

Supplementary Materials

One-Enzyme RTX-PCR for the Detection of RNA Viruses from Multiple Virus Genera and Crop Plants

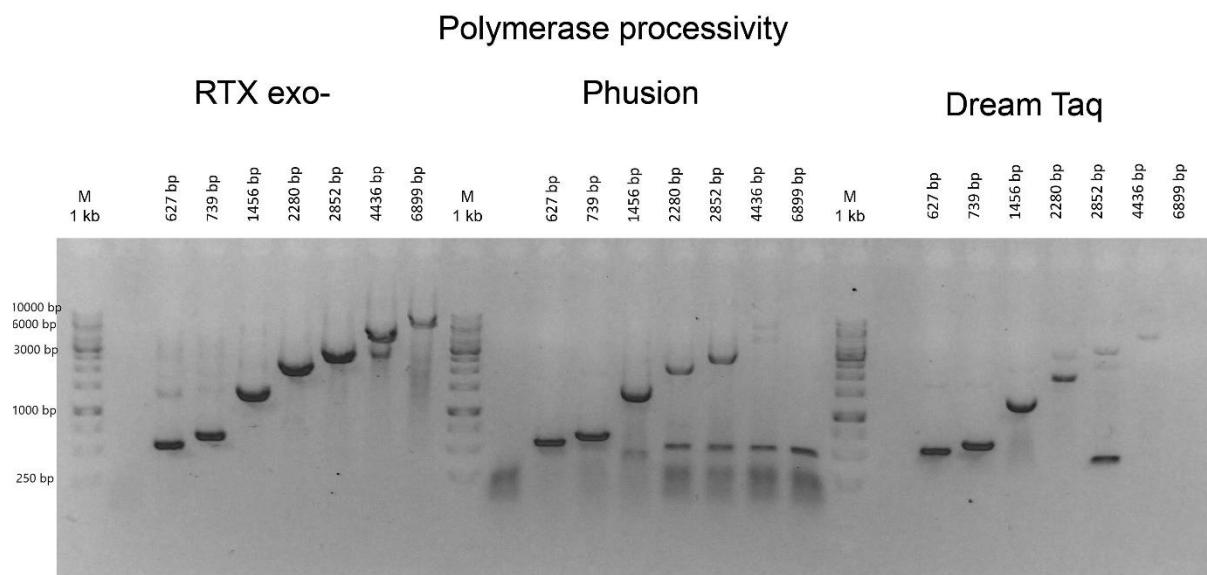


Figure S1. Estimate of processivity of thermostable polymerases. Universal domestication plasmid pUPD2 [56] with cloned inserts of various sizes (from 332 to 6604) were used as a template in PCR with universal sequencing primers pUPD F2/R2 [38]. 30 cycles of PCR program consisting of 30s denaturation at 94°C, 30s annealing at 55°C and short extension for 10s at 72°C were run with 3 different polymerases: RTX exo-, Phusion (ThermoScientific), and DreamTaq (ThermoScientific). Under these settings, the RTXexo- can be used to amplify products up to 5-6 kb in length. The marker is the GeneRuler 1 kb bp Plus DNA ladder from ThermoScientific.

