

**Table S1.** List of primers used in RT-qPCR analysis.

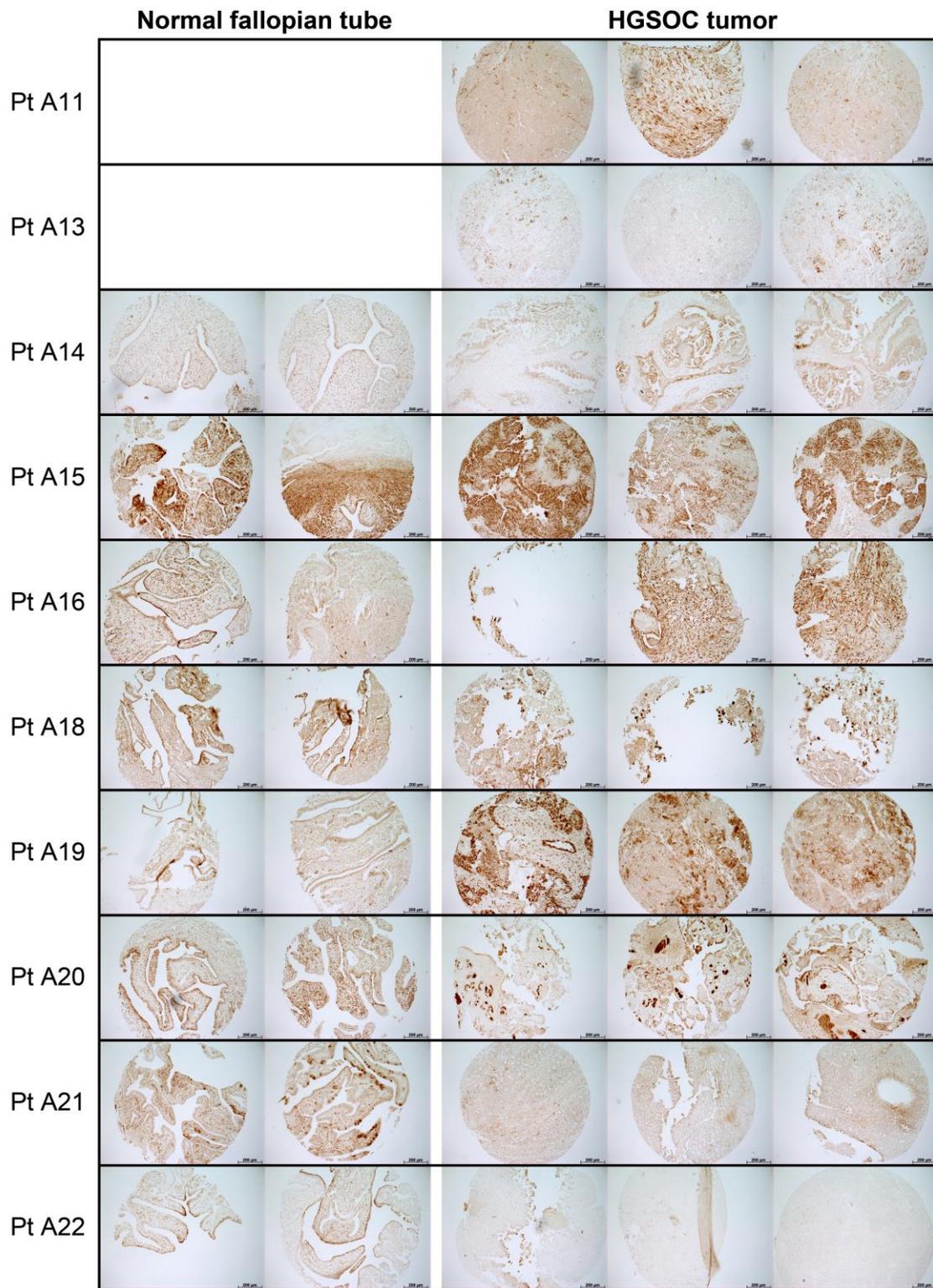
Gene	Forward primer	Reverse primer
<i>TBP</i>	GAGAGTTCTGGGATTGTACC	GGATTATATTCGGCGTTTCG
<i>PHLDA1</i>	GGCAAGACAAGGTTTTGAGG	TCGCAAGTTTTTCAGTAGGGTG
<i>SPRY2</i>	TCCACTCAGCACAAACAC	GATTATGCCATCAGCAACAG
<i>SPRY4</i>	CAACGGCTCTTAGACCAC	CACACTCCTTGCATTTACAC
<i>DUSP4</i>	CCCCACTACACGACCAG	TCCGAGGAGACATTCAACAG
<i>DUSP6</i>	GACGCTCGCTGTTTGTATC	GCTTCTAATCCCTCCCTCC
<i>CCND1</i>	CTGGATGCTGGAGGTCTG	GGTCTCCTTCATCTTAGAGGC
<i>EPHA2</i>	GTCAGCATCAACCAGACAGA	TCCCTTCTTGCGGTAAGTG
<i>EPHA4</i>	CGGAGCGGAGAATGC	TCCTTCCCAGACAGAGTAG
<i>ETV4</i>	AGGAGACATCAAGCAGGAAGGG	CCCAGAGCCTGGCGACC
<i>ETV5</i>	AGCACAAGTTCCTGATGATG	CATAGTTAGCACCAAGAGCC
<i>SOX2</i>	CACATGAACGGCTGGAG	CTGGTCATGGAGTTGTA CTG
<i>NANOG</i>	AACTCTCCAACATCCTGAAC	GTAGGAAGAGTAAAGGCTGG
<i>POU5F1</i>	GGGAAGGTATTCAGCCAAAC	AGAACCACACTCGGACC
<i>KLF4</i>	CACACGGGATGATGCTC	GTCACAGTGGTAAGGTTTCT
<i>ALDH1A1</i>	TGTTAGCTGATGCCGACTTG	TTCTTAGCCCGCTCAACACT
<i>ALDH1A2</i>	CCATTGGAGTGTGTGGACAG	GATGAGGGCTCCCATGTAGA
<i>ALDH1A3</i>	TCTCGACAAAGCCCTGAAGT	GTCCGATGTTTGAGGAAGGA

**Table S2.** Main elements of MEK1/2-ERK1/2 signaling pathway.

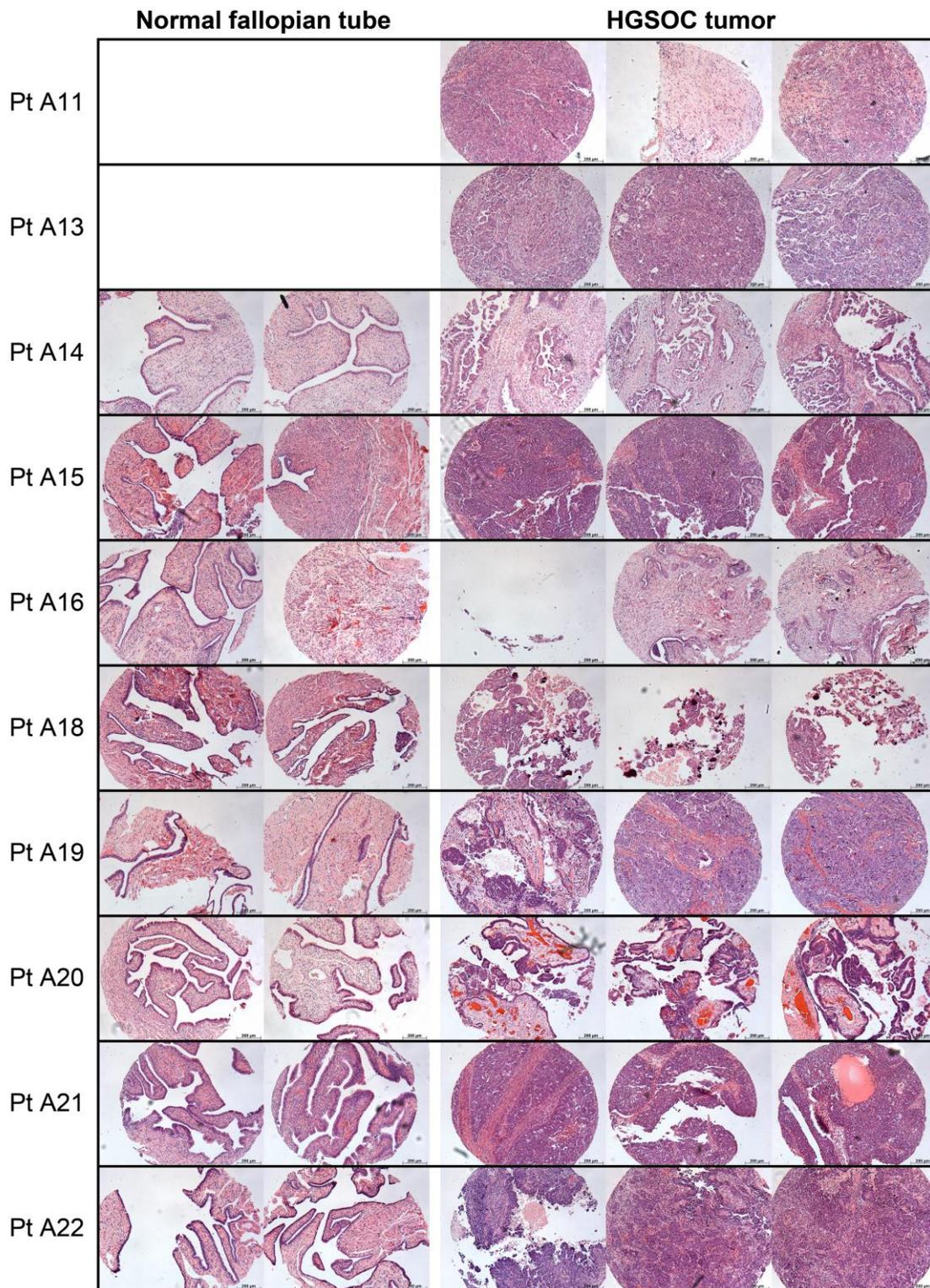
Protein name	UniProt ID	Gene name	NCBI Gene ID	Function	References
GRB2	P62993	<i>GRB2</i>	2885	Adaptor protein linking membrane receptors and RAS-controlled signaling pathways	[85-88]
SOS1	Q07889	<i>SOS1</i>	6654	Promoter of GDP-GTP exchange in RAS proteins	[89]
KRas HRas NRas	P01116 P01112 P01111	<i>KRAS</i> <i>HRAS</i> <i>NRAS</i>	3845 3265 4893	Family of membrane-bound GTPases mediating signal transduction from GRB2 to RAF proteins	[90-93]
BRAF RAF1	P15056 P04049	<i>BRAF</i> <i>RAF1</i>	673 5894	Family of protein kinases activated by Ras proteins and targeting MEK1/2	[91,94-98]
MEK1 MEK2	Q01986 P36507	<i>MAP2K1</i> <i>MAP2K2</i>	5604 5605	Protein kinases, central elements of MAP kinase signaling cascade targeting ERK1/2	[94,99,100]
ERK1 ERK2	P27361 P28482	<i>MAPK3</i> <i>MAPK1</i>	5595 5594	Protein kinases, main effectors of MAP kinase signaling cascade, phosphorylate more than 100 proteins	[94,99,101]

**Table S3.** MEK-responsive gene signature [52].

Gene name	NCBI Gene ID	Function	References
<i>PHLDA1</i>	22822	Apoptosis regulator, may exert both pro- and anti-apoptotic effects in different tumors	[102,103]
<i>SPRY2</i>	10253	Proliferation suppressor, negative feedback regulator of MEK-ERK activity, promotes glioblastoma growth and chemoresistance	[104-106]
<i>SPRY4</i>	81848	Proliferation suppressor, negative feedback regulator of MEK-ERK activity	[107,108]
<i>DUSP4</i>	1846	Proliferation and invasion suppressor, ERK phosphorylation inhibitor	[109,110]
<i>DUSP6</i>	1848	Proliferation suppressor, metastases formation suppressor, ERK phosphorylation inhibitor	[111,112]
<i>CCND1</i>	595	Proliferation stimulator, proto-oncogene	[113,114]
<i>EPHA2</i>	1969	Proliferation, EMT and invasion stimulator, apoptosis stimulator in some systems	[115-117]
<i>EPHA4</i>	2043	Invasion stimulator, marker of poor prognosis	[118,119]
<i>ETV4</i>	2118	Proliferation and invasion stimulator, sustains cell stemness	[120-122]
<i>ETV5</i>	2119	Proliferation stimulator, invasion suppressor, sustains cell stemness	[122-124]



**Figure S1.** Immunohistochemical staining of phosphorylated ERK1/2 in all available clinical samples of HGSOC and normal fallopian tube tissues. Scale bars: 200  $\mu$ m. Pt XXX – patient number XXX as provided by Lifespan Research Laboratories.



