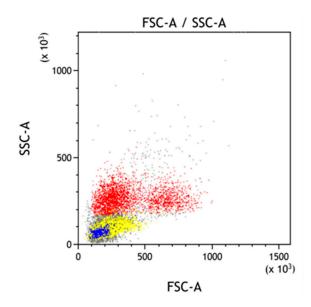


**Supplementary Figure 1A.** Gating strategy. The peripheral blood mononuclear cells were selected as singlet cell by forward-scatter channel (FSC)-Area (A) and -High (H). Subsequent gating in medium side-scatter channel (SSC) and low SSC was performed. Within medium-SSC, the monocytes were identified with anti-CD14-ECD, whereas within low-SSC, T and B lymphocytes were selected with anti-CD3 and anti-CD19, respectively.

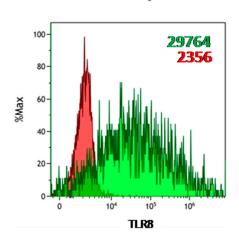
## **Supp Figure 1B**



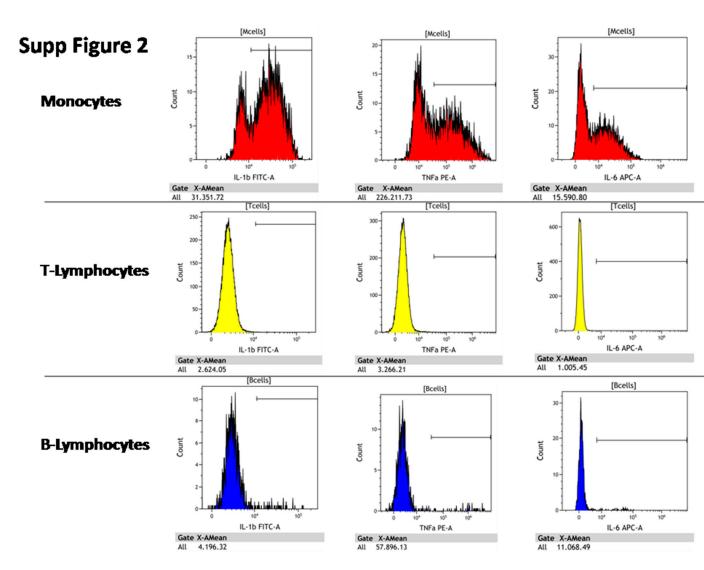
**Supplementary Figure 1B.** Backgating. After backgating, the subpopulations are depicted as Monocytes in red, T-lymphocytes in yellow, and B-lymphocytes in blue.

## **Supp Figure 1C**

## Monocytes



**Supplementary Figure 1C.** TLR8 expression in Monocytes. The monocytes were identified following gating strategy (Supplementary Figure 1A) and mean fluorescence intensity (MFI) of TLR8 expression (green) and isotype control (red) was measured.



**Supplementary Figure 2.** IL-1b, TNF-a, and IL-6 expression after TLR8 agonist treatment in different subpopulations. The raw mean fluorescence intensity in each subpopulation is shown. After 18 hours of culture with single-stranded RNA (ssRNA) 40 (TLR8 agonist) the cells were stained with monoclonal antibodies to identify monocytes (red), T (yellow), and B lymphocytes (blue) with CD14, CD3, and CD19, respectively. Further permeabilization and anti- interleukin-1b (IL-1b, right panels), tumor necrosis factor (TNF)- $\alpha$  in center panels, and IL-6 (on the left panels) were used to measure the expression of soluble factors.