

## Review

# Potentials of Endophytic Fungi in the Biosynthesis of Versatile Secondary Metabolites and Enzymes

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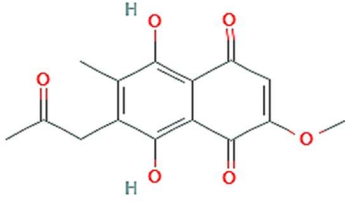
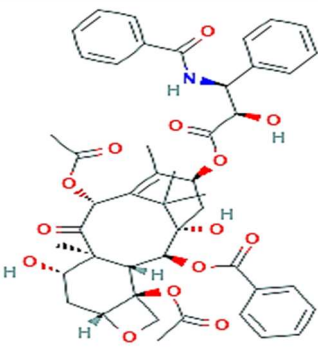
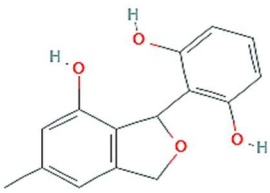
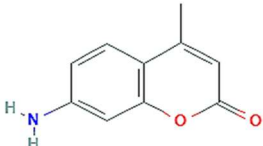
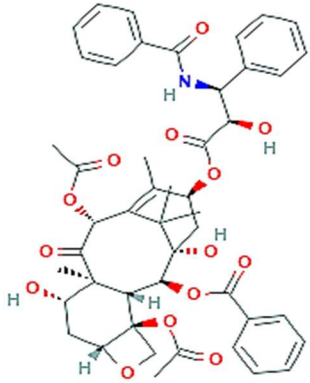
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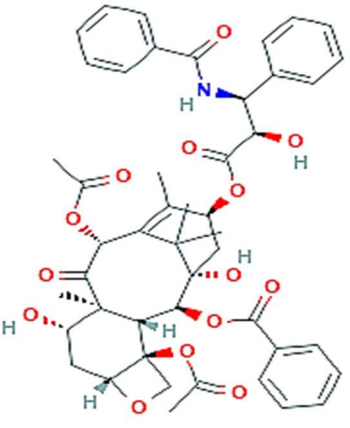

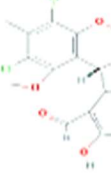
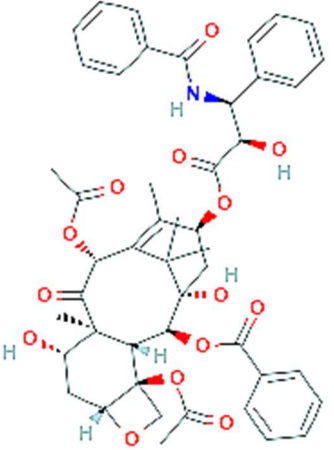
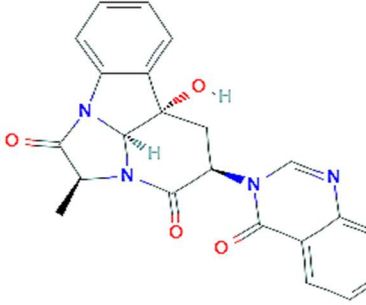
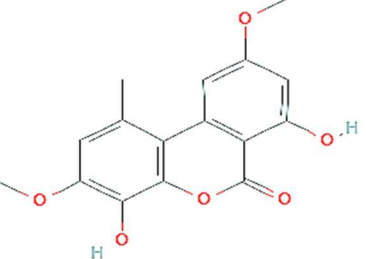
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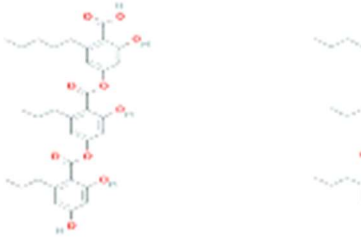
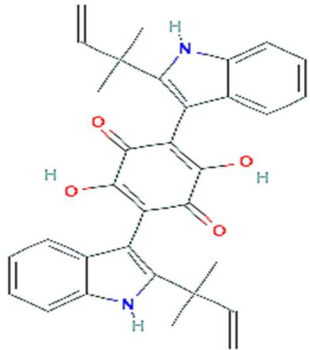
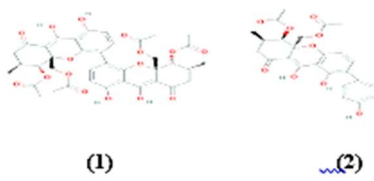
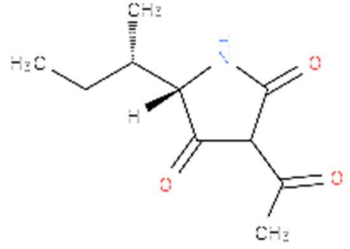
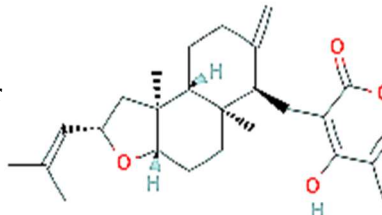
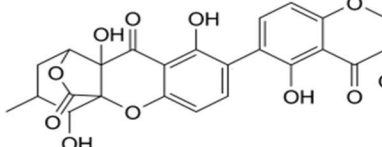


