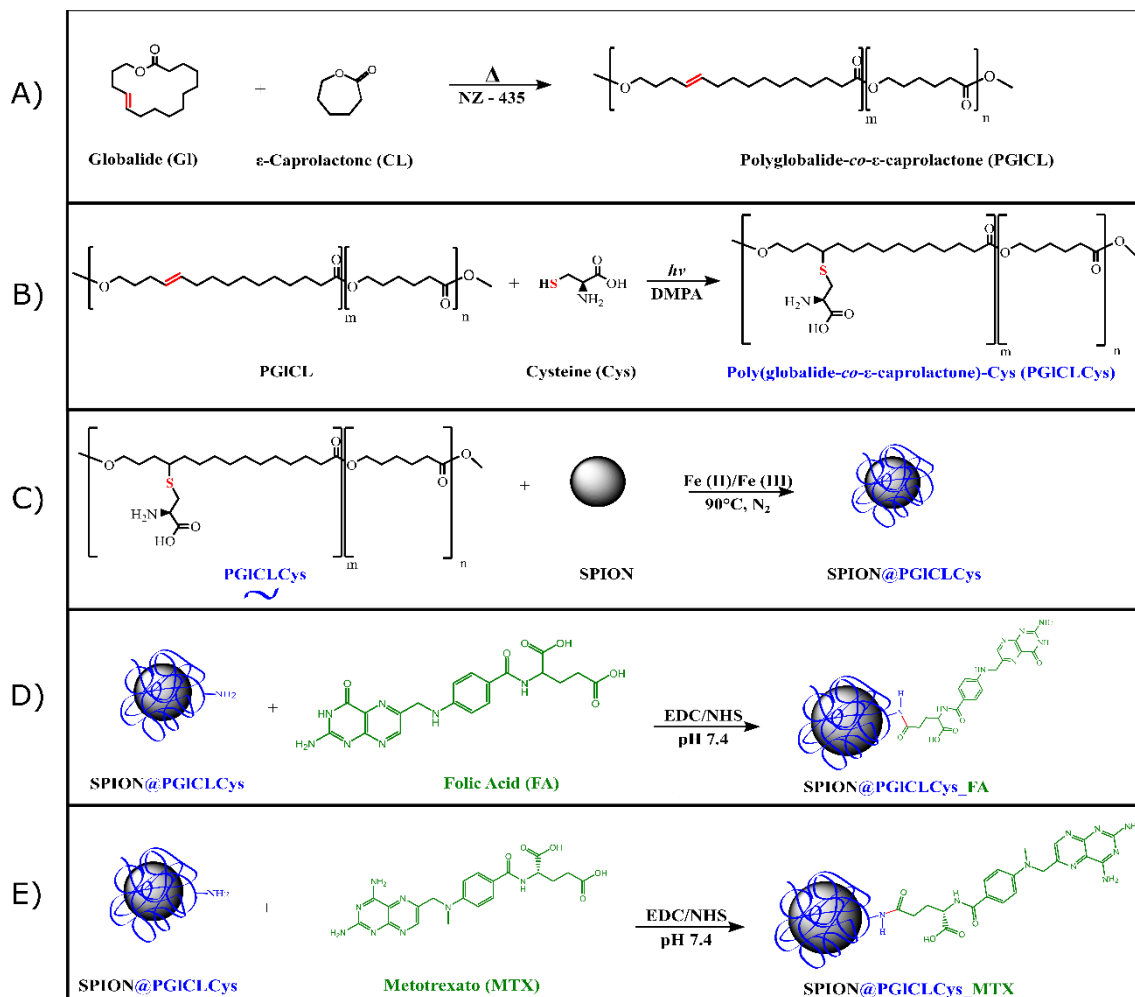


# Coating of SPIONs with a Cysteine-Decorated Copolyester: A Possible Novel Nanoplatfom for Enzymatic Release

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**Figure S1.** Representative scheme of the reaction steps to obtain PGICL and its modification with cysteine followed by coating of SPIONs. A) Enzymatic Ring-Opening Polymerization reaction (e-ROP). B) Modification reaction with the amino acid cysteine (Cys) via thiol-ene reaction to obtain PGICLCys. C) Synthesis and stabilization of SPIONs with PGICLCys. D) Conjugation of SPIONs with FA. E) Conjugation of SPIONs with MTX.