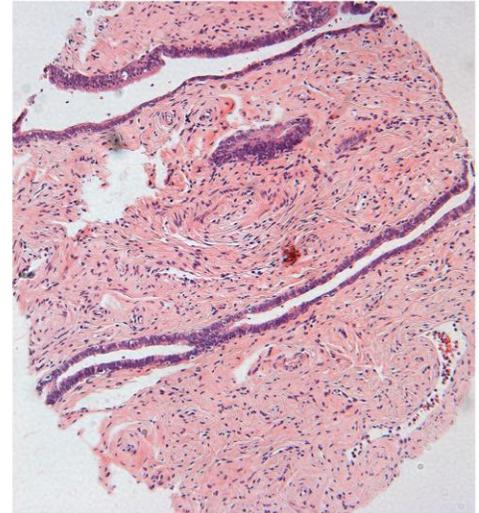
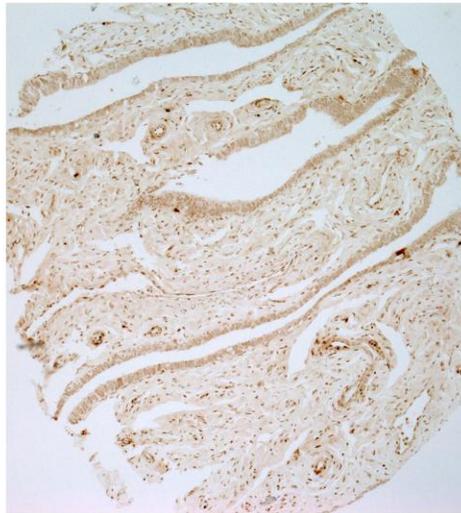
**Figure S2.** Hematoxylin and eosin staining of all available clinical samples of HGSOC and normal fallopian tube tissues. Scale bars: 200  $\mu\text{m}$ . Pt XXX – patient number XXX as provided by Lifespan Research Laboratories.

Pt A19

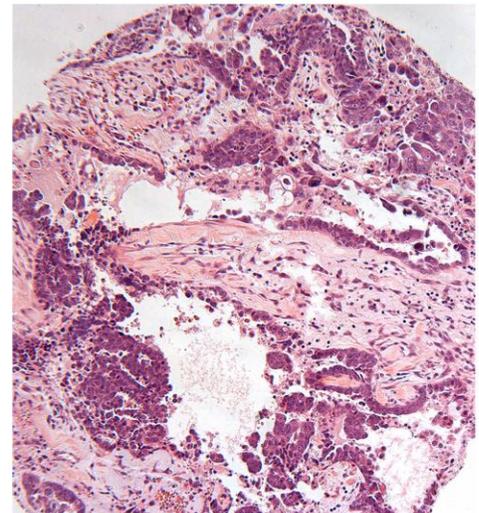
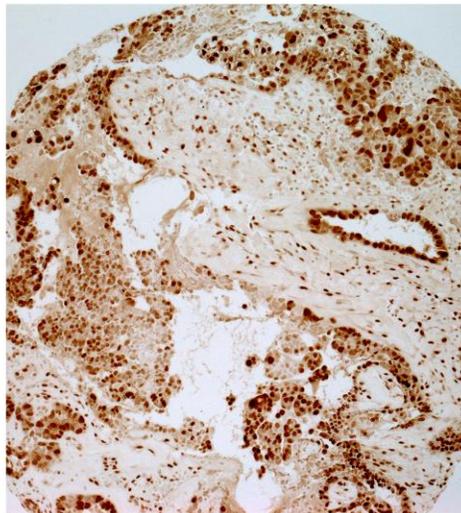
pERK1/2  
(T202/Y204)

