

De-palmitoylation of tissue factor regulates its activity, phosphorylation and cellular functions

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Running title: Regulation of TF function by palmitoylation

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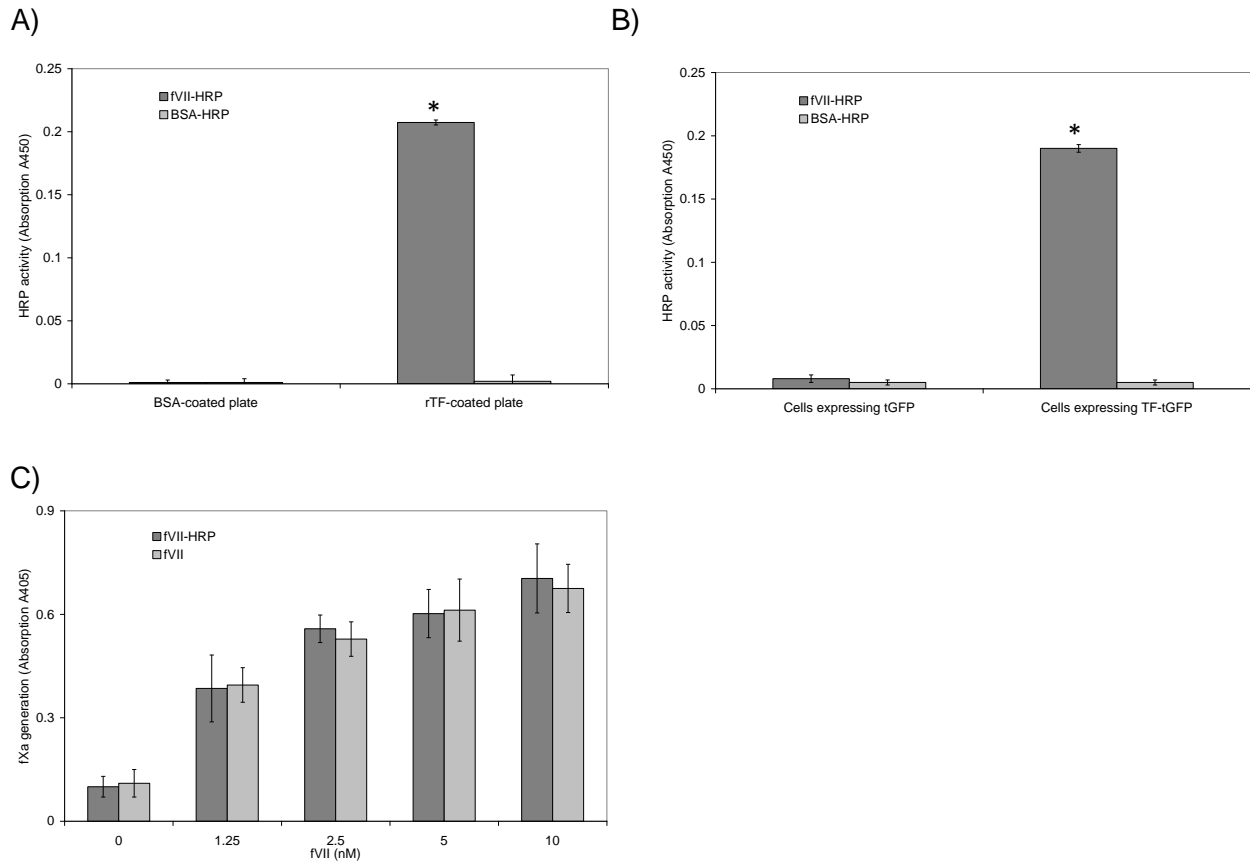
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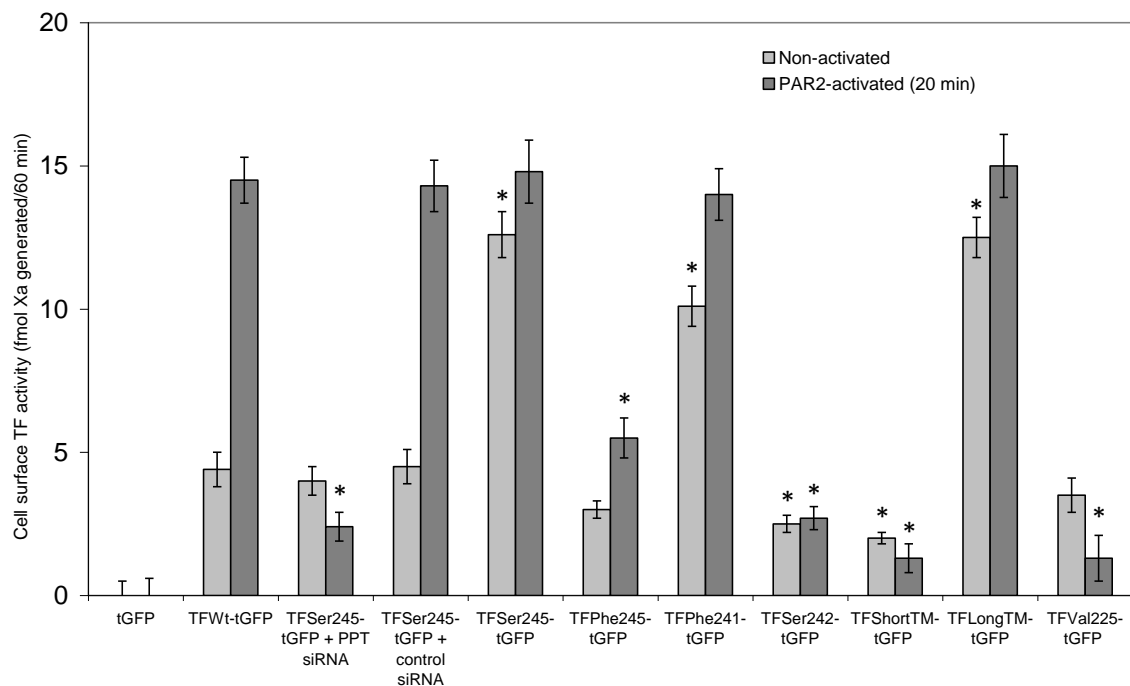
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Supplementary Figure S1 Confirmation of fVIIa-HRP activity and binding to TF