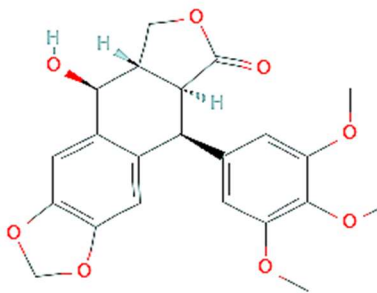
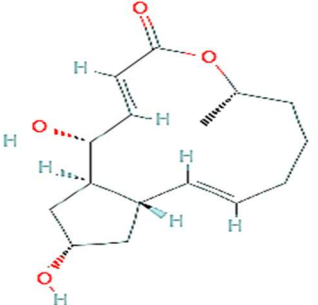
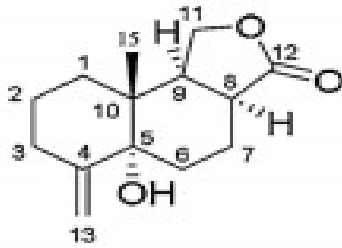
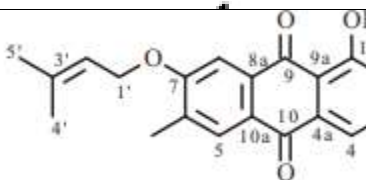
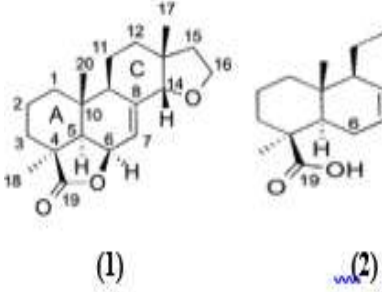
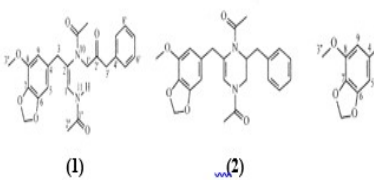
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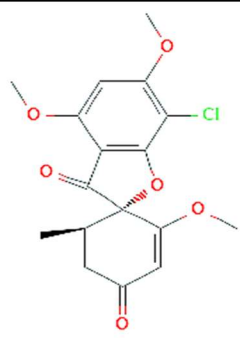
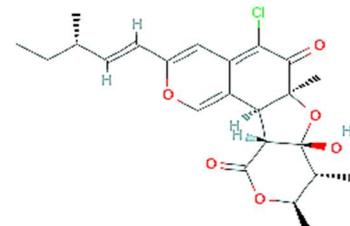
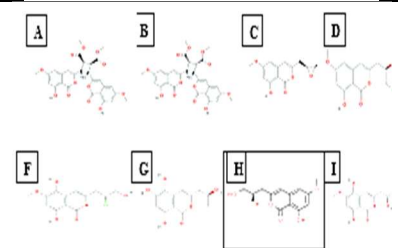
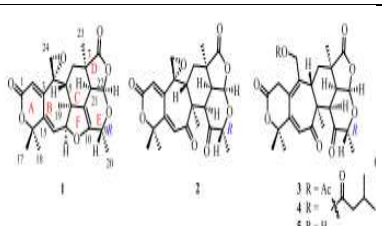
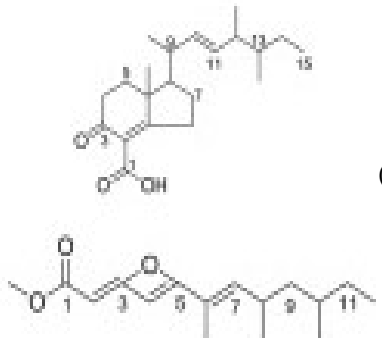
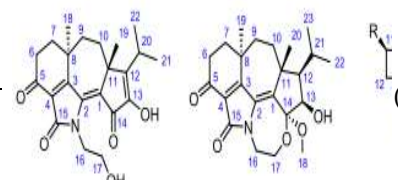
**Table S1.** Medicinal applications of novel secondary metabolites produced by endophytic fungi.


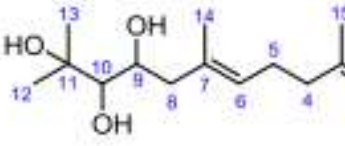
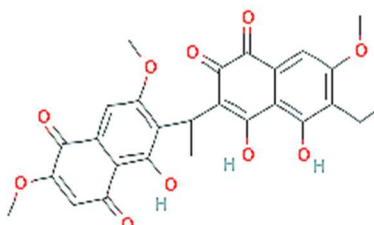
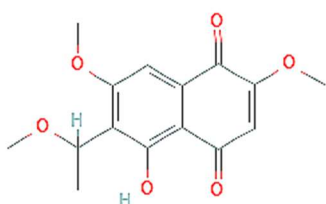
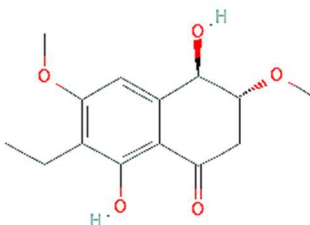
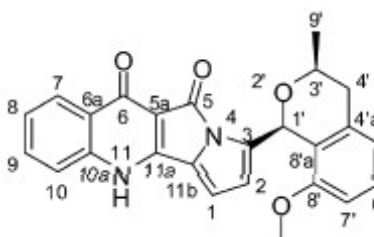
Compounds	Fungal endophytes	Host plants	Biological activities	Structural formulas	References
Javanicin	<i>Chloridium</i> sp.	<i>Azadirachta indica</i>	Antibacterial activity		(Kharwar et al., 2009)
Taxol	<i>Tubercularia</i> sp. TF5	<i>Taxus mairei</i>	Anticancer activity		(Wang et al., 2000)
Pestacin	<i>Pestalotiopsis microspora</i>	-	Antioxidant and antimycotic activities		(Harper et al., 2003)
7-amino-4-methylcoumarin	<i>Xylaria</i> sp.YX-28	<i>Ginkgo biloba</i>	Antimicrobial activity		(Liu et al., 2008)
Camptothecin	-	<i>Camptotheca acuminata</i>	Antineoplastic agent		(Kusari et al., 2009)