Hematoxylin &  
eosin

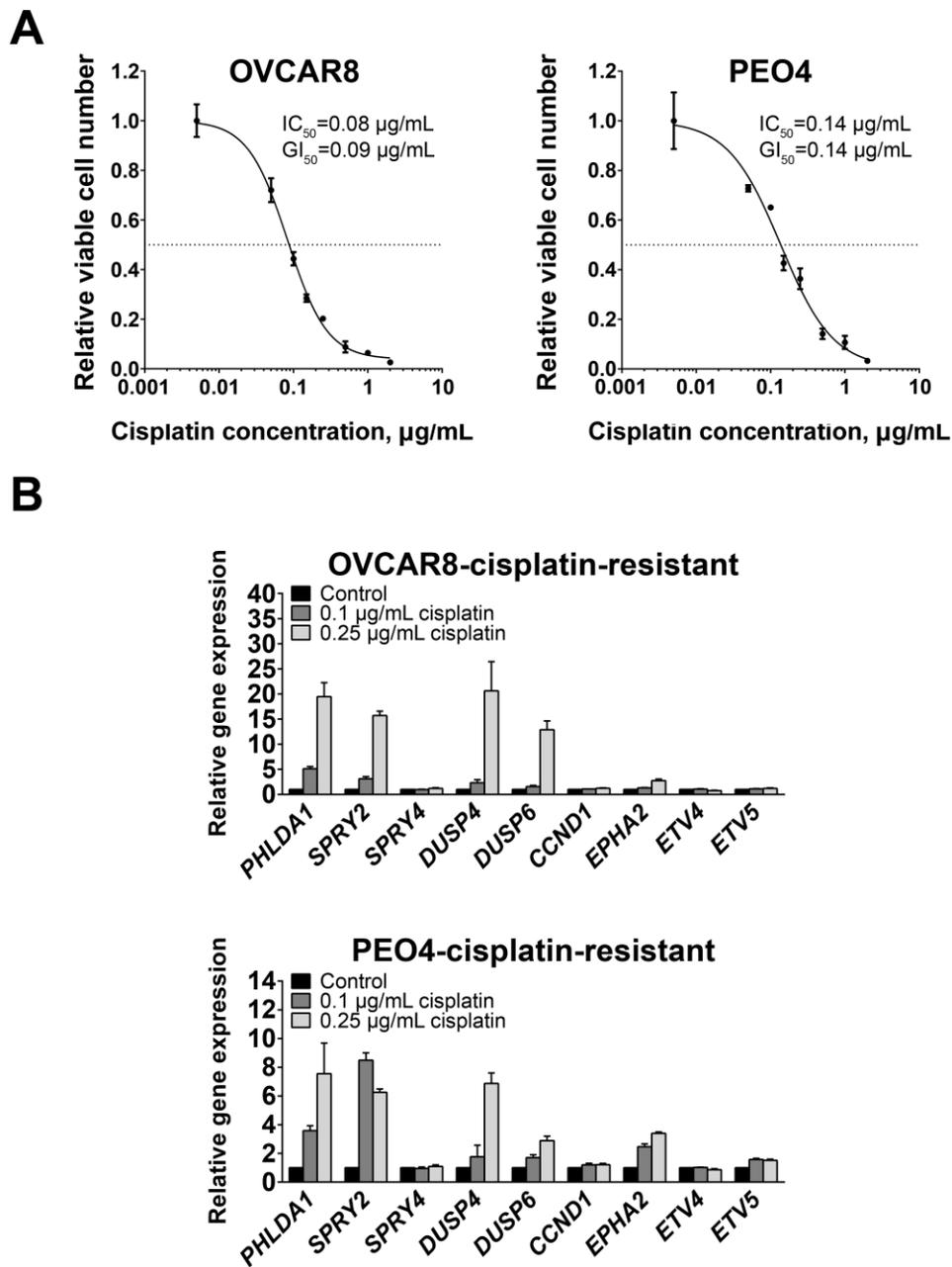
Normal  
fallopian tube



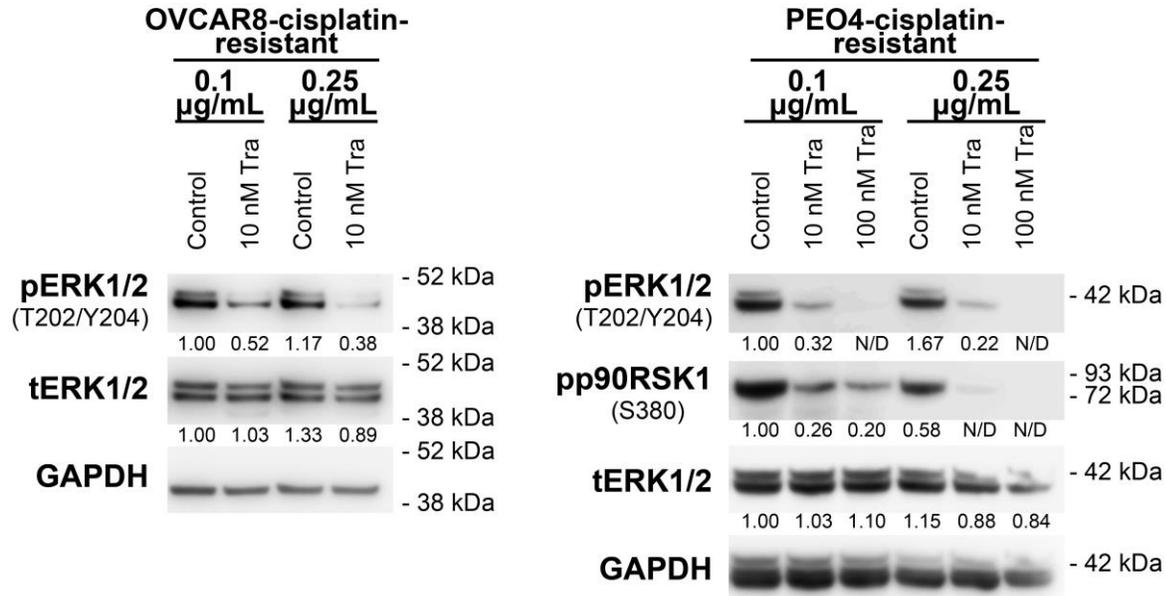
HGSOC  
tumor



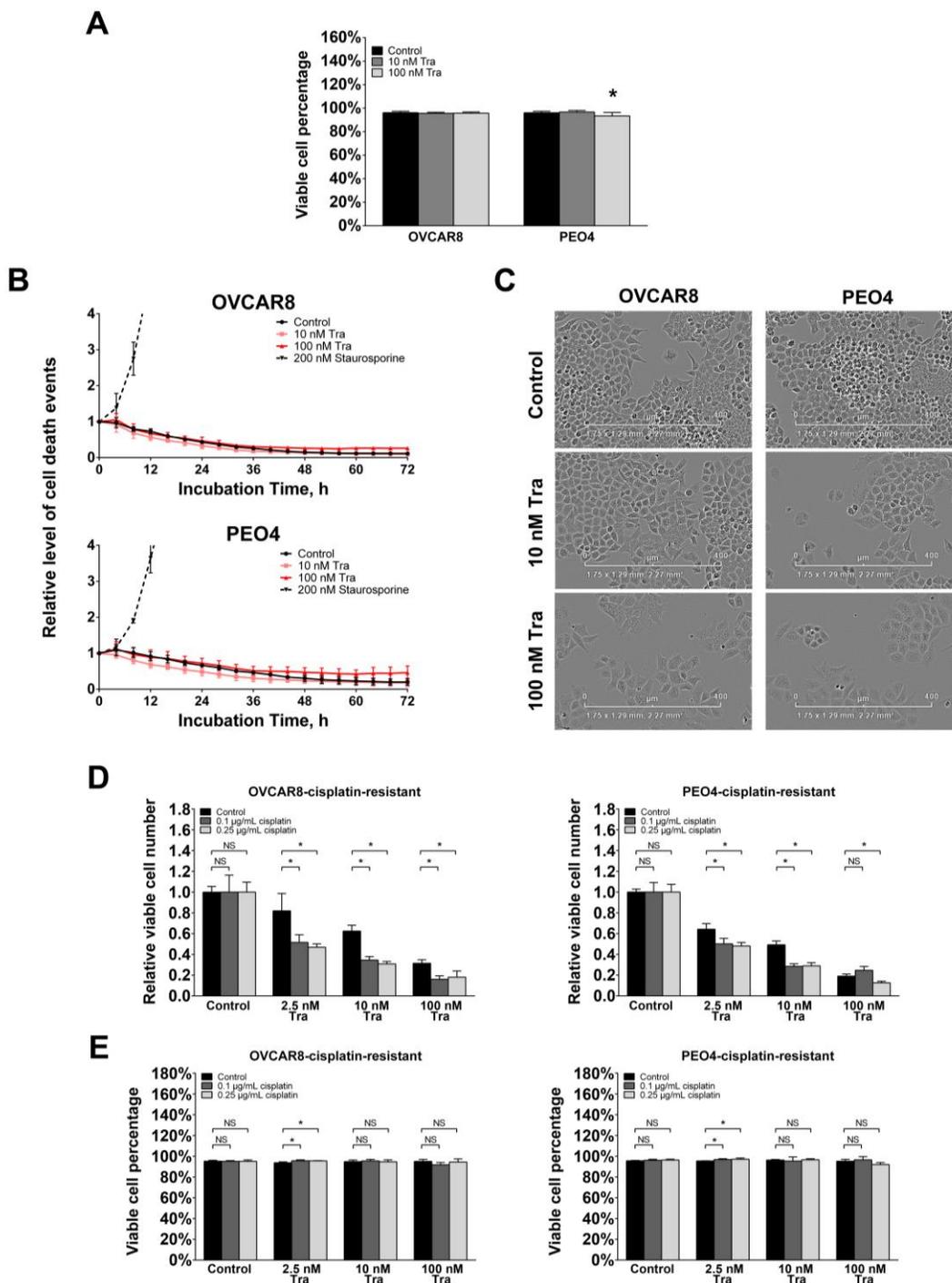
**Figure S3.** High-resolution images of phosphorylated ERK1/2 and hematoxylin and eosin staining for patient A19 clinical samples of HGSOC and normal fallopian tube tissues. Scale bars: 200  $\mu$ m. Pt A19 - patient number A19 as provided by Lifespan Research Laboratories, pERK1/2 – phosphorylated ERK1/2.



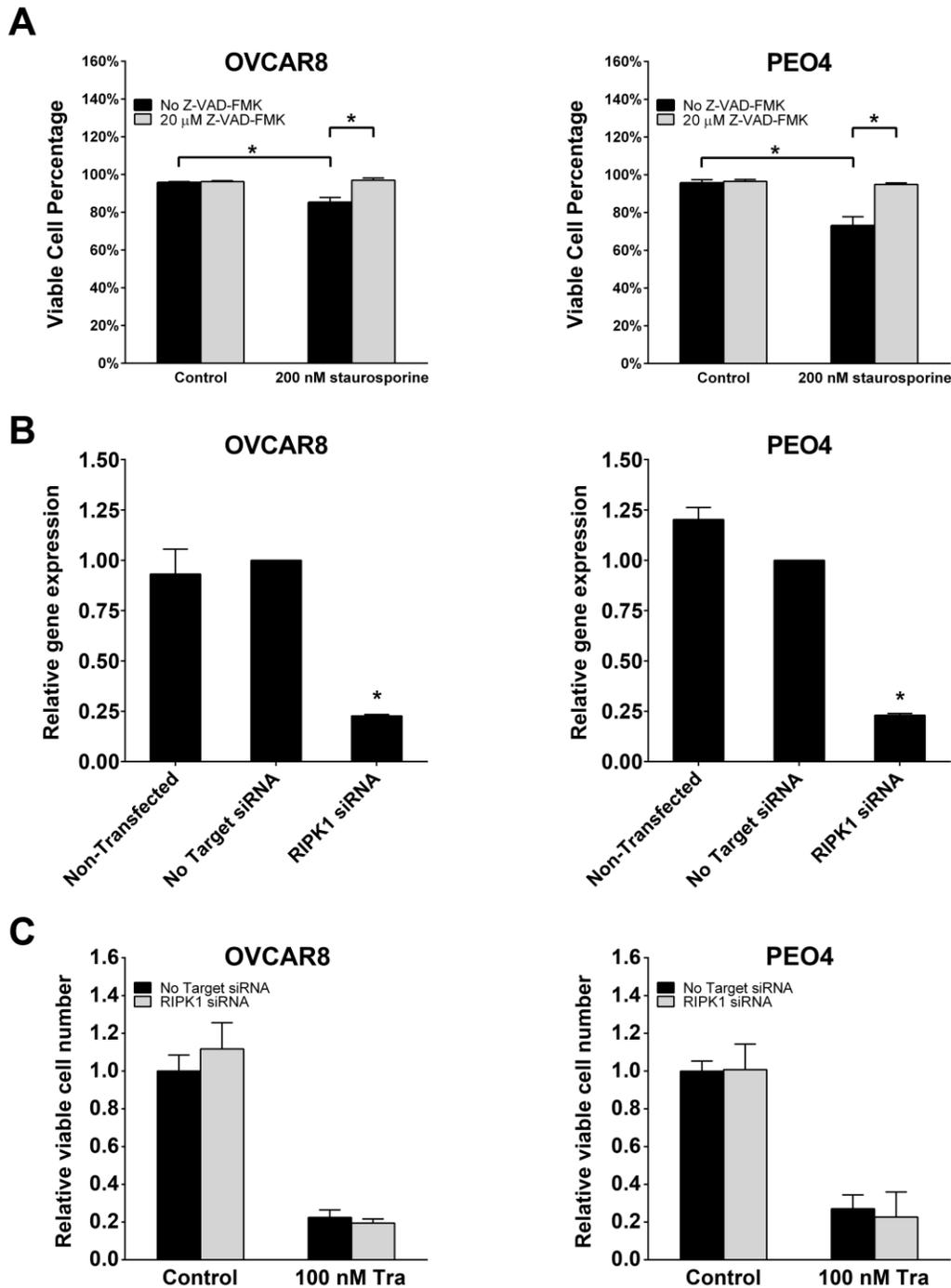
**Figure S4. Response of HGSOc cell lines OVCAR8 and PEO4 to transient or prolonged cisplatin treatment. (A)** Dose-response curves generated using relative viable cell numbers after treatment with various concentrations of cisplatin for 72 hours. Data are normalized to vehicle-treated control samples (not shown) and presented as mean $\pm$ S.D. (N=3). **(B)** Gene expression levels of MEK1/2-responsive genes in cells resistant to the indicated concentrations of cisplatin. Data are normalized to "Control" samples and presented as mean+S.E.M. (N=3, technical replicates).



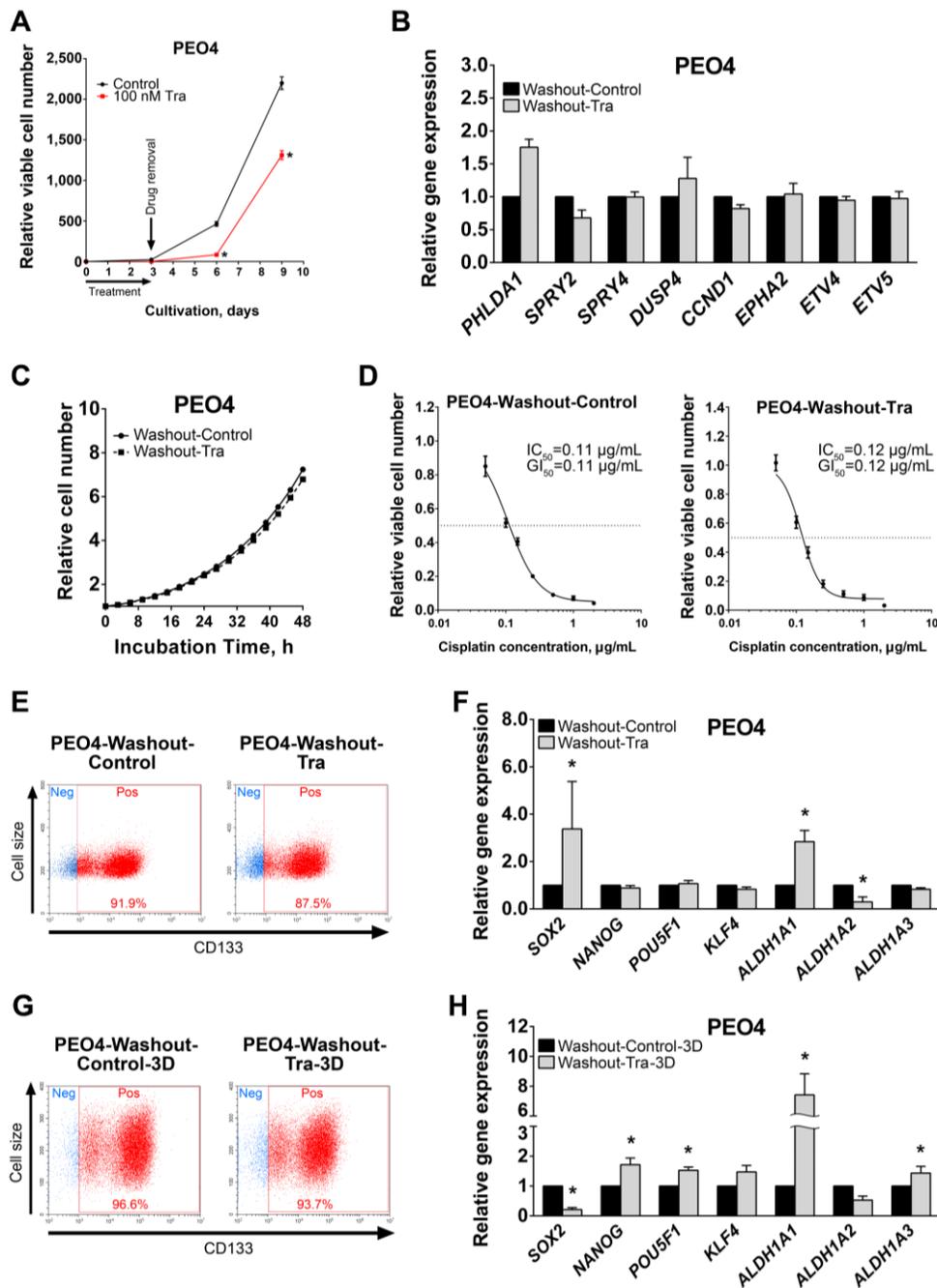
**Figure S5. MEK1/2-ERK1/2 pathway activity changes in cisplatin-resistant HGSOC cell cultures.** Immunoblotting analysis of phosphorylated and total ERK1/2 protein levels in cisplatin-resistant cells treated with trametinib for 24 hours. Numbers under the bands represent relative intensity normalized to GAPDH levels and “0.1 µg/mL-Control” samples. Tra – trametinib, pERK1/2 – phosphorylated ERK1/2, pp90RSK1 – phosphorylated p90RSK1, tERK1/2 – total ERK1/2, N/D – non-detectable signal.