**Table S1.** Thiol-ene reaction conversion calculated based on the consumption of the double bonds present in PGICL chains, determined by  $^1\text{H}$  NMR.

Sample	Integration by corresponding peak (ppm)			Double bond Conversion (%)
	Double Bond (5.40 ppm)	Reference (4.06 ppm)	Cysteine (2.90 – 2.70 ppm)	
PGICL	1.00	1.00	-	-
PGICLCys	0.82	1.00	3.00	18%

**Table S2.** Thermal properties of the polymers, determined by DSC.

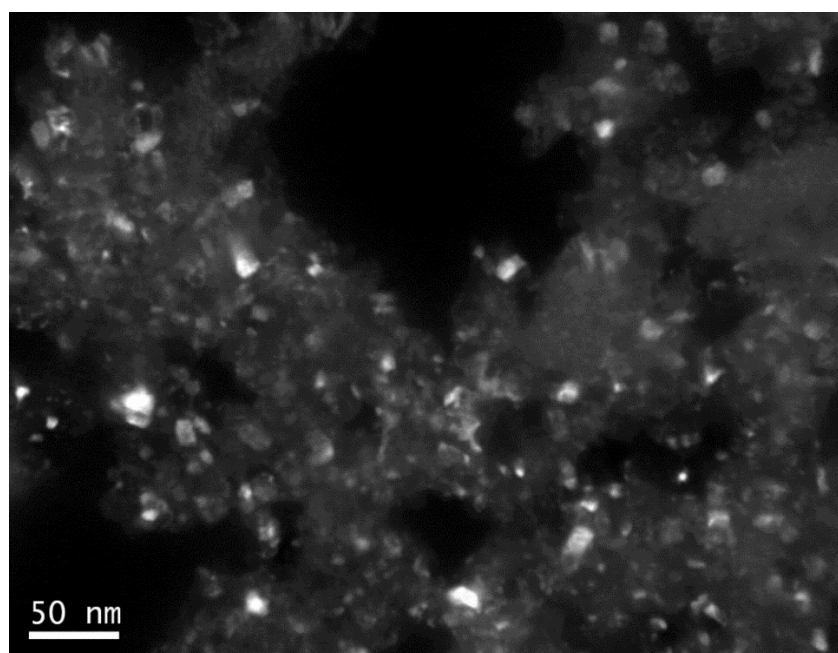
Sample	T <sub>m</sub> (°C)	$\Delta H$ (J/g)	X <sub>c</sub> (%)
PGICL	38	59.22	44
PGICLCys	32	40.6	30

PS: the degree of crystallinity (X<sub>c</sub>) is calculated based on the heat of fusion of a PCL 100% crystalline sample<sup>1</sup>.

Table S3 presents the calculated Gibbs free energy ( $G$ ) and the Gibbs free energy of solvation ( $G_{solv}$ ) in gas-phase, water and n-octanol.  $\Delta G_{solv}$  was calculated as the difference between the Gibbs free energy of solvation and gas-phase Gibbs free energy.

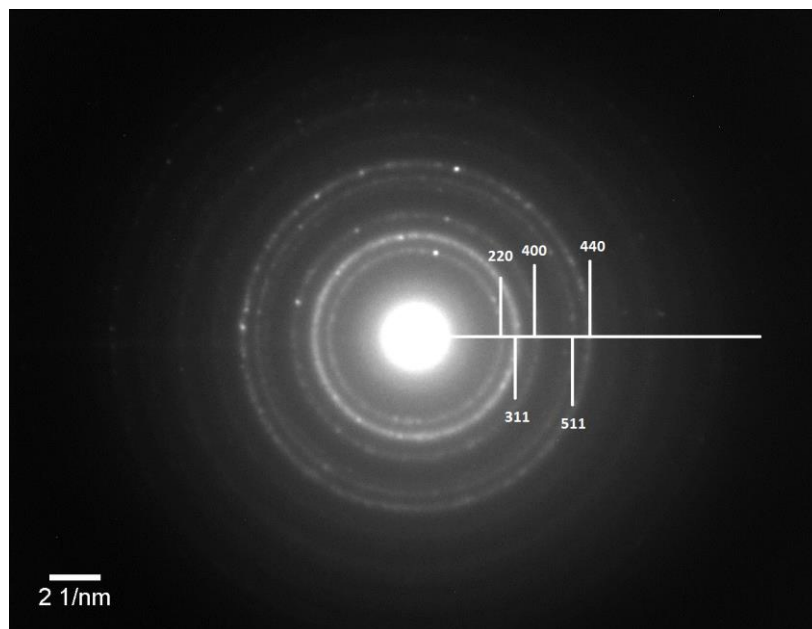
**Table S3.** Gibbs free energy calculated at 1 atm and 25 °C using DFT/B3LYP/6-31G\*\* with water and n-octanol solvents in SMD model.