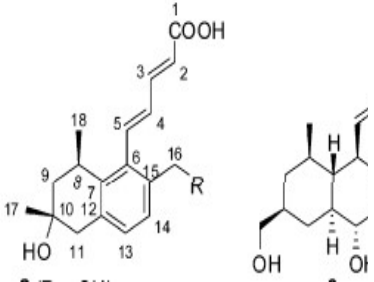
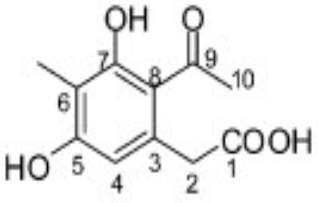
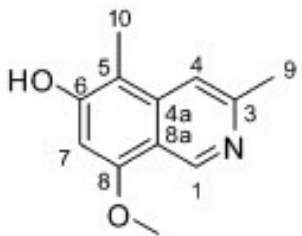
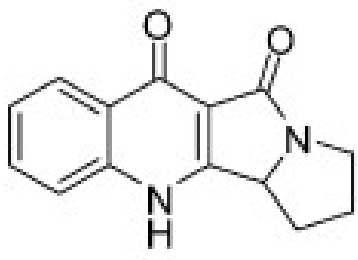
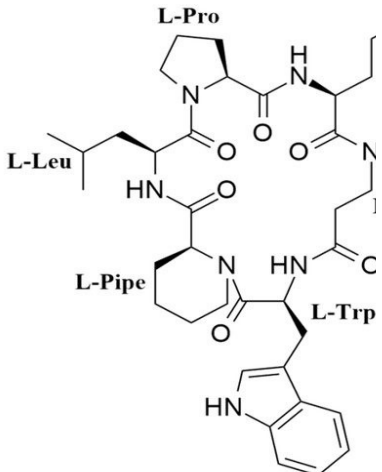
Taxol	<i>Fusarium solani</i> , Tax-3	<i>Taxus chinensis</i>	Anticancerous activity		(Deng et al., 2009)
Pestalachlorides A–C (1-2)	<i>Pestalotiopsis adusta</i>	-	Antifungal activity	  (1) (2)	(Li et al., 2008)
Taxol	<i>Taxomyces andreanae</i>	<i>Taxus brevifolia</i>	Anticancerous activity		(Stierle et al., 1993)
Chaetominine	<i>Chaetomium</i> sp. IFB-E015	<i>Adenophora axilliflora</i>	Cytotoxic activity		(Jiao et al., 2006)
Graphislactone A	<i>Cephalosporium</i> sp. IFB-E001	<i>Trachelospermum jasminoides</i>	Antioxidant activities		(Song et al., 2005)

Cytotoxic Acids A (1) and B (2)	<i>Cytospora</i> sp.	-	Tridepside Inhibitors of hCMV Protease	 <b>(1)</b>	(Guo et al., 2000)
Hinnuliquinone	-	-	Inhibit HIV-1		(Singh et al., 2004)
Phomoxanthones A and B	<i>Phomopsis</i> sp. BCC 1323	-	Antimalarial and antitubercular activities and cytotoxicity	 <b>(1)</b>	(Isaka et al., 2001)
Tenuazonic Acid	<i>Alternaria alternata</i>	-	Antibacterial activity against <i>Mycobacterium tuberculosis</i> H37Rv		(Sonaimuthu et al., 2011)
Subglutinol A	<i>Fusarium subglutinans</i>	<i>Tripterygium wilfordii</i>	Immunosuppressive activity		(Lin et al., 2014)
Ergoflavin	-	<i>Mimosops elengi</i>	Anti-Inflammatory and Anticancer Activity		(Deshmukh et al., 2009)

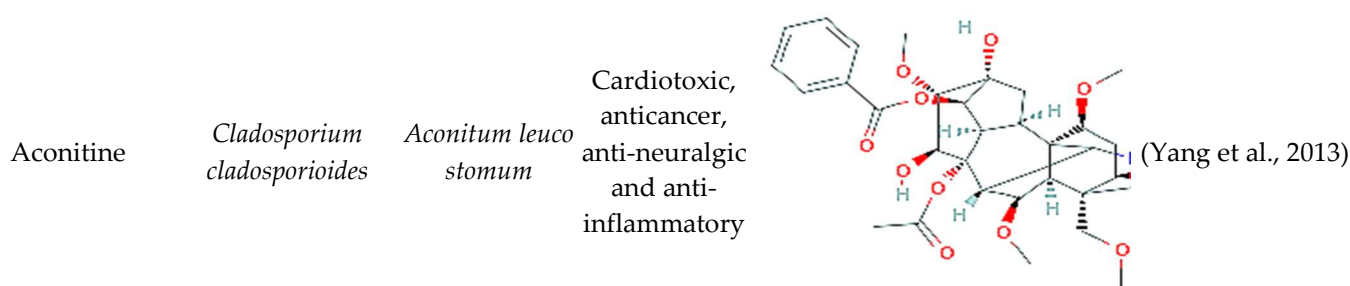
Podophyllotoxin	-	<i>Podophyllum peltatum</i>	Anticancer agent		(Ardalani et al., 2017)
Brefeldin A	<i>Penicillium</i> sp. FKI-7127	-	Antiviral activity		(Raekiansyah et al., 2017)
14-nordrimane	<i>Phoma</i> sp.	<i>Aconitum vilmorinianum</i> .	Antiviral activity		(Liu et al., 2019)
Anthraquinone, 7-( $\gamma,\gamma$ )-dimethylallyloxy macrosporin	<i>Phoma</i> sp. L28.	Mangrove	Antifungal activity		(Huang et al., 2017)
Xylabisboein A and B	<i>Xylaria</i> sp. SNB-GTC2501	<i>Bisboecklera microcephala</i>	Antibacterial activity		(Sorres et al., 2015)
Brasilamides A (1), B (2) and F (3)	<i>Penicillium brasilianum</i>	<i>Melia azedarach</i>	Anticancer activity		(Fill et al., 2010)

Griseofulvin	<i>Xylaria</i> sp.F0010	<i>Abies</i> sp.	Antifungal activity		(PARK et al., 2005)
Chaetomugilin D	<i>Chaetomium globosum</i>	<i>Ginkgo biloba</i>	Antimicrobial activity		(Qin et al., 2009)
Peniisocoumarins A–J	<i>Penicillium commune</i> QQF-3	<i>Kandelia candel</i>	Cytotoxicity and enzyme inhibitory activities		(Cai et al., 2018)
Amestolkolides A–D	<i>Talaromyces amestolkiae</i> YX1	Mangrove	Anti-inflammatory		(Chen et al., 2018)
Fusariumins C and D	<i>Fusarium oxysporum</i> ZZP-R1	<i>Rumex madaio</i>	Antimicrobial activities		(Chen et al., 2019a)
Koninginols A–C	<i>Trichoderma koningiopsis</i> A729	<i>Morinda officinalis</i>	Antitumor activity		(Chen et al., 2019b)

