonospora</i> sp. Tu 6368]	54/69	ACP19363.1
<i>gcnG2</i>	404	O-Glycosyltransferases	SaqGT4[<i>Micromonospora</i> sp. Tu 6368]	58/70	ACP19365.1
<i>gcnG3</i>	376	C-Glycosyltransferase	UrdGT2[<i>Streptomyces fradiae</i> Tue2717]	79/85	AAF00209.1
<i>gcnQ</i>	529	FAD/FMN-containin-g dehydrogenases	AknOx[<i>Streptomyces galilaeus</i> ATCC31615]	56/69	ABI15166.1

Table S2. Bacteria and plasmid used in this study.

Strains	Description	Reference or source
<i>E. coli</i>		
ET12567	<i>dam</i> , <i>dcm</i> , <i>hsdS</i> , <i>cat</i> , <i>tet</i>	[MacNeil, 1992]
BW25113	K12 derivative: <i>araBAD</i> , <i>rhaBAD</i>	[Datsenko, 2000]
Plasmids		
pCR2.1	Amp ^r , Kan ^r , general clone vector	Invitrogen
pIJ790	CmI ^r , including λ -RED (<i>gam</i> , <i>bet</i> , <i>exo</i>) for PCR-targeting	This study
pIJ773	<i>aac(3)IV</i> (ApraR), <i>oriT</i>	This study
pUZ8002	<i>tra</i> , <i>neo</i> , <i>RP4</i>	[Paget, 1999]
Cosmid		
228A	<i>Streptomyces lusitanus</i> SCSIO LR32 genomic library cosmid	This study
pJu4014	228A cosmid derivative where <i>gcnG1</i> was disrupted by <i>aac(3)IV</i> + <i>oriT</i> using primers G1delF and G1delR	This study
pJu4015	228A cosmid derivative where <i>gcnG2</i> was disrupted by <i>aac(3)IV</i> + <i>oriT</i> using primers G2delF and G2delR	This study
pJu4016	228A cosmid derivative where <i>gcnG1G2</i> was disrupted by <i>aac(3)IV</i> + <i>oriT</i> using primers G1delF and G2delR	This study
pJu4017	228A cosmid derivative where <i>gcnG3</i> was disrupted by <i>aac(3)IV</i> + <i>oriT</i> using primers G3delF and G3delR	This study

Table S3. The primer pairs used for mutant construction.

Gene target	Primer code	Primer pairs used for inactivation (5'-3')
<i>gcnG1</i>	G1delF	GTCTGTTCACCATCTTTCCGGCGACGGCGCACCTGTATattccggggatccgctcgacc
	G1delR	GACCATCTCGTCCCACATGACCAGCGGCATCACCAGCTGttaggctggagctgcttc
<i>gcnG2</i>	G2delF	TTCGAGGTGAACGGGATGCTGCTGGAGGGCTGGAGCCGattccggggatccgctcgacc
	G2delR	CAGCGGGAAGTGGTCCGCCAGCTGCGGAAGCACCAACTGttaggctggagctgcttc
<i>gcnG1</i> <i>G2</i>	G1delF	GTCTGTTCACCATCTTTCCGGCGACGGCGCACCTGTATattccggggatccgctcgacc
	G2delR	CAGCGGGAAGTGGTCCGCCAGCTGCGGAAGCACCAACTGttaggctggagctgcttc
<i>gcnG3</i>	G3delF	ACCACCGATCTGCCGATCCGGCACCTTCATCACCACCGACattccggggatccgctcgacc
	G3delR	CTTGGAATGAGGAGTTGCGGTACACGGCGTTCAGACCTgtaggctggagctgcttc

Table S4. The primers used for PCR confirmation of the double-crossover mutants.

Gene	Primer code	Primer pairs designed to verify the mutant strains (5'-3')	Fragment replaced	Length of desired PCR fragments	
				Wild strain	Mutant strain
<i>gcnG1</i>	G1tF	CTTCGCAGCCCTCAATCTCCA	999	1449	1819
	G1tR	GTACGAGCTGCTTGTGCATCT			
<i>gcnG2</i>	G2tF	ACCCATTTCTTCCGTTGGTG	717	1035	1687
	G2tR	GTCGGACAGCAGCTCGTCCAG			
<i>gcnG1G2</i>	G1tF	CTTCGCAGCCCTCAATCTCCA	2201	2668	1836
	G2tR	GTCGGACAGCAGCTCGTCCAG			
<i>gcnG3</i>	G3tF	ATGGTCCCGTACATCACGTCG	687	1044	1726
	G3tR	TCCTCTCGGTTTCGTGTCGTCT			

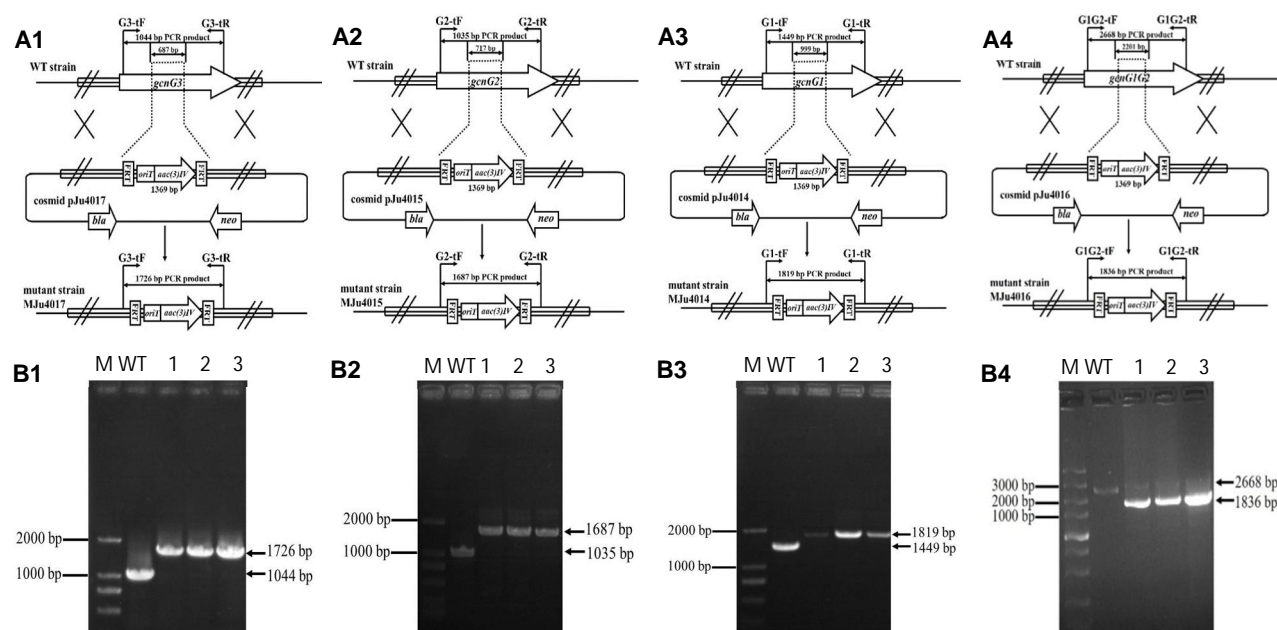


Figure S1. Gene inactivation of *gcnG3~gcnG1*, and *gcnG1G2*. A1~A4: construction of $\Delta gcnG3$, $\Delta gcnG2$, $\Delta gcnG1$, and $\Delta gcnG1G2$ mutants; B1~B4: gel electrophoresis of PCR products, DNA templates were from DNA marker DL 2000 (Takara, lane M), *Streptomyces lusitanus* SCSIO LR32 (lane WT), and $\Delta gcnG3$, $\Delta gcnG2$, $\Delta gcnG1$, and $\Delta gcnG1G2$ mutant strains (lanes 1, 2, 3), respectively.

Figure S2. Procedure for isolation of compound from the mutant $\Delta gcnG3$.

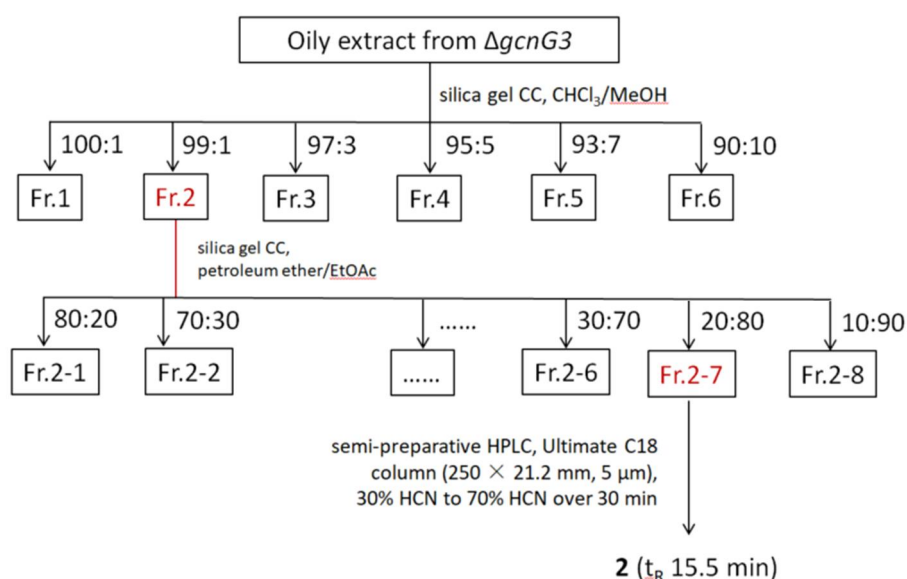


Figure S3. Procedure for isolation of compounds from the mutant $\Delta gcnG2$.