Compound	Gas-phase	Water		n-octanol	
	$G$ (Kcal/mol)	$G_{solv}^W$ (Kcal/mol)	$\Delta G_{solv}^W$ (Kcal/mol)	$G_{solv}^O$ (Kcal/mol)	$\Delta G_{solv}^O$ (Kcal/mol)
PGICL	344.3180	348.5650	4.2473	341.0100	3.3081
PGICLCys	407.4900	416.9260	9.4357	412.0820	4.5921
PGICLCys_FA	609.0233	615.9962	6.9728	614.8832	5.8599

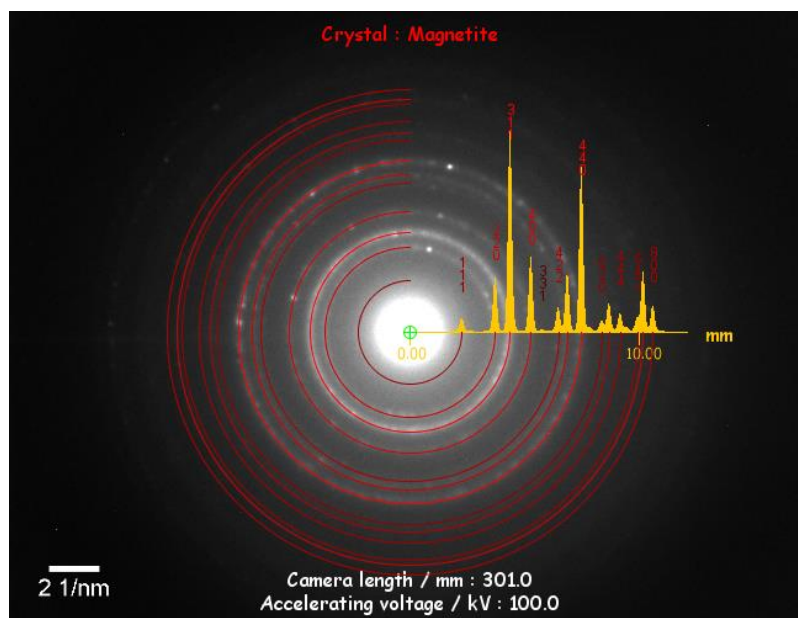


**Figure S2.** TEM dark field image showing individual crystalline particles (DF-TEM) of SPION@PGICLCys.

The dark field TEM image (Figure S2) was crucial for the identification of individual nanoparticles, enabling the measurement of the size of nanoparticles. Since the samples tends to agglomerate, the conventional bright field TEM image shows superposed nanoparticles. The dark field technique otherwise makes visible only certain particles with particular geometrical orientation glow facilitating the identification of single-particle boundaries.



**Figure S3.** Selected area electron diffraction (SAED) image showing indexed diffraction rings corresponding to magnetite crystallographic planes of SPION@PGICLCys.



**Figure S4.** SAED image with Indexed diffraction pattern of magnetite and simulated diffraction ring pattern matching the results for the SPION@PGICLCys sample.

X-ray diffraction (XRD) analysis was used to characterize the crystallite size, phase, and crystallization of SPIONS. According to the half-maximum full width (FWHM) of (311) reflections, the mean size of the  $\text{Fe}_3\text{O}_4$  nanocrystalline particles was calculated as 7.662 nm. In addition, the crystal cell dimension of (311) reflections were calculated to be

$a = 0.83615$  nm using for quantitative analysis the Rietveld approach and the GSAS software.