11-hydroxy-15-drimeneoic acid	<i>Trichoderma koningiopsis</i> A729	<i>Morinda officinalis</i>	Antitumor activity		(Chen et al., 2019b)
Koninginol D	<i>Trichoderma koningiopsis</i> A729	<i>Morinda officinalis</i>	Antitumor activity		(Chen et al., 2019b)
Neofusnaphthoquinone A	<i>Neofusicoccum austral</i> SYSU-SKS024	Mangrove	Indoleamine 2, 3-dioxygenase (IDO) inhibitory activity		(Cui et al., 2018)
6-(1-methoxyethyl)-2,7-dimethoxyjuglone	<i>Neofusicoccum austral</i> SYSU-SKS024	Mangrove	Indoleamine 2, 3-dioxygenase (IDO) inhibitory activity		(Cui et al., 2018)
(3R,4R)-3-methoxybotryosphaerone D	<i>Neofusicoccum austral</i> SYSU-SKS024	Mangrove	Indoleamine 2, 3-dioxygenase (IDO) inhibitory activity		(Cui et al., 2018)
Citriquinochroman	<i>Penicillium citrinum</i>	<i>Ceratonia siliqua</i>	Cytotoxic activity		(El-Neketi et al., 2013)

Tanzawaic acids G and H	<i>Penicillium citrinum</i>	<i>Ceratonia siliqua</i>	-		(El-Neketi et al., 2013)
6-methylcurvulinic acid	<i>Penicillium citrinum</i>	<i>Ceratonia siliqua</i>	-		(El-Neketi et al., 2013)
8-methoxy-3,5-dimethylisoquinolin-6-ol	<i>Penicillium citrinum</i>	<i>Ceratonia siliqua</i>	-		(El-Neketi et al., 2013)
1,2,3,11b-tetrahydroquinolactide	<i>Penicillium citrinum</i>	<i>Ceratonia siliqua</i>	-		(El-Neketi et al., 2013)
Pipicolisporin	<i>Nigrospora oryzae</i>	<i>Triticum aestivum</i>	Antimalarial and Antitrypanosome activities		(Fernández-Pastor et al., 2021)





Ardalani, H., Avan, A., and Ghayour-Mobarhan, M. (2017). Podophyllotoxin: a novel potential natural anticancer agent. *Avicenna J. Phytomedicine* 7, 285.

Cai, R., Wu, Y., Chen, S., Cui, H., Liu, Z., Li, C., et al. (2018). Peniisocoumarins A–J: Isocoumarins from *Penicillium commune* QQF-3, an Endophytic Fungus of the Mangrove Plant *Kandelia candel*. *J. Nat. Prod.* 81, 1376–1383. doi:10.1021/acs.jnatprod.7b01018.

Chen, J., Bai, X., Hua, Y., Zhang, H., and Wang, H. (2019a). Fusariumins C and D, two novel antimicrobial agents from *Fusarium oxysporum* ZZP-R1 symbiotic on *Rumex madaio* Makino. *Fitoterapia* 134, 1–4. doi:10.1016/j.fitote.2019.01.016.

Chen, S., Ding, M., Liu, W., Huang, X., Liu, Z., Lu, Y., et al. (2018). Anti-inflammatory meroterpenoids from the mangrove endophytic fungus *Talaromyces amestolkiae* YX1. *Phytochemistry* 146, 8–15.

Chen, S., Li, H., Chen, Y., Li, S., Xu, J., Guo, H., et al. (2019b). Three new diterpenes and two new sesquiterpenoids from the endophytic fungus *Trichoderma koningiopsis* A729. *Bioorganic Chem.* 86, 368–374. doi:10.1016/j.bioorg.2019.02.005.

Cui, H., Zhang, H., Liu, Y., Gu, Q., Xu, J., Huang, X., et al. (2018). Ethylnaphthoquinone derivatives as inhibitors of indoleamine-2, 3-dioxygenase from the mangrove endophytic fungus *Neofusicoccum austral* SYSU-SKS024. *Fitoterapia* 125, 281–285.

Deng, B. W., Liu, K. H., Chen, W. Q., Ding, X. W., and Xie, X. C. (2009). *Fusarium solani*, Tax-3, a new endophytic taxol-producing fungus from *Taxus chinensis*. *World J. Microbiol. Biotechnol.* 25, 139–143.

Deshmukh, S. K., Mishra, P. D., Kulkarni-Almeida, A., Verekar, S., Sahoo, M. R., Periyasamy, G., et al. (2009). Anti-inflammatory and anticancer activity of ergoflavin isolated from an endophytic fungus. *Chem. Biodivers.* 6, 784–789.

El-Neketi, M., Ebrahim, W., Lin, W., Gedara, S., Badria, F., Saad, H.-E. A., et al. (2013). Alkaloids and polyketides from *Penicillium citrinum*, an endophyte isolated from the Moroccan plant *Ceratonia siliqua*. *J. Nat. Prod.* 76, 1099–1104.

