

# The Influence of Short Motifs on the Anticancer Activity of HB43 Peptide

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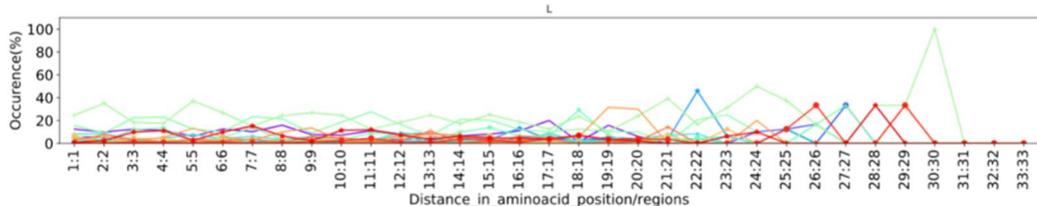
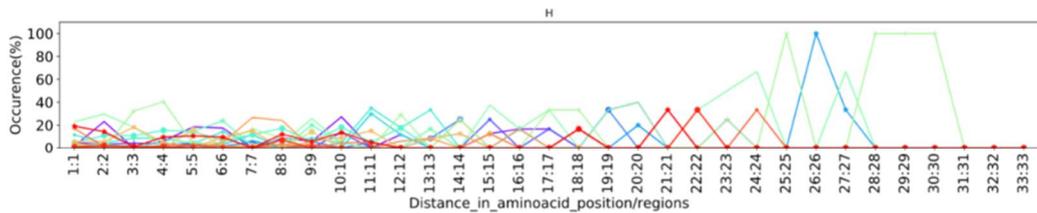
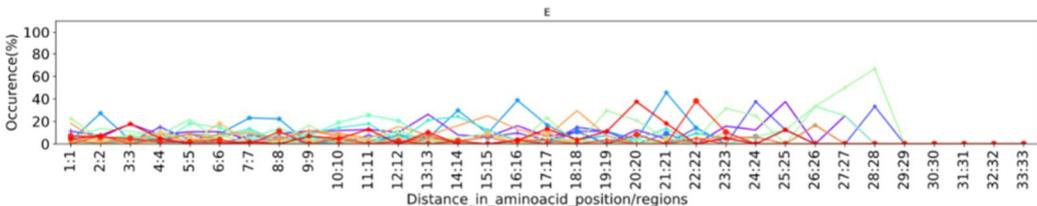
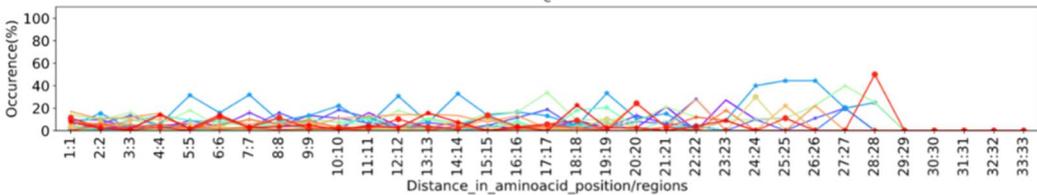
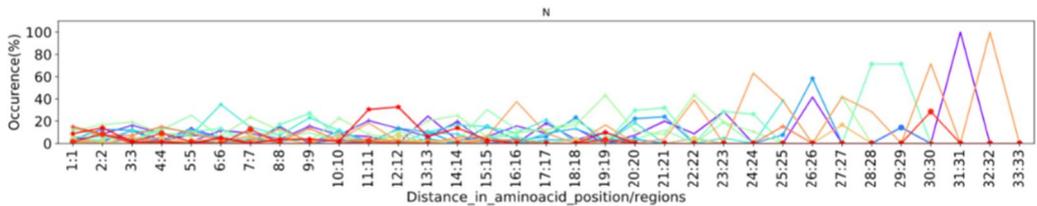
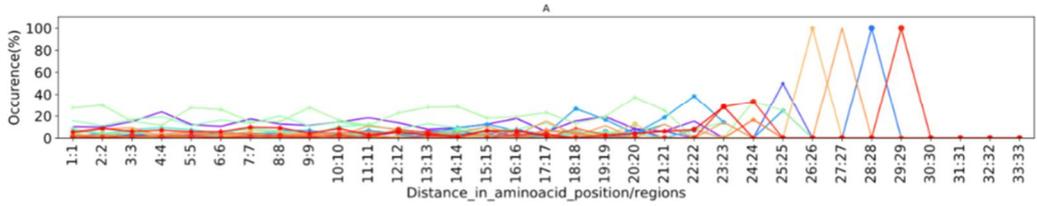
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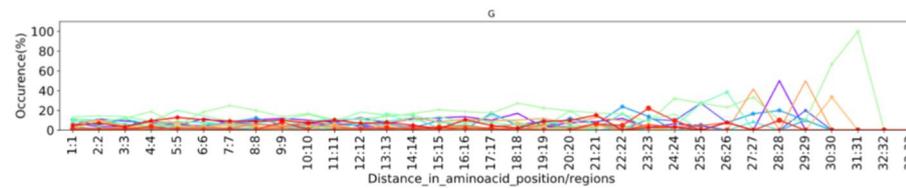
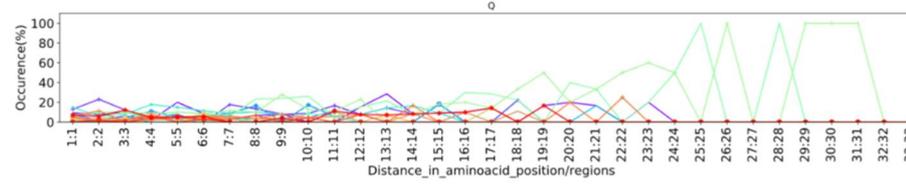
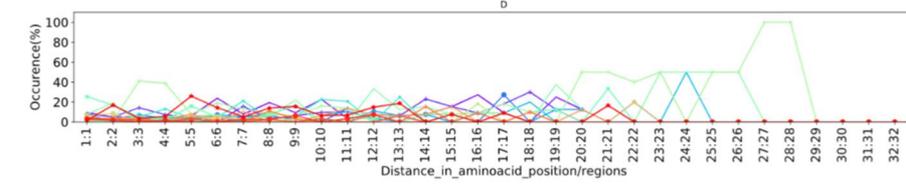
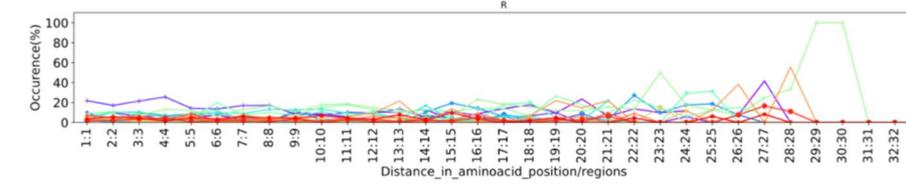
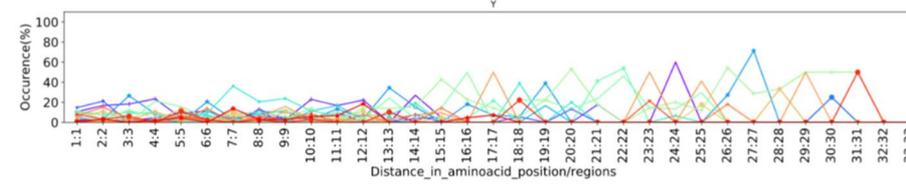
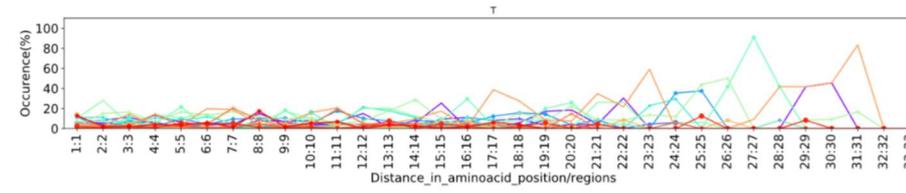
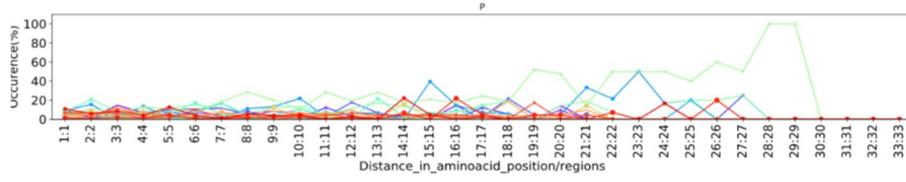
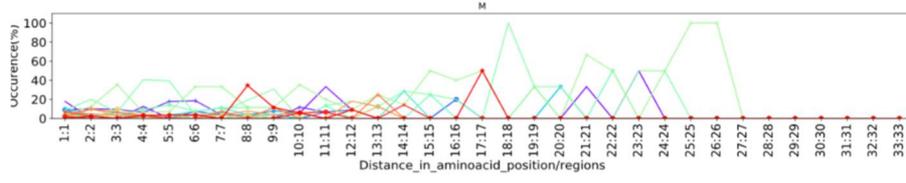
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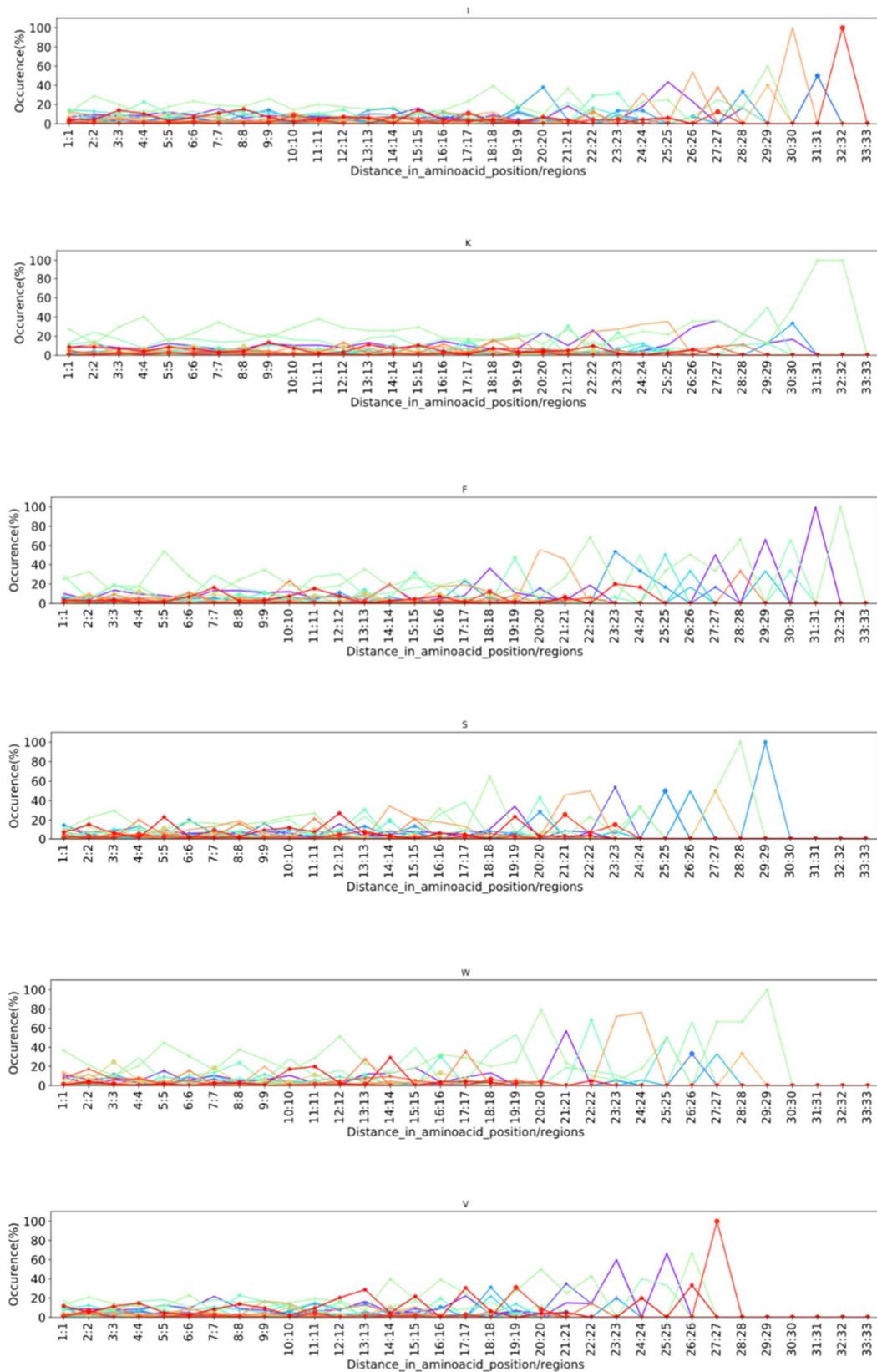
**Motifs:**  
**Position-independent probability to find a residue type at a certain distance from a reference residue type**

