

Figure S1: Compliance with the intervention.

Compliance with the intervention was assessed as the proportion (%) of BJ shots consumed by each participant in each intervention group. P; participant, HN; high NO<sub>3</sub>-, MN; Medium NO<sub>3</sub>-, LN; Low NO<sub>3</sub>-, PL; Placebo.

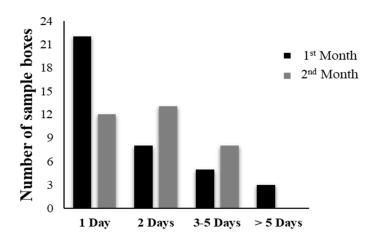


Figure S2: Distribution of number of days between samples being posted by the participants and receipt by the researcher.

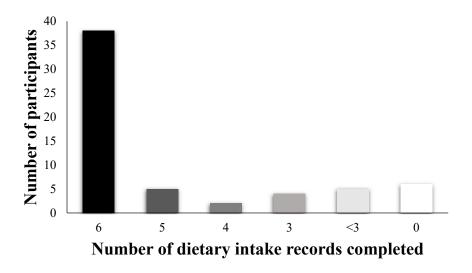


Figure S3: Number of dietary intake records completed by participants. Participants used Intake24 software to record their dietary intakes every two weeks during the trial.

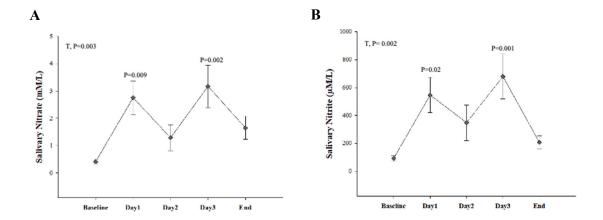


Figure S4: Mean of salivary NO<sub>3</sub><sup>-</sup> (A) and NO<sub>2</sub><sup>-</sup> (B) concentrations in LN group (low NO<sub>3</sub><sup>-</sup> dose; 70 ml of BJ every alternate days).

Statistical analysis using one factor (time) repeated measure ANOVA. Saliva samples were collected on three consecutive days at week 4 and week 8. Data collected on the week4 and week 8 were combined. Data are expressed as mean  $\pm$  SEM, (n = 50).

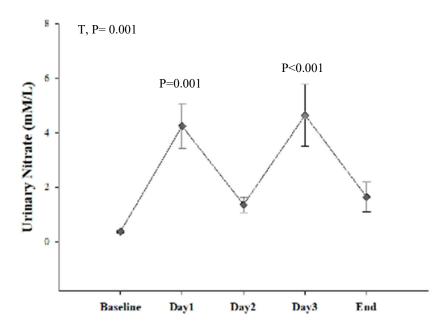


Figure S5: Mean of urinary NO<sub>3</sub><sup>-</sup> concentrations in the LN group (low NO<sub>3</sub><sup>-</sup> dose; 70 ml of BJ every alternate days).

Statistical analysis using one factor (time) repeated measure ANOVA. Urine samples were collected on three consecutive days at week 4 and week 8. Data collected on the week 4 and week 8 were combined Data are expressed as mean  $\pm$  SEM, (n = 50).

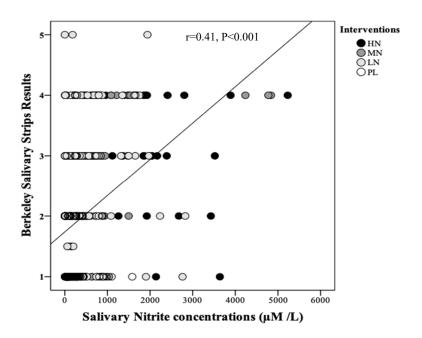


Figure S6: Scatterplot of Pearson correlation between Berkeley salivary  $NO_2^-$  strips readings and salivary  $NO_2^-$  concentrations.

Scatterplot showing Pearson correlation between Berkeley salivary NO<sub>2</sub><sup>-</sup> strips readings and salivary NO<sub>2</sub><sup>-</sup> concentrations measured by chemiluminescence; n=50. Berkeleysalivary strips results provided by the manufacturer (1=Depleted, 2=Low, 3=Threshold, 4=Target and 5=High).

