Supplementary materials



Figure S1. Morphological phenotypes of tobacco overexpressing *LhSLRL*.

- (a) Phenotypes of 5-month-old wild-type and transgenic plants.
- (b) Phenotypes of the 1-month-old wild-type and transgenic plants.
- (c) Measurement of plant height in wild-type and transgenic lines (a). Error bars: \pm SD. *P <
- 0.05; **P < 0.01.
- (d) Number of roots in wild-type and transgenic lines (b). Scale bars: 10 cm (a) and 1 cm (b).



Figure 2. Morphological phenotypes of Arabidopsis overexpressing LhRGA.

Phenotypes of the 60-day-old wild-type and transgenic plants. Scale bar = 1 cm.



Figure S3. Sampling method.

(a) The stems were collected from the top of the 25-year-old Liriodendron hybrids by scaffolding.

(b) Liriodendron hybrids is a shallow-rooted tree species, and its lateral roots are mainly

distributed in the 0-20cm soil layer. The lateral roots of Liriodendron hybrids were used as q-PCR

materials.



Figure S4. PCR testing for Arabidopsis overexpressing LhSLRL.

(a) Perform PCR using scf7 180-F and scf7 180-R. CK+: plasmid DNA, CK-: Wild-

type DNA.

(b) Perform PCR using 35S-F and 35S: LhSLRL-R. CK+: plasmid DNA, CK-: Wild-

type DNA.



Figure S5. PCR testing for Arabidopsis overexpressing LhRGA.

(a) Perform PCR using LOC 32933-F and LOC 32933-R. CK+: plasmid DNA, CK-:

Wild-type DNA.

(b) Perform PCR using 35S-F and 35S: LhRGA-R. CK+: plasmid DNA, CK-: Wild-

type DNA.



Figure S6. RT-PCR analysis of wild types and Arabidopsis/tobacco overexpressing

LhDELLAs.

- (a) RT-PCR analysis of wild types and *Arabidopsis* overexpressing *LhSLRL*.
- (b) RT-PCR analysis of wild types and *Arabidopsis* overexpressing *LhRGA*.
- (c) RT-PCR analysis of wild types and tobacco overexpressing *LhSLRL*.



Figure S7. PCR testing for tobacco overexpressing *LhSLRL*.

(a) Perform PCR using scf7 180-F and scf7 180-R. CK+: plasmid DNA, CK-: Wild-

type DNA.

(b) Perform PCR using 35S-F and 35S: LhSLRL-R. CK+: plasmid DNA, CK-: Wild-

type DNA.



Figure S8. The thermal profile of qRT-PCR

Family	species	DELLA	The serial number	
	Brachypodium distachyon	SLN1	XP_003560731.1	
	Oryza sativa	SLR1	BAE96289.1	
Poaceae		D8	NP_001130629.2	
Barnhart	Zea mays	D9	NP_001296780.1	
	Hordeum vulgare	SLN1	Q8W127.1	
	Triticum aestivum	Rht	AGE81922.1	
		GAI	NP_172945.1	
		RGA	NP_178266.1	
Draggiogogg	Arabidopsis thaliana	RGL1	NP_176809.1	
Drassicaceae		RGL2	NP_186995.1	
Duillett		RGL3	NP_197251.1	
		RGA1	XP_009101333.1	
	Brassica campestris	RGA2	Q5BN22.1	
Constantia		GAIP	Q6EI06.1	
Cucurbitaceae	Cucurbita maxima	GAIP-B	Q6EI05.1	
Vitaceae Juss.	Vitaceae Juss. Vitis vinifera		AEK06229.1	
Malvaceae Iuss	Gossprium hirsutum	SLR1a	AAY28970.1	
Ivialvaceae Juss.	Gossyptum nirsutum	SLR1b	AAO62757.1	
Amborellaceae	Amboralla trichopoda	GAI1	XP_006826893.1	
Pichon	Атоогена тепороца	RHT-1	XP_006829673.3	
Nymphaeaceae	Nymphaea colorata	GAI	XP_031491894.1	
	Nymphaea coloraia	GAI1	XP_031497443.1	
Solanaceae Juss.	Solanum lycopersicum	GAI	NP_001234365.1	
Funariaceae	Dhugoomituella patong	DELLAa	XP_024361039.1	
	r nyscomittetta patens	DELLAb	XP_024403888.1	
Selaginellaceae Willk.	Selaginella kraussiana	DELLA	ABU63412.1	
		SLR1-like	XP_010249953.1	
Malumhanaaaa	N 1	GAI-like	XP_010254273.1	
Nelumbonaceae	Nelumbo nucifera	GAI1	XP_010264458.1	
		GAI1-like	XP_010274729.1	
		DELLA1	PIA63880.1	
Ranunculaceae	Aquilegia coerulea	DELLA2	PIA26570.1	
		DELLA3	PIA65400.1	