e.g. if an Alanine (A) is present one peptide of the family, the graph relative to A reports on the probability to find each residue type n positions (or regions) apart.

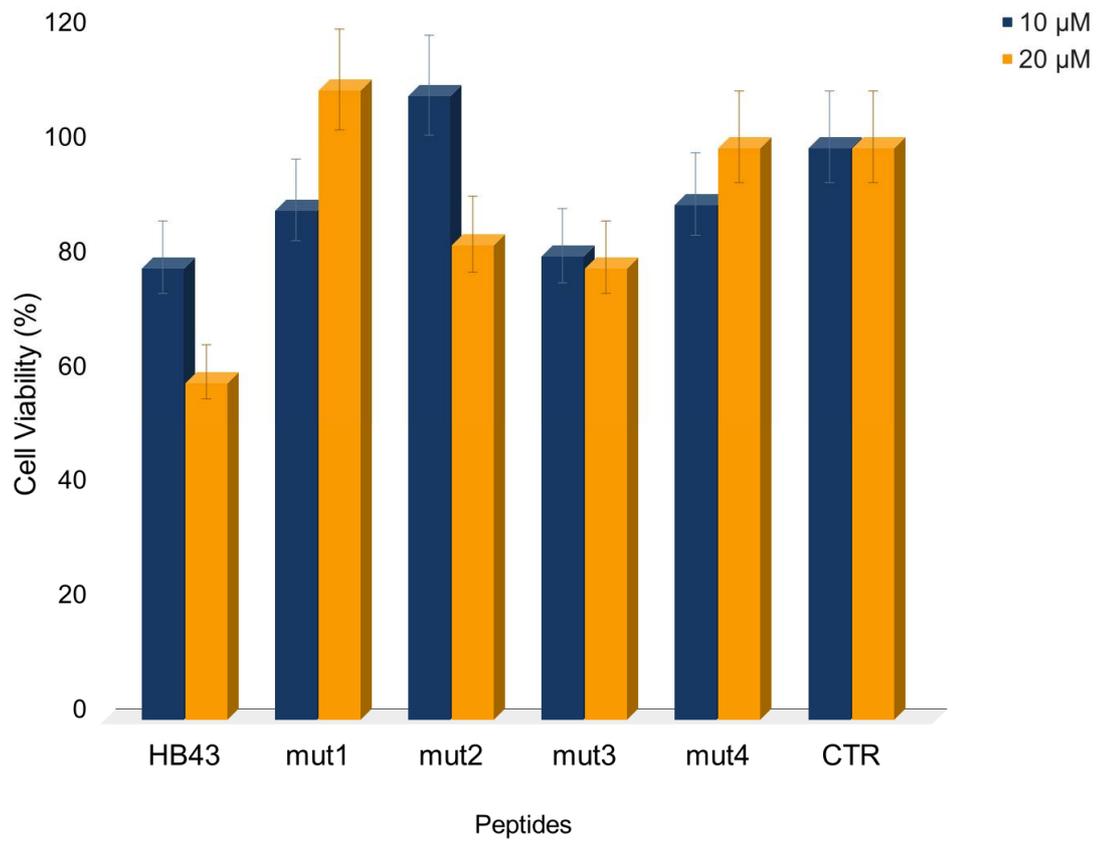
- (A) Alanine
- (R) Arginine
- (N) Asparagine
- (D) Aspartate
- (C) Cysteine
- (Q) Glutamine
- (E) Glutamate
- (G) Glycine
- (H) Histidine
- (I) Isoleucine
- (L) Leucine
- (K) Lysine
- (M) Methionine
- (F) Phenylalanine
- (P) Proline
- (S) Serine
- (T) Threonine
- (W) Tryptofan
- (Y) Tyrosine
- (V) Valine