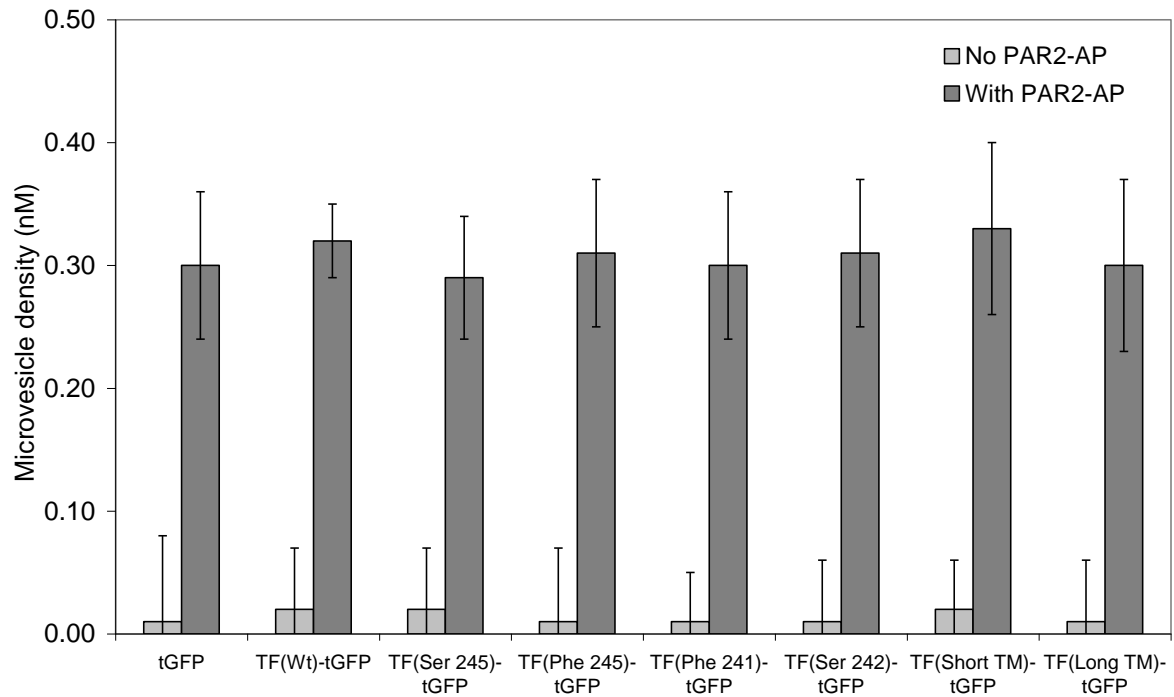
Factor VIIa-HRP and BSA-HRP conjugates were prepared using the Lightning-link HRP kit. Samples (20 nM) were incubated for 15 min in A) 48-well plates pre-coated with recombinant Innovin TF (13 ng/ml) in BSA (1 % w/v) or the vehicle solution, or B) with transfected-HDBEC expressing either TF-tGFP or tGFP. The plates were washed with PBS and the HRP activity determined using the One-solution TMB substrate (200 μ l). (n= 3, * = p<0.05 vs. BSA-HRP sample). C) Equal amounts of fVIIa-HRP and unconjugated-fVIIa (0-10 nM) were added to HDBEC expressing TF-tGFP and incubated for 10 min following which fXa-generation was measured. (n= 5).

Supplementary Figure S2 Analysis of the association of TF and PAR2 in the presence and absence of fVIIa, examined in HCAEC



Human coronary artery endothelial cells (HCAEC; 5×10^4) were co-transfected with combinations of pCMV-Ac-TF-tGFP and PPT-siRNA or control siRNA. HCAEC (5×10^4) were also transfected to express TF_{Wt}-tGFP, TF_{Ser245}-tGFP, TF_{Phe245}-tGFP, TF_{ShortTM}-tGFP, TF_{LongTM}-tGFP, TF_{Phe241}-tGFP, TF_{Ser242}-tGFP or TF_{Val225}-tGFP. Sets of cells were activated using PAR2-agonist peptide and fXa-generation was measured after 20 min. (n= 3, * = p<0.05 vs. the respective cells expressing TF_{Wt}-tGFP).

Supplementary Figure S3 Quantification of HDBEC-derived microvesicles using the Zymuphen Assay



HDBEC (5×10^4) were transfected with the TF variants as shown and one set of cells were activated using PAR2-AP (20 μ M). The density of released microvesicles was then measured using the Zymuphen Microparticle Assay Kit. (n= 4).