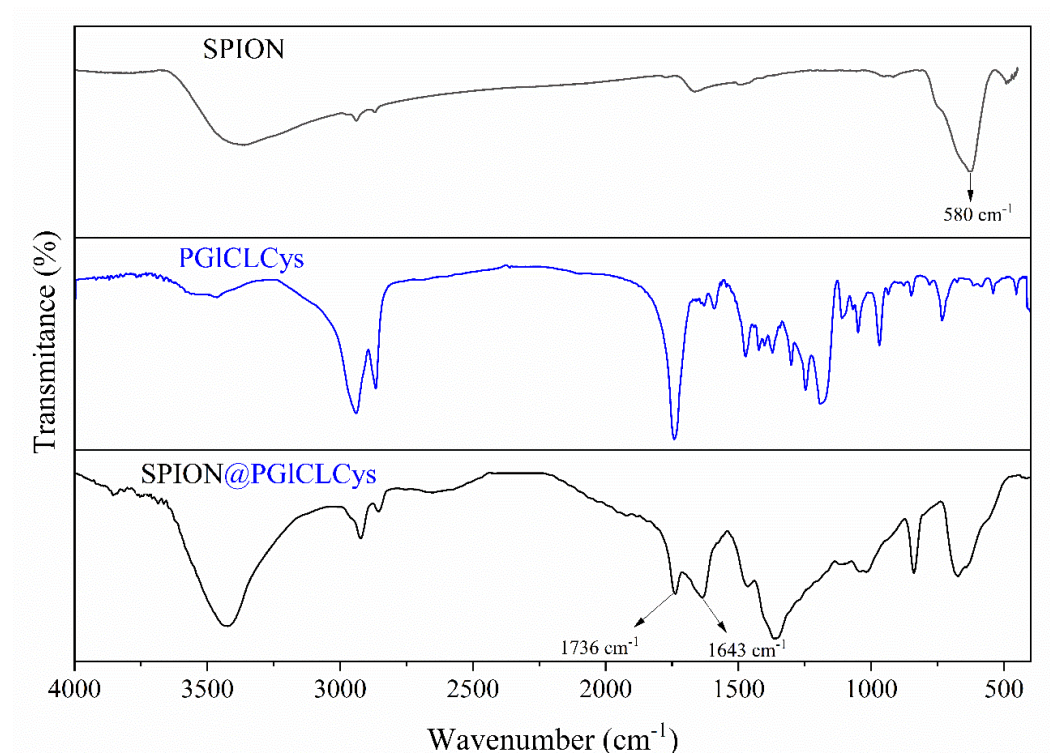
**Table S4.** Interplanar distance.

Plane (hkl)	Measured Distance d (nm)	Theoretical Distance <sup>2</sup> d (nm)
220	0.299	0.297
311	0.255	0.253
400	0.212	0.210
511	0.162	0.162
440	0.149	0.148