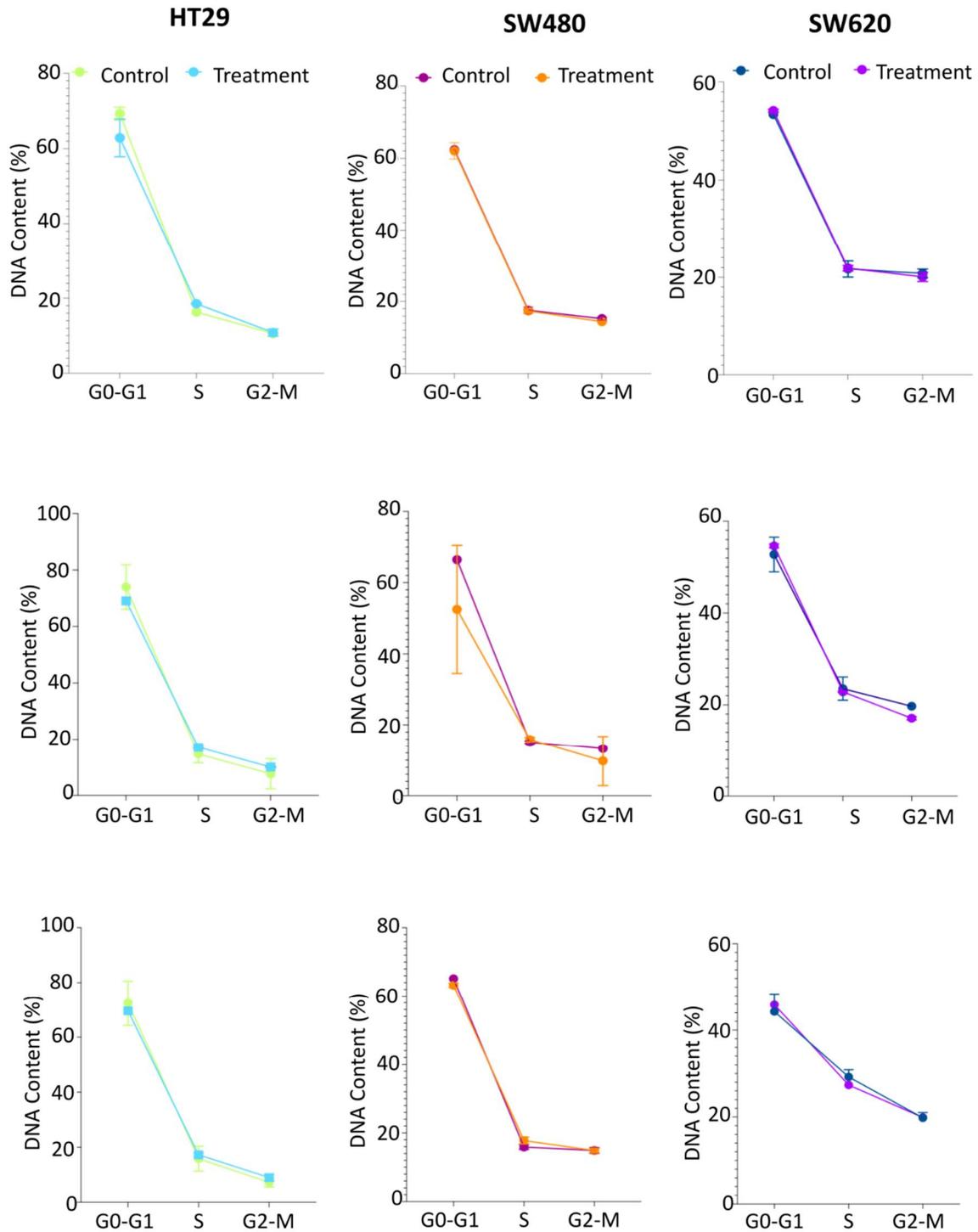




**Figure S1.** Analysis of the amino acid composition of HB43-related family generated by ADAPTABLE web server.



**Figure S2.** Cell viability of MDA-MB 231 breast cancer cells treated with HB43 and mutants.



**Figure S3.** Analysis of cell percentages in each cell cycle phase for colon cancer cells under study, HT29 (**left**), SW480 (**center**), and SW620 (**right**). Cells were treated with peptides HB43/*mut3* (5 μM), *mut2* (20 μM) and incubated for 12 h. Each data point is expressed as mean ± SD (n = 2).

**Table S1.**  $^1\text{H}$  and  $^{13}\text{C}$  NMR assignment of *mut3* 0.8 mM in 10 mM phosphate buffer pH 6.6, 10%  $\text{D}_2\text{O}$ , 278 K

	<b>Phe 1</b>	<b>Ala 2</b>	<b>Lys 3</b>	<b>Leu 4</b>	<b>Leu 5</b>	<b>Ala 6</b>	<b>Lys 7</b>
$^1\text{H}$	NH=X	NH= 8.47	NH= 8.62	NH= 8.59	NH= 8.41	NH= 8.55	NH= 8.54
	$\alpha$ =4.19	$\alpha$ = 4.33	$\alpha$ = 4.24	$\alpha$ = 4.33	$\alpha$ = 4.33	$\alpha$ = 4.27	$\alpha$ = 4.24
	$\beta_1$ = 3.1	$\beta$ = 1.37	$\beta_1$ = 1.79	$\beta$ = 1.63	$\beta$ = 1.63	$\beta$ = 1.39	$\beta_1$ = 1.79
	$\beta_2$ = 3.25		$\beta_2$ = 1.79	$\gamma$ = 1.66	$\gamma$ = 1.66		$\beta_2$ = 1.79
	$\delta$ = 7.30		$\gamma$ = 1.48	$\delta_1$ = 0.96	$\delta_1$ = 0.96		$\gamma$ = 1.48
	$\epsilon$ = 7.41		$\delta$ = 1.71	$\delta_2$ = 0.89	$\delta_2$ = 0.89		$\delta$ = 1.71
	$\zeta$ = 7.38		$\epsilon$ = 3.00				$\epsilon$ = 3.00
$^{13}\text{C}$	$\alpha$ =57.22	$\alpha$ = 52.66	$\alpha$ = 56.64	$\alpha$ = 55.09	$\alpha$ = 55.09	$\alpha$ = 52.64	$\alpha$ = 56.64
	$\beta$ = 40.2	$\beta$ = 19.41	$\beta$ = 33.23	$\beta$ = 42.34	$\beta$ = 42.34	$\beta$ = 19.26	$\beta$ = 33.23
	$\delta$ = 132.36		$\gamma$ = 25.08	$\gamma$ = 26.99	$\gamma$ = 26.99		$\gamma$ = 25.08
	$\epsilon$ = 131.98		$\delta$ = 29.35	$\delta_1$ = 25.13	$\delta_1$ = 25.13		$\delta$ = 29.35
	$\zeta$ = 130.78		$\epsilon$ = 42.11	$\delta_2$ = 23.45	$\delta_2$ = 23.45		$\epsilon$ = 42.11
	<b>Leu 8</b>	<b>Ala 9</b>	<b>Arg 10</b>	<b>Arg 11</b>	<b>Leu 12</b>	<b>Leu 13</b>	
$^1\text{H}$	NH= 8.51	NH= 8.72	NH= 8.50	NH= 8.60	NH= 8.61	NH= 8.43	
	$\alpha$ = 4.33	$\alpha$ = 4.33	$\alpha$ = 4.29	$\alpha$ = 4.29	$\alpha$ = 4.33	$\alpha$ = 4.33	
	$\beta$ = 1.63	$\beta$ = 1.39	$\beta_2$ = 1.81	$\beta_2$ = 1.81	$\beta$ = 1.634	$\beta$ = 1.63	
	$\gamma$ = 1.66		$\beta_3$ = 1.81	$\beta_3$ = 1.81	$\gamma$ = 1.66	$\gamma$ = 1.66	
	$\delta_1$ = 0.96		$\gamma$ = 1.66	$\gamma$ = 1.66	$\delta_1$ = 0.96	$\delta_1$ = 0.96	
	$\delta_2$ = 0.89		$\delta$ = 3.21	$\delta$ = 3.21	$\delta_2$ = 0.89	$\delta_2$ = 0.89	
			NH <sub>1</sub> = 7.01	NH <sub>1</sub> = 7.68			
			NH <sub>2</sub> = 6.56	NH <sub>2</sub> = 6.27			
$^{13}\text{C}$	$\alpha$ = 55.09	$\alpha$ = 52.37	$\alpha$ = 56.45	$\alpha$ = 56.45	$\alpha$ = 55.09	$\alpha$ = 55.09	
	$\beta$ = 42.34	$\beta$ = 19.26	$\beta$ = 30.82	$\beta$ = 30.82	$\beta$ = 42.34	$\beta$ = 42.34	
	$\gamma$ = 26.99		$\gamma$ = 27.43	$\gamma$ = 27.43	$\gamma$ = 26.99	$\gamma$ = 26.99	
	$\delta_1$ = 25.13		$\delta$ = 43.45	$\delta$ = 43.45	$\delta_1$ = 25.13	$\delta_1$ = 25.13	
	$\delta_2$ = 23.45				$\delta_2$ = 23.45	$\delta_2$ = 23.45	

**Table S2.** <sup>1</sup>H NMR assignment of *mut3* 0.8 mM (90% 10 mM phosphate buffer at pH 6.6, 10% D<sub>2</sub>O) in the presence of 50 mM DPC micelles at 278 K