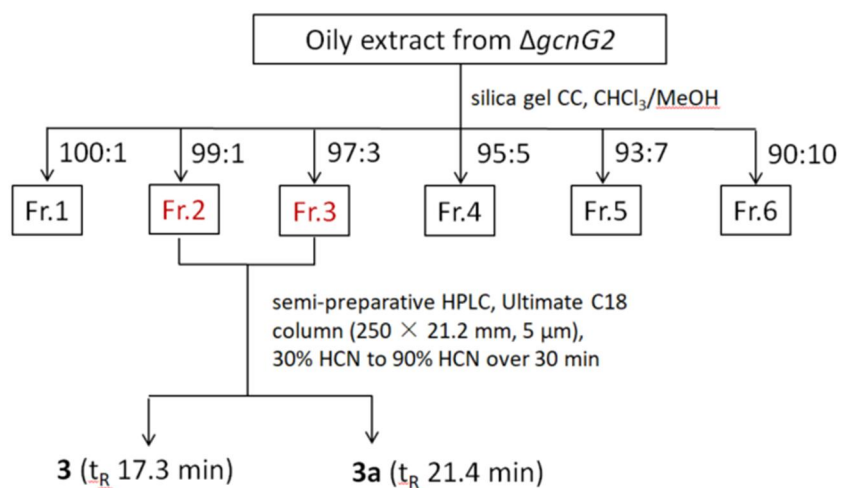


Figure S4. Procedure for isolation of compound from the mutant $\Delta gcnG1$.

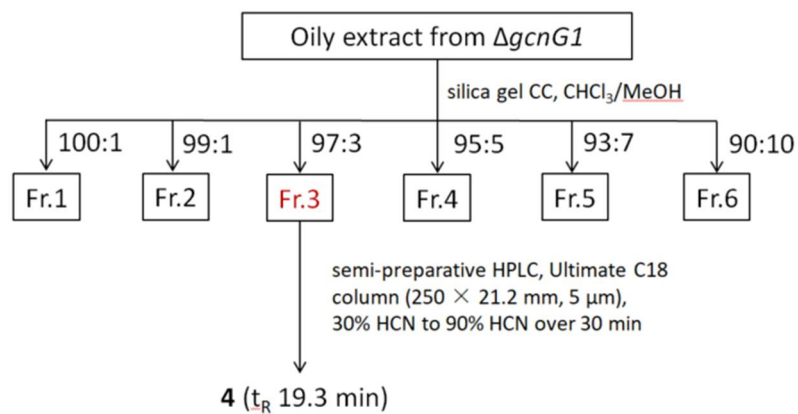


Figure S5. Procedure for isolation of compound from the mutant $\Delta gcnG1G2$.

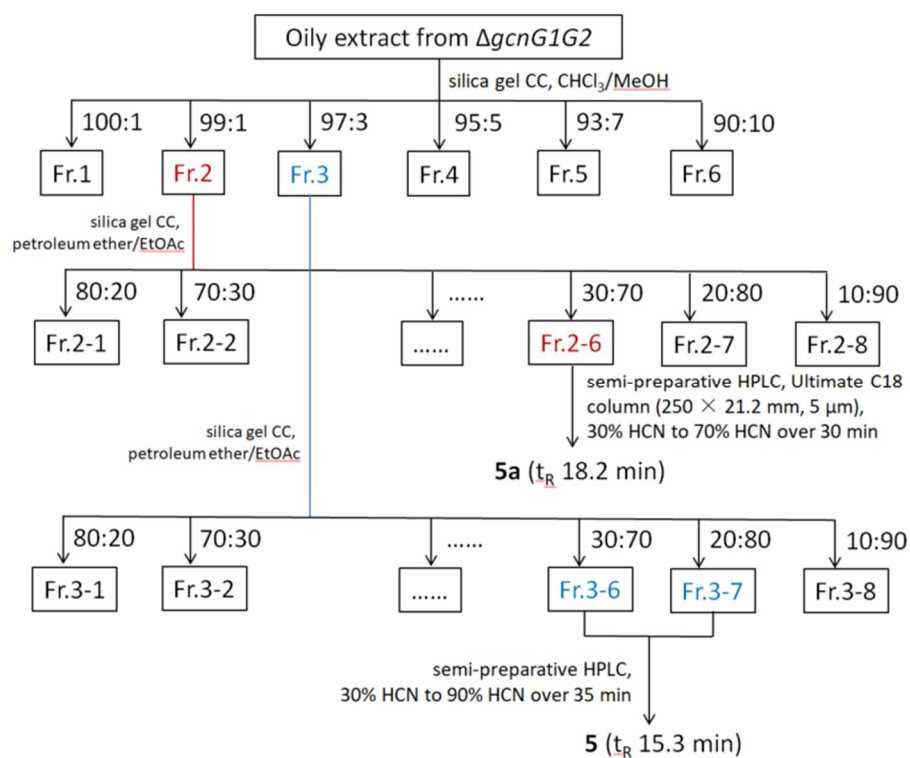


Figure S6. The UV spectrum of grincamycin U (**3a**).

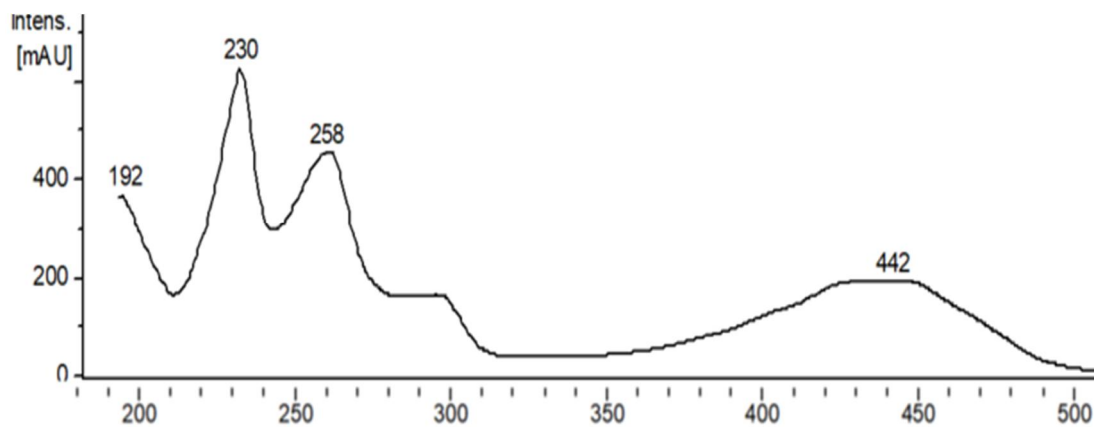


Figure S7. The (-)HR-ESI-MS spectrum of grincamycin U (**3a**).

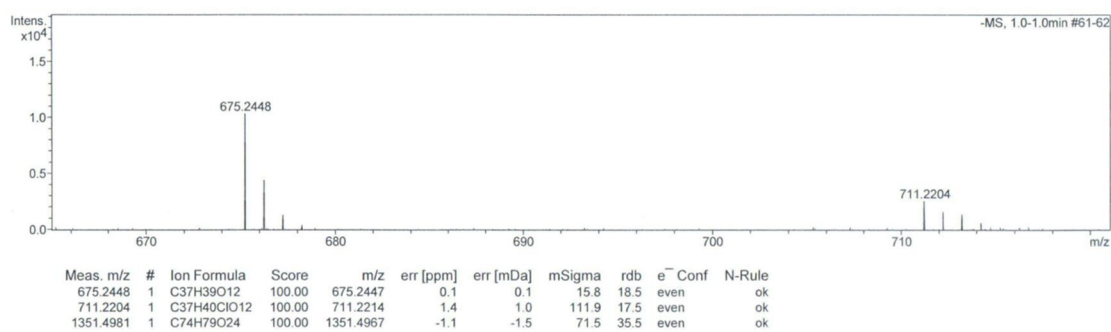


Figure S8. The ^1H NMR (700 MHz) spectrum of grincamycin U (**3a**) in CD_3OD .