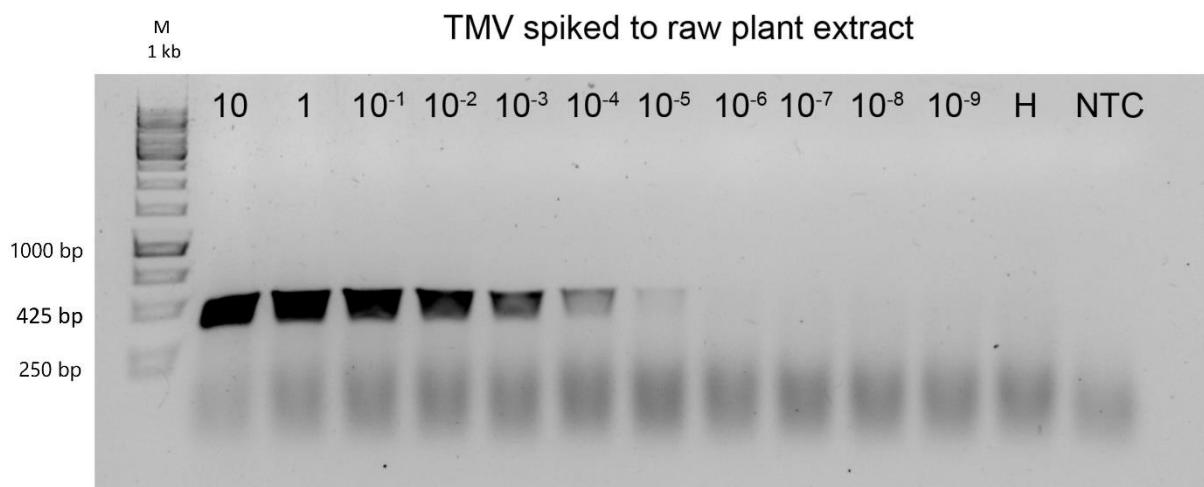


Figure S2. RT-PCR virus detection in crude extracts of *Nicotiana benthamiana*. Purified TMV was added to the extract of uninfected healthy plants to a final concentration of 10 ng/ μ l, and a 10-fold dilution series was prepared using extracts of healthy plants. 1 μ l of the extracts was used as a template for RT-PCR using the RTX enzyme. Numbers indicate ng of pure TMV virus in each

reaction. H= extract from uninfected control plants, NTC = non-template control. The marker is the GeneRuler 1 kb bp Plus DNA ladder from ThermoScientific.

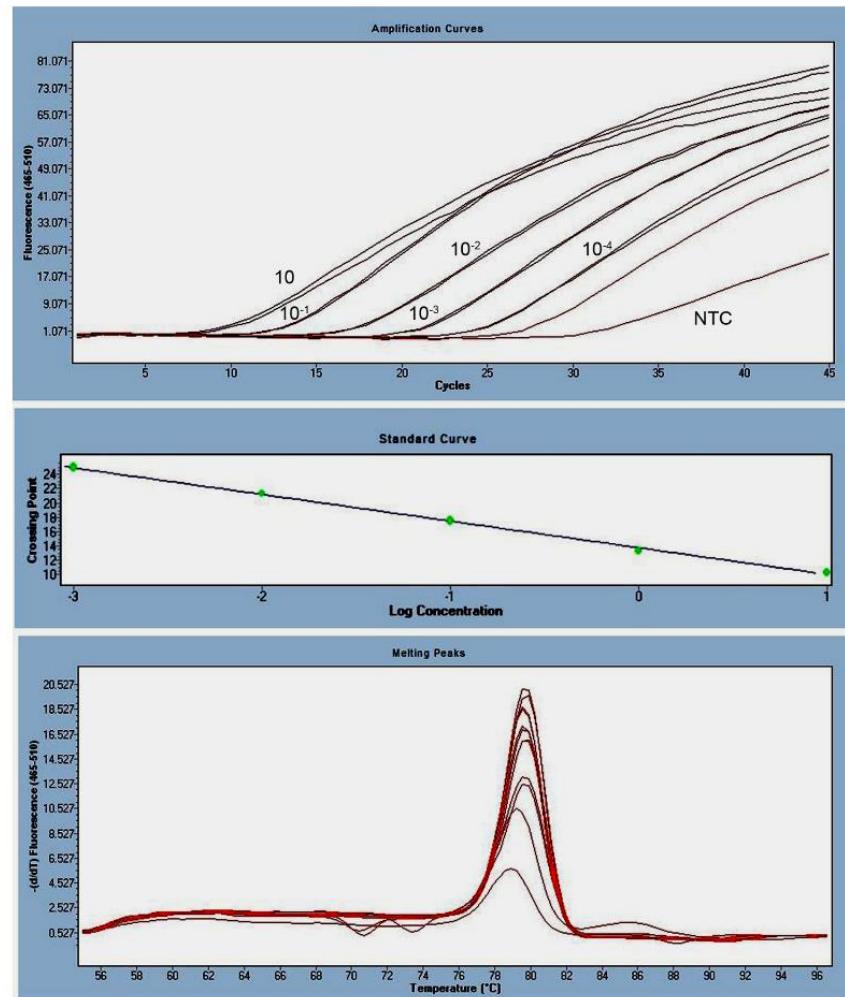


Figure S3. RT-qPCR reactions with the RTX exo- enzyme. Calibration curve constructed with the purified TMV preparation 10-fold serially diluted at 2.5×10^{-5} ng/reaction. Reactions were run in duplicates. The upper panel shows the real-time fluorescence measurements, the middle panel shows the standard calibration curve, and the lower panel shows the melting curve analysis of amplified fragments. .

Table S1. List of primers used in this work.