Table S1 DELLA proteins of different species

gene	Primer name	Primer sequence (5'-3')
LhSLRL	scf7 180-F	ATCTGCACATCCAAACACACCATTC
	scf7 180-R	ACAGCTTATTCTATTGTATCCCGAG
LhRGA	LOC 32933-F	TACATATATAGGCATGAACTCGTCC
	LOC 32933-R	TCAACCCGAGTCACTCTCTCCCGAA

Table S2 Primers used for gene cloning

Table S3 Primers used for qRT-PCR, RT-PCR, and PCR testing

Primer name		Primer sequence (5'-3')
Lh18SrRNA-F		TGTCGAGCAGGAAGCAGATCACAAC
Lh18SrRNA-R		TTGTTATTTATTGTCACTACCTCCC
	qRT-LhSLRL-F	TGTCGAGCAGGAAGCAGATCACAA
	qRT-LhSLRL-R	GCCCAGGTGGAGTGGCTTGAAG
	qRT-LhRGA-F	TCGACCGGTTCAATGAATCACTGC
	qRT-LhRGA-R	CGTCTCATGTCGTTCCACTCGCTC
	AtACTIN2-F	CAGTGTCTGGATCGGTGGTT
	AtACTIN2-R	TGAACGATTCCTGGACCTGC
	1RT-LhSLRL-F	GAAATCGCATTGGCTGGGTC
	1RT-LhSLRL-R	GCTTGATTGGCGGTGAAGTG
	RT-LhRGA-F	AGAACATCAGGACAGTCGCC
	RT-LhRGA-R	TCCTGACCGTTGTACATGGC
	NtACTIN-F	TGAGCTTCCAGATGGGCAAAT
	NtACTIN-R	AGCCACCACTAAGCACAACG

2RT-LhSLRL-F CCAAGCTCTCGGAGATCGAC

2RT-LhSLRL-R GTTCTGTGGGTCCGAGAAGG

35S-F TGAAGATAGTGGAAAAGGAAGGTG

35S: LhSLRL - R TGAGACGGTGGTGATTGGTG

35S: LhRGA - R GCAATCCGAGCCGTAAAACTGATG

Table S4 Identity matrix for LhSLRL/LhRGA and five DELLA proteins in

Arabidopsis

Amino acid identity	AtGAI	AtRGA	AtRGL1	AtRGL2	AtRGL3
LhSLRL	39.74%	40.07%	37.95%	37.46%	35.83%
LhRGA	55.54%	55.37%	52.12%	51.63%	50.00%

>LhSLRL

ATGGGACCATACGACACGGCCATCTCCCCCACCACCACCAACTCCTCACC ATGCGGCCCATCCAAGCTCTCGGAGATCGACGGCCTCCTCGCAGGGGCGG GCTACCACATCCGGTCGTCGGATCTACGCCACGTGGCTCAGAGGCTGGAG CATCTGGAGTCAGCGATGGTCAACCAAGCTCCCGAGAACATCTCCAACCT AGCCACAGAGGCCGTACACTACAACCCCTCCGACCTCGCGTCCTGGGTCG AGTCCATGCTCTCCGAGTTCCACCAATCACCACCGTCTCAGCTCCTCCCC ACCTTCTCGGACCCACAGAACCCCTCCTCGATGCACGACACGTGGCAACC CGACACCTGCACGGACCCGCTGCAGCAGCAGCAGCTGACGGTCATGGCGG CACTGGAGGAGGAGGACTCCGGCATCCGGCTCGTTCACCTGCTGATGACT TGCGCCGAGTCGATGCAACGTGGCGAAATCGCATTGGCTGGGTCTCTGAT CGACGAGATGCGCCTCCTCCTGACACGCGTCAACACCGGATGCGGCATCG GCAAGGTCGCGAGCTACTTCATAGACGCGCTAAGCCGGCGGCTCTTCTCA CCGGCCCAGGCCGGGCCGGATTGCTCCGGTTCGGCCCTTGAGAACGAGAT CCTCTACCACCACTTCTACGAGGCTTGCCCCTACCTCAAATTCGCCCACT TCACCGCCAATCAAGCCATCCTCGAGGCGTTCCACGGCCACGATTGCGTC CCAGGCCCTCGCGCTCCGCCCGGGCGGCCGCCGTTGCTCCGCCTGACCG GCATCGGTCCACCATCCCCGGACGGCCGCGACTCCCTCCGCGAAATCGGC CTCCGGCTGGCCGAGTTGGCTCGGTCCGGTCCGGCTCCGGTTTACCTTCCG CGGCGTCGCAGCGTCCCGGCTCGACGACGTCAAGCCGGGGATGCTCCAGG TGGGCCCTAGGGAAGCCGTGGCTGTCAATTCGGTAATGCAGCTCCACCGG CTGCTTGGATCGGACCAGAACCGTGGCCCCGCACCGATCGACTCAGTACT