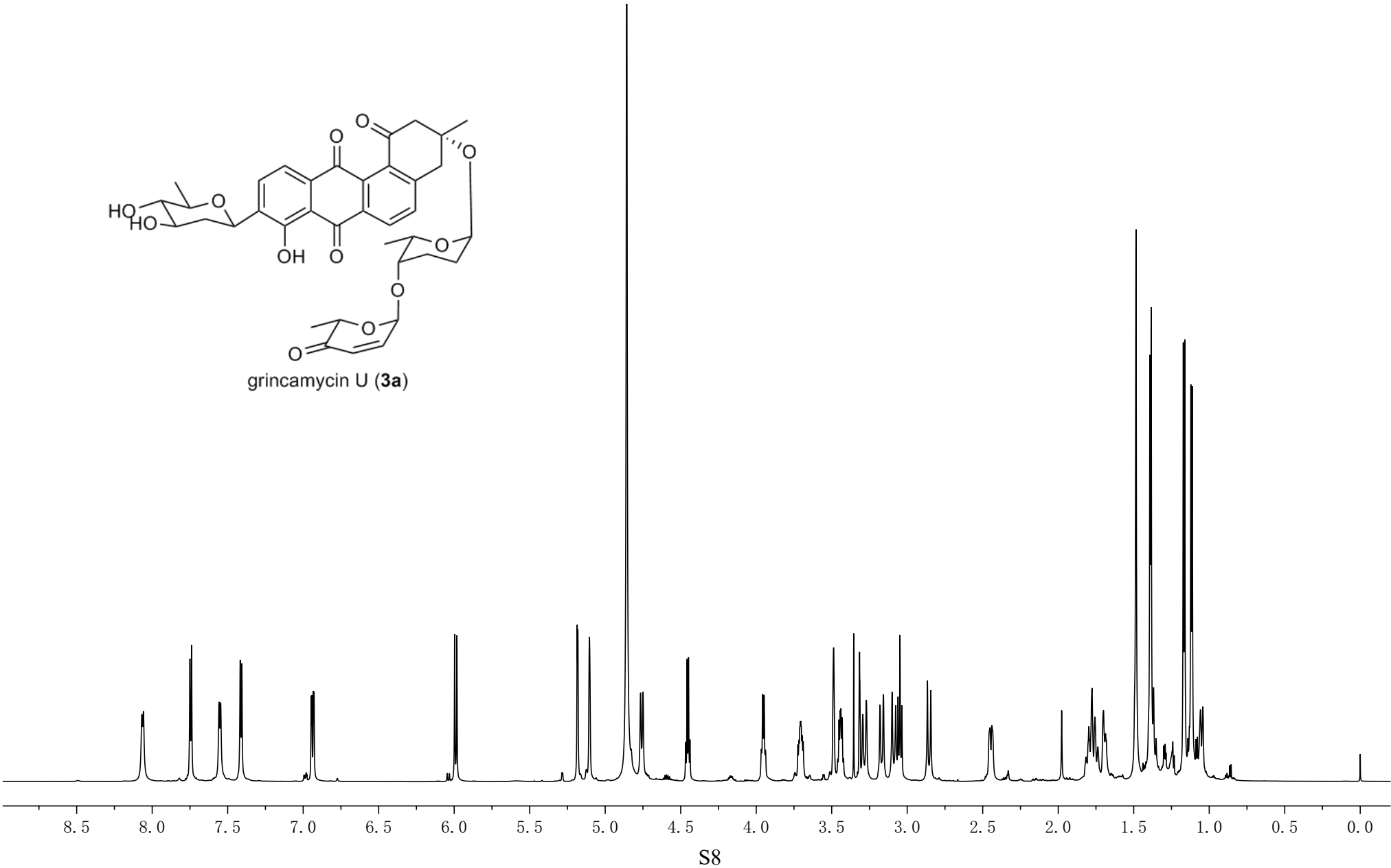


Figure S9. The ^{13}C NMR (175 MHz) spectrum of grincamycin U (**3a**) in CD_3OD .

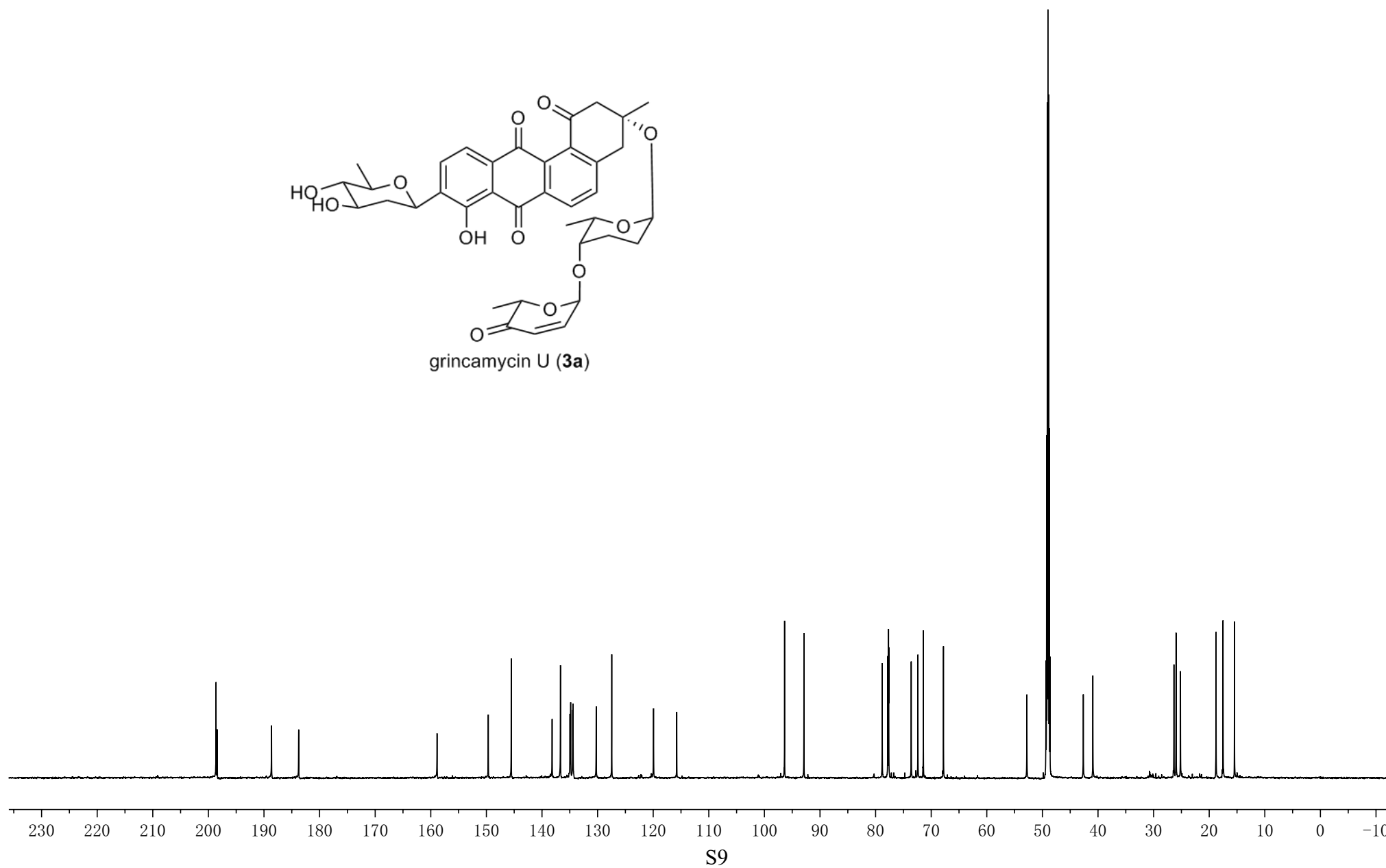


Figure S10. The DEPT-135 NMR (175 MHz) spectrum of grincamycin U (**3a**) in CD₃OD.