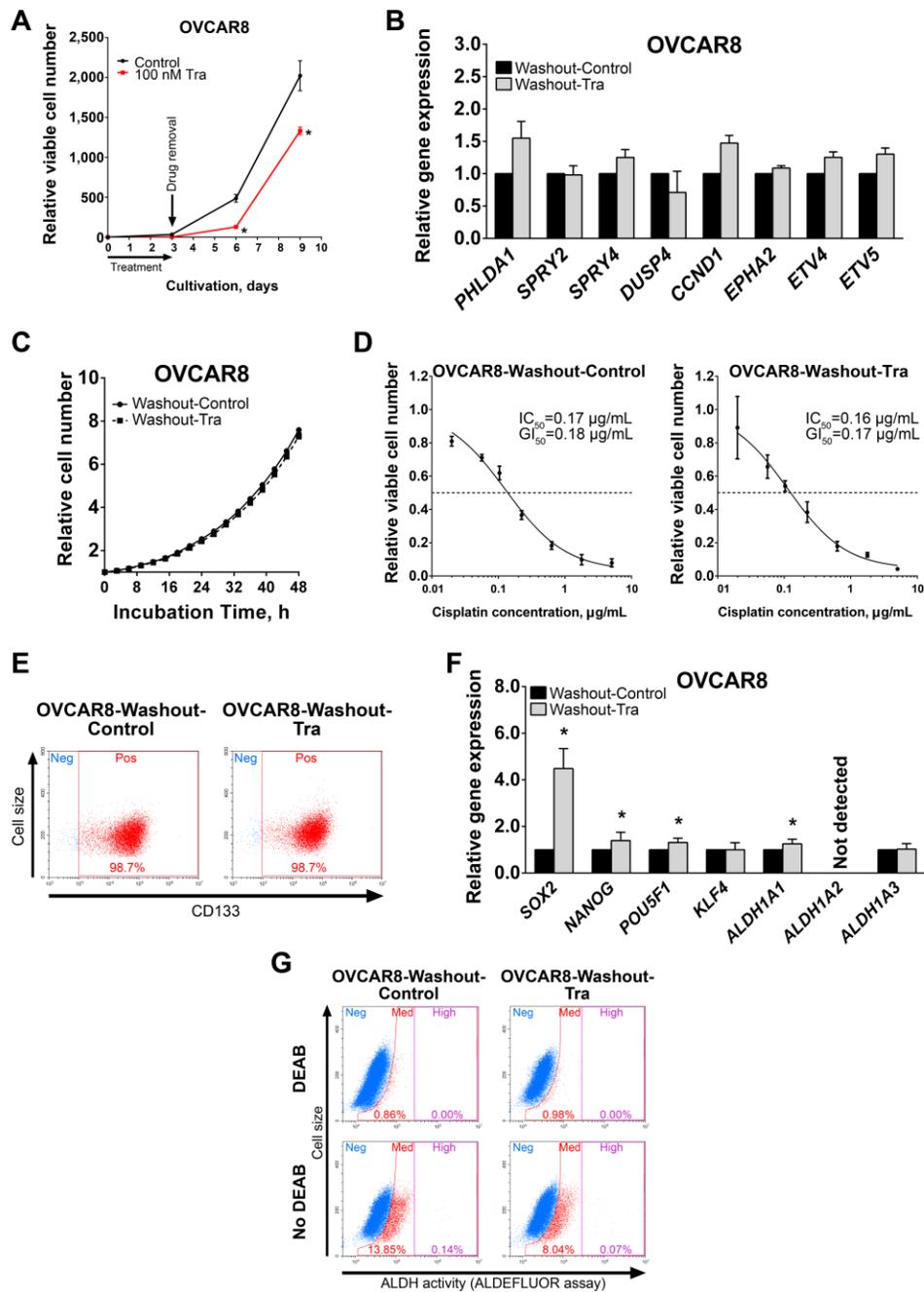
**Figure S6. Changes in functional characteristics of HGSOc cell cultures caused by trametinib treatment.** (A) Viable cell percentages after treatment with trametinib for 72 hours. Data are presented as mean+S.D. (N=6, Mann-Whitney U-test). (B) Cytotoxic effect of trametinib treatment detected using the CytoTox reagent. Staurosporin is used as a positive control. Fluorescence level for each time point is normalized to the area covered by cells and starting value; data are presented as mean±S.D. (N=4, Mann-Whitney U-test). (C) Cell morphology and confluence after treatment with trametinib for 72 hours. Scale bars: 400 µm. (D) Viable cell numbers of cisplatin-resistant cells after treatment with trametinib for 72 hours. Data are normalized to “Control” samples for each cell culture and presented as mean+S.D. (N=4, Mann-Whitney U-test). (E) Viable cell percentages of cisplatin-resistant cells after treatment with trametinib for 72 hours. Data are presented as mean+S.D. (N=4, Mann-Whitney U-test). Tra – trametinib, NS – non-significant difference.



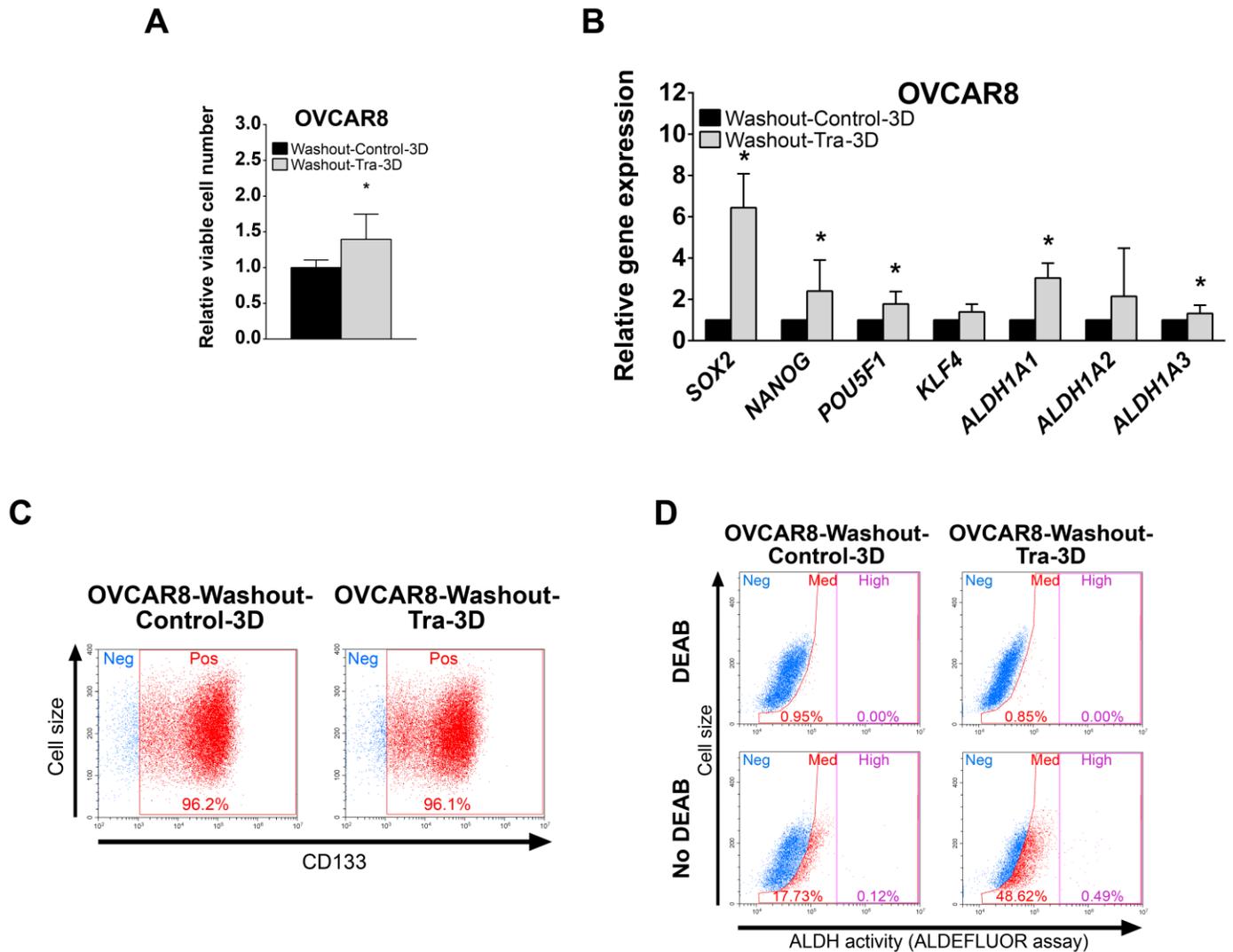
**Figure S7. Effect of cell death-related treatment of HGSOc cells. (A)** Viable cell percentages of cells treated with pan-caspase inhibitor Z-VAD-FMK and staurosporine for 24 hours. Data are normalized to “Control, No Z-VAD-FMK” sample and presented as mean+S.D. (N=3, one-tailed Mann-Whitney U-test, \* –  $p = 0.05$ ). **(B)** Gene expression levels of *RIPK1* gene in cells transfected with anti-*RIPK1* siRNA. Data are normalized to “No Target siRNA” samples and presented as mean+S.D. (N=3, two-tailed Student’s T-test with Welch’s correction). **(C)** Viable cell numbers of cells with siRNA-mediated *RIPK1* knockdown treated with trametinib for 72 hours. Data are normalized to “Control, No Target siRNA” sample and presented as mean+S.D. (N=3, no statistical analysis performed due to insufficient number of replicate and clear lack of differences). Tra – trametinib, siRNA – short interfering RNA.



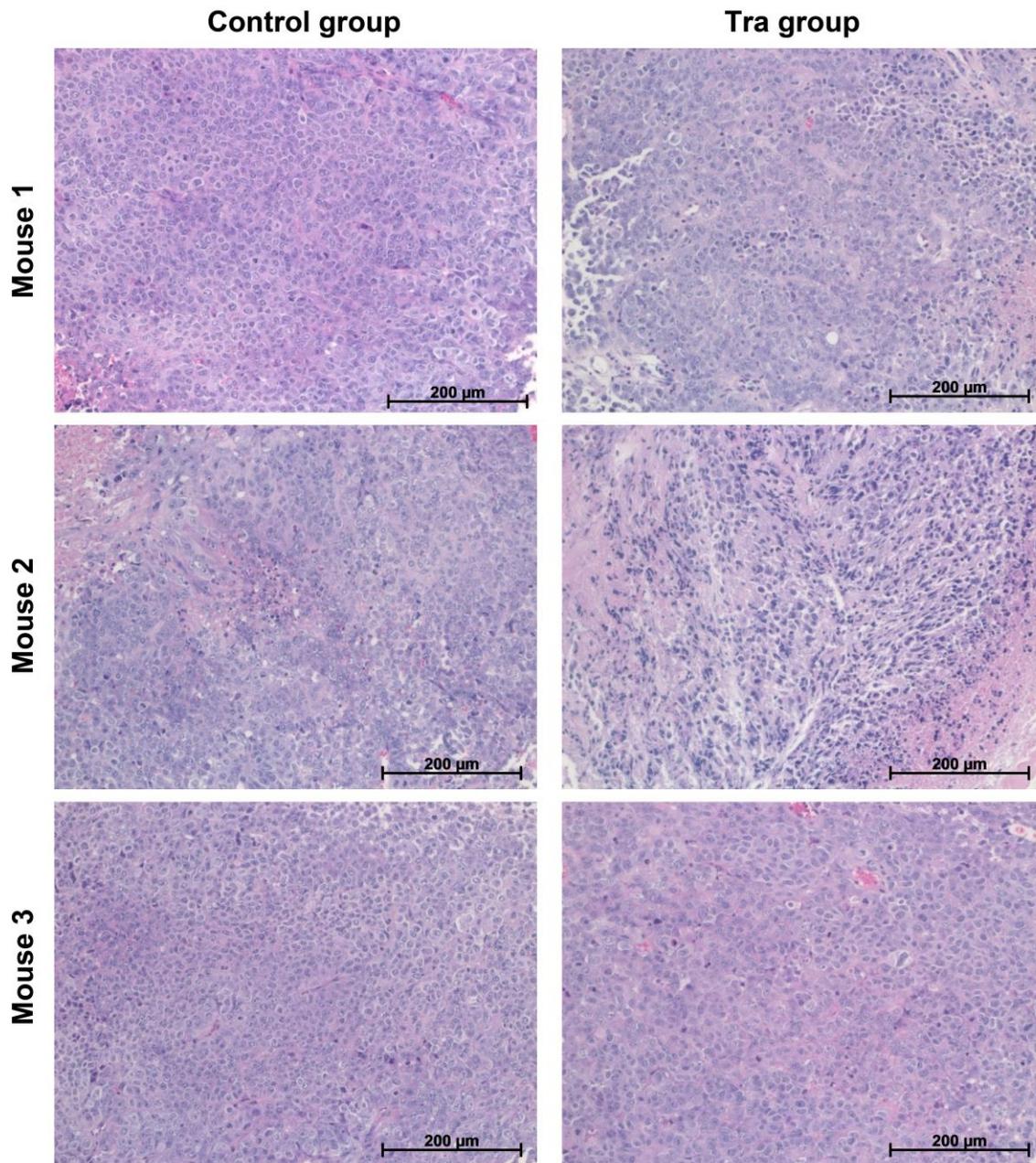
**Figure S8. Effects of trametinib washout upon properties of PEO4 cells.** (A) Cell growth kinetics during the initial establishment of PEO4-Washout cells. Data are normalized to starting cell number and presented as mean±S.D. (N=6, Mann-Whitney U-test). (B) Gene expression levels of MEK1/2-responsive genes. Data are normalized to “Washout-Control” samples and presented as mean+S.E.M. (N=3, technical replicates). (C) Growth kinetics of cells in standard conditions. Data are normalized to starting cell number and presented as mean±S.D. (N=4, Mann-Whitney U-test). (D) Dose-response curves generated using relative viable numbers of cells after treatment with various concentrations of cisplatin for 72 hours. Data are normalized to vehicle-treated control samples (not shown) and presented as mean±S.D. (N=3). (E) Expression of the CD133 surface marker in cells grown in standard conditions. Gates indicate CD133-negative (“Neg”) and CD133-positive (“Pos”) cell subpopulations. (F) Gene expression levels of stemness-related genes. Data are normalized to “Washout-Control” samples and presented as mean+S.D. (N=3, two-tailed Student’s T-test with Welch’s correction). (G) Expression of the CD133 surface marker in cells grown in non-adherent conditions. Gates indicate CD133-negative (“Neg”) and CD133-positive (“Pos”) cell subpopulations. (H) Gene expression levels of stemness-related genes in cells grown in non-adherent conditions. Data are normalized to “Washout-Control” samples and presented as mean+S.D. (N=3, two-tailed Student’s T-test with Welch’s correction). Tra – trametinib.