<b>Phe 1</b>	<b>Ala 2</b>	<b>Lys 3</b>	<b>Leu 4</b>	<b>Leu 5</b>	<b>Ala 6</b>	<b>Lys 7</b>
NH=X	NH= 7.9	NH= 8.79	NH= 7.94	NH= 8.18	NH= 8.24	NH= 7.79
$\alpha$ =3.98	$\alpha$ = 3.92	$\alpha$ = 4.05	$\alpha$ = 4.16	$\alpha$ = 4.19	$\alpha$ = 4.04	$\alpha$ = 4.03
$\beta_1$ = 3.13	$\beta$ = 1.43	$\beta_1$ = 1.85	$\beta$ = 1.82	$\beta$ = 1.72	$\beta$ = 1.55	$\beta_1$ = 2.01
$\beta_2$ = 3.14		$\beta_2$ = 1.85	$\gamma$ = 1.47	$\gamma$ = 1.50		$\beta_2$ = 2.01
$\delta$ = 7.27		$\gamma$ = 1.45	$\delta_1$ = 0.91	$\delta_1$ = 0.91		$\gamma$ = 1.55
$\epsilon$ = 7.32		$\delta$ = 1.60	$\delta_2$ = 0.91	$\delta_2$ = 0.91		$\delta$ = 1.70
$\zeta$ = ?		$\epsilon$ = 3.30				$\epsilon$ = ?
<b>Leu 8</b>	<b>Ala 9</b>	<b>Arg 10</b>	<b>Arg 11</b>	<b>Leu 12</b>	<b>Leu 13</b>	
NH= 8.28	NH= 8.75	NH= 8.14	NH= 7.79	NH= 8.14	NH= 8.01	
$\alpha$ = 4.06	$\alpha$ = 3.92	$\alpha$ = 4.16	$\alpha$ = 4.17	$\alpha$ = 4.17	$\alpha$ = 4.14	
$\beta$ = 1.79	$\beta$ = 1.51	$\beta_2$ = ?	$\beta_2$ = ?	$\beta$ = 1.86	$\beta$ = 1.84	
$\gamma$ = 1.49		$\beta_3$ = ?	$\beta_3$ = ?	$\gamma$ = 1.51	$\gamma$ = 1.55	
$\delta_1$ = 0.92		$\gamma$ = ?	$\gamma$ = ?	$\delta_1$ = 0.95	$\delta_1$ = 0.92	
$\delta_2$ = 0.97		$\delta$ = ?	$\delta$ = ?	$\delta_2$ = 0.91	$\delta_2$ = 0.92	
		NH <sub>1</sub> = ?	NH <sub>1</sub> = ?			
		NH <sub>2</sub> = ?	NH <sub>2</sub> = ?			

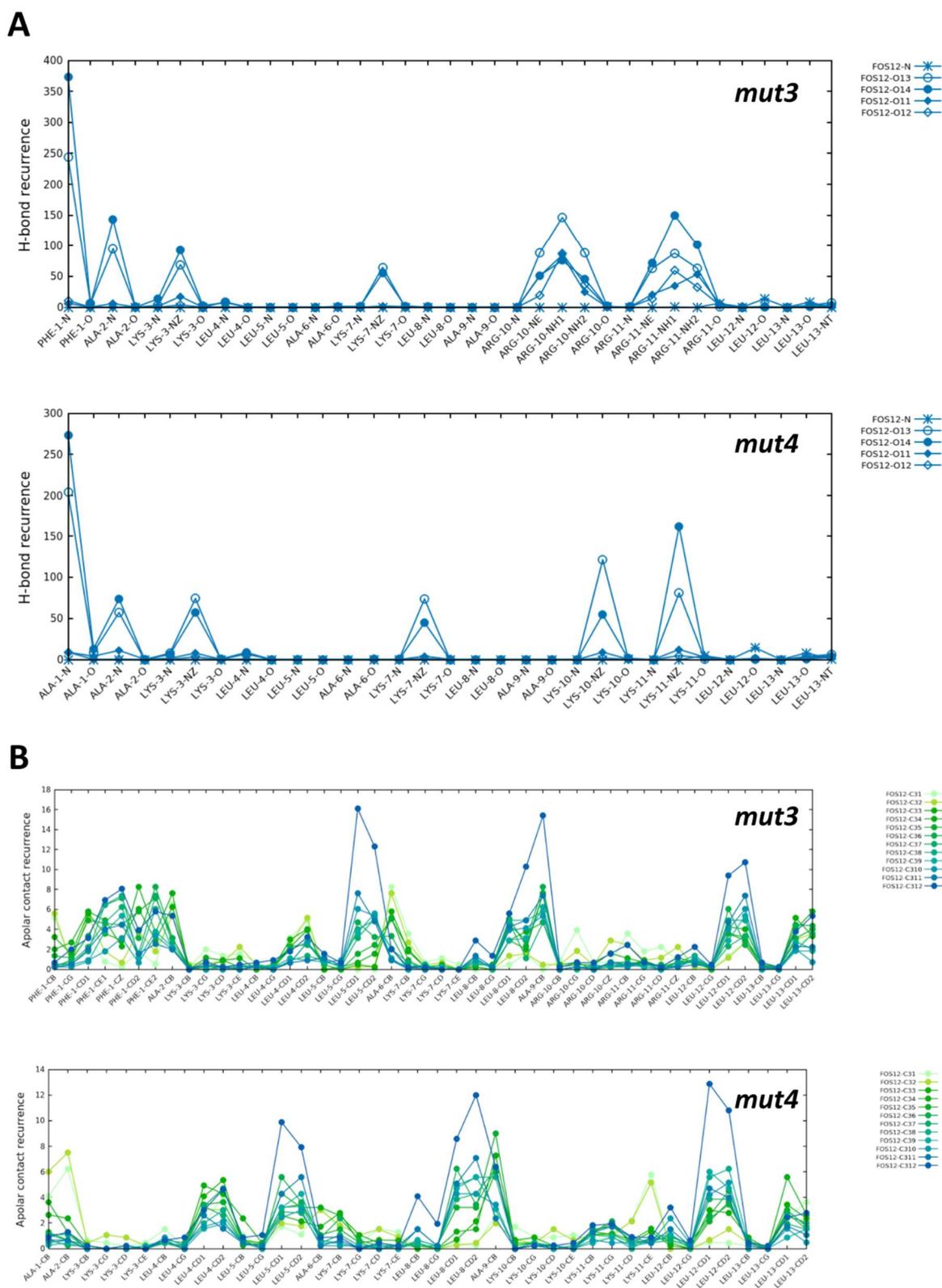
Missing values are due to overlap or exchange with the solvent (exchangeable protons). Severe broadening is observed for lysine and arginine side chains preventing their full assignment.

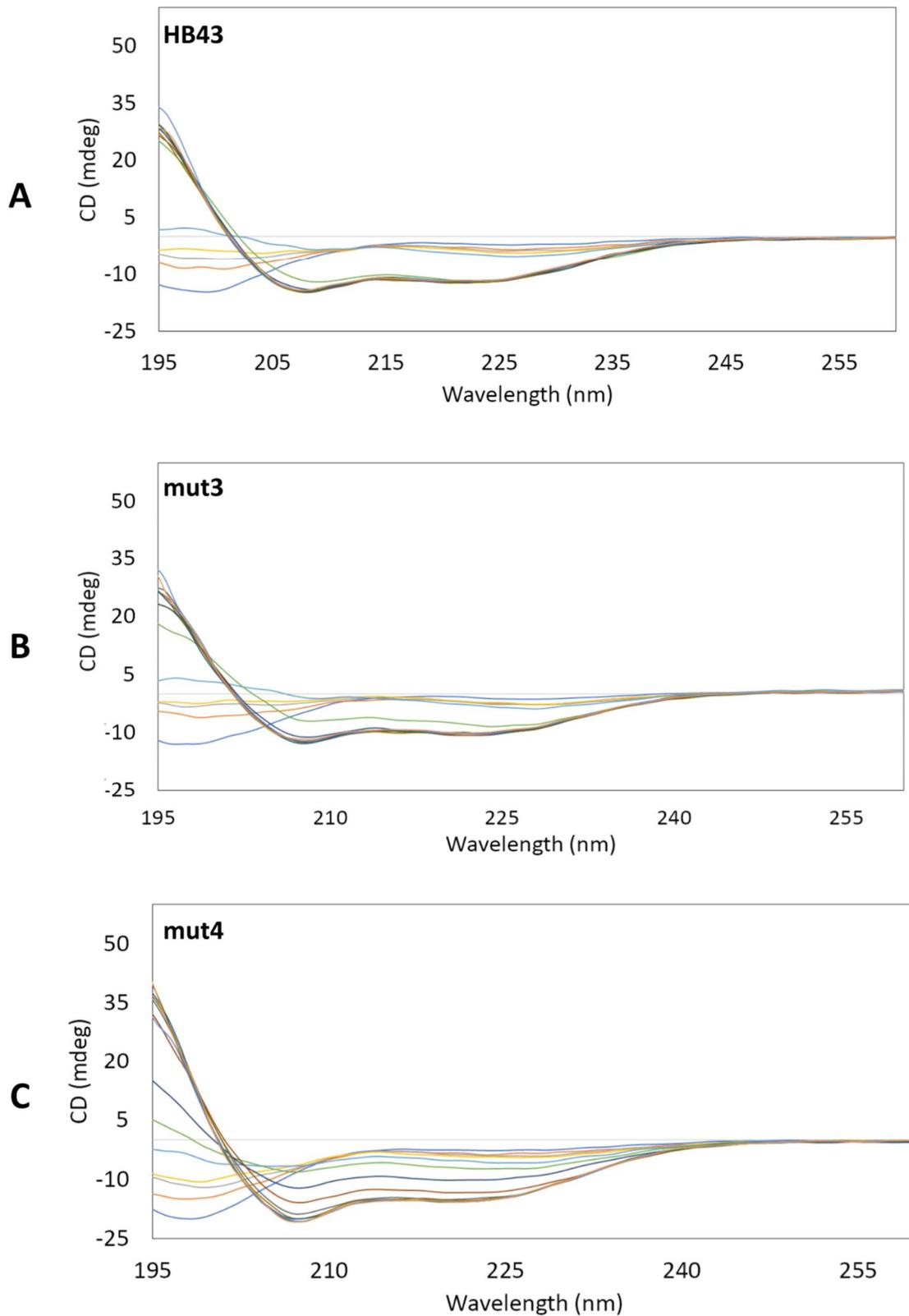
**Table S3.**  $^1\text{H}$  and  $^{13}\text{C}$  NMR assignment of *mut4* 0.8 mM in 10 mM phosphate buffer pH 6.6, 10%  $\text{D}_2\text{O}$ , 278 K