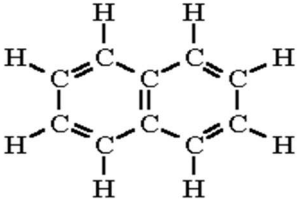
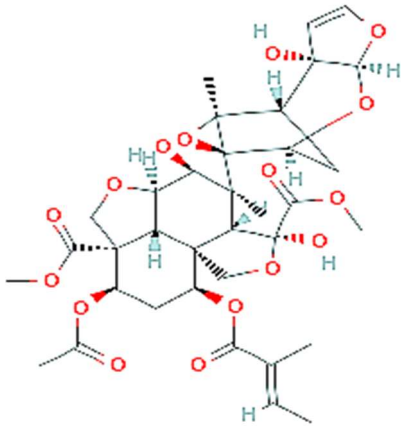
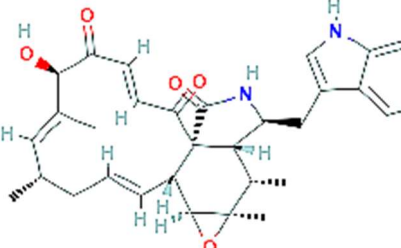
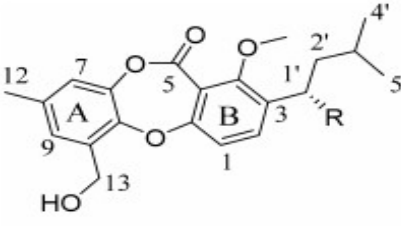
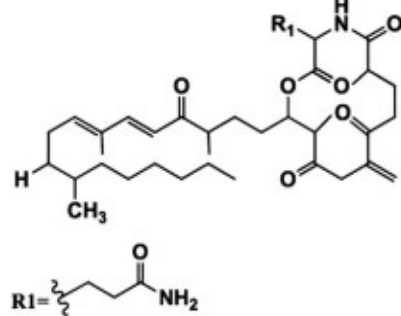
Fernández-Pastor, I., González-Menéndez, V., Annang, F., Toro, C., Mackenzie, T. A., Bosch-Navarrete, C., et al. (2021). Pipecolisporin, a Novel Cyclic Peptide with Antimalarial and Antitrypanosome Activities from a Wheat Endophytic *Nigrospora oryzae*. *Pharmaceutics* 14, 268.

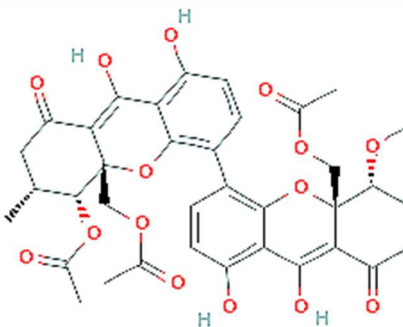
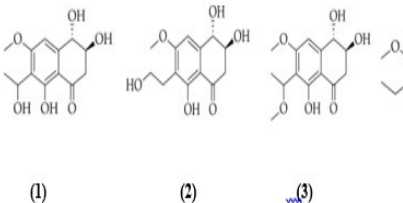
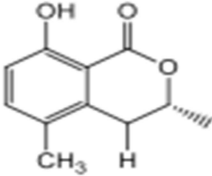
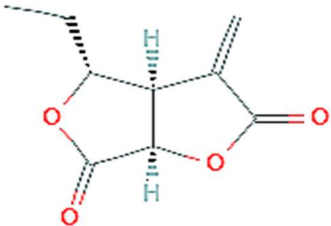
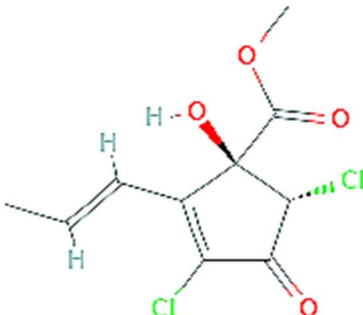
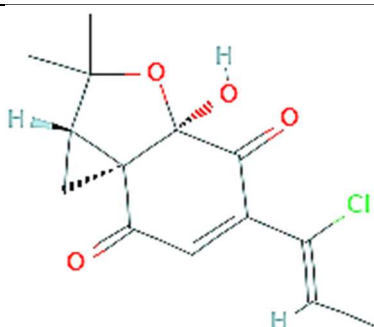
Fill, T. P., Silva, B. F. D., and Rodrigues-Fo, E. (2010). Biosynthesis of phenylpropanoid amides by an endophytic *penicillium brasilianum* found in root bark of *Melia azedarach*. *J. Microbiol. Biotechnol.* 20, 622–629.