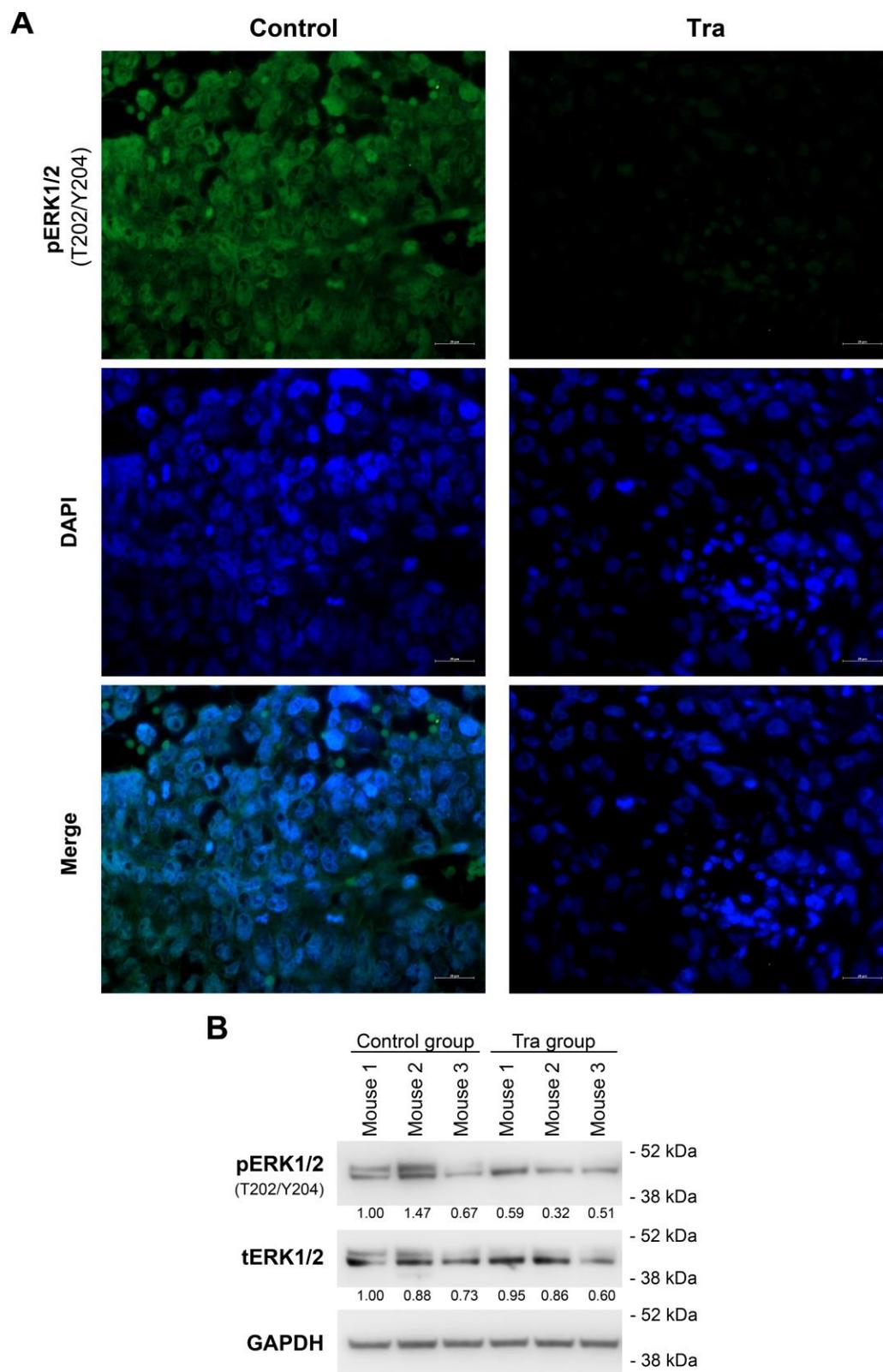
**Figure S9. Effects of trametinib washout upon properties of OVCAR8 cells.** (A) Cell growth kinetics during the initial establishment of OVCAR8-Washout cells. Data are normalized to starting cell number and presented as mean±S.D. (N=6, Mann-Whitney U-test). (B) Gene expression levels of MEK1/2-responsive genes. Data are normalized to “Washout-Control” samples and presented as mean+S.E.M. (N=3, technical replicates). (C) Growth kinetics of cells in standard conditions. Data are normalized to starting cell number and presented as mean±S.D. (N=4, Mann-Whitney U-test). (D) Dose-response curves generated using relative viable numbers of cells after treatment with various concentrations of cisplatin for 72 hours. Data are normalized to vehicle-treated control samples (not shown) and presented as mean±S.D. (N=3). (E) Expression of the CD133 surface marker in cells grown in standard conditions. Gates indicate CD133-negative (“Neg”) and CD133-positive (“Pos”) cell subpopulations. (F) Gene expression levels of stemness-related genes. Data are normalized to “Washout-Control” samples and presented as mean+S.D. (N=3, two-tailed Student’s T-test with Welch’s correction). (G) ALDEFLUOR analysis of cells after cultivation in non-adherent 3D conditions for 7 days. DEAB was used as ALDH inhibitor to define background ALDEFLUOR signal and set proper gates for ALDH-positive cells. Gates indicate cell subpopulations displaying negative (“Neg”), medium (“Med”), or high (“High”) levels of ALDH activity. Tra – trametinib.



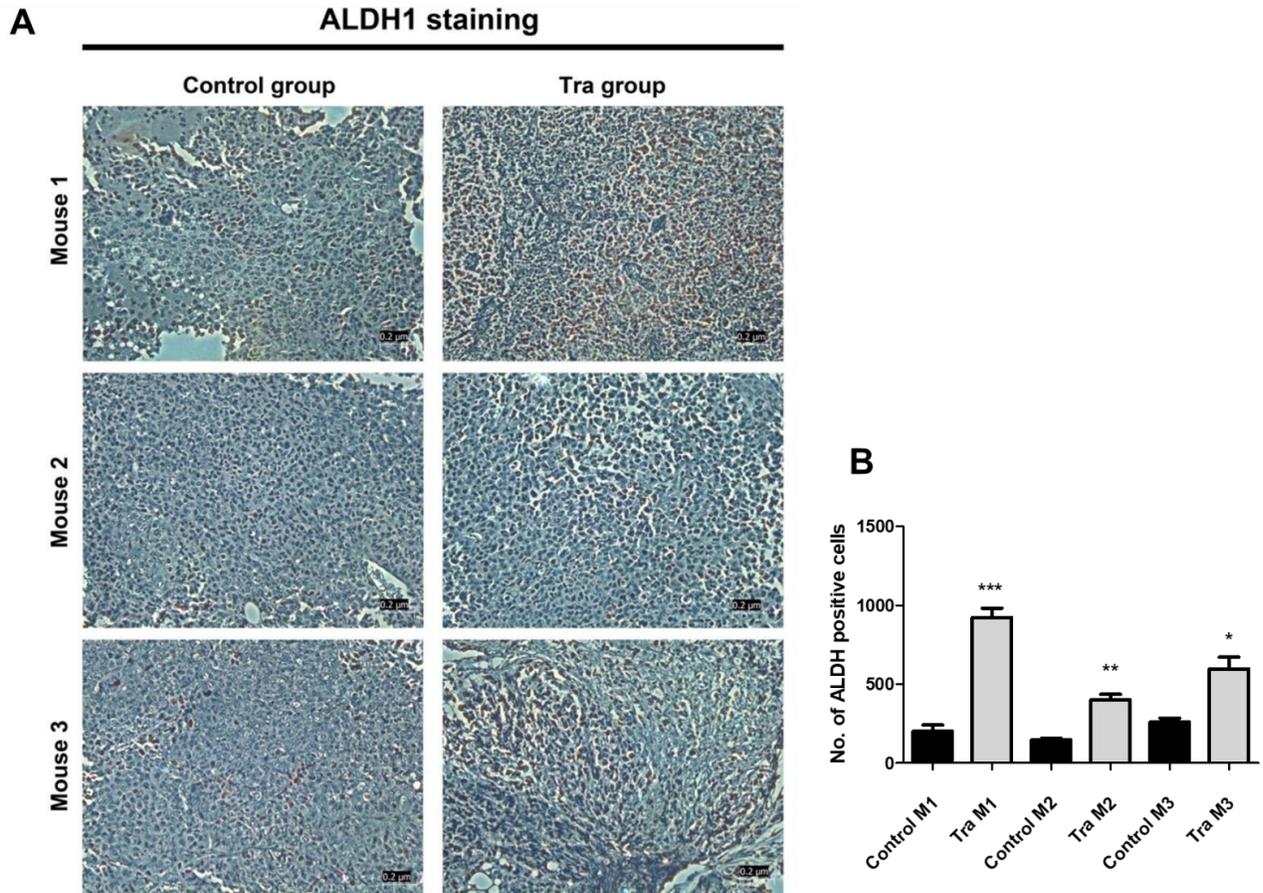
**Figure S10. Stemness-related properties of OVCAR8-Washout cells grown in non-adherent 3D conditions. (A)** Viable cell numbers after cultivation in non-adherent 3D conditions for 7 days. Data are normalized to “Washout-Control-3D” samples and presented as mean+S.D. (N=12, Mann-Whitney U-test). **(B)** Gene expression levels of stemness-related genes in cells grown in non-adherent conditions. Data are normalized to “Washout-Control” samples and presented as mean+S.D. (N=3, two-tailed Student’s T-test with Welch’s correction). **(C)** Expression of the CD133 surface marker in cells grown in non-adherent conditions. Gates indicate CD133-negative (“Neg”) and CD133-positive (“Pos”) cell subpopulations. **(D)** ALDEFLUOR analysis of cells after cultivation in non-adherent 3D conditions for 7 days. DEAB was used as ALDH inhibitor to define background ALDEFLUOR signal and set proper gates for ALDH-positive cells. Gates indicate cell subpopulations displaying negative (“Neg”), medium (“Med”), or high (“High”) levels of ALDH activity. Tra – trametinib.



**Figure S11.** Hematoxylin and eosin staining of PEO4 xenograft tissue samples. Scale bars: 200  $\mu\text{m}$ . Tra – trametinib.



**Figure S12. Effects of trametinib treatment on MEK1/2 pathway activity in HGSOC *in vivo*.** (A) High-resolution images of immunofluorescent staining of phosphorylated ERK1/2 (green) in PEO4 xenograft tissue samples. Cell nuclei were counterstained with DAPI (blue). Scale bars: 20  $\mu$ m. (B) Immunoblotting analysis of phosphorylated ERK1/2 levels in PEO4 xenograft tissue samples. Numbers under the bands represent relative intensity normalized to GAPDH levels and Control Mouse 1 sample. Tra – trametinib.



**Figure S13. Effects of trametinib treatment on ALDH1 expression in HGSOc xenografts.** A) Immunohistochemical staining of ALDH1 in PEO4 xenograft tissue samples with hematoxylin counterstaining. Scale bars: 0.2  $\mu$ m. Tra – trametinib. B) No. of ALDH positive cells quantified by image J software. (N=5, two-tailed Student's T-test with Welch's correction). Tra – trametinib.