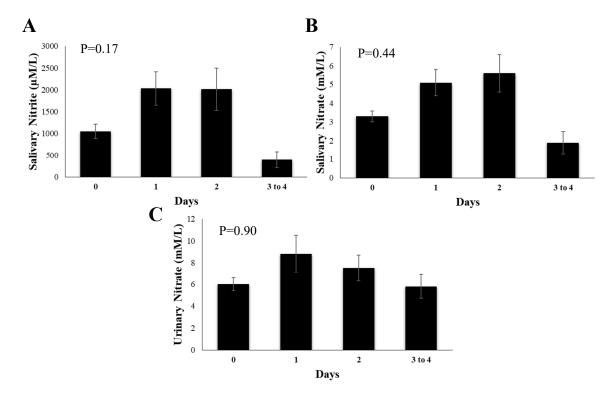
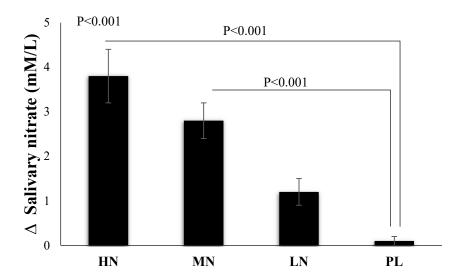


Figure S7: Salivary  $NO_2^-$ ,  $NO_3^-$  and urinary NO3- (C) concentrations. Mean salivary  $NO_2^-$  (A) and  $NO_3^-$  (B) and urinary  $NO_3^-$  (C) concentrations in samples received after a different number of days. Results presented in this analysis are based on combined data collected during the two interim monthly visits and across the 4 intervention groups. One-way ANOVA was used to test differences in biomarker concentrations between different days. Data are expressed as mean  $\pm$  SEM.



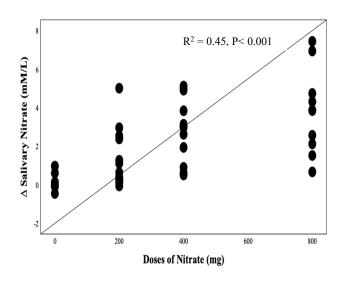


Figure S8: Mean of salivary NO<sub>3</sub>-concentrations after removing data from one participant.

# Feedback Questionnaire

## Nitrate, Cognition and Cerebral Blood Flow

Da	nte:	ID:		-
1.	Why did you join this study?			
2.	What were your expectations	from the stud	dy?	
3.	Would you like to be approac Yes □ No □	hed for anoth	ner study similar to tl	his?
	If no, why?			
	[1] Duration			
	[2] Beetroot juice taste			
	[3] Side Effects			
	[4] Other			
4_	· Would you join another stud	y if it had a k	onger duration (>3m	onths)?
•		y II it iiaa a k	onger duration (* 5 inc	onensy.
	Yes □ No □	. 1	1101	
	If yes, what is the longes	t duration of the	he study you could fol	low?months
N	<u> utritional supplementa</u>	<u>ition</u>		
1	. Do you eat or drink beetroo	t regularly?	Yes □	No □
	If yes, how many times a w			

If no, why?			
Would you recommend the If yes, why?			
Do you prefer having raw		oot juice if you	
Why?			
Which one of the followin	g interventions did you	ı follow?	
[1] One bottle per day			
[2] One bottle every 2 d [3] 2bottles per day	lays 🗆		
Was it difficult to drink tl	his dose of beetroot juic	ce during the st	tudy?
Yes □ No			
If Yes, please explain why?			
Have you felt any differen	uce as a result of the be		
If so, please explain?			
If any, what was the most	inconvenient aspect of	taking the bee	troot juice?
Would you drink beetroo If No, why?	t juice regularly?	Yes □	No □
[1] Side effects $\Box$			

ave you forgotten/missed any dose	s?	Yes □	No □
What was the reason?			
Measurement protocols			
s the ONT protocol (collection of s	saliva samples	s) easy to foll	low?
Yes □ No □			
If no, why?			
<ul><li>[1] Too detailed</li><li>[2] Too many samples required</li><li>[3] Inconvenient</li><li>[5] Fasting</li><li>[6] Other</li></ul>			
l you have any difficulties with ON Yes □ No □	1 Conceion?		
If yes, why?			
[1] Chewing cotton balls			
[2] Collecting Saliva			
[3] Other			
you have any difficulties recording	ng the time at	which you to	ook vour heet
ee every day?	s me ame at		our jour beet
Yes □ No □			