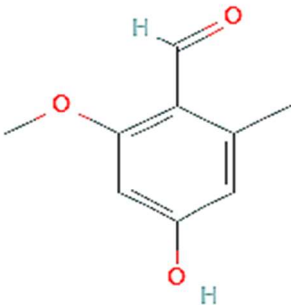
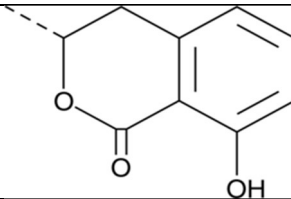
- Guo, B., Dai, J.-R., Ng, S., Huang, Y., Leong, C., Ong, W., et al. (2000). Cytonic Acids A and B: Novel Tridepside Inhibitors of hCMV Protease from the Endophytic Fungus *Cytonaema* Species. *J. Nat. Prod.* 63, 602–604. doi:10.1021/np990467r.
- Harper, J. K., Arif, A. M., Ford, E. J., Strobel, G. A., Porco Jr, J. A., Tomer, D. P., et al. (2003). Pestacin: a 1, 3-dihydro isobenzofuran from *Pestalotiopsis microspora* possessing antioxidant and antimycotic activities. *Tetrahedron* 59, 2471–2476.
- Huang, S., Xu, J., Li, F., Zhou, D., Xu, L., and Li, C. (2017). Identification and Antifungal Activity of Metabolites from the Mangrove Fungus *Phoma* sp. L28. *Chem. Nat. Compd.* 53, 237–240. doi:10.1007/s10600-017-1961-z.
- Isaka, M., Jaturapat, A., Rukseree, K., Danwisetkanjana, K., Tanticharoen, M., and Thebtaranonth, Y. (2001). Phomoxanthonones A and B, novel xanthone dimers from the endophytic fungus *Phomopsis* species. *J. Nat. Prod.* 64, 1015–1018.
- Jiao, R. H., Xu, S., Liu, J. Y., Ge, H. M., Ding, H., Xu, C., et al. (2006). Chaetominine, a cytotoxic alkaloid produced by endophytic *Chaetomium* sp. IFB-E015. *Org. Lett.* 8, 5709–5712.
- Kharwar, R. N., Verma, V. C., Kumar, A., Gond, S. K., Harper, J. K., Hess, W. M., et al. (2009). Javanicin, an antibacterial naphthaquinone from an endophytic fungus of neem, *Cloridium* sp. *Curr. Microbiol.* 58, 233–238.
- Kusari, S., Zühlke, S., and Spiteller, M. (2009). An endophytic fungus from *Camptotheca acuminata* that produces camptothecin and analogues. *J. Nat. Prod.* 72, 2–7.
- Li, E., Jiang, L., Guo, L., Zhang, H., and Che, Y. (2008). Pestalachlorides A–C, antifungal metabolites from the plant endophytic fungus *Pestalotiopsis adusta*. *Bioorganic Med. Chem.* 16, 7894–7899.
- Lin, R., Kim, H., Hong, J., and Li, Q.-J. (2014). Biological Evaluation of Subglutinin A As a Novel Immunosuppressive Agent for Inflammation Intervention. *ACS Med. Chem. Lett.* 5, 485–490. doi:10.1021/ml4004809.
- Liu, S.-S., Jiang, J.-X., Huang, R., Wang, Y.-T., Jiang, B.-G., Zheng, K.-X., et al. (2019). A new antiviral 14-nordrimane sesquiterpenoid from an endophytic fungus *Phoma* sp. *Phytochem. Lett.* 29, 75–78.
- Liu, X., Dong, M., Chen, X., Jiang, M., Lv, X., and Zhou, J. (2008). Antimicrobial activity of an endophytic *Xylaria* sp. YX-28 and identification of its antimicrobial compound 7-amino-4-methylcoumarin. *Appl. Microbiol. Biotechnol.* 78, 241–247.
- PARK, J.-H., CHOI, G.-J., LEE, S.-W., LEE, H.-B., KIM, K.-M., JUNG, H.-S., et al. (2005). Griseofulvin from *Xylaria* sp. strain F0010, an endophytic fungus of *Abies holophylla* and its antifungal activity against plant pathogenic fungi. *J. Microbiol. Biotechnol.* 15, 112–117.
- Qin, J.-C., Zhang, Y.-M., Gao, J.-M., Bai, M.-S., Yang, S.-X., Laatsch, H., et al. (2009). Bioactive metabolites produced by *Chaetomium globosum*, an endophytic fungus isolated from *Ginkgo biloba*. *Bioorganic Med. Chem. Lett.* 19, 1572–1574. doi:10.1016/j.bmcl.2009.02.025.
- Raekiansyah, M., Mori, M., Nonaka, K., Agoh, M., Shiomi, K., Matsumoto, A., et al. (2017). Identification of novel antiviral of fungus-derived brefeldin A against dengue viruses. *Trop. Med. Health.* 45, 32. doi:10.1186/s41182-017-0072-7.
- Singh, S. B., Ondeyka, J. G., Tsipouras, N., Ruby, C., Sardana, V., Schulman, M., et al. (2004). Hinnuliquinone, a C2-symmetric dimeric non-peptide fungal metabolite inhibitor of HIV-1 protease. *Biochem. Biophys. Res. Commun.* 324, 108–113. doi:10.1016/j.bbrc.2004.08.234.

- Sonaimuthu, V., Parihar, S., Prakash Thakur, J., Luqman, S., Saikia, D., Chanotiya, C. S., et al. (2011). Tenuazonic acid: a promising antitubercular principle from *Alternaria alternate*. *Microbiol. Res.* 2, 63–65.
- Song, Y. C., Huang, W. Y., Sun, C., Wang, F. W., and Tan, R. X. (2005). Characterization of Graphislactone A as the Antioxidant and Free Radical-Scavenging Substance from the Culture of *Cephalosporium* sp. IFB-E001, an Endophytic Fungus in *Trachelospermum jasminoides*. *Biol. Pharmaceut. Bull.* 28, 506–509. doi:10.1248/bpb.28.506.
- Sorres, J., Nirma, C., Touré, S., Eparvier, V., and Stien, D. (2015). Two new isopimarane diterpenoids from the endophytic fungus *Xylaria* sp. SNB-GTC2501. *Tetrahed. Lett.* 56, 4596–4598. doi:10.1016/j.tetlet.2015.06.022.
- Stierle, A., Strobel, G., and Stierle, D. (1993). Taxol and taxane production by *Taxomyces andreanae*, an endophytic fungus of Pacific yew. *Science* 260, 214–216.
- Wang, J., Li, G., Lu, H., Zheng, Z., Huang, Y., and Su, W. (2000). Taxol from *Tubercularia* sp. strain TF5, an endophytic fungus of *Taxus mairei*. *FEMS Microbiol. Lett.* 193, 249–253.
- Yang, K., Liang, J., Li, Q., Kong, X., Chen, R., and Jin, Y. (2013). *Cladosporium cladosporioides* XJ-AC03, an aconitine-producing endophytic fungus isolated from *Aconitum leucostomum*. *World J. Microbiol. Biotechnol.* 29, 933–938.

**Table S2.** Agricultural applications of novel secondary metabolites produced by endophytic fungi.

Compounds	Fungal endophytes	Host plants	Biological activities	Structural formulas	References
Naphtalene	<i>Muscodor vitigenus</i>	<i>Paullinia paullinioides</i>	Insect repellent activity		(Daisy et al., 2002)
Azadirachtin	<i>Azadirachta indica</i>	-	Insecticidal activity		(Verma et al., 2014)
Chaetoglobosin A	<i>Chaetomium</i> spp.	-	Antifungal activity		(Ortega et al., 2020)
Talaromyones A (1) and B (2)	<i>Talaromyces stipitatus</i> SK-4	<i>Acanthus ilicifolius</i>	Antibacterial activity	 <p>1: R = OH 2: R = OAc</p>	(Cai et al., 2017)
Fusaristin A	<i>Phomopsis</i> sp.	-	Antimicrobial activity	 <p>R1 = <math>\text{CH}_2\text{CH}_2\text{CH}_2\text{CONH}_2</math></p>	(Lim et al., 2010)