# Uncropped Immunoblotting Images

Figure 2A.

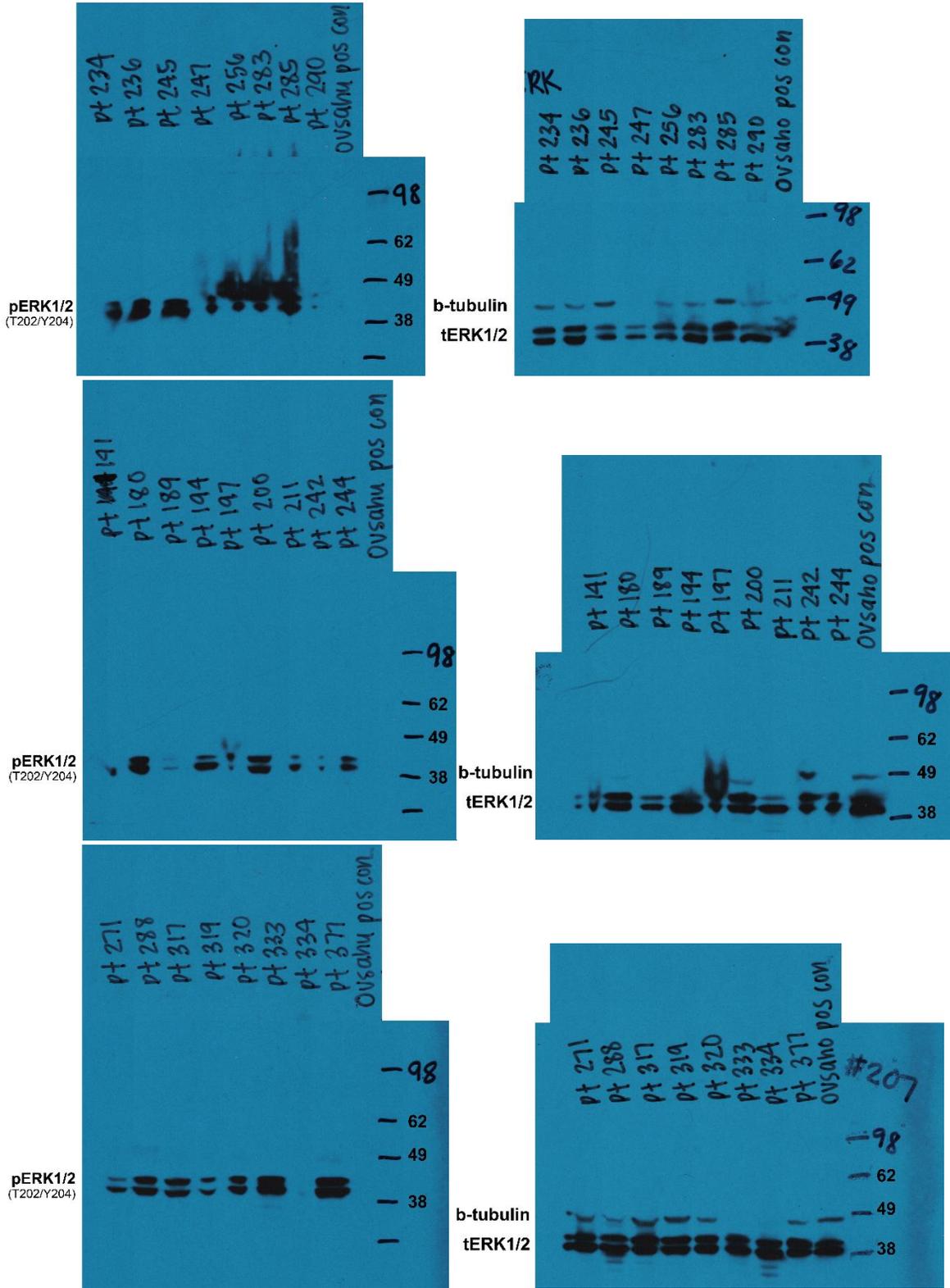


Figure 2A (Continued).

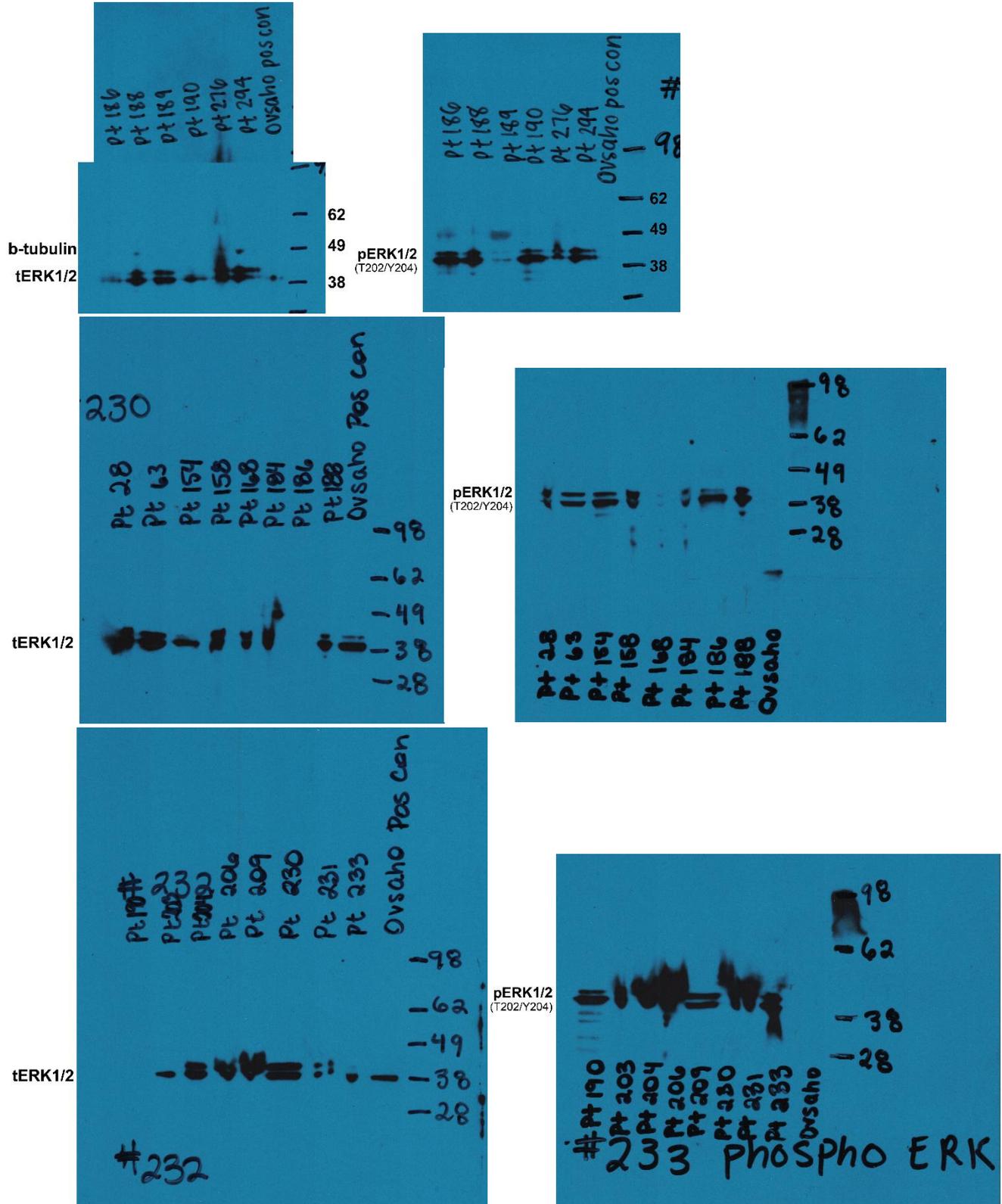


Figure 2A (Continued).

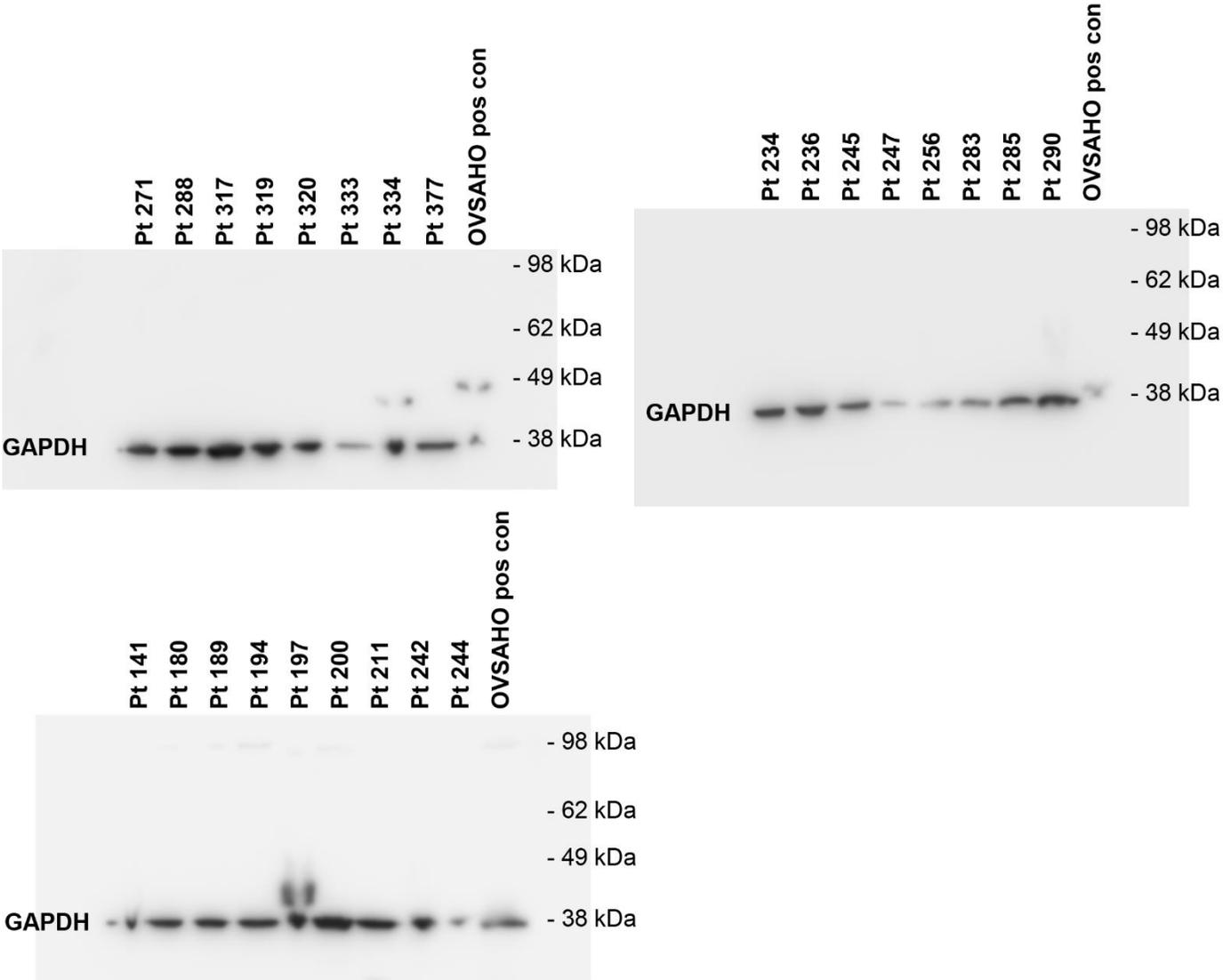


Figure 2C.