ATGAAAAGAGAACATCAGGACAGTCGCCGCAGCATGGCAACGACTGGGA AGAGCAAGATATTAGAACCTGACGCAGGCGAGGACGAGCTCCTTGCTGTC CTCGGCTACAAGGTCCGCTCCTCTGACATGGCTGACGTGGCTCAGAAGCTC GAGCAGCTGGAGATGGCCATGTACAACGGTCAGGAAGATGGTATTTCCCA TCGAATCCATGCTCTCCGAATTCAATACGCCATCCGAACCGGCCATCTCC AAGCAACAGCAGCCGCATCAGTTTTACGGCTCGGATTGCAGCGACCTGCG GGCCATTCCCGGCGAGGTTGTTTATGGGAGAGGGGAATTAGATTTGGGCG AGCCAAGAGAAGAAGAAACGGATGAGATTGGGGGACTGCAGTGGCGGCCGC AGCTGAGTCGGCTCGCCCAGTCGTCCTGGTCGACTCACAGGAGACCGGGA TCCGACTCGTTCATTCGCTGATTGCCTGTGCCGAGGCCGTCCACCGGGACG ATCTCGAATCGGCAGAGGCACTGGTGAAGCAGATCGGGCTGCTGGCCGCC TCGCAGGGGGGGGGCGATGAGGAAGGTTGCGATGTACTTCGCCGAGGCACT TGCCCGGCGGATTTACAAAATTAAACCACGGGATGCGCTGGATTCGTTCT CTGACATCCTGCAAATGCACTTCTATGAATCCTGCCCATACCTGAAATTC GCCCATTTCACTGCGAATCAGGCCATCCTTGAGGCTTTTGAGGGGGAAGAG TCGCGTCCATGTGATCGATTTCAGCATGAAGCAGGGGATGCAGTGGCCTG CACTGATTCAGGCACTTGCGCTGCGCCCTGGCGGTCCACCAATATTCCGG CTGACGGGCATTGGCCCGCCGCAGCCTGACGATACGGACACCCTCCAGCA TGTGGGAATGAAGCTGGCGCAGCTAGCGGATACTATCAATGTGGAATTCA GTTACAGAGGAATCGTAGCGGACAGTCTTGCTGATGTTGAAGCTTTCATG CTCGATGCGCATTCGGGAGGCGATGATGAGGTTGTGGCAGTGAATTCAGT GTTCGAACTTCATCGTCTGTTGGCCCACCCAGGGGGCAATCGACAGGATGC TGGCGACTGTGAAGGCGGTGCAGCCAAAGATCGTGACGATCGTCGAGCAG GAGGCGGACCACAATGGGCCGGATTTCCTCGACCGGTTCAATGAATCACT GCATTACTACTCAACGATGTTCGATTCGCTTGAAGGATGCGGGATGTCGC TGCCGGACGGCAAGGACCAGCTGATGTCGGAAGTGTATCTCGGGACGCAG GACGCTGGCACGGTGGCAGGGCCGGATGGGCGCGGCCGGGTTCGTGCCGG TGCACCTGGGCTCGAACGCGTTCAAGCAGGCAAGCATGCTGCTGGCACTG TTCGGGGGGCGGTGAAGGTTACAGGGTGGAGGAGAACAATGGGTGTTTGAT GCTGGGCTGGCACACACGGCCTCTCATAGCAACTTCAGCTTGGCAAATCG TCGAACAGGAGAGTGATCACTGA