Phomoxanthone A	<i>Phomopsis</i> sp	<i>Costus</i> sp	Antimicrobial activity		(Elsässer et al., 2005)
Botryosphaerones A-D (1-4)	<i>Botryosphaeria australis</i>	<i>Sonneratia apetala</i>	Cytotoxic and antimicrobial activities	 (1) (2) (3)	(Xu et al., 2011)
5-Methylmellein	<i>Xylaria</i> sp. PSUG12	<i>Garcinia hombroniana</i>	Insecticidal activity		(Rukachaisirikul et al., 2013)
4-epi-ethiosolide	<i>Pezicula</i> sp.	Coniferous tree	Fungicidal and herbicidal activities		(Schulz et al., 1995)
(+)-cryptosporiopsin	<i>Pezicula</i> sp.	Coniferous tree	Fungicidal and herbicidal activities		(Schulz et al., 1995)
(-)-mycorrhizin A	<i>Pezicula</i> sp.	Coniferous tree	Fungicidal and herbicidal activities		(Schulz et al., 1995)

4-Hydroxy-2-methoxy-6-methylbenzaldehyde	<i>Pezicula</i> sp.	Coniferous tree	Fungicidal and herbicidal activities		(Schulz et al., 1995)
(R)-mellein	<i>Pezicula</i> sp.	Coniferous tree	Fungicidal and herbicidal activities		(Schulz et al., 1995)

Cai, R., Chen, S., Long, Y., Li, C., Huang, X., and She, Z. (2017). Depsidones from *Talaromyces stipitatus* SK-4, an endophytic fungus of the mangrove plant *Acanthus ilicifolius*. *Phytochem. Lett.* 20, 196–199. doi:10.1016/j.phytol.2017.04.023.

Daisy, B. H., Strobel, G. A., Castillo, U., Ezra, D., Sears, J., Weaver, D. K., et al. (2002). Naphthalene, an insect repellent, is produced by *Muscodor vitigenus*, a novel endophytic fungus. *Microbiology* 148, 3737–3741.

Elsässer, B., Krohn, K., Flörke, U., Root, N., Aust, H.-J., Draeger, S., et al. (2005). X-ray Structure Determination, Absolute Configuration and Biological Activity of Phomoxanthone A. *Eur. J. Org. Chem.* 2005, 4563–4570. doi:10.1002/ejoc.200500265.

Lim, C.-S., Kim, J.-Y., Choi, J.-N., Ponnusamy, K., Jeon, Y.-T., Kim, S.-U., et al. (2010). Identification, fermentation, and bioactivity against *Xanthomonas oryzae* of antimicrobial metabolites isolated from *Phomopsis longicolla* S1B4. *J. Microbiol. Biotechnol.* 20, 494–500.

Ortega, H. E., Torres-Mendoza, D., and Cubilla-Rios, L. (2020). Patents on Endophytic Fungi for Agriculture and Bio- and Phytoremediation Applications. *Microorganisms* 8, 1237.

Rukachaisirikul, V., Buadam, S., Sukpondma, Y., Phongpaichit, S., Sakayaroj, J., and Hutadilok-Towatana, N. (2013). Indanone and mellein derivatives from the *Garcinia*-derived fungus *Xylaria* sp. PSU-G12. *Phytochem. Lett.* 6, 135–138.

Schulz, B., Sucker, J., Aust, H. J., Krohn, K., Ludewig, K., Jones, P. G., et al. (1995). Biologically active secondary metabolites of endophytic *Pezicula* species. *Mycol. Res.* 99, 1007–1015.

Verma, V. C., Prakash, S., Singh, R. G., and Gange, A. C. (2014). “Host-mimetic metabolomics of endophytes: looking back into the future,” in *Advances in Endophytic Research* (Springer), 203–218.

Xu, Y.-H., Lu, C.-H., Zheng, Z.-H., and Shen, Y.-M. (2011). New Polyketides Isolated from *Botryosphaeria australis* Strain ZJ12-1A. *Helv. Chimica. Acta.* 94, 897–902.

**Table S3.** Industrial applications of extracellular enzymes produced by endophytic fungi.

Compounds	Fungal endophytes	Host plants	Biological activities	References
Amylase	<i>Streptosporangium</i> sp.	<i>Zea mays</i>	Starch hydrolysis	(Stamford et al., 2002)
Amylase	<i>Cylindrocephalum</i> sp.	<i>Alpinia calcarata</i>	Starch hydrolysis	(Sunitha. et al., 2012)
Lipase	<i>Fusarium oxysporum</i>	<i>Croton oblongifolius</i>	Catalyze a wide range of chemical reactions in aqueous and non-aqueous environments	(Panuthai et al., 2012)
Protease	<i>Fusarium</i> sp. CPCC 480097	<i>Chrysanthemum</i>	Proteins degradation	(Wu et al., 2009)
Lipase	<i>Cercospora kikuchii</i>	-	Catalyze the hydrolysis of triacylglycerols to fatty acids, mono- and diacylglycerols, and glycerol	(Costa-Silva et al., 2011)
Cellulase	<i>Xylaria</i> sp.	<i>Opuntia ficus-indica</i>	Hydrolysis of cellulose	(Bezerra et al., 2012)
Pectinase	<i>Penicillium glandicola</i>	<i>Opuntia ficus-indica</i>	Hydrolysis of pectine	(Bezerra et al., 2012)
Cellulase	<i>Penicillium</i> sp.	<i>Centella asiatica</i>	Cellulose degradation	(Devi et al., 2012)
Cellulase	<i>Fusarium oxysporum</i>	<i>Baccharis dracunculifolia</i>	Catalyze cellulolysis	(Onofre et al., 2013)
Amylase	<i>Aspergillus</i>	-	Starch hydrolysis	(Khan et al., 2017)
Xylanase	<i>Alternaria alternata</i>	<i>Croton oblongifolius</i>	Degrade $\beta$ -1, 4-xylan	(Wipusaree et al., 2011)
Xylanase	<i>Aspergillus terreus</i>	<i>Memora peregrine</i>	Degrade $\beta$ -1, 4-xylan	(Sorgatto et al., 2012)
Lipase	<i>Cercospora kikuchii</i>	<i>Tithonia diversifolia</i>	Catalyze the hydrolysis of triacylglycerols	(Costa-Silva et al., 2011)
Lipase	<i>Fusarium oxysporum</i>	<i>Croton oblongifolius</i>	Catalyze the hydrolysis of triacylglycerols	(Amirita et al., 2012)
Pectinase	<i>Penicillium chrysogenum</i>	<i>Asclepias sinaica</i>	Hydrolyze pectin substances	(Fouda et al., 2015)
Pectinase	<i>Piriformospora indica</i>	-	Hydrolyze pectin substances	(Heidarizadeh et al., 2018)
Phytase	<i>Azospirillum</i> spp.	<i>Oryza sativa</i>	Phytates degradation	(Mehdipour-Moghaddam et al., 2010)
Protease	<i>Alternaria alternata</i>	-	Proteins degradation	(Rajput et al., 2016)
Chitinase	<i>Streptomyces griseus</i>	<i>Citrus reticulata</i>	Chitin hydrolysis	(Quecine et al., 2008)