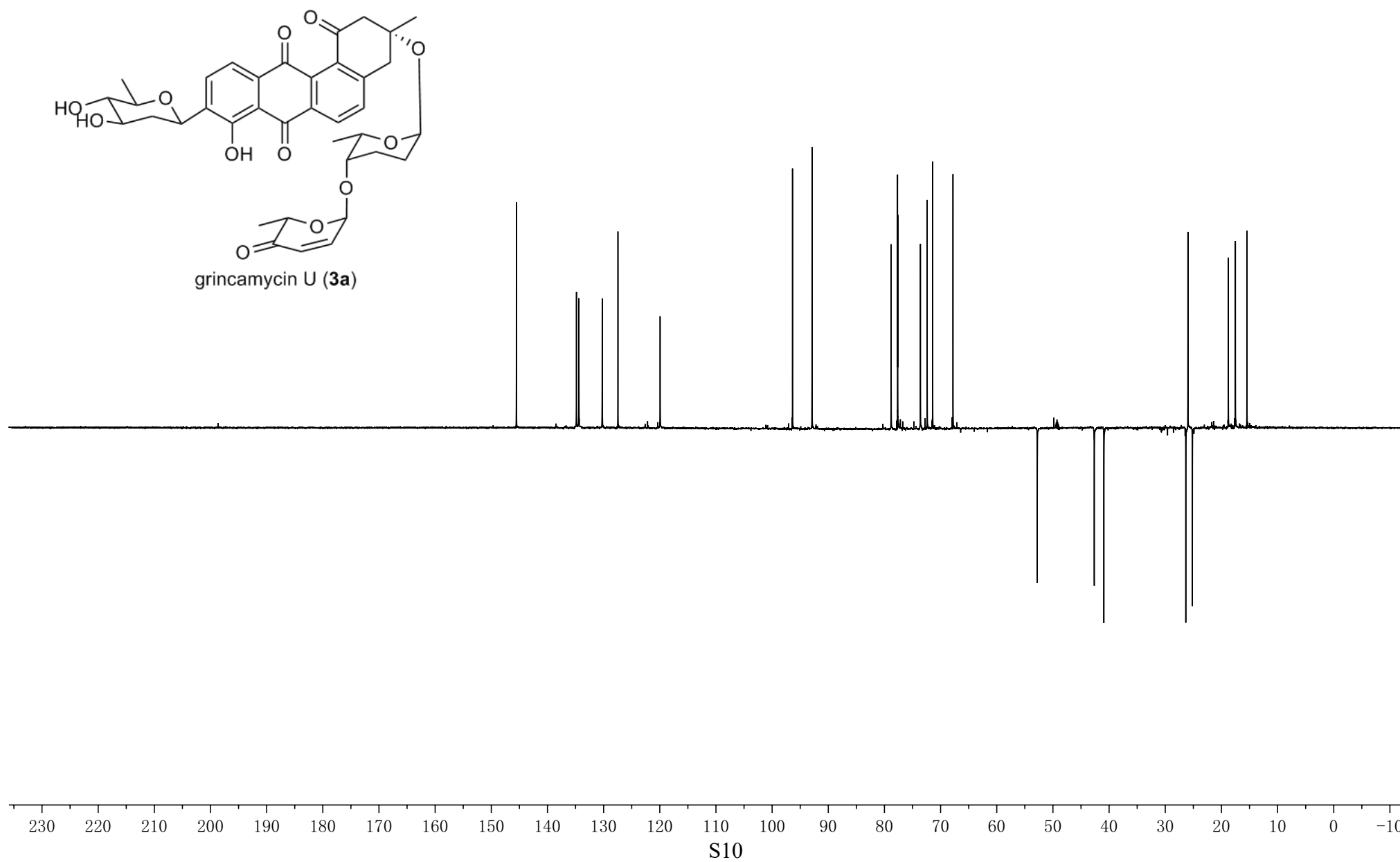


Figure S11. The COSY NMR spectrum of grincamycin U (**3a**) in CD₃OD.

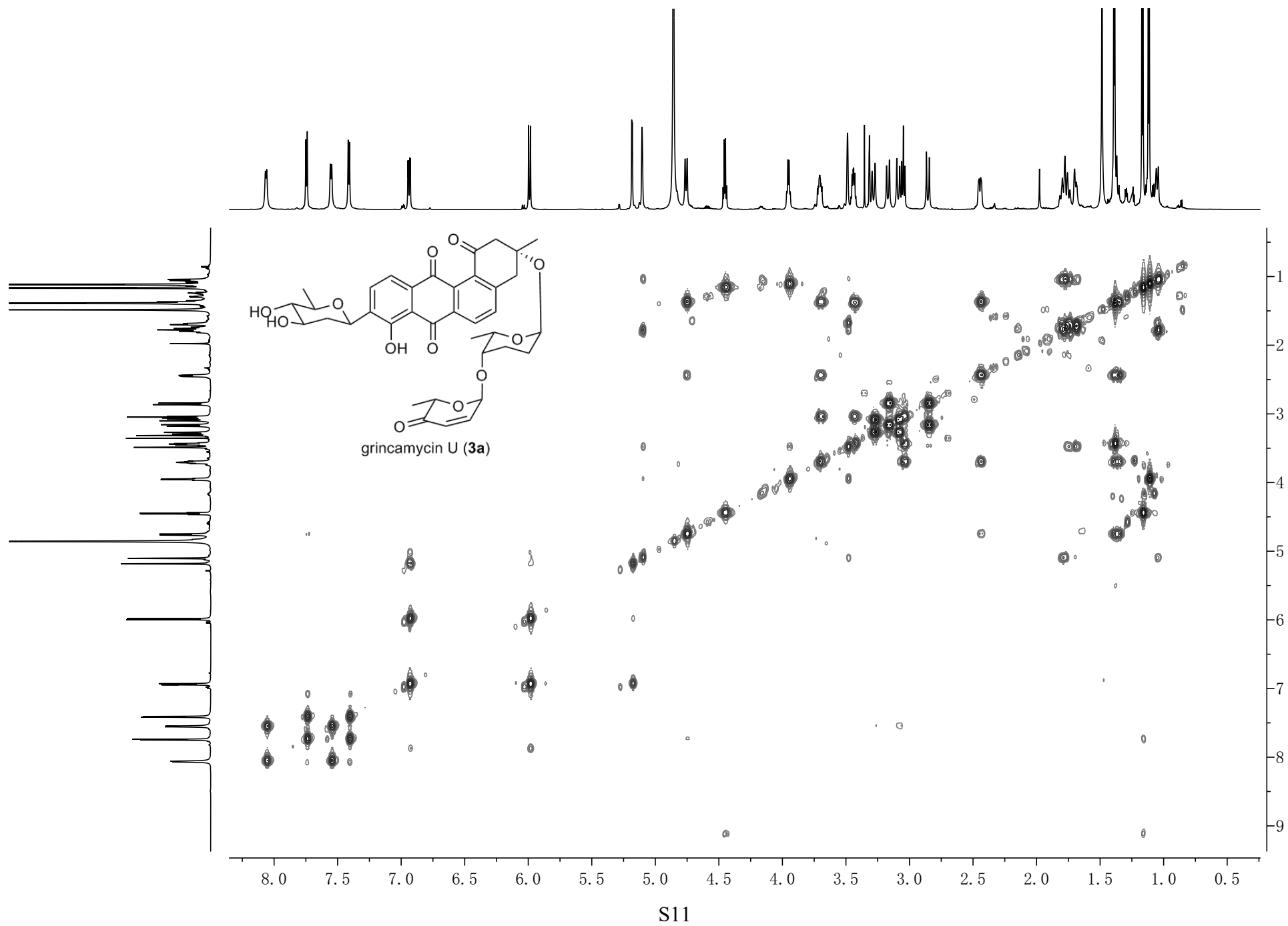


Figure S12. The HMQC NMR spectrum of grincamycin U (**3a**) in CD₃OD.