Virus	Genus	Primer Name	Primer sequence (5'-3')	Nucleotide Position	PCR Fragment bp	Reference
Tobacco mosaic virus (TMV)	Tobamovirus	TMV-MP-detAS	TGCAAGCCTGATCGACATAG	5507-5526		in this study
		TMV-MP-detS	TCTGTTAGCCGGTTGGTC	5102-5121	425	in this study
		Gb2-MP-F	GCGCCGTCTCGCTCGA ATGGCTCTAGTGTAAAGGAAAA*	4903-4926	624 + 16	in this study
		TMV-Rep-BsaI-S	GGAGTTCAGAAGATCTTTGTGATG	4740-4752	787	in this study
		TMV-LB	GAAACACTGTGATCATTGCTGC	4426-4447	1101	in this study
		TMV-rep-4080S	CCGTTGTTAGTGAGCTTACTAGGC	4080-4104	1447	in this study
		TMV-rep-3148S	CTCTGCTTCAAGAGGGTATTCAAG	3148-3171	2379	in this study
		GB-TMV-3_4 F	GCGCCGTCTCGCTCGAATG CTTCATGATGAATTGGGAAAAG*	2728-2752	2799 + 19	in this study
		TMV-rep-2223S	GCTGCGGTGTCGAATCTCGTCAAG	2223-2246	3304	in this study
		TMV-rep-1396S	TGGACAAATCTTGTACAATCCT	1396-1419	4131	in this study
		TMV qFor1	ATTAGACCCGCTAGTCACAGCAC	5939-2961		in this study
		TMV qRev1	TAGAGTAGACGACGCCAACGGTG	6050-6071	131	in this study
Potato virus X (PVX)	Potexvirus	PVX-F3	AAGCTCCACAGGAAACACAG	1612-1631		in this study
		PVX-B3	GGTTGGGAGTGTGAGTTCTT	1790-1809	198	in this study
Potato virus Y (PVY)	Potyvirus	GB2-YCPF1	GCGCCGTCTCGCTCGA ATGGGAAATGACACAATCGATG*	8571-8589		in this study
		GB2-YCPR1	GCGCCGTCTCGCTCAA AGCTCACATGTTCTGACTCCAAG*	9354-9374	804 + 19 + 19	in this study
		Mor1	AGGAGGAAGCACTAAGAAG	8591-8609	332	[17]
		Mor2	CAAACCATAAGCCCATTCACT	8903-8923		[17]
		MF05-21-R	CAATACGTAAGTGCACACCCCG	5654-5632		[18]
Turpin mosaic virus (TuMV)	Potyvirus	MF05-22-F	CAGGGCGTGAGTGGGGCTC	5334-5354	320	[18]
		TRSV-Pr-F	GGGGTGCTTACTGGCAAGG	3195-3213	521	[19]
		TRSV-R	ACTTGTGCCAGGAGAGCTA	3716-3695		[20]
		Nibfrg1F	GCACGGCAGTCTACGCACAAAC	7174-7195	679	in this study
		Nibfrg1R	CTGGTAACGACTCCAACAAGCG	7911-7890		in this study
		TuMV-full-CPF	GTGTTATCACCAGGCAGGTG	8731-8751	896	[21]
		TuMV-full-CPR	GTCTACCAGCATAACAACCCAC	9627-9606		[21]

