





Simple In-House Fabrication of Microwells for Generating Uniform Hepatic Multicellular Cancer Aggregates and Discovering Novel Therapeutics

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Figure S1. Schematic of microwell prototyping. **(A)** Cell culture plates were ablated by CO₂ laser to create microwell structures on the substrate; **(B)** Staggered microwell arrangement; **(C)** Illustration of the top and side views of the concave microwells and recast region due to laser ablation.



Figure S2. Schematic diagram of a 96-well plate prototype with microwells for the comparison with 2D MCAs and drug screening. (**A**) The prototype of size-controlled microwells in a 96-well multi-well plate was generated using CO₂ laser ablation; (**B**) MCAs are formed in four days, at which time, DOX was added at a range of concentrations to each well of the 96-well plate and incubated for 12 h; (**C**) On the fifth day, the supernatant was aspirated and 20 μ L/well of CellTiter-Blue Reagent and medium were added at the appropriate levels such that the final volume of each well was 100 μ L; (**D**) The supernatant with CellTiter-Blue was transferred to an opaque 96-well plate to minimize background fluorescence; (**E**) Fluorescence was recorded by Luminoskan Ascent at excitation/emission wavelengths of 560/590 nm.



Figure S3. Formation of MCAs combined with ConA-conjugated SCHSs for photothermal treatment. (**A**) MCAs are formed in a 12-well plate with prototype microwells; (**B**) Illustration of an MCA formed in a microwell (M_{0@10}); (**C**) A schematic diagram of ConA-conjugated SCHSs is shown; (**D**) ConA-conjugated SCHSs are associated with MCAs and exposed to a near-infrared laser.



Figure S4. Formation of hepatic MCAs in different parametric microwells. Cell cultures are shown 1, 3, and 5 days after cell seeding in microwells fabricated by laser power of 5 W (M_{0@5}) and 10 W (M_{0@1}).





Figure S5. Optimization of the proportion of cells binding with ConA-SCHSs. (**A**) The proportion of the conjugation between ConA and SCHSs was identified and evaluated with LIVE/DEAD cell stain; (**B**) Quantification of the relative fluorescence reflects the ratio of live to dead cells at different proportions.