

Supplementary Materials: Global Autozygosity is Associated with Cancer Risk, Mutational Signature and Prognosis

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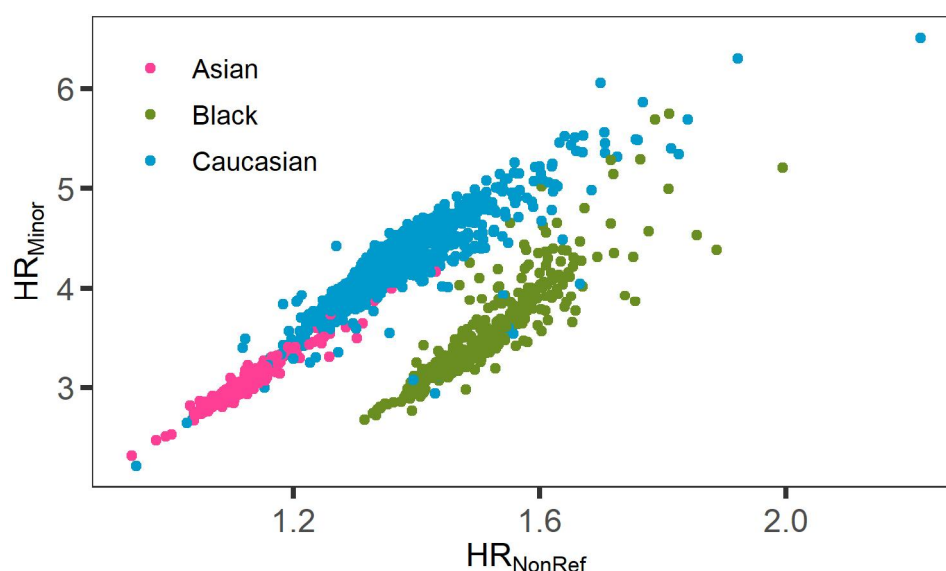


Figure S1. Scatter plot for HR_{NonRef} VS HR_{Minor} . Three clear clusters based on race can be observed. This suggests that HR is strongly associated with race.

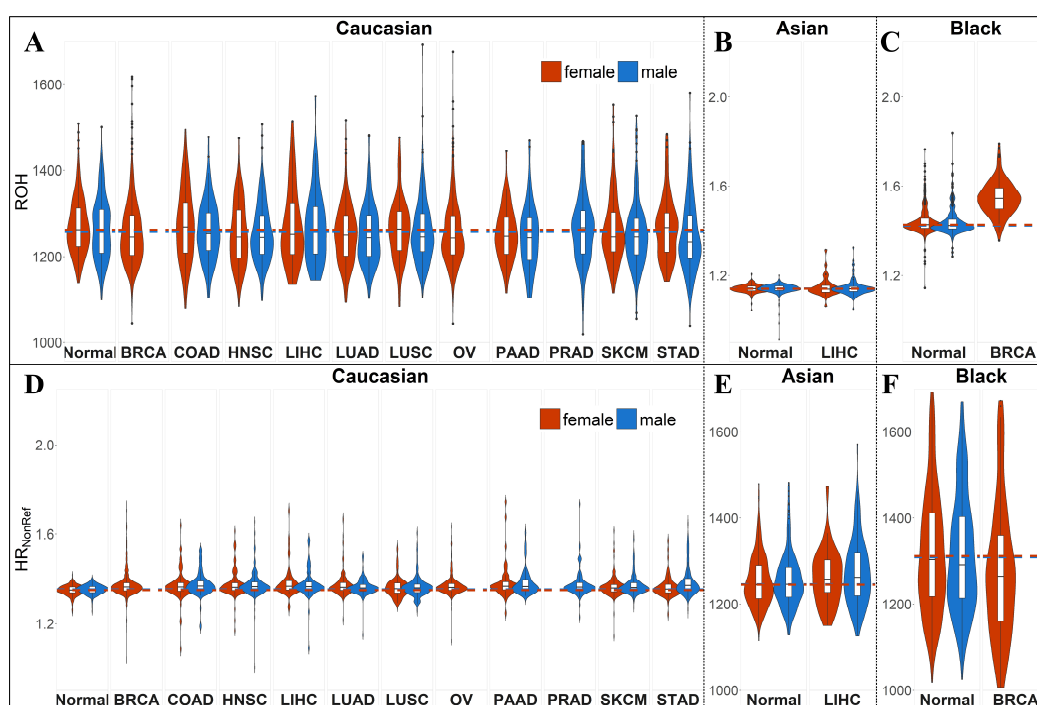


Figure S2. Comparison of HR_{NonRef} and ROH between sex across all three races tested. This is equivalent to Figure 3 from the main text without but removed data from sex chromosomes X and Y. The sex difference observed in Figure 3 were mostly gone.