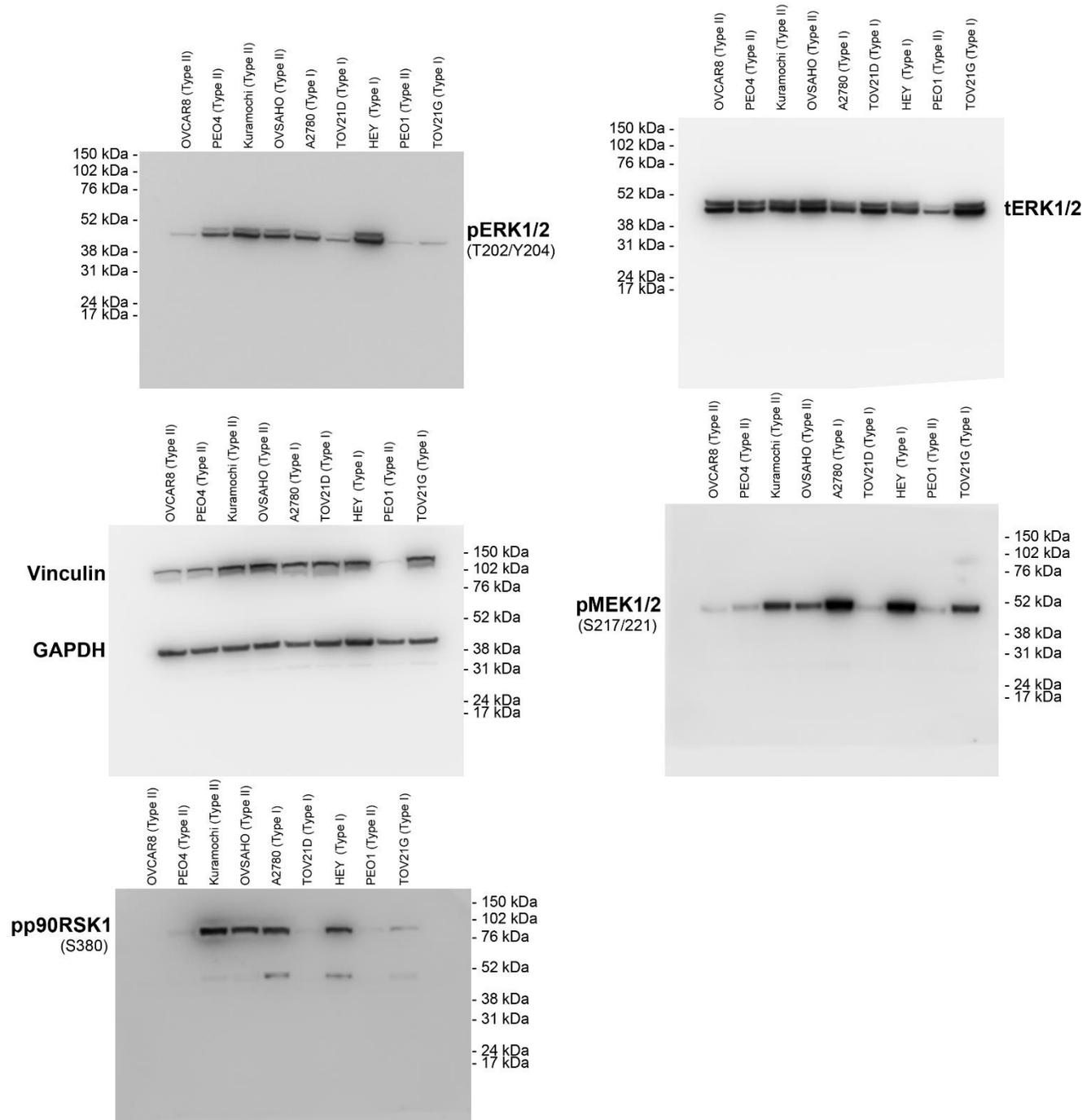




Figure 3A (Continued).

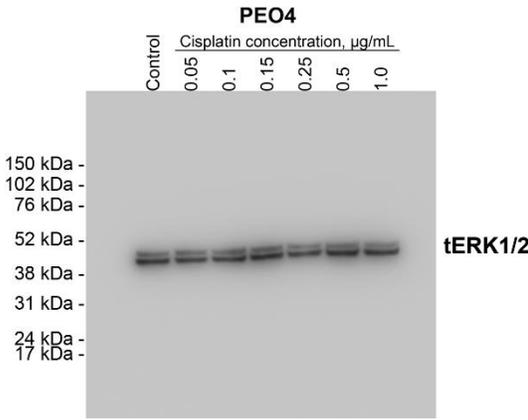
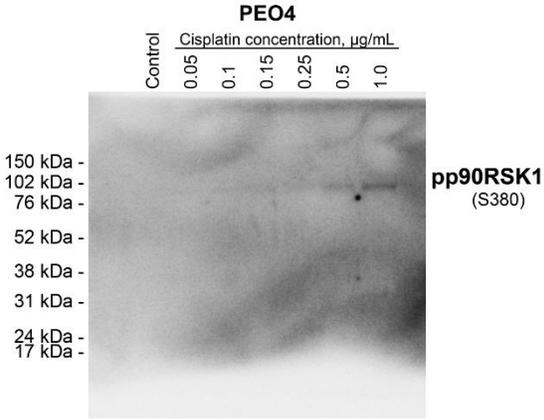
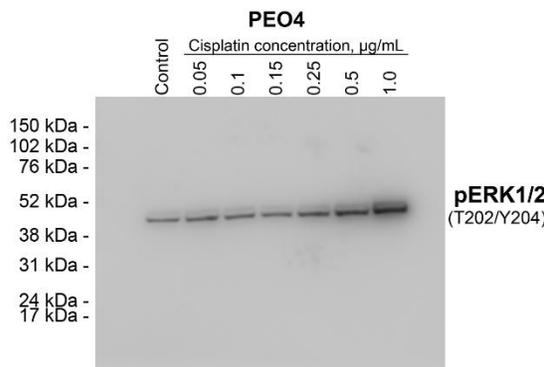
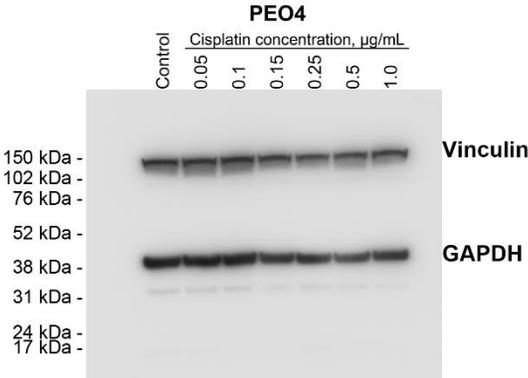


Figure 3B.

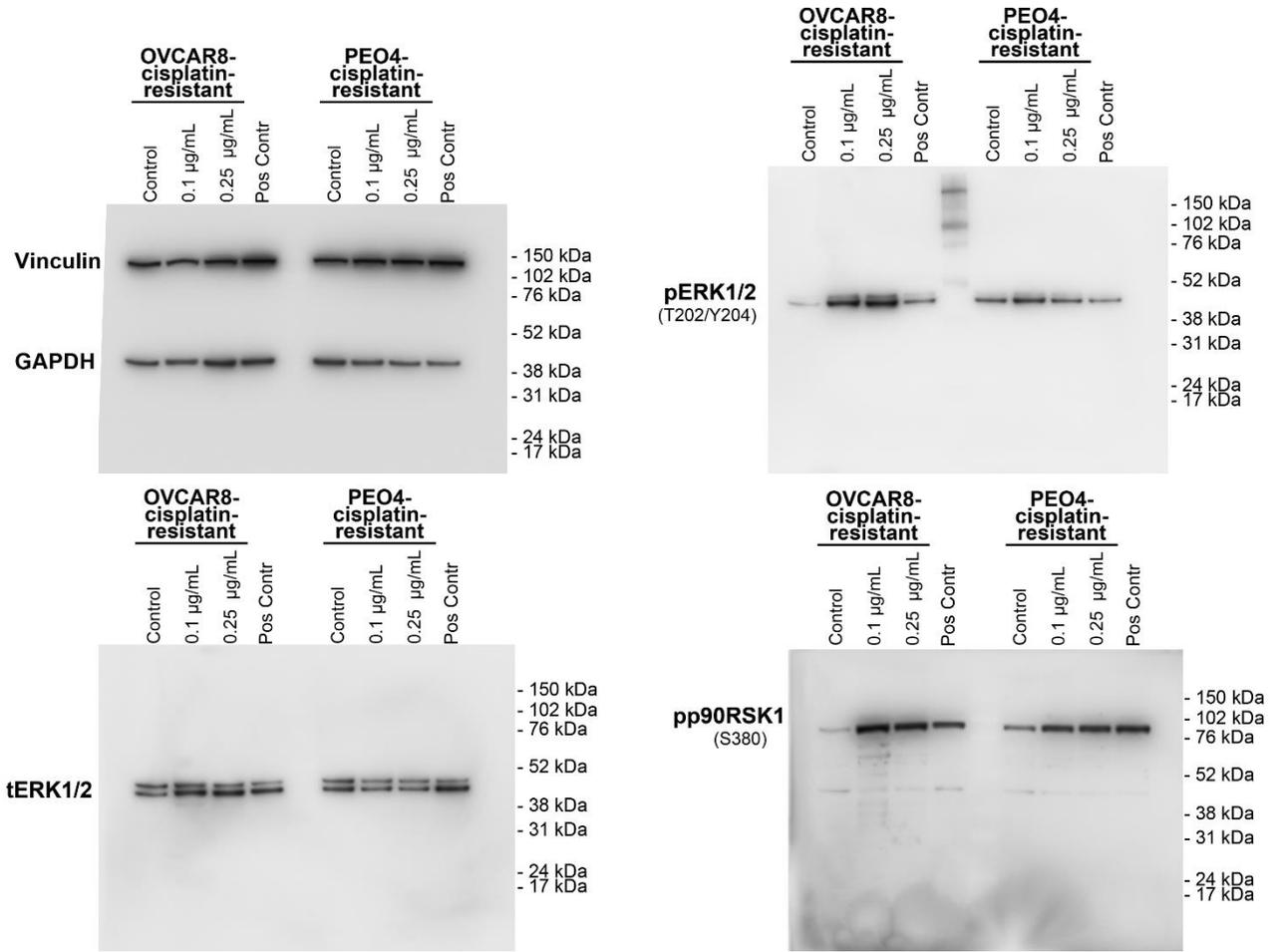


Figure 3C.

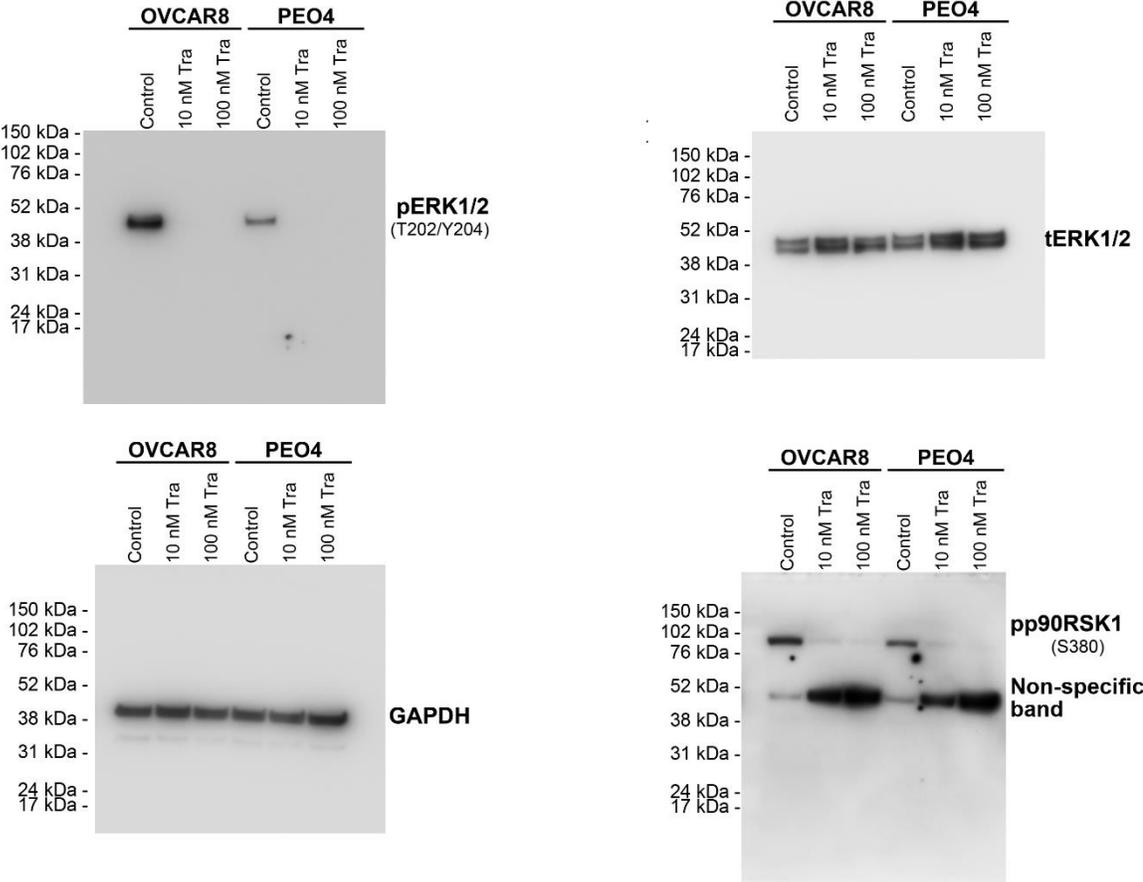


Figure 5C.