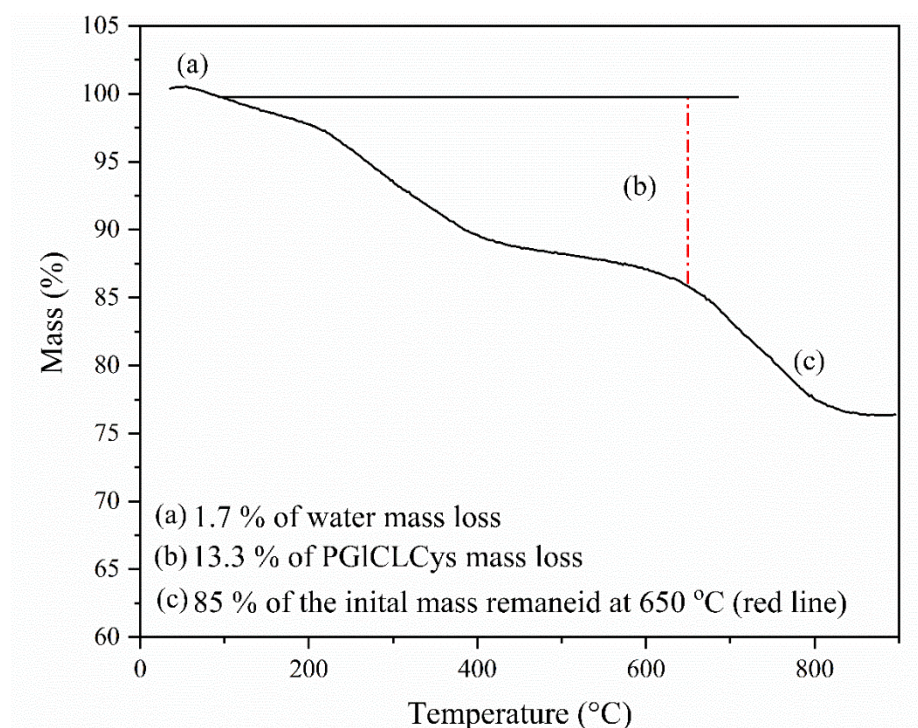
Table S4 shows the measured interplanar distances obtained by transmission electron microscopy - selected area electron diffraction (TEM-SAED). The values are compared with theoretical values of magnetite<sup>2</sup> and a good agreement is observed.

To calculate the unit cell, the plane distance, calculated with the equation for cubic cells, was used:

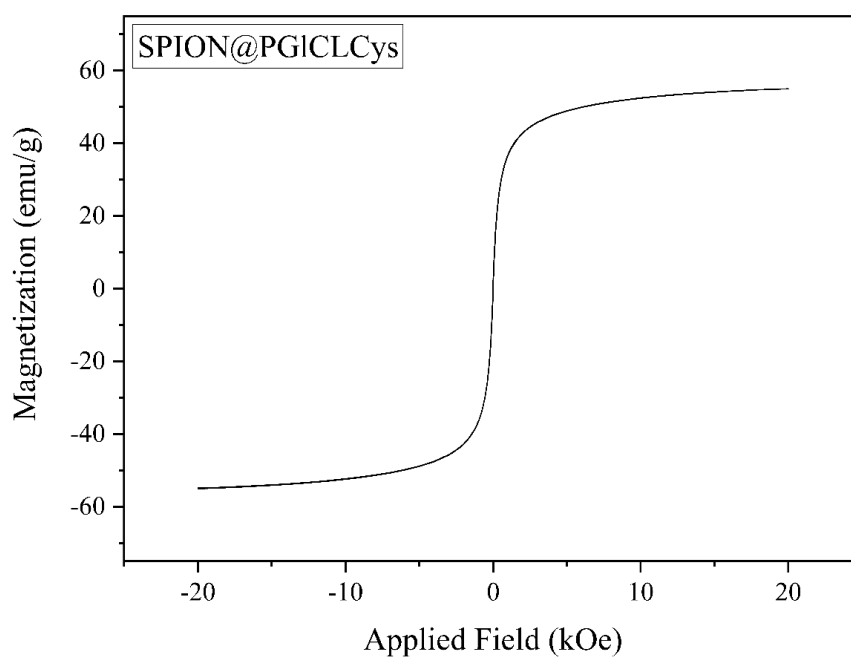
$$d_{(hkl)} = \frac{a_0}{\sqrt{h^2 + k^2 + l^2}} \quad (S1)$$