		TuMV-F1qPCR	GTTCAACGCGGAAGCAGTTT	9071-9090	208	in this study
		TuMV-R2qPCR	GCCTAAATGTGGGTTGGCG	9279-9260		in this study
		luteoviruses-F	GCTCTAGAATTGTTAACGCTCG	generic primer	610	[22]
		luteoviruses-R	CACCGCGTCIACCTATTIGGTTG	generic primer		[22]
Turnip yellows virus (TuYV)	Polerovirus	TuYV-full-CPR	CCGGGTTCCCTCGTCTACCTA	4095-4076	947	in this study
		TuYV-full-CPF	CACGTACCGAAATCGTTAACG	3148-3130		in this study
		TuYVR-K2	ATTGGTCCTCGGCAACGTCG	4114-4126	966	[23]
		BYcpF	CCACTAGAGAGGTGGTGAAT	2840-2859	640	[24]
		BYcpR	CCGATGTTGAGGAGTCTACC	3479-3460		[24]
Barley yellow dwarf virus (BYDV)	Luteovirus	BYDV-PVinterF	GTTGAGTTAACGTCACACGC	3182 - 3201	294	[25]
		BYDV-Yan-Ra	TGTTGAGGAGTCTACCTATTG	3475 - 3454		[25]
		BY5661R	TGCCGAACTGCTCTTCGAGTG	5661-5640		in this study
		BY4836F	ATCCTGGAAACAGGCAGAAC	4836-4856	825	in this study
		WSMVcoatPRv	GAAACTGTGCGTGTCTCCC	9197-9178		[26]
		WSMVcoatPFv	GCGTGTACGAATCGAGTGA	8161-8181	1014	[26]
Wheat streak mosaic virus (WSMV)	Tritimovirus	WSM8166	GAGAGCAAATACTGCGTGTACG	8166-8186		[27]
		WSM8909	GCATAATGGCTCGAAGTGATG	8909-8929	743	[27]
		WSMVspeFv	GCCTCGACA CGGGAGCTA	8397-8417		[28]
		WSMVspeRv	ACC CATCCAGGAAGCAAGG	8754-8736	354	[28]
		ASPV-A	ATAGCCGCCCCGGTTAGGTT	9237-9256		[29]
		ASPV-C	CTCTTGAACCAGCTGATGCC	8993-9012	264	[29]
Apple stem pitting virus (ASPV)	Foveavirus	ASPV forward	CWAAYCCWTTGAAACTGG	8312-8220		[30]
		ASPV reverse	GCTTGGCTCCAAYYTTTC	9151-9134		[30]
		ASPV sense	ATGTCTGGAACCTCATGCTGAA	8869-8895		[31]
		ASPV antisense	TTGGGATCAACTTACTAAAAAGCATAA	9211-9238	370	[31]
		AGUV-2	GGAATTTCACACGACTCTAACCTCC	6345-6371		[32]
		AGUV-U	CCCGCTGTTGGATTGATACACCTC	5873-5897	499	[32]
Apple stem grooving virus (ASGV)	Capillovirus	ASGV forward	GTTTCCAAGACGTGCTTC	5644-5661	819	[30]
		ASGV reverse	ACACTAACCGGAAATGC	6446-6463		[30]
		ASGV sense	GTTTCCAAGACGTGCTTC	6039-6064		[31]
		ASGV antisense	ACAACTAACCGGAAATGC	6286-6311	272	[31]
		PPV-F3	GGAATGTGGGTGATGATGG	9 153–9 171		[33]
Plum pox virus (PPV)	Potyvirus	PPV-RR	CTCTTCTGTTCCGACGTTTC	9 475–9 497	345	[33]
		mM3	CATTCCATTAACCTCCAAAAGAC	8 786–8 808		[34]
		mD5	TATGTCACATAAAGGCGTTCTC	8 207–8 228	605	[34]

		PPV-RR	CTCTTCTTGTGTTCCGACGTTTC	9 475–9 497	965	[33]
		RecJF	AATGATATTGATGATAGCCTTGAC	8532–8556		[35]
		8RACLS	GCCTACAAATTAGGTGAGAGGCTC	5 851–5 832	290	[36]
		5FACLS	TTCCAATGGATCATGAGGTC	5564–5 585		[36]
Apple chlorotic leaf spot virus (ACLSV)	Trichovirus	ACLSVFrII	CAAGAGAATTTCAGTTGCTCG	6745–6766		[30]
		ACLSV antisense	AAGTCTACAGGCTATTATTATAAGTCTAA	7507–7536	791	[31]
		ACLSV sense	TTCATGGAAAGACAGGGGCAA	6860–6880		[31]
		ACLSV antisense	AAGTCTACAGGCTATTATTATAAGTCTAA	7507–7536	647	[31]
		PDVdpuF	CCGAGTGGATGCTTCACG	1353–1370	220	[35]
		PDVdpR	CCTTAATGAGTCGT	1572–1557		[35]
Prune dwarf virus (PDV)	Iilarvirus	PDVcpF	CTTCCAACTTCGACTGTTG	1143–1162		[37]
		PDVcpR	TCATCCAATGACTATTTATCC	1830–1809	687	[37]
		PDVrna2F	TGATGACGTCGAAGAGCAAG	550–569		[37]
		PDVrna2R	GGGACAAGTCGAAAGAGCAG	949–968	418	[37]
General primers	pUPD2 plasmid	pUDP2-R2	GAGGAAGCCTGCATAACG	n.a.	Various	[38]
		pUDP2-F2	CCCGATCAACTCGAGTGCCA	n.a.		[38]

* nontemplate bases in italics