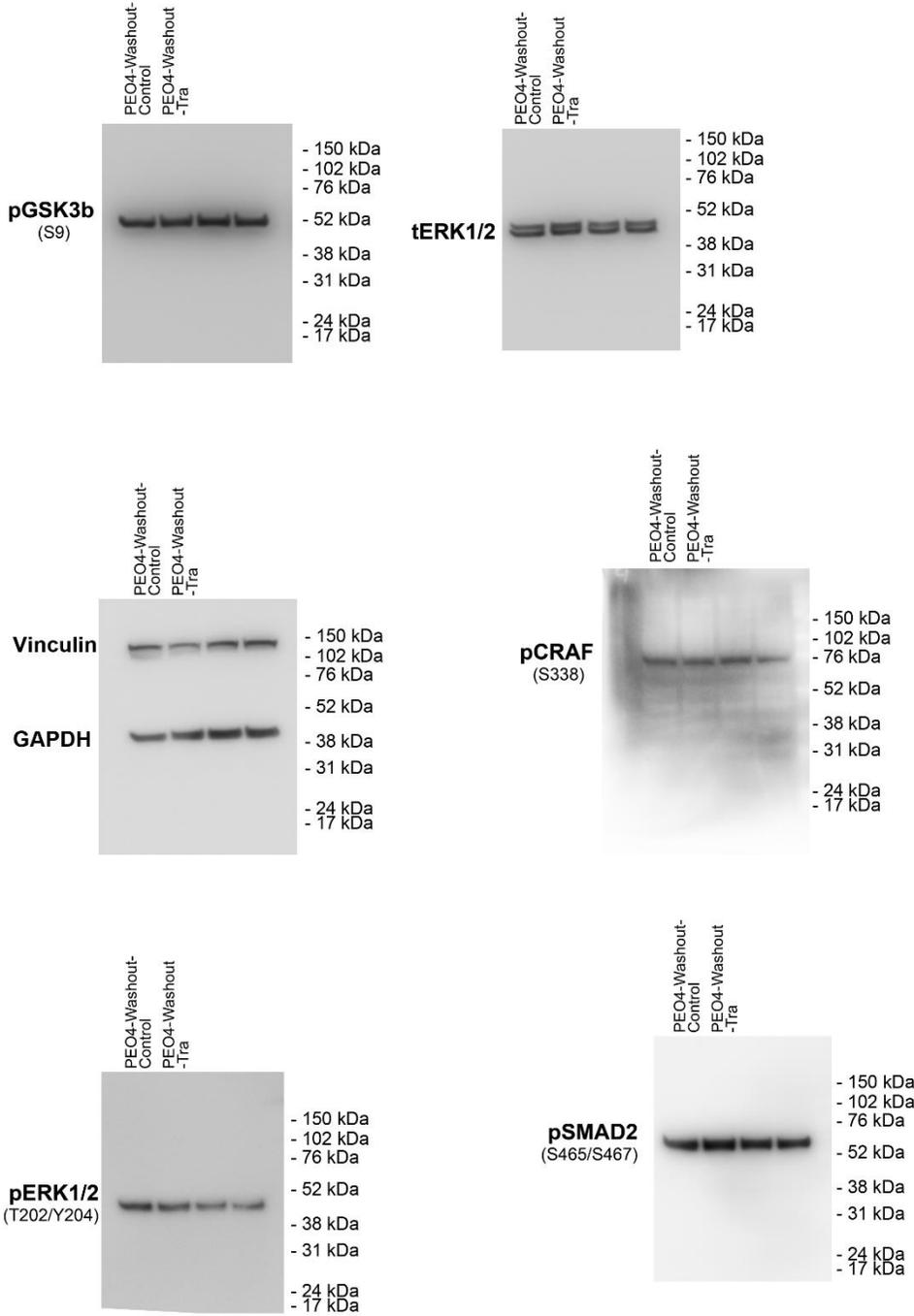


Figure S5.

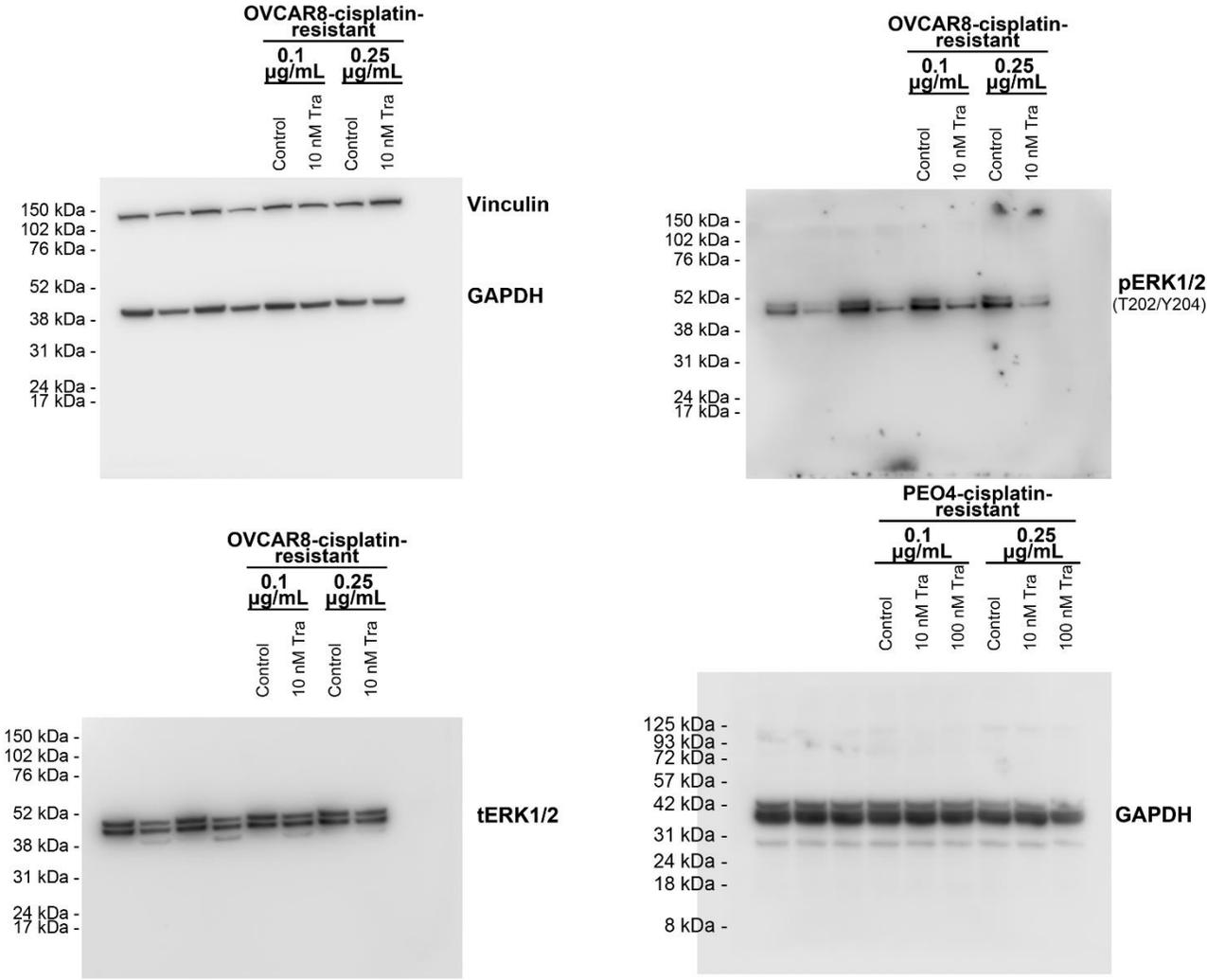


Figure S5 (Continued).

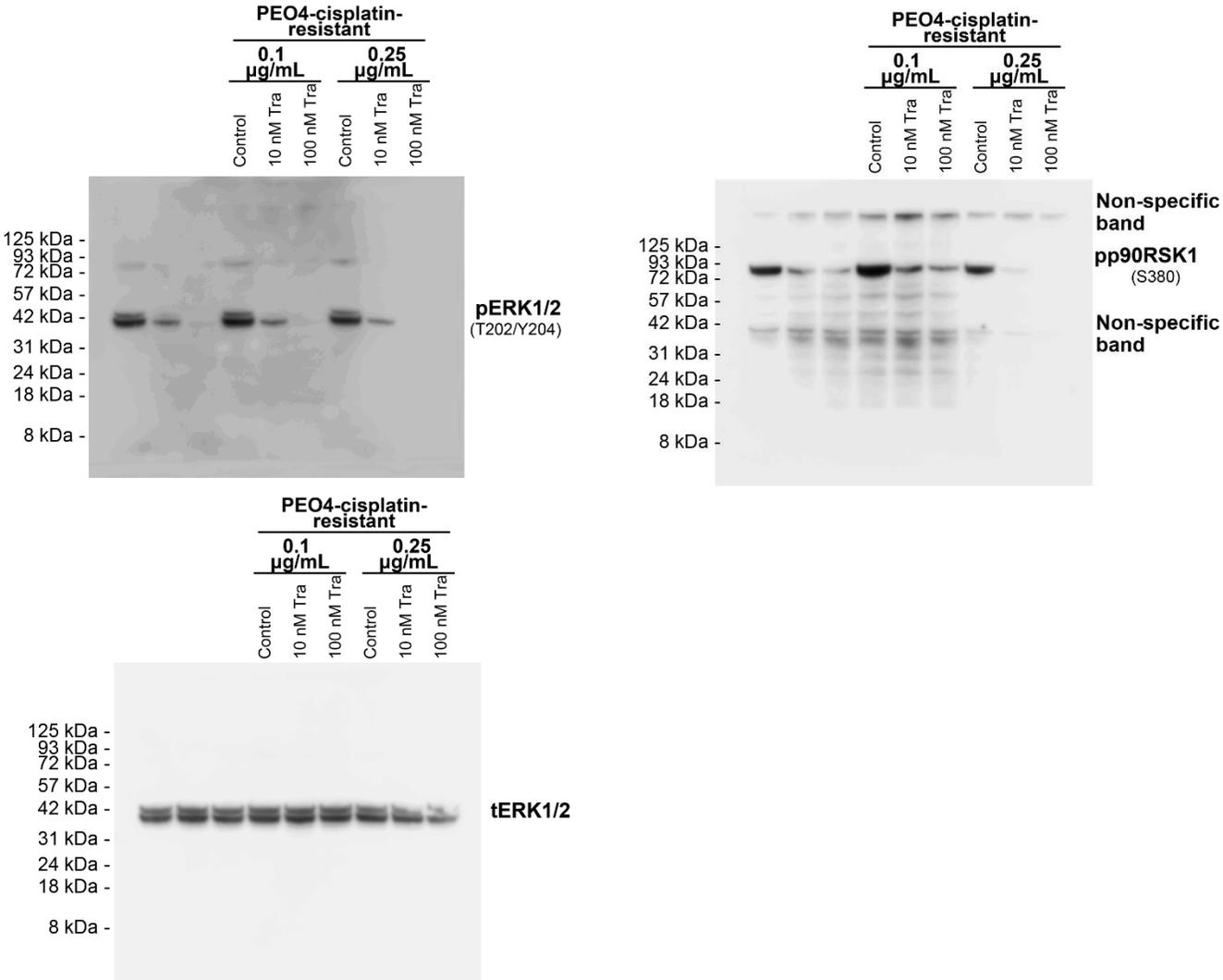


Figure S12B.