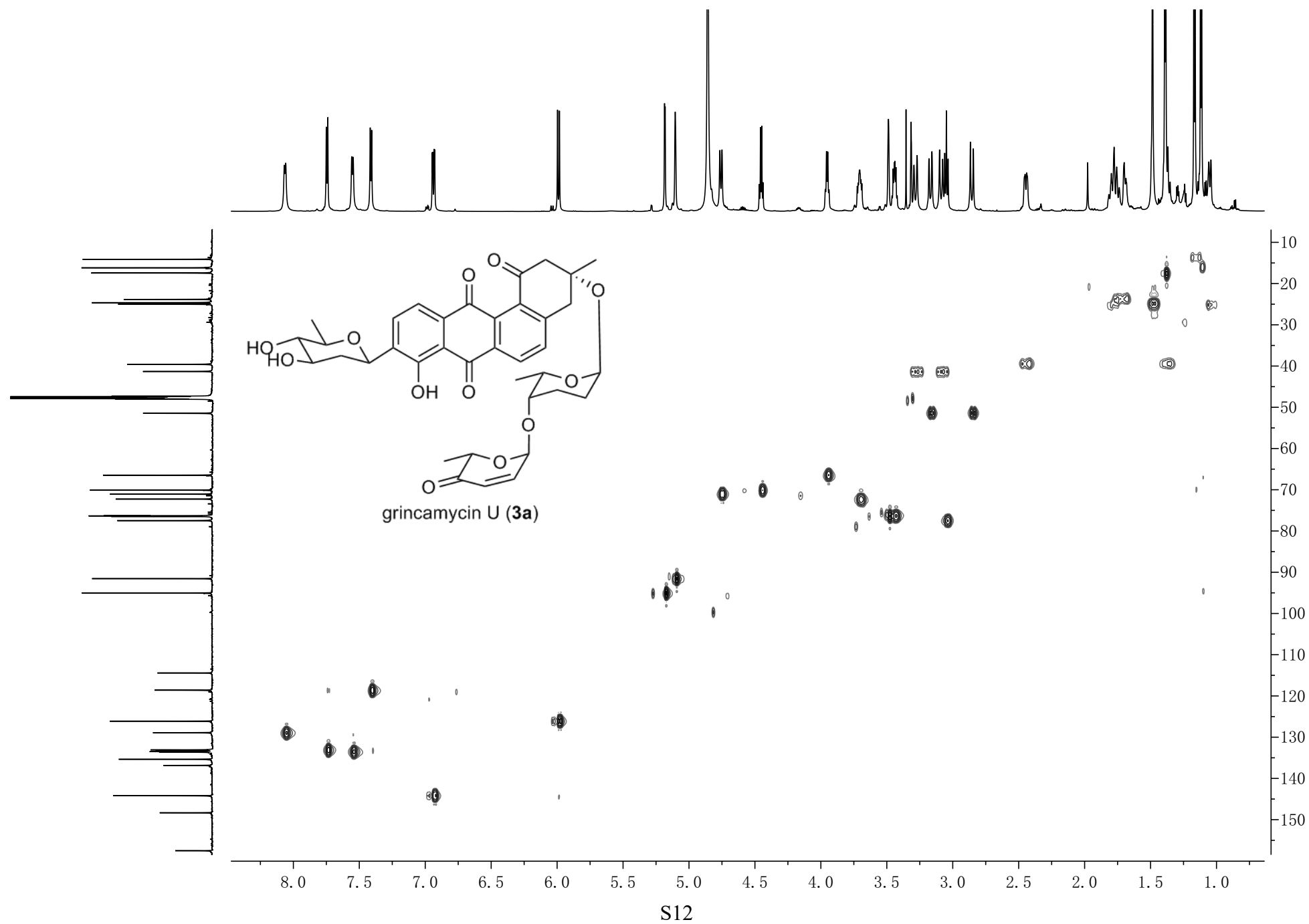


Figure S13. The HMBC NMR spectrum of grincamycin U (**3a**) in CD₃OD.

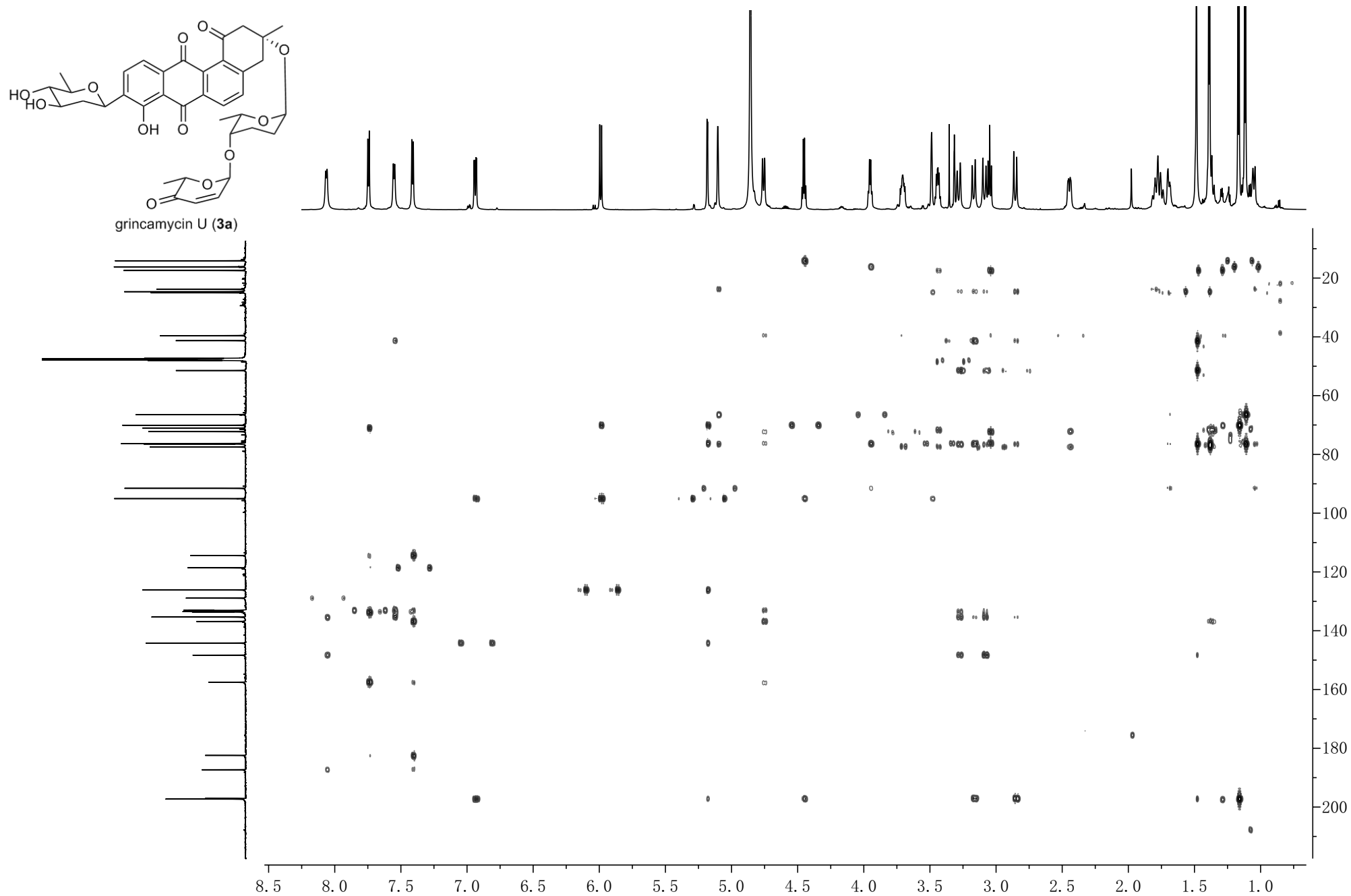


Figure S14. The NOESY NMR spectrum of grincamycin U (**3a**) in CD₃OD.

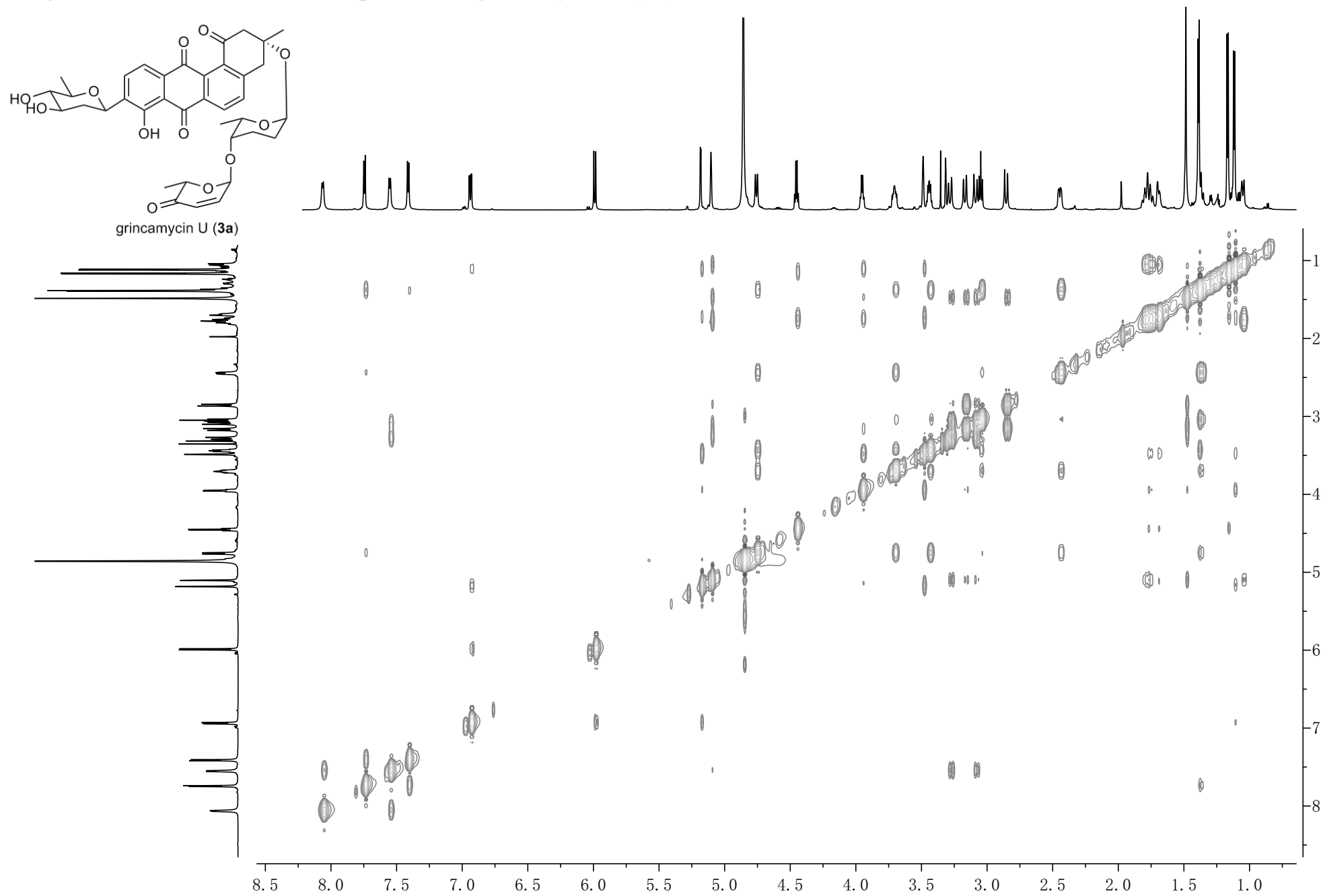


Figure S15. The UV spectrum of grincamycin V (**3'**).