	<b>Ala 1</b>	<b>Ala 2</b>	<b>Lys 3</b>	<b>Leu 4</b>	<b>Leu 5</b>	<b>Ala 6</b>	<b>Lys 7</b>
$^1\text{H}$	NH= X	NH= 8.80	NH= 8.72	NH= 8.47	NH= 8.58	NH= 8.47	NH= 8.55
	$\alpha = 4.07$	$\alpha = 4.31$	$\alpha = 4.25$	$\alpha = 4.35$	$\alpha = 4.35$	$\alpha = 4.30$	$\alpha = 4.28$
	$\beta = 1.53$	$\beta = 1.39$	$\beta_1 = 1.78$	$\beta = 1.62$	$\beta = 1.62$	$\beta = 1.39$	$\beta_1 = 1.73$
			$\beta_2 = 1.78$	$\gamma = 1.66$	$\gamma = 1.66$		$\beta_2 = 1.73$
			$\gamma = 1.47$	$\delta_1 = 0.95$	$\delta_1 = 0.95$		$\gamma = 1.46$
			$\delta = 1.70$	$\delta_2 = 0.89$	$\delta_2 = 0.89$		$\delta = 1.70$
			$\epsilon = 3.01$				$\epsilon = 3.01$
$^{13}\text{C}$	$\alpha = 51.79$	$\alpha = 52.58$	$\alpha = 56.73$	$\alpha = 55.07$	$\alpha = 55.35$	$\alpha = 52.65$	$\alpha = 56.62$
	$\beta = 19.72$	$\beta = 19.37$	$\beta = 33.43$	$\beta = 42.53$	$\beta = 42.53$	$\beta = 19.37$	$\beta = 33.31$
			$\gamma = 25.08$	$\gamma = 27.20$	$\gamma = 27.20$		$\gamma = 25.08$
			$\delta = 29.42$	$\delta_1 = 25.07$	$\delta_1 = 25.07$		$\delta = 29.42$
			$\epsilon = 42.26$	$\delta_2 = 23.66$	$\delta_2 = 23.66$		$\epsilon = 42.26$
	<b>Leu 8</b>	<b>Ala 9</b>	<b>Lys 10</b>	<b>Lys 11</b>	<b>Leu 12</b>	<b>Leu 13</b>	
$^1\text{H}$	NH= 8.58	NH= 8.67	NH= 8.56	NH= 8.45	NH= 8.52	NH= 8.49	
	$\alpha = 4.35$	$\alpha = 4.26$	$\alpha = 4.26$	$\alpha = 4.26$	$\alpha = 4.36$	$\alpha = 4.33$	
	$\beta = 1.62$	$\beta = 1.39$	$\beta_1 = 1.78$	$\beta_1 = 1.78$	$\beta = 1.62$	$\beta = 1.62$	
	$\gamma = 1.66$		$\beta_2 = 1.78$	$\beta_2 = 1.78$	$\gamma = 1.66$	$\gamma = 1.66$	
	$\delta_1 = 0.95$		$\gamma = 1.47$	$\gamma = 1.47$	$\delta_1 = 0.95$	$\delta_1 = 0.95$	
	$\delta_2 = 0.89$		$\delta = 1.70$	$\delta = 1.70$	$\delta_2 = 0.89$	$\delta_2 = 0.89$	
			$\epsilon = 3.01$	$\epsilon = 3.01$			
$^{13}\text{C}$	$\alpha = 55.35$	$\alpha = 52.65$	$\alpha = 56.61$	$\alpha = 56.49$	$\alpha = 55.10$	$\alpha = 55.21$	
	$\beta = 42.53$	$\beta = 19.37$	$\beta = 33.31$	$\beta = 33.43$	$\beta = 42.53$	$\beta = 42.53$	
	$\gamma = 27.204$		$\gamma = 25.08$	$\gamma = 25.08$	$\gamma = 27.20$	$\gamma = 27.20$	
	$\delta_1 = 25.07$		$\delta = 29.42$	$\delta = 29.42$	$\delta_1 = 25.07$	$\delta_1 = 25.07$	
	$\delta_2 = 23.66$		$\epsilon = 42.26$	$\epsilon = 42.26$	$\delta_2 = 23.66$	$\delta_2 = 23.66$	

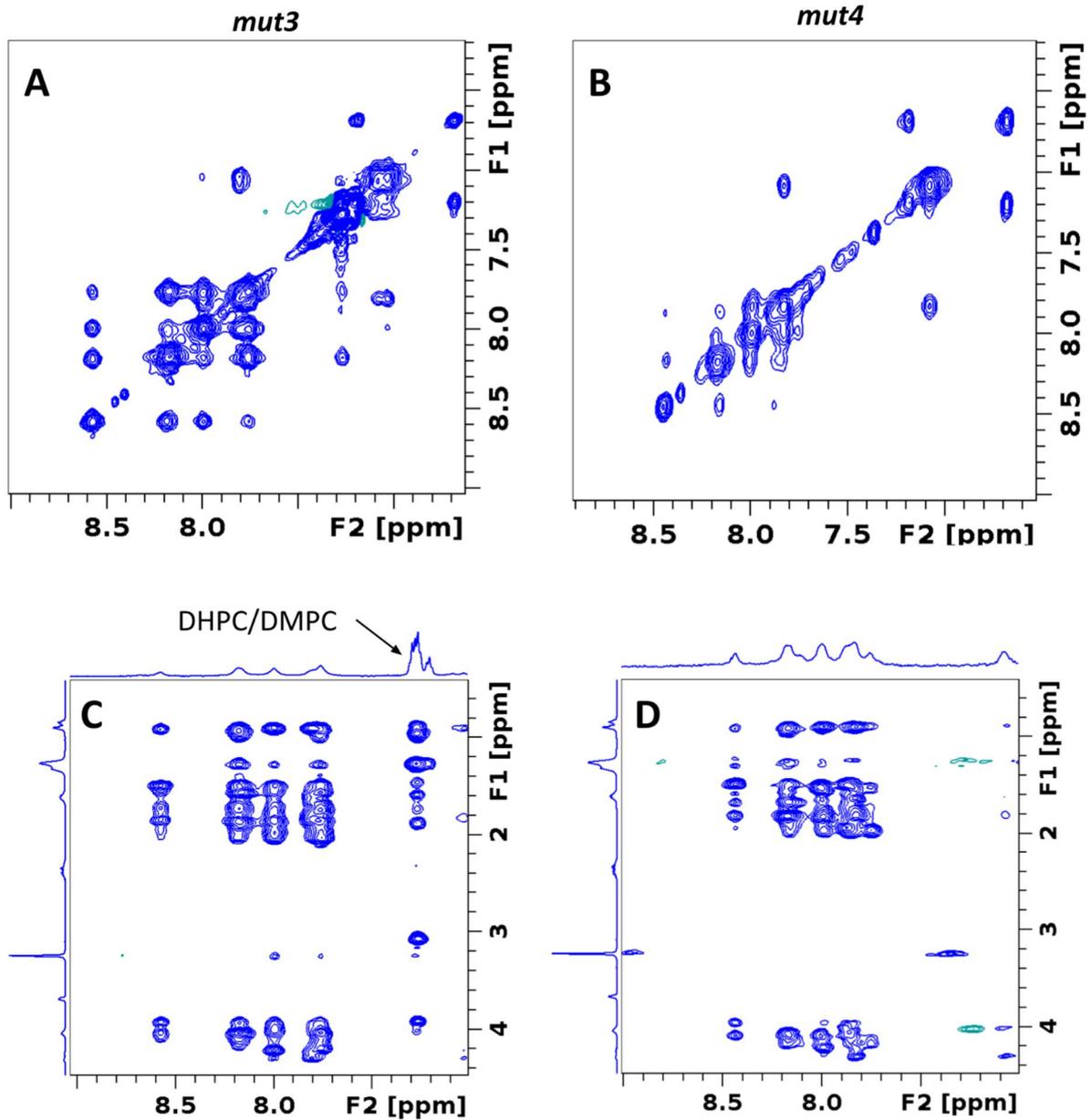
**Table S4.**  $^1\text{H}$  NMR assignment of *mut4* 0.8 mM (90% 10 mM phosphate buffer at pH 6.6, 10%  $\text{D}_2\text{O}$ ) in the presence of 50 mM DPC micelles at 278 K