Amirita, A., Sindhu, P., Swetha, J., Vasanthi, N. S., and Kannan, K. P. (2012). Enumeration of endophytic fungi from medicinal plants and screening of extracellular enzymes. *World J. Sci. Technol.* 2, 13–19.

Bezerra, J. D. P., Santos, M. G. S., Svedese, V. M., Lima, D. M. M., Fernandes, M. J. S., Paiva, L. M., et al. (2012). Richness of endophytic fungi isolated from *Opuntia ficus-indica* Mill. (Cactaceae) and preliminary screening for enzyme production. *World J. Microbiol. Biotechnol.* 28, 1989–1995. doi:10.1007/s11274-011-1001-2.

Costa-Silva, T. A., Nogueira, M. A., Souza, C. R. F., Oliveira, W. P., and Said, S. (2011). Lipase Production by Endophytic Fungus *Cercospora kikuchii*: Stability of Enzymatic Activity after Spray Drying in the Presence of Carbohydrates. *Drying Technol.* 29, 1112–1119. doi:10.1080/07373937.2011.573153.

Devi, N. N., Prabakaran, J. J., and Wahab, F. (2012). Phytochemical analysis and enzyme analysis of endophytic fungi from *Centella asiatica*. *Asian Pac. J. Trop. Biomed.* 2, S1280–S1284.

- Fouda, A. H., Hassan, S. E.-D., Eid, A. M., and Ewais, E. E.-D. (2015). Biotechnological applications of fungal endophytes associated with medicinal plant *Asclepias sinaica* (Bioss.). *Ann. Agric. Sci.* 60, 95–104.
- Heidarizadeh, M., Rezaei, P. F., and Shahabivand, S. (2018). Novel pectinase from *Piriformospora indica*, optimization of growth parameters and enzyme production in submerged culture condition. *Turk. J. Biochem.* 43, 289–295.
- Khan, A. L., Shahzad, R., Al-Harrasi, A., and Lee, I.-J. (2017). “Endophytic microbes: a resource for producing extracellular enzymes,” in *Endophytes: Crop productivity and protection* (Springer), 95–110.
- Mehdipour-Moghaddam, M. J., Emtiazi, G., Bouzari, M., Mostajeran, A., and Salehi, Z. (2010). Novel phytase and cellulase activities in endophytic *Azospirilla*. *W. Appl. Sci. J.* 10, 1129–1135.
- Onofre, S. B., Mattiello, S. P., da Silva, G. C., Groth, D., and Malagi, I. (2013). Production of cellulases by the endophytic fungus *Fusarium oxysporum*. *J. Microbiol. Res.* 3, 131–134.
- Panuthai, T., Sihanonth, P., Piapukiew, J., Sooksai, S., Sangvanich, P., and Karnchanatat, A. (2012). An extracellular lipase from the endophytic fungi *Fusarium oxysporum* isolated from the Thai medicinal plant, *Croton oblongifolius* Roxb. *Afr. J. Microbiol. Res.* 6, 2622–2638.
- Quecine, M. C., Araujo, W. L., Marcon, J., Gai, C. S., Azevedo, J. L., and Pizzirani-Kleiner, A. A. (2008). Chitinolytic activity of endophytic *Streptomyces* and potential for biocontrol. *Lett. Appl. Microbiol.* 47, 486–491. doi:10.1111/j.1472-765X.2008.02428.x.
- Rajput, K., Chanyal, S., and Agrawal, P. K. (2016). Optimization of protease production by endophytic fungus, *Alternaria alternata* isolated from gymnosperm tree-Cupressus torulosa D Don. *World J. Pharm. Pharmaceut. Sci.* 5, 1034–1054.
- Sorgatto, M., Guimarães, N. C. A., Zanoelo, F. F., Marques, M. R., Peixoto-Nogueira, S. C., and Giannesi, G. G. (2012). Purification and characterization of an extracellular xylanase produced by the endophytic fungus, *Aspergillus terreus*, grown in submerged fermentation. *Afr. J. Biotechnol.* 11, 8076–8084.
- Stamford, T. L. M., Stamford, N. P., Coelho, L., and Araujo, J. M. (2002). Production and characterization of a thermostable glucoamylase from *Streptosporangium* sp. endophyte of maize leaves. *Bioresour. Technol.* 83, 105–109.
- Sunitha., V. H., Ramesha., A., Savitha., J., and Srinivas., C. (2012). Amylase production by endophytic fungi *Cylindrocephalum* sp. isolated from medicinal plant *Alpinia calcarata* (Haw.) Roscoe. *Braz. J. Microbiol.* 43, 1213–1221. doi:10.1590/S1517-83822012000300049.
- Wipusaree, N., Sihanonth, P., Piapukiew, J., Sangvanich, P., and Karnchanatat, A. (2011). Purification and characterization of a xylanase from the endophytic fungus *Alternaria alternata* isolated from the Thai medicinal plant, *Croton oblongifolius* Roxb. *Afr. J. Microbiol. Res.* 5, 5697–5712.
- Wu, B., Wu, L., Chen, D., Yang, Z., and Luo, M. (2009). Purification and characterization of a novel fibrinolytic protease from *Fusarium* sp. CPCC 480097. *J. Ind. Microbiol. Biotechnol.* 36, 451–459.