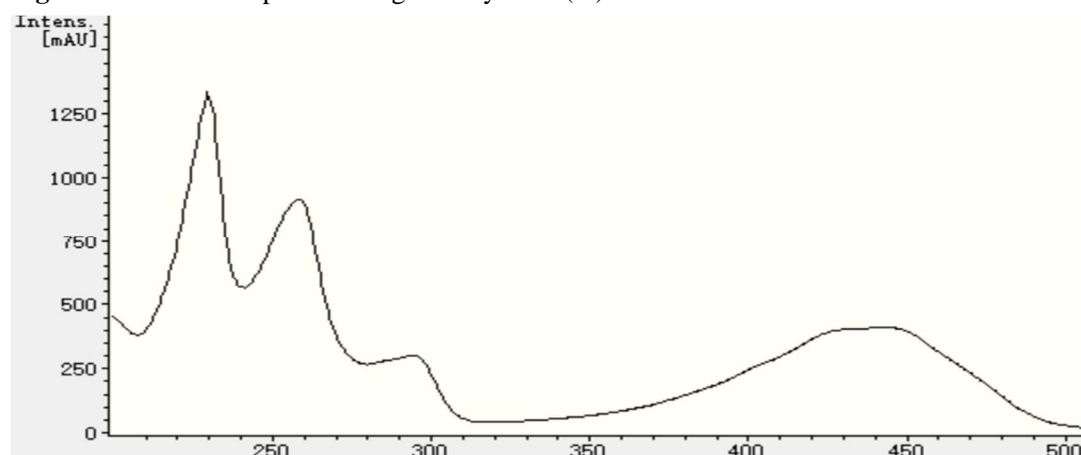


Figure S16. The (-)HR-ESI-MS spectrum of grincamycin V (**3'**).

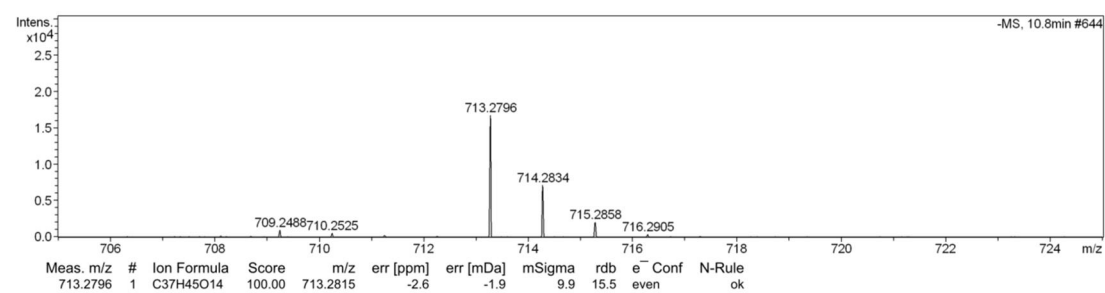


Figure S17. The ^1H NMR (700 MHz) spectrum of grincamycin V (**3'**) in CD_3OD .

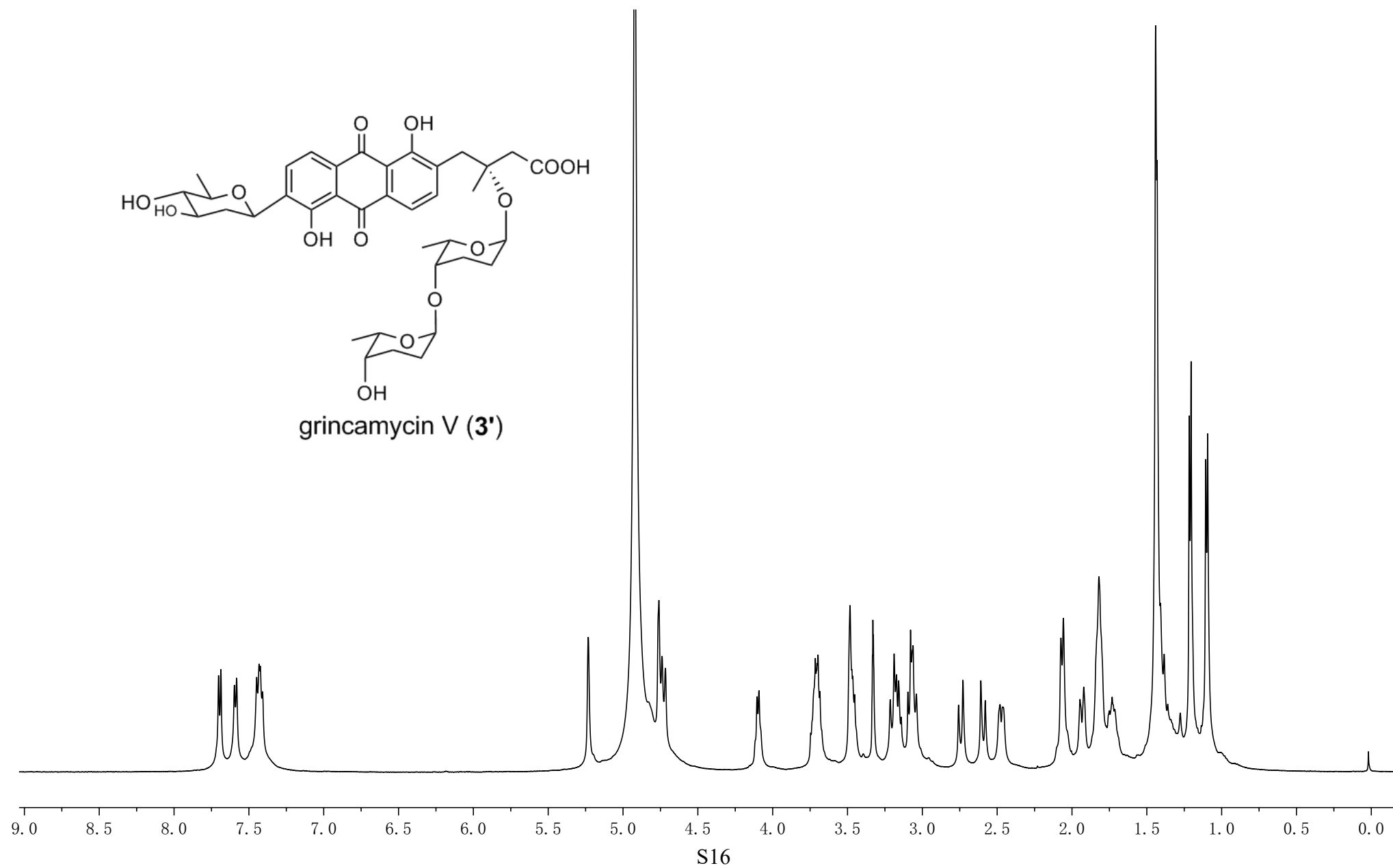


Figure S18. The ^{13}C NMR (175 MHz) spectrum of grincamycin V (**3'**) in CD_3OD .