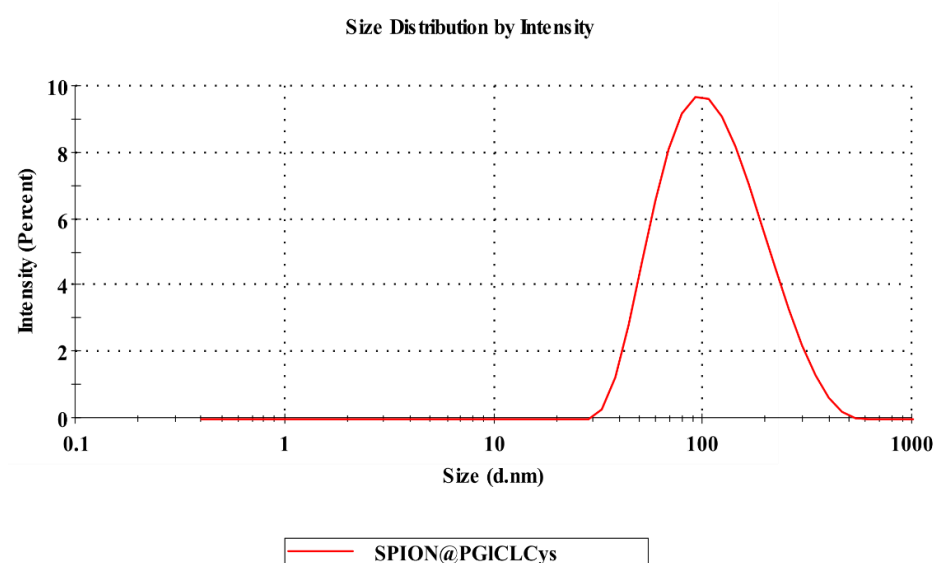
**Figure S5.** FT-IR spectra for SPIONs, and modified copolymer (PGICLCys) and after coating of magnetic nanoparticles (SPION@PGICLCys).



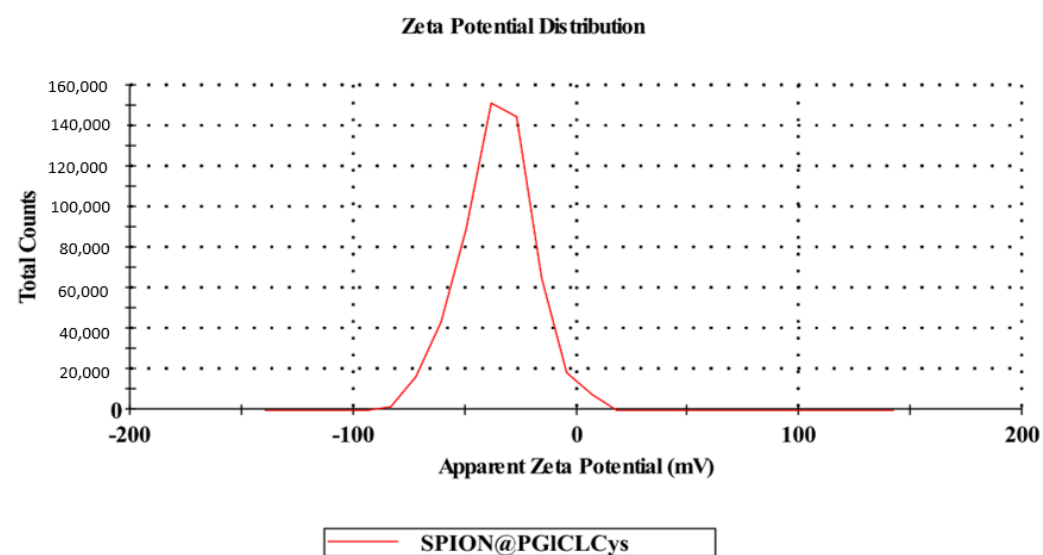
**Figure S6.** Thermogravimetric analysis of SPION@PGI/CLCys.