<b>Ala 1</b>	<b>Ala 2</b>	<b>Lys 3</b>	<b>Leu 4</b>	<b>Leu 5</b>	<b>Ala 6</b>	<b>Lys 7</b>
NH= X	NH= 8.12	NH= 8.8	NH= 8.39	NH= 8.32	NH= 8.32	NH= 7.89
$\alpha = 4.13$	$\alpha = 4.06$	$\alpha = 4.16$	$\alpha = 4.14$	$\alpha = 4.06$	$\alpha = 4.09$	$\alpha = 4.05$
$\beta = 1.52$	$\beta = 1.53$	$\beta_1 = 1.91$	$\beta = 1.87$	$\beta = 1.85$	$\beta = 1.54$	$\beta_1 = 2.04$
		$\beta_2 = 1.91$	$\gamma = 1.68$	$\gamma = ?$		$\beta_2 = 2.04$
		$\gamma = 1.51$	$\delta_1 = ?$	$\delta_1 = 0.92$		$\gamma = 1.53$
		$\delta = 1.71$	$\delta_2 = ?$	$\delta_2 = 0.92$		$\delta = 1.70$
		$\epsilon = ?$				$\epsilon = ?$
<b>Leu 8</b>	<b>Ala 9</b>	<b>Lys 10</b>	<b>Lys 11</b>	<b>Leu 12</b>	<b>Leu 13</b>	
NH= 8.32	NH= 8.63	NH= 7.95	NH= 7.75	NH= 8.09	NH= 7.99	
$\alpha = 4.06$	$\alpha = 3.93$	$\alpha = 3.97$	$\alpha = 4.13$	$\alpha = 4.16$	$\alpha = 4.27$	
$\beta = 1.84$	$\beta = 1.50$	$\beta_1 = 1.95$	$\beta_1 = 2.02$	$\beta = 1.84$	$\beta = 1.82$	
$\gamma = ?$		$\beta_2 = 1.95$	$\beta_2 = 2.02$	$\gamma = ?$	$\gamma = 1.82$	
$\delta_1 = 0.92$		$\gamma = 1.52$	$\gamma = ?$	$\delta_1 = 0.91$	$\delta_1 = 0.92$	
$\delta_2 = 0.92$		$\delta = 1.72$	$\delta = ?$	$\delta_2 = 0.91$	$\delta_2 = 0.92$	
		$\epsilon = ?$	$\epsilon = ?$			

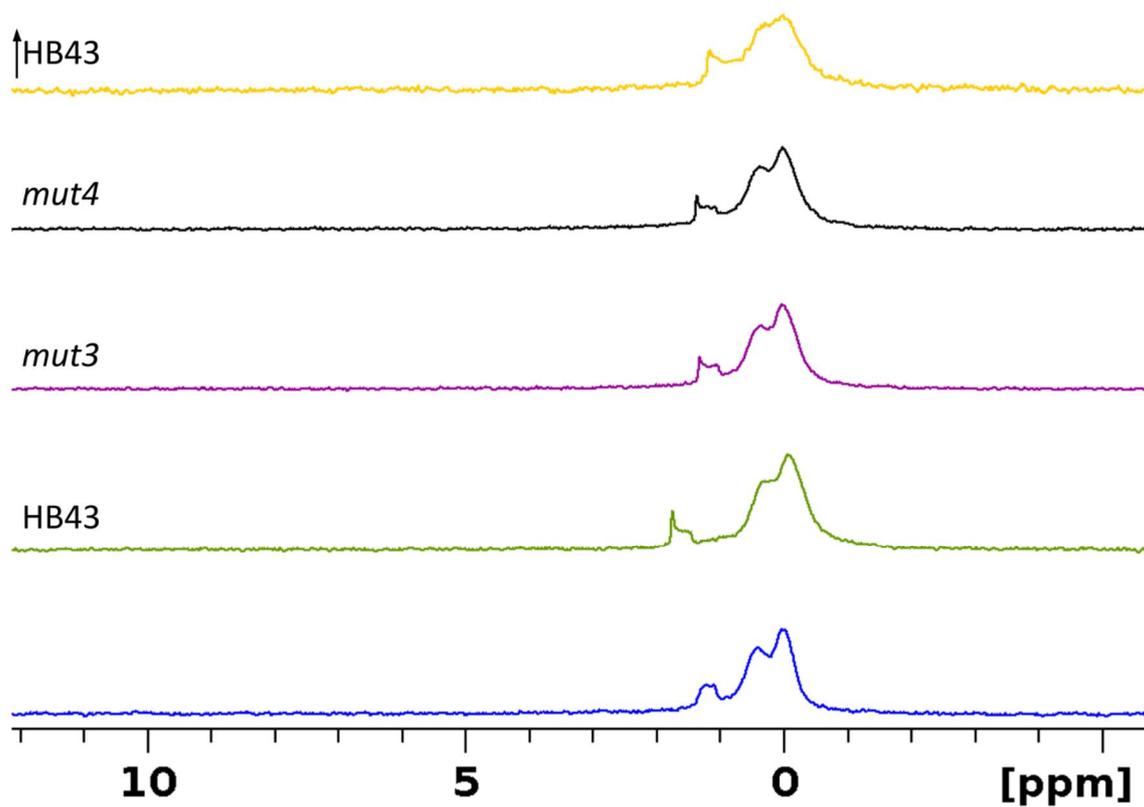




**Figure S5.** CD spectra of HB43 (A), *mut3* (B), and *mut4* (C) 10  $\mu$ M in 10 mM phosphate buffer at pH 6.6, in the absence (light green) and in the presence of increasing amounts of POPC/POPS SUVs (0,  $6.67 \times 10^6$ ,  $1.33 \times 10^5$ ,  $2.00 \times 10^5$ ,  $3.33 \times 10^5$ ,  $4.67 \times 10^5$ ,  $6.00 \times 10^5$ ,  $7.33 \times 10^5$ ,  $8.67 \times 10^5$ ,  $1.00 \times 10^4$ ,  $1.13 \times 10^4$ ,  $1.27 \times 10^4$ ,  $1.40 \times 10^4$ , and  $1.53 \times 10^4$  M).

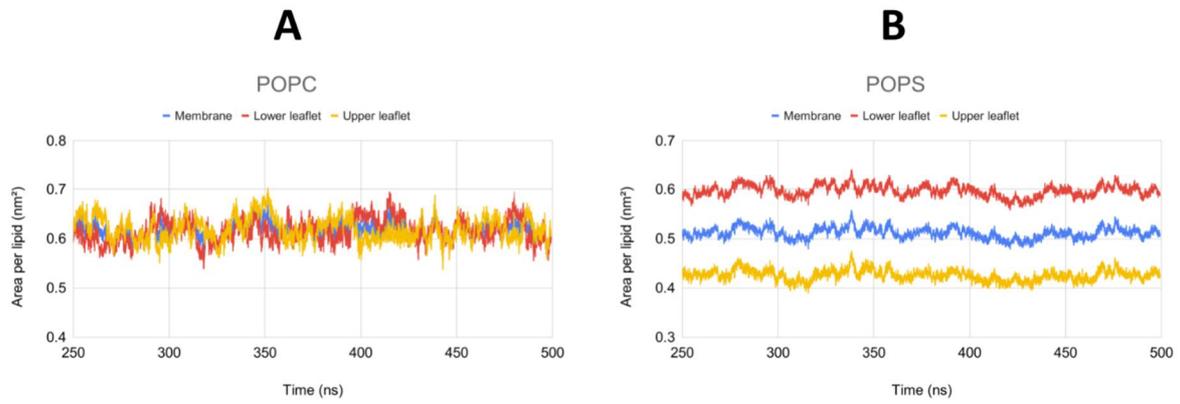


**Figure S6.** Amide and aromatic regions of  $^1\text{H},^1\text{H}$ -NOESY NMR spectrum of *mut3* and *mut4* 1.6 mM in 10 mM phosphate buffer at pH 6.6 (blue) and 310 K, in the presence of DMPC/DHPC isotropic bicelles at a total lipid concentration of 100 mM. (A,B) Amide region of *mut3* (A) and *mut4* (B) showing meaningful NOEs. (C,D) side-chain spectral regions of *mut3* (C) where aromatic signals of phenylalanine (Phe1) clearly show cross-peaks with the lipid chains of bicelles, a phenomenon not observed for *mut4* (D) due to the absence of this residue.