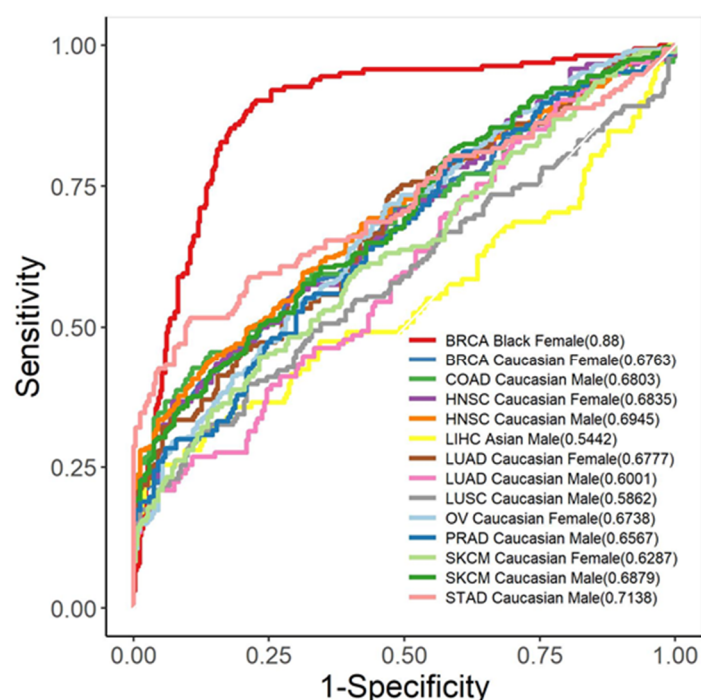


Figure S3. ROC curves based on the cancer risk analysis in Table 1.

Table S1. Cancer types encompassed in The Cancer Genome Atlas.

Cancer Abbreviation	Cancer Full Name	Sample Size
BRCA	Breast Invasive Carcinoma	883
COAD	Colon Adenocarcinoma	264
HNSC	Head and Neck Squamous Cell Carcinoma	476
LIHC	Liver Hepatocellular Carcinoma	299
LUAD	Lung Adenocarcinoma	340
LUSC	Lung Squamous Cell Carcinoma	250
OV	Ovarian Serous Cystadenocarcinoma	410
PAAD	Pancreatic Adenocarcinoma	143
PRAD	Prostate Adenocarcinoma	134
READ	Rectum Adenocarcinoma	84
SKCM	Skin Cutaneous Melanoma	455
STAD	Stomach Adenocarcinoma	319

Table S2. COSMIC mutational signatures (v3).

Signature	Etiology
SBS1	An endogenous mutational process initiated by spontaneous or enzymatic deamination of 5-methylcytosine to thymine which generates G:T mismatches in double stranded DNA. Failure to detect and remove these mismatches prior to DNA replication results in fixation of the T substitution for C.
SBS2	Attributed to activity of the AID/APOBEC family of cytidine deaminases on the basis of similarities in the sequence context of cytosine mutations caused by APOBEC enzymes in experimental systems. APOBEC3A is probably responsible for most mutations in human cancer, although APOBEC3B may also contribute (these differ in the sequence context two bases 5' to the mutated cytosine, see 1,536 mutation classification signature extraction). SBS2 mutations may be generated directly by DNA replication across uracil or by error prone polymerases replicating across abasic sites generated by base excision repair removal of uracil.
SBS3	Defective homologous recombination-based DNA damage repair which manifests predominantly as small indels and genome rearrangements due to abnormal double strand break repair but also in the form of this base substitution signature.
SBS4	Associated with tobacco smoking. Its profile is similar to the mutational spectrum observed in experimental systems exposed to tobacco carcinogens such as benzo[a]pyrene. SBS4 is, therefore, likely due to direct DNA damage by tobacco smoke mutagens.
SBS5	Unknown SBS5 mutational burden is increased in bladder cancer samples with ERCC2 mutations and in many cancer types due to tobacco smoking.