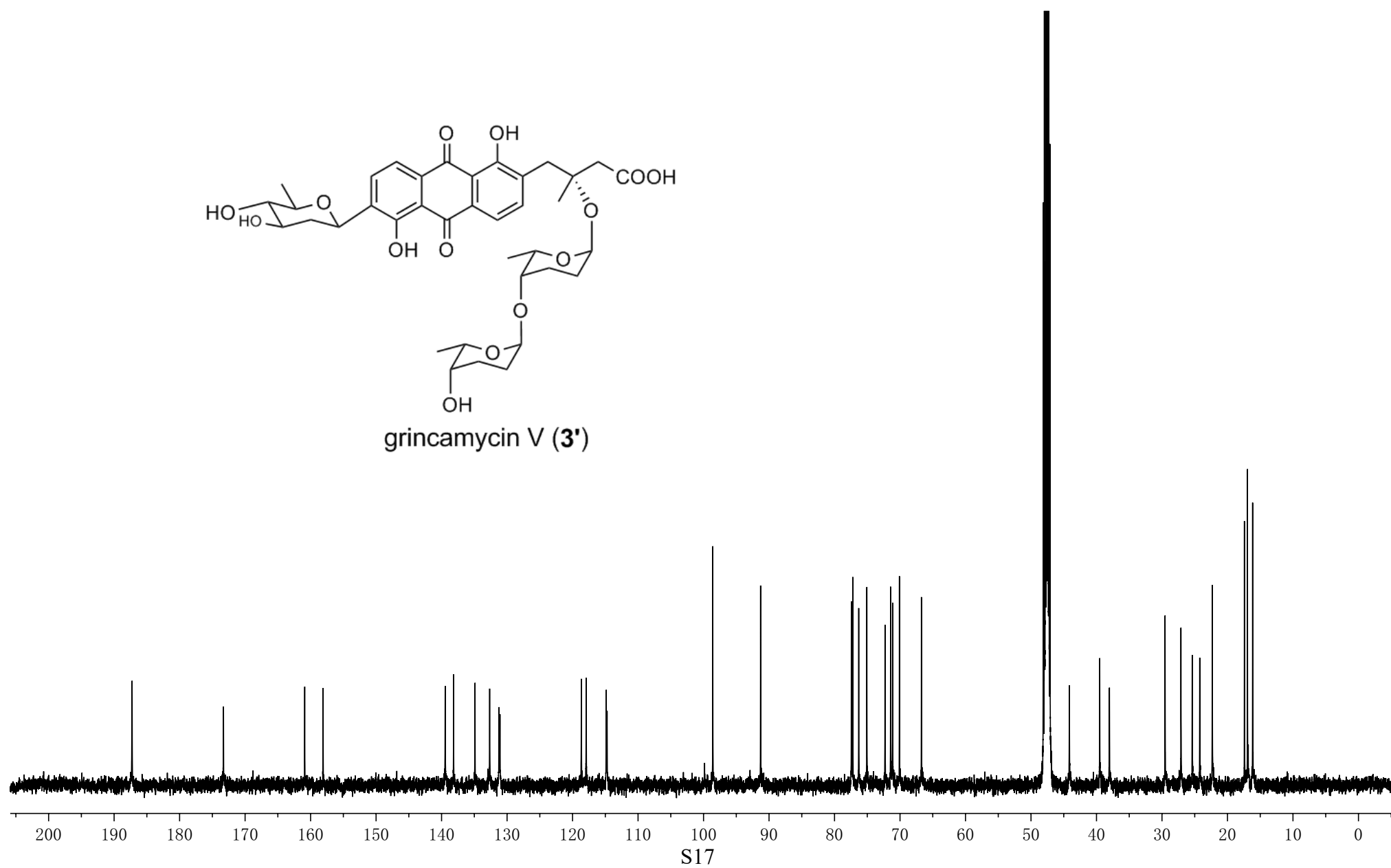


Figure S19. The DEPT-135 NMR spectrum of grincamycin V (**3'**) in CD₃OD.

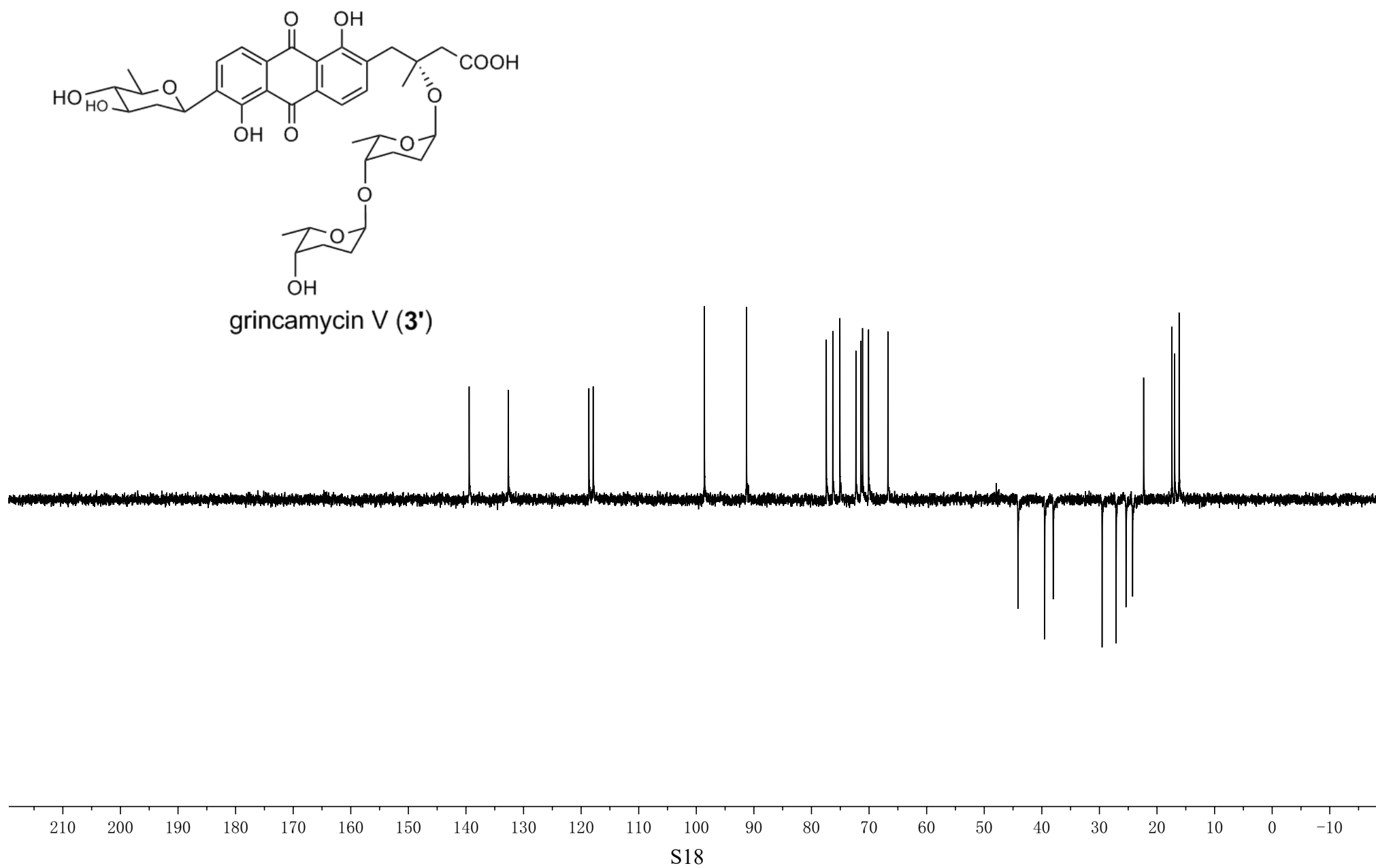


Figure S20. The NOESY NMR spectrum of grincamycin V (3') in CD₃OD.

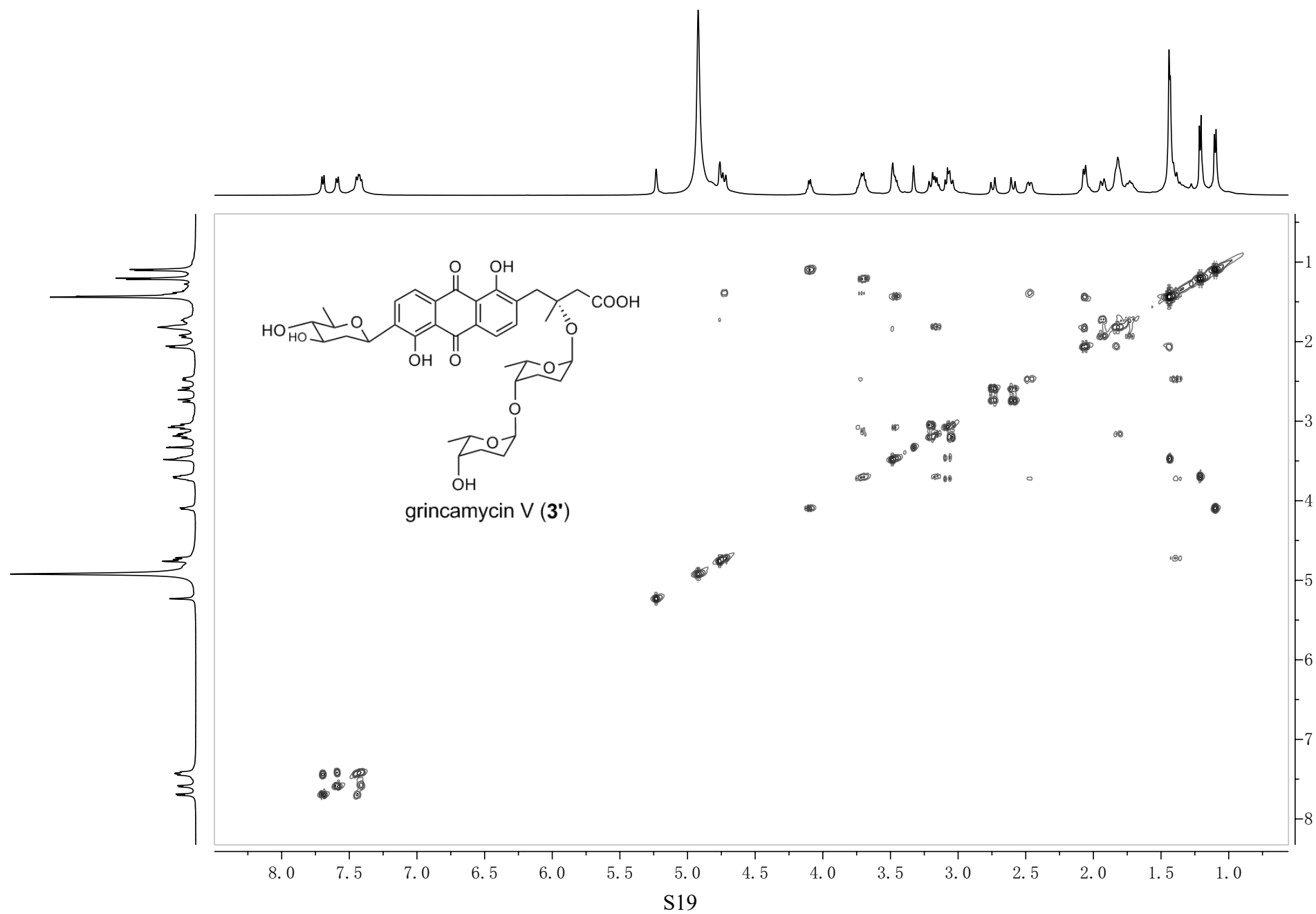


Figure S21. The HMQC NMR spectrum of grincamycin V (**3'**) in CD₃OD.