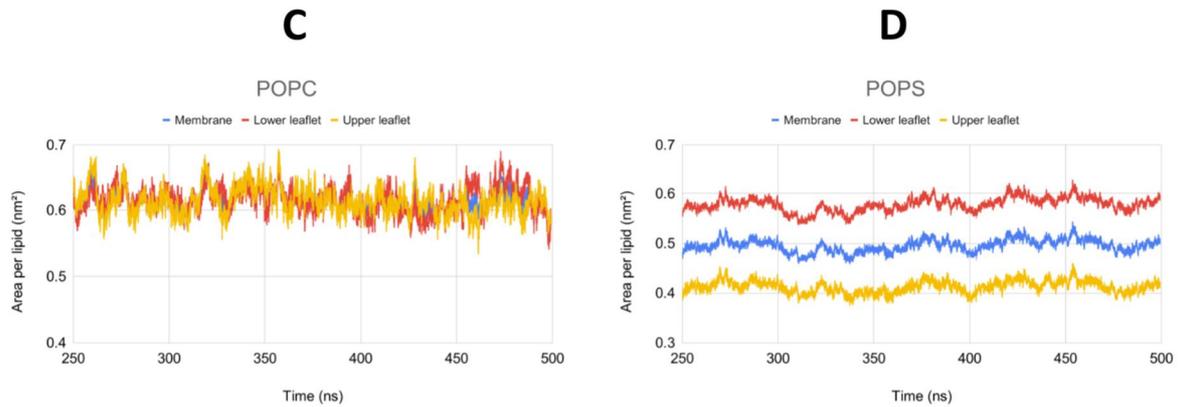


**Figure S7.** MAS <sup>31</sup>P spectra of POPC/POPS (1:1) liposomes in the absence (blue) and the presence of HB43 (green), *mut3* (magenta), *mut4* (black), and a more concentrated sample of HB43 (yellow).

## *mut3*

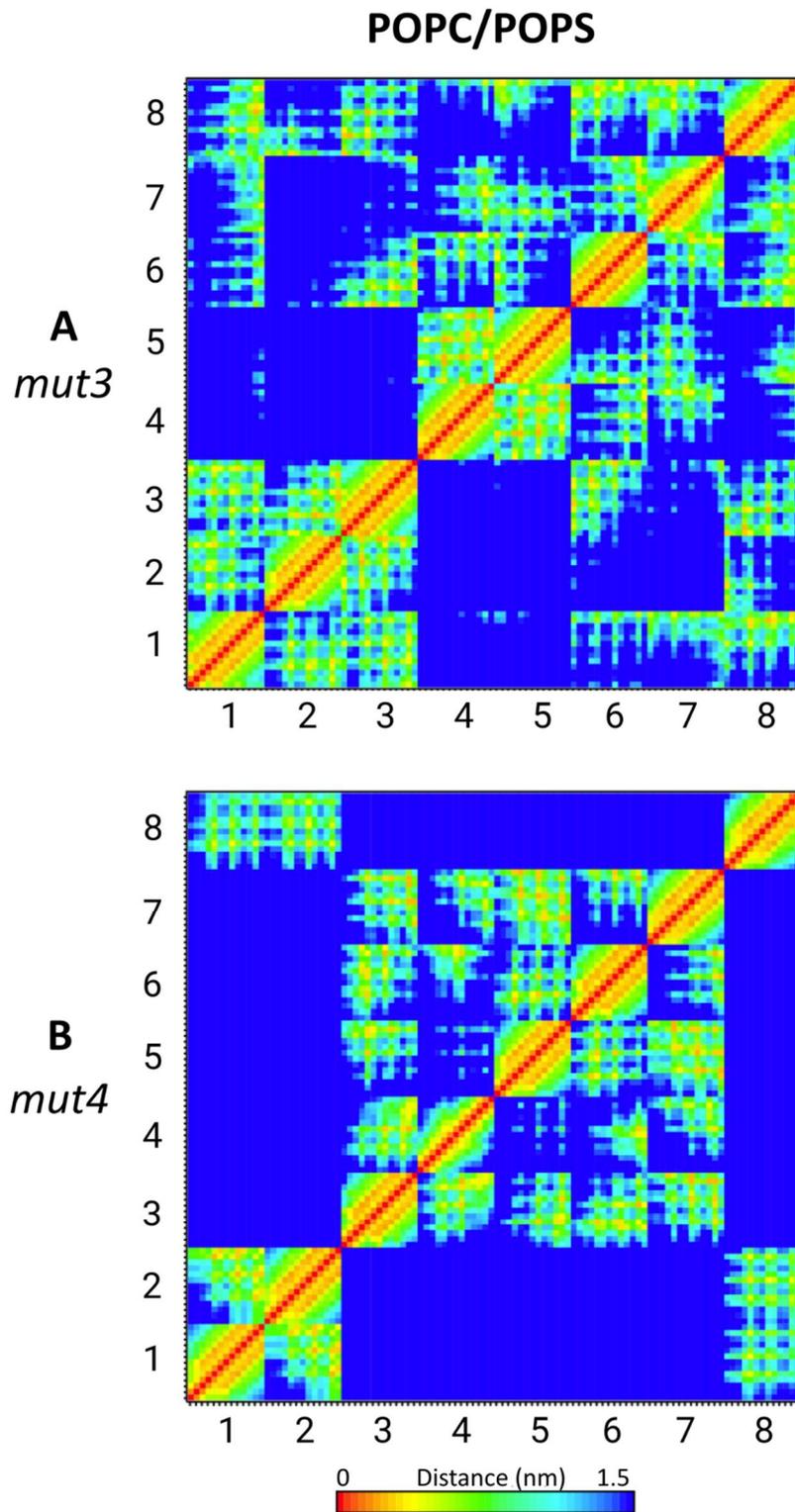


## *mut4*

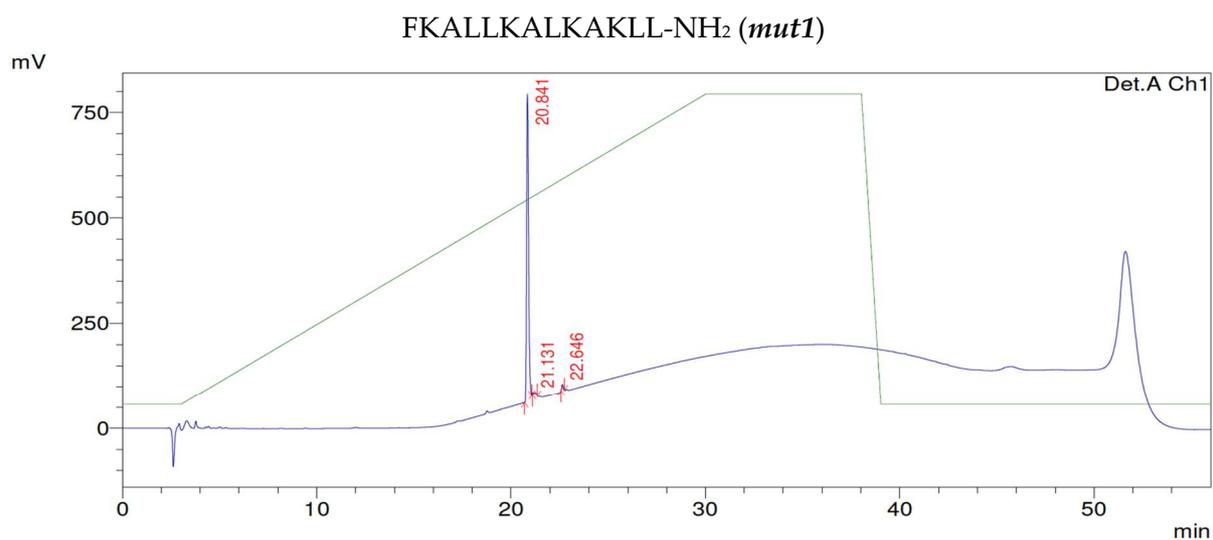


**Figure S8.** Area per lipid (nm<sup>2</sup>) in bilayers containing POPC and POPS as calculated from MD simulations in the presence of eight peptides of *mut3* (A,B) and *mut4* (C,D). The average value is shown in blue, while the upper and lower leaflet are shown in yellow and red, respectively.





**Figure S10.** Contact maps in simulated POPC/POPS systems when eight peptides of *mut3* (A) and *mut4* (B) are present.

