

## **Materials and methods**

### **Reagents**

PTSO (93% purity) has been developed, isolated and provided by DOMCA S.A. (Granada, Spain). Diets were provided by Scientific Animal Food & Engineering (Augy, France). All the other reagents have been purchased from Merck Life Science S.L.U. (Madrid, Spain), unless another provider is specified.

### **Glucose tolerance test and plasma determinations**

50% glucose solution in water was administered intraperitoneally (2 g/kg) and a blood sample was obtained from the tail vein at 0, 15, 30, 60 and 120 min. Glucose was measured with a Contour Next glucometer (Ascensia, Barcelona, Spain).

Glucose and cholesterol were measured by colorimetric methods using Spinreact kits (Spinreact, S.A., Girona, Spain). Plasma insulin was quantified by ELISA (Alpco Diagnosis, Salem, MA, USA), following the instructions provided by the manufacturers.

The Insulin Resistance Index was calculated according to the homeostatic model (HOMA-IR) and plasmatic lipopolysaccharide (LPS) levels were quantified by the Pierce™ Chromogenic Endotoxin Quant Kit (Thermo Scientific, Inc., Waltham, MA, USA).

### **NADPH oxidase activity**

The aortic rings were incubated for 30 minutes at 37°C in HEPES-containing physiological salt solution (pH 7.4). Aortic O<sub>2</sub><sup>-</sup> production was stimulated by addition of NADPH (100 μM). Rings were then placed in tubes containing physiological salt solution, with or without NADPH, and lucigenin was injected automatically at a final concentration of 5 μmol/L to avoid known artifacts when used at higher concentration. NADPH oxidase activity were determined by measuring luminescence over 200 s in a scintillation counter (Lumat LB 9507, Berthold, Germany) in 5-s intervals and was calculated by subtracting the basal values from those in the presence of NADPH. Vessels were then dried, and dry weight was determined. NADPH oxidase activity is expressed as relative luminescence units (RLU)/min/mg dry aortic ring.

## Flow cytometry

Both tissues were isolated excised, washed and cut into small pieces ( $\leq 3 \text{ mm}^3$ ) using sterile scissors or scalpel blades. The fat minced tissue was resuspended in a HBSS solution containing 1 mg/mL collagenase Type I. These were transferred into a 50 mL tube and placed in a 37 °C water bath for 30 min. After that, the digested tissue solution was filtered using 100  $\mu\text{m}$  cell strainers. The pellet was resuspended in HBSS to wash it before passing it through a 70  $\mu\text{m}$  cell strainer. Then, the red blood cells were removed using a red blood cell lysis buffer for 5 minutes at room temperature. Cells were centrifuged and the pellet was resuspended in HBSS medium to wash it. The harvested cells were stained for different markers (Miltenyi Biotec GmbH, Bergisch Gladbach, Germany), unless otherwise stated.  $2 \times 10^6$  cells were counted and submitted to Fc $\gamma$ R blocking. After that, the cells were stained in the surface with Viability Dye (eFluor 780, REF. 65-0865-14; eBioscience, San Diego, USA), anti-CD3 (AmCyan, Clone 145-2C11), anti-CD4 (APC-Vio 770, Clone REA1211), anti-CD11b (APC-Vio 770, Clone REA592), anti-CD11c (FITC, Clone N418) and LyC6 (PE-Vio770, Clone 1G7.G10) for 15 min at 4 °C in the dark. The cells were then fixed with the Flow Cytometry Fixation Buffer (R&D Systems®, Catalog #FC004) Data collection was performed using a flow cytometer CANTO II (BD Biosciences, New Jersey, USA) and analyzed by Flowjo (Flowjo LLC, Ashland, Oregon, USA).