**Figure S7.** VSM analysis of SPION@PGI/CLCys.

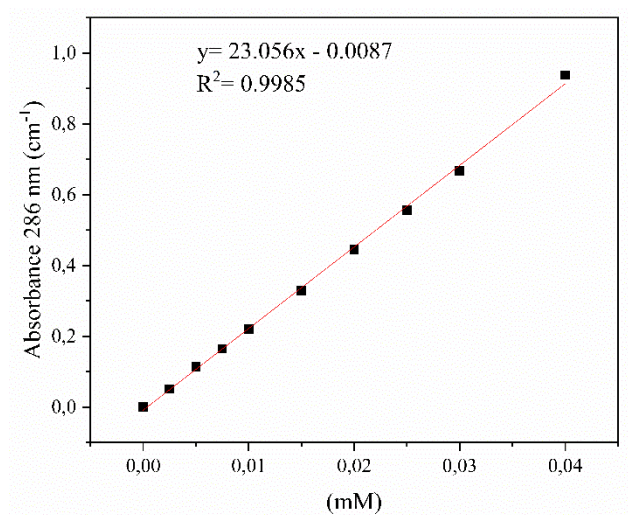


**Figure S8.** DLS analysis of SPION@PGICLCys.

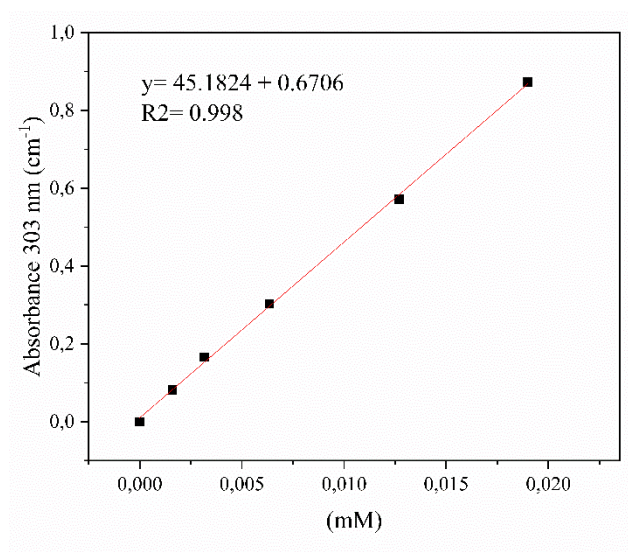


**Figure S9.** Zeta potential surface analysis of SPION@PGICLCys ( $\zeta = -35.4\text{mV}$ ) dispersed in buffer solution (pH = 8.0).

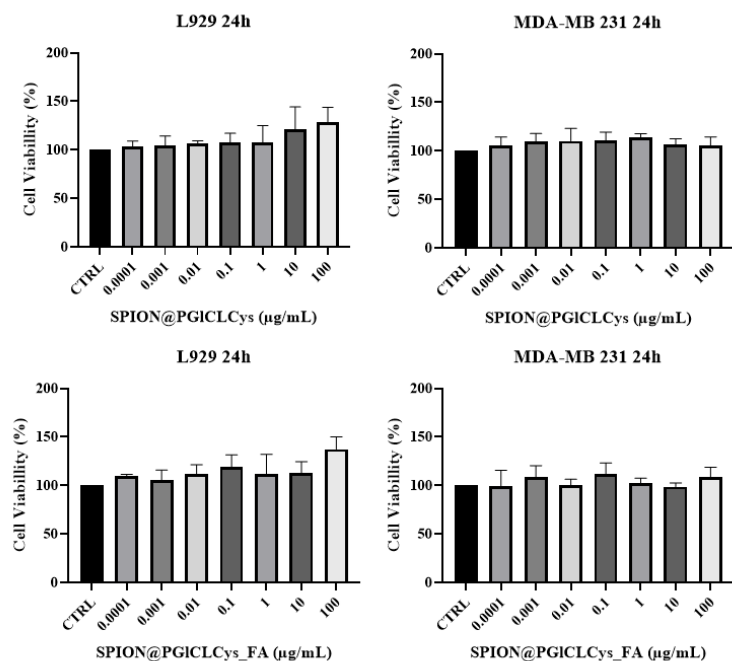
A)



