# Supplementary Materials: Transport-Mediated Oxaliplatin Resistance Associated with Endogenous Overexpression of MRP2 in Caco-2 and PANC-1 Cells 

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(A)


Marker \%Gated X-Med X-AMean X-GMean

| $\square$ All | 100.00 | 0.68 | 0.82 | 0.67 |
| :--- | :--- | :--- | :--- | :--- |
| $\square$ All | 100.00 | 1.98 | 2.12 | 1.93 |

(C)


Marker \%Gated X-Med X-AMean X-GMean

| All | 100.00 | 0.87 | 0.88 | 0.81 |
| :--- | :--- | :--- | :--- | :--- |
| All | 100.00 | 1.44 | 1.52 | 1.43 |

(B)


| Marker $\%$ Gated | X-Med | X-AMean | X-GMean |  |
| :--- | ---: | ---: | ---: | ---: |
| $\square$ All | 100.00 | 0.79 | 0.81 | 0.76 |
| $\square$ All | 100.00 | 1.45 | 1.51 | 1.43 |

(D)


Figure S1. MRP2 surface staining flow cytometry histogram data in Caco-2 cells. Data represent flow cytometry histogram of cell surface staining using the ant-MRP2 primary antibody and isotype control IgG2a on Caco-2 and siRNA transfected Caco-2 cells. Graphs show the fluorescence intensity in (A) Caco-2 cells stained with MRP2 antibody and IgG2a stained cells; and (B, C, D) ABCC2-siRNAs transfected Caco-2 cell compared with control-siRNA cells. Both the primary antibody and isotype control were labelled with Alexa Fluor 488 secondary antibody. The $x$-axis is the fluorescence signal intensity in the FL1 blue laser channel displayed in a liner $\log$ scale. The $y$-axis represents the cell counts.
(A)


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(C)
(B)

(D)

|  | Marker X-Med X-AMean X-GMean |  | Marker $\%$ Gated X-Med X-AMean X-GMean |  |  |  |  |  |
| :--- | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| $\square$ All | 0.80 | 0.84 | 0.71 | $\square$ All | 100.00 | 0.94 | 0.97 | 0.84 |
| All | 1.12 | 1.20 | 1.11 | $\square$ All | 100.00 | 1.14 | 1.26 | 1.15 |

Figure S2. MRP2 surface staining flow cytometry histogram data in PANC-1 cells. Data represent flow cytometry histogram of cell surface staining using the ant-MRP2 primary antibody and isotype control IgG2a on PANC-1 and siRNA transfected PANC-1 cells. Graphs show the fluorescence intensity in (A) PANC-1 cells stained with MRP2 antibody and IgG2a stained cells; and (B, C, D) ABCC2-siRNAs transfected PANC-1 cell compared with control-siRNA cells. Both the primary antibody and isotype control were labelled with Alexa Fluor 488 secondary antibody. The $x$-axis is the fluorescence signal intensity in the FL1 blue laser channel displayed in a liner $\log$ scale. The $y$-axis represents the cell counts.


Figure S3. Oxaliplatin-induced apoptosis rate in MRP2 silenced Caco-2 cells. Caco-2 cells transfected with control and ABCC2-siRNAs were treated with oxaliplatin at different concentrations ( 0,25 and $100 \mu \mathrm{M}$ ) for 2 hrs . Cells were then incubated in blank complete medium for 48 hrs and subsequently stained with Annexin-V-FITC and PI. The fluorescence intensity was measured by flow cytometry. Viable cells (V) are both Annexin-V and PI negative. At an early stage of apoptosis (Ap), the cells bind with only Annexin-V. At the late stage of apoptosis (N), the cells bind with both Annexin-V FITC and PI.


Figure S4. Oxaliplatin-induced apoptosis rate in MRP2 silenced PANC-1 cells. PANC-1 cells transfected with control and ABCC2-siRNAs were treated with oxaliplatin at different concentrations ( 25 and $100 \mu \mathrm{M}$ ) for 2 hrs . Cells were then incubated in blank complete medium for 48 hrs and subsequently stained with Annexin-V-FITC and PI and their fluorescence was measured by flow cytometry.


Figure S5. Effects of control and ABCC2-siRNA transfection on the Cp values of the reference gene GAPDH in Caco-2 (A) and PANC-1 (B) cells. All data were expressed as mean $\pm$ SEM from three independent experiments. No significant differences were detected from Dunnett's post hoc test that followed one-way ANOVA for comparisons of all ABCC2-siRNA samples to the negative control.
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