SBS6	SBS6 is associated with defective DNA mismatch repair and is found in microsatellite unstable tumours.
SBS7a	SBS7a/SBS7b/SBS7c/SBS7d are found in cancers of the skin from sun exposed areas and are thus likely to be due to exposure to ultraviolet light. SBS7a may possibly be the consequence of just one of the two major known UV photoproducts, cyclobutane pyrimidine dimers or 6-4 photoproducts. However, there is currently no evidence for this hypothesis and it is unclear which of these photoproducts may be responsible for SBS7a.
SBS7b	SBS7a/SBS7b/SBS7c/SBS7d are found in cancers of the skin from sun exposed areas and are likely to be due to exposure to ultraviolet light. SBS7b may possibly be the consequence of just one of the two major known UV photoproducts, cyclobutane pyrimidine dimers or 6-4 photoproducts. However, there is no evidence for this hypothesis and it is unclear which of these photoproducts may be responsible for SBS7b.
SBS7c	SBS7a/SBS7b/SBS7c/SBS7d are found in cancers of the skin from sun exposed areas and are likely to be due to exposure to ultraviolet light. SBS7c is possibly the consequence of translesion DNA synthesis by enzymes with propensity to insert T, rather than A, opposite ultraviolet induced thymidine and cytidine photodimers. The preponderance of T>A rather than T>C mutations may reflect the heavier burden of thymidine compared to cytidine dimers induced by UV light.
SBS7d	SBS7a/SBS7b/SBS7c/SBS7d are found in cancers of the skin from sun exposed areas and are likely to be due to exposure to ultraviolet light. SBS7d is possibly the consequence of translesion DNA synthesis by error-prone polymerases with greater propensity to insert G, rather than A, opposite UV light induced thymidine and cytidine photodimers.
SBS8	Unknown
SBS9	May be due in part to mutations induced during replication by polymerase eta as part of somatic hypermutation in lymphoid cells.
SBS10a	Polymerase epsilon exonuclease domain mutations.
SBS10b	Polymerase epsilon exonuclease domain mutations.
SBS11	SBS11 exhibits a mutational pattern resembling that of alkylating agents. Patient histories indicate an association between previous treatment with the alkylating agent temozolomide and SBS11 mutations.
SBS12	Unknown
SBS13	Attributed to activity of the AID/APOBEC family of cytidine deaminases on the basis of similarities in the sequence context of cytosine mutations caused by APOBEC enzymes in experimental systems. APOBEC3A is probably responsible for most mutations in human cancer, although APOBEC3B may also contribute (these differ in the sequence context two bases 5' to the mutated cytosine, see 1536 mutation classification signature extraction). SBS13 mutations are likely generated by error prone polymerases (such as REV1) replicating across abasic sites generated by base excision repair removal of uracil.
SBS14	Concurrent polymerase epsilon mutation and defective DNA mismatch repair.
SBS15	Defective DNA mismatch repair.
SBS16	Unknown
SBS17a	Unknown
SBS17b	Unknown
SBS18	Possibly damage by reactive oxygen species.
SBS19	Unknown
SBS20	Concurrent POLD1 mutations and defective DNA mismatch repair.
SBS21	DNA mismatch repair deficiency.
SBS22	Aristolochic acid exposure. Found in cancer samples with known exposures to aristolochic acid and the pattern of mutations exhibited by the signature is consistent with that observed in experimental systems of aristolochic acid exposure.
SBS23	Unknown
SBS24	Aflatoxin exposure. SBS24 has been found in cancer samples with known exposures to aflatoxin and the pattern of mutations exhibited by the signature is consistent with that observed in experimental systems exposed to aflatoxin.
SBS25	Unknown However, some Hodgkin's cell line samples in which the signature has been found were from patients exposed to chemotherapy and it is possible that SBS25 is due to chemotherapy treatment.
SBS26	Defective DNA mismatch repair.
SBS28	Unknown
SBS29	SBS29 has been found in cancer samples from individuals with a tobacco chewing habit.
SBS30	SBS30 is due to deficiency in base excision repair due to inactivating mutations in NTHL1.
SBS31	Prior chemotherapy treatment with platinum drugs.
SBS32	Prior treatment with azathioprine to induce immunosuppression. Associated mutation classes and signatures
SBS33	N/A
SBS34	Unknown
SBS35	Prior chemotherapy treatment with platinum drugs.
SBS36	Defective base excision repair, including DNA damage due to reactive oxygen species, due to biallelic germline or somatic MUTYH mutations.
SBS37	Unknown

SBS38	Unknown	Found only in ultraviolet light associated melanomas suggesting potential indirect damage from UV-light.
SBS39	Unknown	
SBS40	Unknown	
SBS41	Unknown	
SBS42		Occupational exposure to haloalkanes.
SBS44		Defective DNA mismatch repair.
SBS84		Activity of activation-induced cytidine deaminase (AID).
SBS85		Indirect effects of activation-induced cytidine deaminase (AID) induced somatic mutagenesis in lymphoid cells.



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