4. Did you have any difficulties recording your 24h recall diet and using the software (INTAKE 24) every two weeks?

Yes □	No □
If yes, why?	
_	saliva, urine and saliva strips every 4 weeks for 3 days acceptable for
•	No □
If No, why	
Did you feel an	y difficulty to post them?
Yes □	No □
If yes, explain v	why
•	ne general procedure of vascular assessment acceptable for you?
r es 🗀	NO L
If no, why?	
Did you find the you?	ne general procedure of cerebral blood flow assessment acceptable for
Yes □	No □
If no, why?	
Did you find the for you?	ne cognitive function assessment using COMPASS software acceptable
Yes □	No □
If no, why?	
	Was collecting you? Yes □ If No, why  Did you feel are Yes □ If yes, explain vertical the Yes □ If no, why?  Did you find the Yes □ If no, why?  Did you find the you? Yes □ If no, why?

10	10. Would be acceptable for you if the tests to assess cognitive function last more than				
30	30 minutes?				
	Yes □ No □				
<u>St</u>	zudy design				
1.	Was the number of visits appropriate?				
	[1] Appropriate □ [2] Too many □				
	If 2, please explain why:				
2.	What were the main factors which motivated you to participate and complete this				
	idy?				
	[1] Commitment to the study				
	[2] Health benefits of beetroot				
	[3] Challenges of the cognitive function tasks				
	4 Interested in nutritional research				
	[5] Other				
	Do you have any other comment?				

Thank you for your time!

### **Supplemental methods**

#### Ozone-based chemiluminescence

Before analysis, plasma samples were deproteinised using cold ethanol that was first chilled to 0 °C. 250 μl of each plasma sample was placed in a 1.5 ml microcentrifuge tube and 500 μl of cold ethanol was added and the tubes were then vortexed for 10 seconds. The tubes were then centrifuged at 14,000 RPM for 5 minutes. The supernatant was removed to a pre-labelled tube. Urine and saliva samples were diluted to 1:100 in deionized water prior to the analysis. Deproteinised plasma and diluted saliva samples were used for the determination of NO<sub>3</sub> and NO<sub>2</sub><sup>-</sup>, while diluted urine samples were used for the determination of NO<sub>3</sub><sup>-</sup> by ozone based chemiluminescence (NOA 280i, Analytix, UK). The standard curve for both NO<sub>3</sub><sup>-</sup> and NO<sub>2</sub><sup>-</sup> was created to calculate the concentrations of NO<sub>3</sub><sup>-</sup> and NO<sub>2</sub><sup>-</sup>. The diluted samples were injected into the purge vessel with an injection volume ranging from 10 µl to 100 µl. The quality of the peaks was checked visually. Samples were re-diluted with a higher dilution factor (1:20) if no peaks were detected. The area under the curve of each peak was used to calculate the concentration of NO<sub>3</sub><sup>-</sup> and NO<sub>2</sub><sup>-</sup>. Samples were analysed in singlicate and analyses were repeated for samples with low quality peaks. The quality of peaks was checked visually based on height, fronting and tailing. The calculations were based on the AUCs calculated automatically for each peak by the software which minimise the risk of between-operator differences in the calculation of the results.

#### Salivary nitrite strips

Berkeley strips (Berkeley Test®, CA, USA) were used as per the manufacturer's guidelines. Specifically, participants were asked to place the test strip with the 'saliva here' side on the tongue and swab it over a 10 s period covering different areas including the dorsal surface of the tongue. The two ends of the strip were folded and pressed gently for 10 s. The colour of the NO test pad was then allowed to develop over a 45 s period. The intensity of the colour was compared with a colour chart included in the product package (Depleted, Low, Threshold, Target and High), with darker colours corresponding to higher NO2<sup>-</sup> concentrations. For the purpose of quantitative analysis, categorical colours were given a categorical number from 1 to 5, where 1 corresponding to "Depleted" and 5 corresponding to "High". Participants were asked to repeat a similar procedure at home as required by the study protocol.