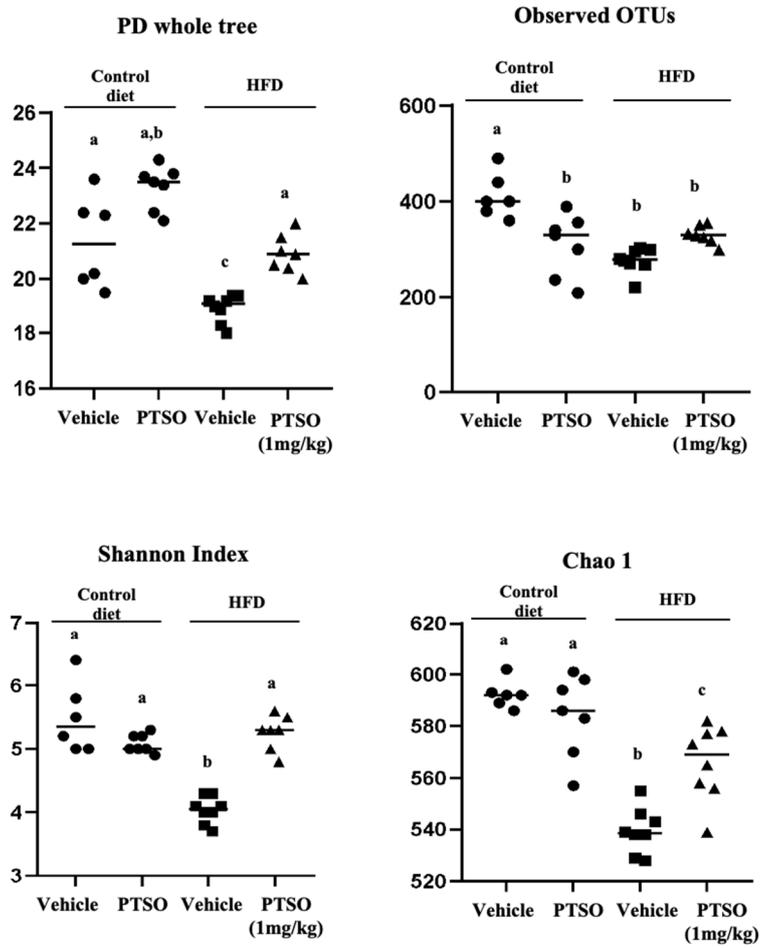
## Tables

**Table S1.** qPCR primer sequences.

Gene	Organism	Sequence 5'- 3'	Annealing T °C
<i>Adiponectin</i>	Mouse	FW: GCGAGAGTTTCTGGCAGAGT RV: CCGAGATAGCTGCCAGAGTT	60
<i>Ampk</i>	Mouse	FW: GACTTCCTTCACAGCCTCATC RV: CGCGCGACTATCAAAGACATACG	60
<i>Gapdh</i>	Mouse	FW: CCATCACCATCTTCCAGGAG RV: CCTGCTTCACCACCTTCTTG	60
<i>Glut 4</i>	Mouse	FW: GAGAATACAGCTAGGACCAGTG RV: TCTTATTGCAGCAGCGCCTGAG	62
<i>Il-1<math>\beta</math></i>	Mouse	FW: TGATGAGAATGACCTCTTCT RV: CTTCTTCAAAGATGAAGGAAA	60
<i>Il-6</i>	Mouse	FW: TAGTCCTTCCTACCCCAATTTCC RV: TTGGTCCTTAGCCACTCCTTCC	60

<i>Jnk-1</i>	Mouse	FW: GATTTTGGACTGGCGAGGACT RV: TAGCCCATGCCGAGAATGA	60
<i>Jnk-2</i>	Mouse	FW: TTGTGCTGCTTTTGATACAGTTCTTGGG RV: CTGGAAAGAGCTCTTCAAATTTGAT	60
<i>Leptin</i>	Mouse	FW: AGATCCCAGGGAGGAAAATG RV: TGAAGCCCAGGAATGAAGT	60
<i>Leptin R</i>	Mouse	FW: GCTATTTTGGGAAGATGT RV: TGCCTGGGCCTCTATCTC	60
<i>Muc-2</i>	Mouse	FW: GCAGTCCTCAGTGGCACCTC RV: CACCGTGGGGCTACTGGAGAG	60
<i>Muc-3</i>	Mouse	FW: CGTGGTCAACTGCGAGAATGG RV: CGGCTCTATCTCTACGCTCTCC	60
<i>Occludin</i>	Mouse	FW: ACGGACCCTGACCACTATGA RV: TCAGCAGCAGCCATGTACTC	56
<i>Ppar-<math>\alpha</math></i>	Mouse	FW: TCGAGGAAGGCACTACACCT RV: TCTTCCCAAAGCTCCTTCAA	60
<i>Ppar-<math>\gamma</math></i>	Mouse	FW: ACGATCTGCCTGAGGTCTGT RV: CATCGAGGACATCCAAGACA	60
<i>Tlr4</i>	Mouse	FW: GCCTTTCAGGGAATTAAGCTCC RV: AGATCAACCGATGGACGTGTAA	60
<i>Tnf-<math>\alpha</math></i>	Mouse	FW: AACTAGTGGTGCCAGCCGAT RV: CTTACAGAGCAATGACTCC	60
<i>Zo-1</i>	Mouse	FW: GGGGCCTACACTGATCAAGA RV: TGGAGATGAGGCTTCTGCTT	56

## Figures



**Figure S1.** Impact of PTSO administration on microbiome diversity (PD whole tree, Observed OTUs, Shannon index and Chao1). Groups with different letters statistically differ ( $P < 0.05$ ).