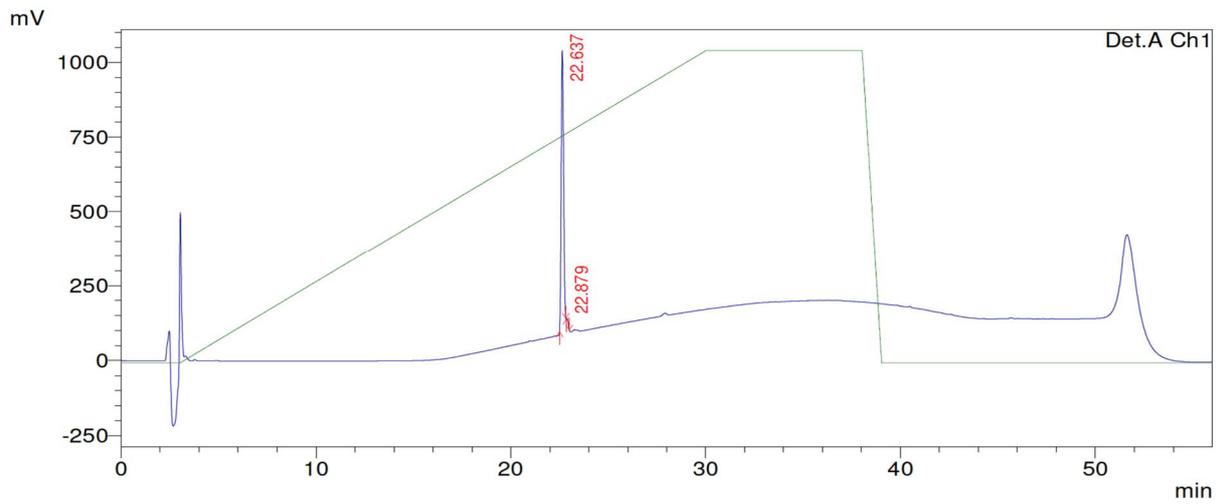


Detector A Ch1 210nm

Peak Purity Index	Ret. Time	Area	Height	Area %	Height %
	20.841	5065200	725793	97.472	97.346
	21.131	52512	5500	1.011	0.738
	22.646	78835	14284	1.517	1.916
		5196547	745577	100.000	100.000

**Figure S11.** Analytical purity of *mut1*. HPLC C12 column (Phenomenex® C12, Jupiter 4  $\mu$  Proteo, 90 Å, 250  $\times$  4.6 mm) using a mixture of aqueous 0.1% (*v/v*) TFA (**A**) and 0.1% (*v/v*) TFA in acetonitrile (**B**) as the mobile phase (flow rate of 1 mL/min) and employing UV detection at 210 nm.

FAKLLAKLAKLLK-NH<sub>2</sub> (*mut2*)

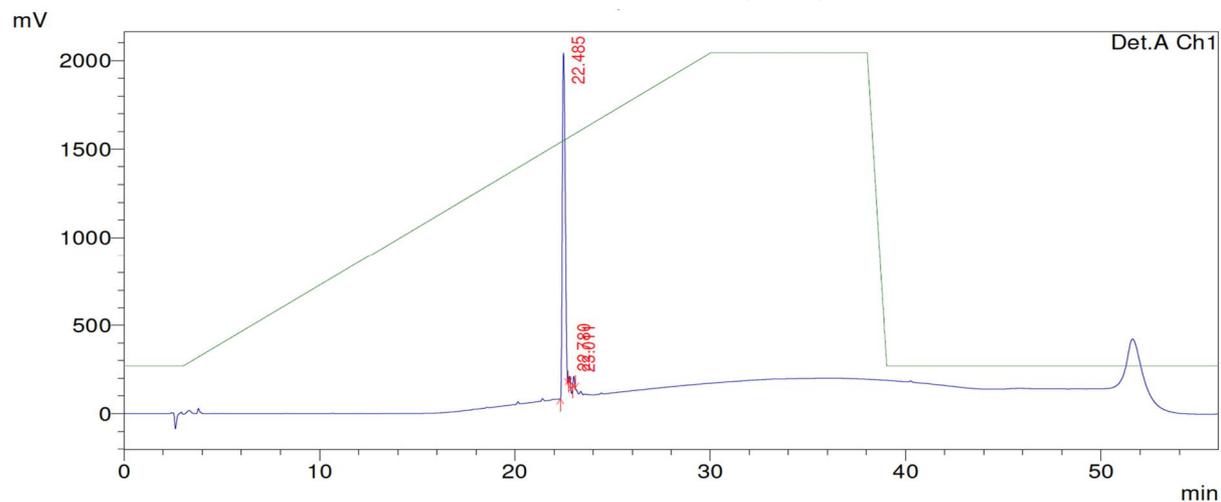


Detector A Ch1 210nm

Peak Purity Index	Ret. Time	Area	Height	Area %	Height %
	22.637	7822238	924470	99.316	98.977
	22.879	53894	9558	0.684	1.023
		7876132	934028	100.000	100.000

**Figure S12.** Analytical purity of *mut2*. HPLC C12 column (Phenomenex® C12, Jupiter 4  $\mu$  Proteo, 90 Å, 250  $\times$  4.6 mm) using a mixture of aqueous 0.1% (*v/v*) TFA (**A**) and 0.1% (*v/v*) TFA in acetonitrile (**B**) as the mobile phase (flow rate of 1 mL/min) and employing UV detection at 210 nm.

FAKLLAKLARLL-NH<sub>2</sub> (*mut3*)

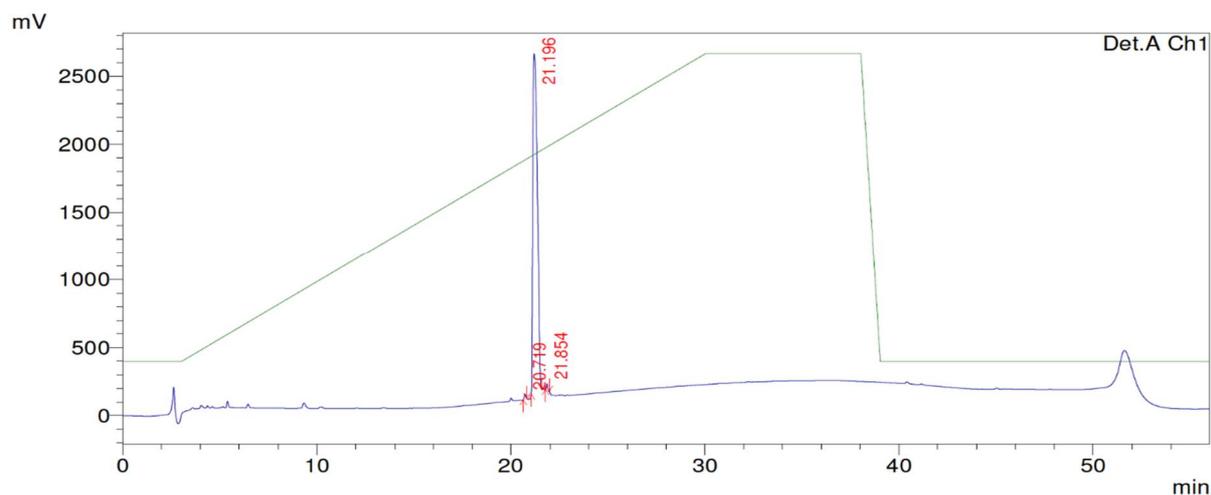


Detector A Ch1 210nm

Peak Purity Index	Ret. Time	Area	Height	Area %	Height %
	22.485	19782282	1922920	97.975	95.341
	22.780	140362	35185	0.695	1.745
	23.011	268462	58779	1.330	2.914
		20191106	2016884	100.000	100.000

**Figure S13.** Analytical purity of *mut3*. HPLC C12 column (Phenomenex® C12, Jupiter 4  $\mu$  Proteo, 90 Å, 250  $\times$  4.6 mm) using a mixture of aqueous 0.1% (*v/v*) TFA (**A**) and 0.1% (*v/v*) TFA in acetonitrile (**B**) as the mobile phase (flow rate of 1 mL/min) and employing UV detection at 210 nm.

AAKLLAKLAKLL-NH<sub>2</sub> (*mut4*)



Detector A Ch1 210nm

Peak Purity Index	Ret. Time	Area	Height	Area %	Height %
	20.719	198306	38021	0.477	1.472
	21.196	41137147	2498505	98.977	96.734
	21.854	226985	46343	0.546	1.794
		41562438	2582869	100.000	100.000

**Figure S14.** Analytical purity of *mut4*. HPLC C12 column (Phenomenex® C12, Jupiter 4  $\mu$  Proteo, 90 Å, 250  $\times$  4.6 mm) using a mixture of aqueous 0.1% (*v/v*) TFA (A) and 0.1% (*v/v*) TFA in acetonitrile (B) as the mobile phase (flow rate of 1 mL/min) and employing UV detection at 210 nm.