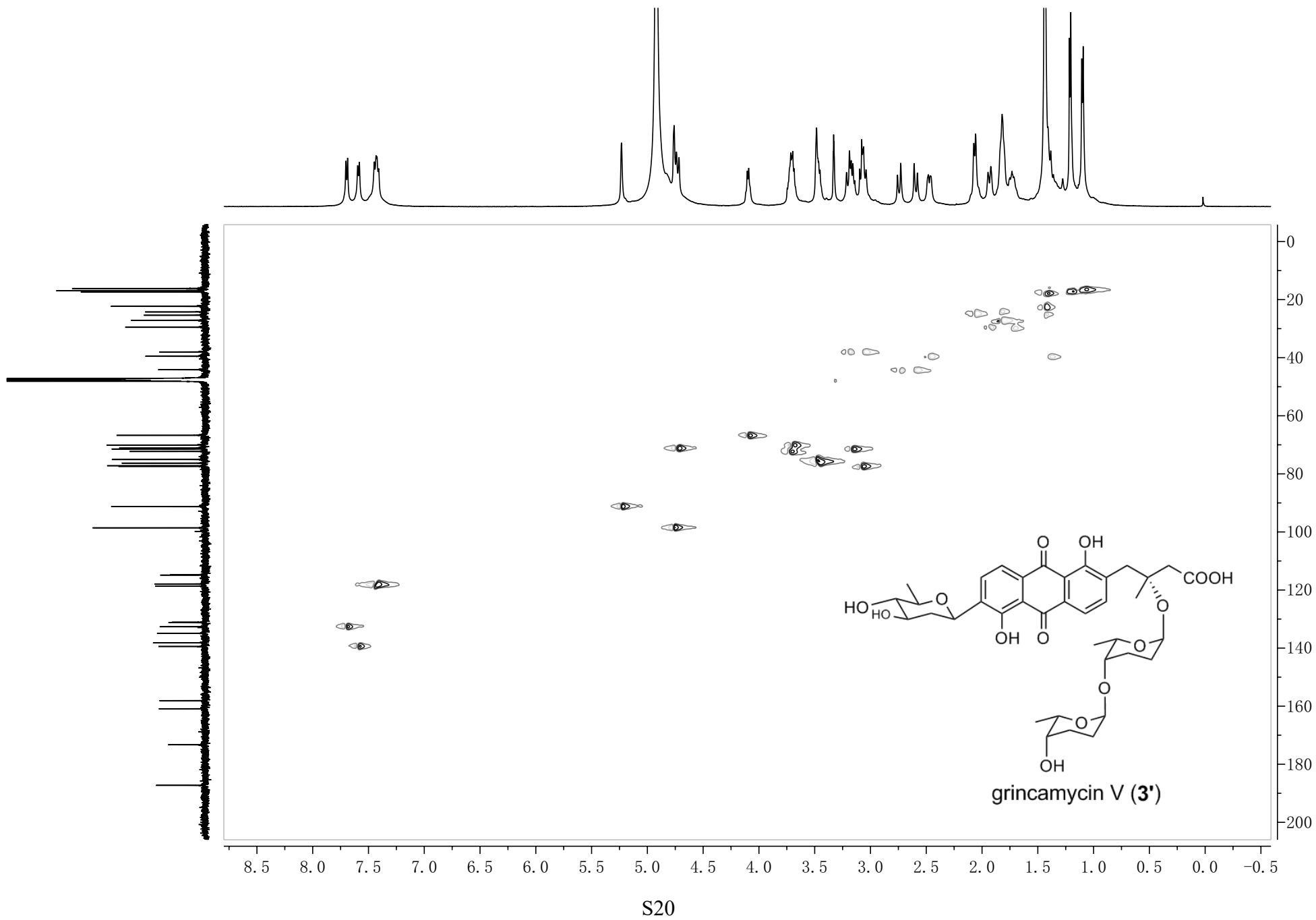


Figure S22. The HMBC NMR spectrum of grincamycin V (**3'**) in CD₃OD.

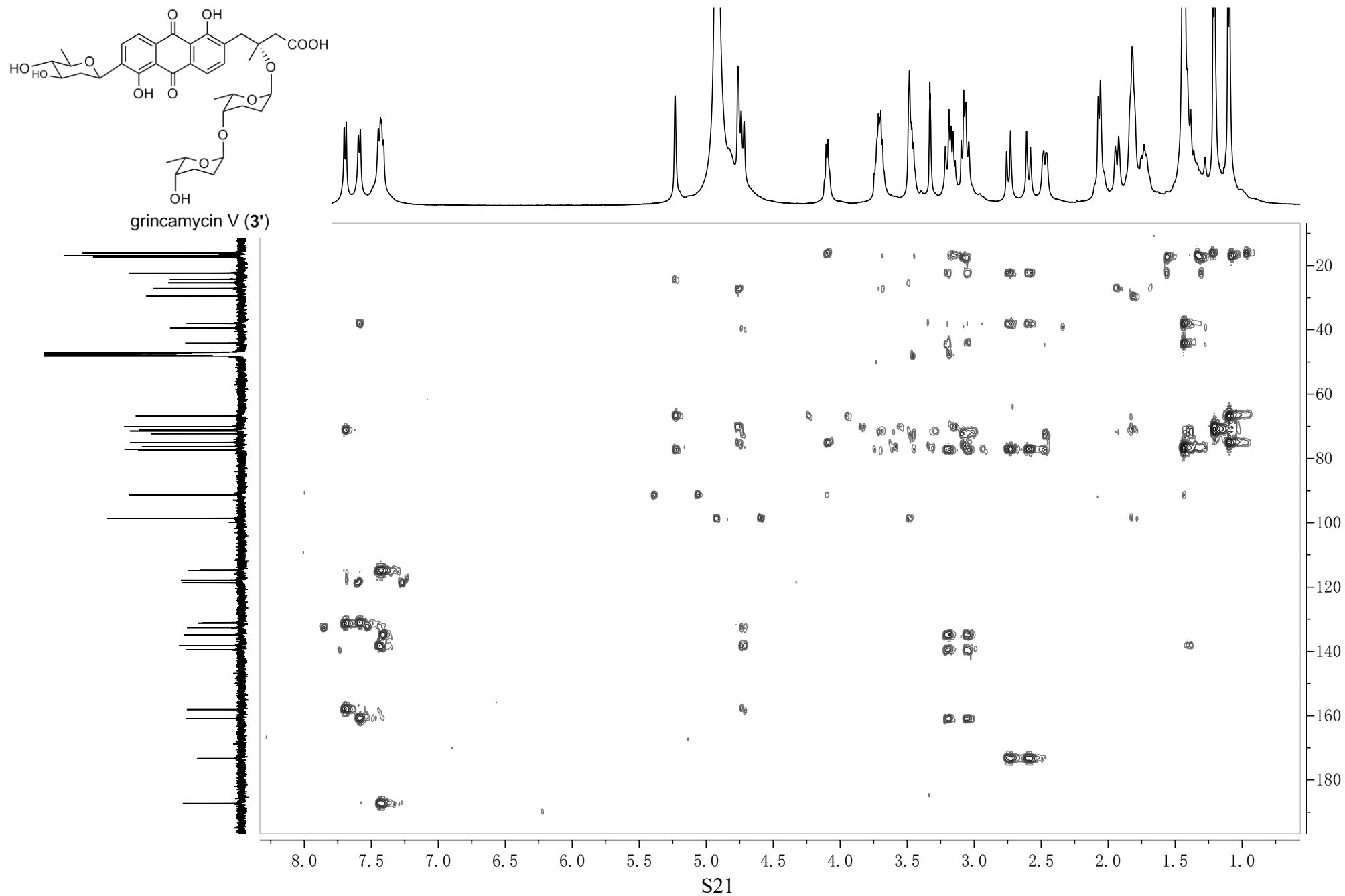


Figure S23. The NOESY NMR spectrum of grincamycin V (**3'**) in CD₃OD.