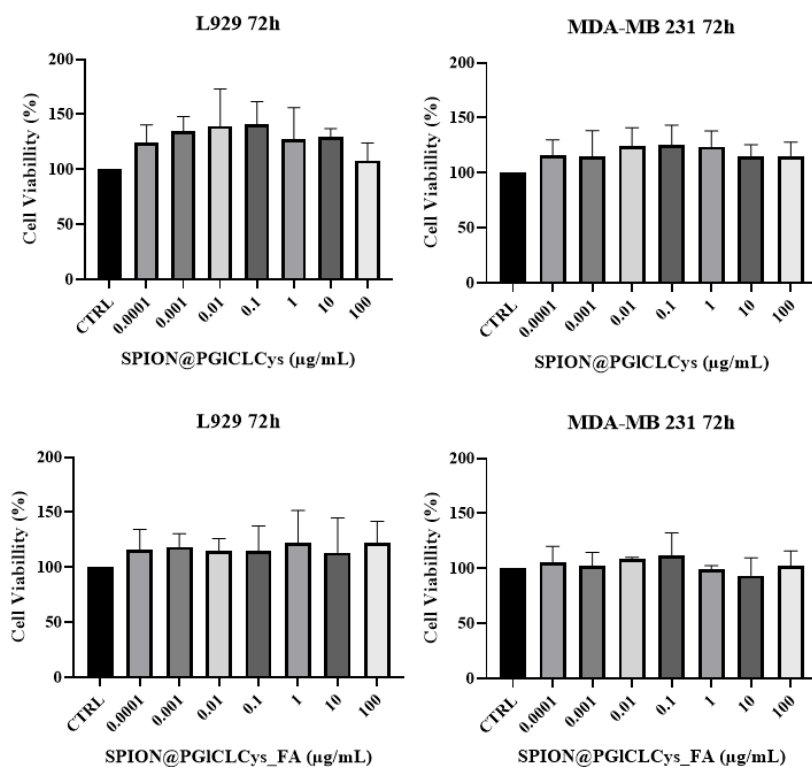
B)



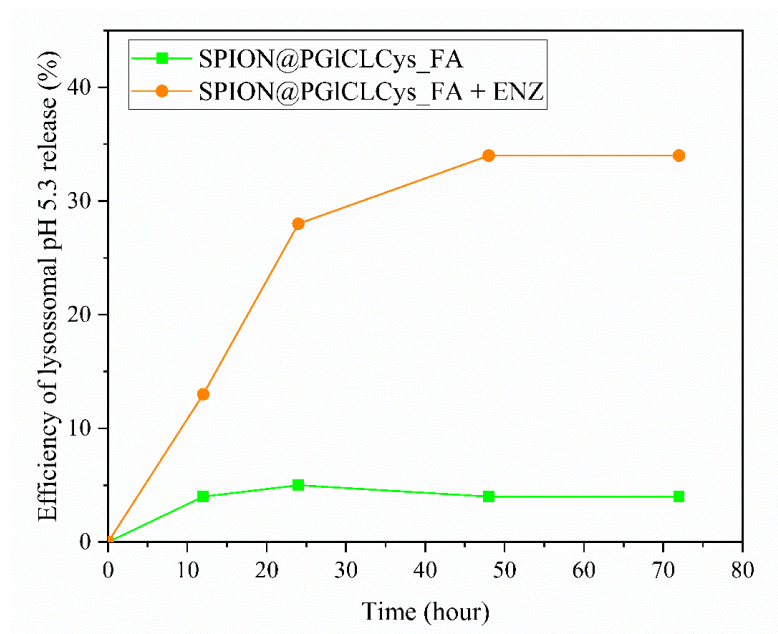
**Figure S10.** UV-vis calibration curve for: (A) folic acid (FA), and (B) methotrexate (MTX) in buffer solution (pH = 8.0).



**Figure S11:** MTT assay of SPION@PGICLCys and SPION@PGICLCys\_FA showing cells viability as a function of nanoparticles concentration (0.0001 to 100  $\mu\text{g}\cdot\text{mL}^{-1}$ ) for 24 h. All SPIONs tested at different concentrations did not exert difference (ANOVA) in relations to the control group  $n=3$ .



**Figure S12:** MTT assay of SPION@PGICLCys and SPION@PGICLCys\_FA showing cells viability as a function of nanoparticles concentration (0.0001 to 100  $\mu\text{g}\cdot\text{mL}^{-1}$ ) for 72 h. All SPIONs tested at different concentrations did not exert difference (ANOVA) in relations to the control group  $n = 3$ .



**Figure S13.** FA assay at lysosomal pH (pH 5.3).

## References

- 1 V. Crescenzi, G. Manzini, G. Calzolari and C. Borri, *Eur. Polym. J.*, **1972**, 8, 449–463.
- 2 M. E. Fleet, *Acta Crystallogr. Sect. B Struct. Crystallogr. Cryst. Chem.*, **1981**, 37, 917–920.