

Alternative splicing, epigenetic modifications and cancer: A dangerous triangle, or a hopeful one?

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SUPPLEMENTARY MATERIALS

Table S1. Writers, readers and erasers of DNA methylation.

Writers and their functions [1]	Readers [2]	Erasers and their mechanism [3]
DNMT1 Responsible for <i>de novo</i> methylation	Methyl binding domain (MDB)-containing proteins MeCP1 MeCP2	Ten-eleven translocation (TET) enzymes TET1 TET2 TET3
DNMT3A	MBD1	
DNMT3B Use hemimethylated DNA as substrate. Function: maintenance of methylation in mitotic division	MBD2 MBD3 MBD4	Catalyse oxidative removal of methyl group <i>via</i> 5- hydroxymethylcytosine
DNMT3L		
DNMT3B3 No DNA methyl transferase activity; only regulatory roles		

Table S2. Writers, readers and erasers of histone acetylation.

Writers [4–6]	Readers [7–11]	Erasers [12–14]
<i>GCN5/PCAF family (GNAT)</i> GCN5 (KAT2A), PCAF (KAT2B), HAT1 (KAT1), ELP3 (KAT9), ATF2	Bromodomain <i>BRD family I</i> PCAF, GCN5L2, FALZ/BPTF, CECR2, BAZ1A	<i>CLASS I family</i> HDAC1, HDAC2, HDAC3, HDAC8
<i>MYST family</i> MOF/MYST1 (KAT8), MOZ/MYST3 (KAT6A), MORF/MYST4 (KAT6B), HBO1/MYST2 (KAT7), TIP60 (KAT5)	<i>BRD family II</i> BRD2, BRD3, BRD4, BRDT	<i>CLASS IIa family</i> HDAC4, HDAC5, HDAC7, HDAC9a, HDAC9b
<i>p300/CBP family</i> CBO (KAT3B), CBP (KAT3A)	<i>BRD family III</i> EP300, CREBBP, WDR9, PHIP, BRD8B, BAZ1B, BRWD3	<i>CLASS IIb family</i> HDAC6, HDAC10
<i>SRC family</i> SRC-1 (KAT13A), ACTR/SRC-3 (KAT13B), TIF-2 /NCOA2(KAT13C), CLOCK (KAT13D)	<i>BRD family IV</i> ATAD2B/KIAA1240	<i>CLASS IV family</i> HDAC11
<i>TRANSCRIPTION CO-ACTIVATORS family</i> TAF1/TAFII250 (KAT4), FIIIC220/ GTF3C1	<i>BRD family V</i> TRIM66, TRIM33, TRIM24/TIF1 α , SP100, SP110, SP140, SP140L, LOC93349, BAZ2A, BAZ2B	<i>SIRTUINS</i> SIRT1-7
	<i>BRD family VI</i> MLL, TRIM28	
	<i>BRD family VII</i> ZMYND11, ZMYND8, TAF1, TAF1L, WDR9d1, BRWD3d1, PHIPd1	
	<i>BRD family VIII</i> ASH1L, SMARCA2, SMARCA4, PBRM1/PB1	
	Tandem-PHD domain MOZ (KAT6A), MORF (KAT6B), DPF1 (BAF complex), DPF2 (BAF complex), DPF3 (BAF complex)	
	YEATS domain AF9 (YEATS3), YAF9 (YEATS4), YEATS2, ENL (YEATS1)	

Details on writers, readers and erasers can be found at <http://weram.biocuckoo.org/>

Table S3. Writers, readers and erasers of histone methylation.

Writers [11,13,15]	Readers [8,11,13]	Erasers [11,13]
K-HMT		
SET domain	Chromodomain	K-HDM
<i>SUV39 family</i>	CHD1, HP1, CDY1, PC1/PC2/PC/LHP1, MSL3, MRG15, CBX1-8, MPP8, Tip60,	FAD-dependent amine oxidase
SUV39H1, SUV39H2, G9A, GLP, ESET, CLLL8		<i>LSD1 family</i>
<i>SET1 family</i>		LSD1(KDM1A), LSD2 (KDM1B)
MLL1, MLL2, SET1A, SET1B, MLL4, MLL3, EZH2, EZH1	ING1-5, BPTF, RAG2, TAF3, ICBP90, PYGO, CHD4, UHRF1 (ICBP90), DPF3, KDM5A (JARID1A), KDM5C (JARID1C), KDM7D (JMJD1D), KMT2A (MLL1), KDM7B (PHF8), KDM7C (PHF2)	Fe²⁺ and α-KG dependent dioxygenase
<i>SET2 family</i>		<i>KDM2 family</i>
ASH1, NSD1, NSD2, NSD3, HYPB/HIF1		KDM2A, KDM2B
<i>RIZ family</i>	Tudor domain	<i>KDM3 family</i>
PRDM1/BLIMP, PDRM2/RIZ, PRDM4/PFM1	JMJD2A, 53BP1, PHF1, PHF19, PHF20, TDRD3	KDM3A-3C (JMJD1A-1C)
<i>SMYD family</i>		<i>KDM4 family</i>
SMYD1, SMYD3		KDM4A-4D
<i>SUV4-20 family</i>		<i>KDM5 family</i>
SUV4-20H1, SUV4-20H2	DNMT3A, BRPF1, PDP1, HDGF2, PSIP1 (LEDGF)	KDM5A-5D (JARID1A-1D)
<i>SET7/9 family</i>		<i>KDM6 family</i>
SET7, SET9	PHF20L1, SFMBT, L3MBTL1/2, MBTD1	KDM6A(UTX), KDM6B(JMJD3), KDM6C(UTY)
Non-SET domain		<i>KDM7 family</i>
DOT1L		KDM7A(JMJD1D), KDM7B(PHF8), KDM7C(PHF2)
R-HMT		<i>KDM8 family</i>
<i>PRMT Type-I family</i>		KDM8 (JMJD5)
PRMT1, PRMT2, PRMT3, PRMT4 (CARM1), PRMT6, PRMT8	ELP2 (STATIP1), EED, WDR5, TBL1X, TBL1XR1, L3MBTL2 (LIN-61)	R-HDM
<i>PRMT Type-II family</i>	14-3-3 domain	PAD4/PADI4, JMJD6
PRMT5, PRMT9	14-3-3	
<i>PRMT Type-III family</i>	ZF-CW domain	
PRMT7	ZCWPW1	
	Ankyrin domain	
	G9A/GLP	

Details on writers, readers and erasers can be found at <http://weram.biocuckoo.org/>

Table S4. Epidrugs approved or under clinical trial.

Inhibitors of DNA methylation		Inhibitors of HDACs		Inhibitors of bromodomains		Inhibitors of PRMTs	
Name	status	Name	status	name	status	name	status
Azacitidine	approved	Panobinostat	approved	I-BET762	CT	GSK3326595	CT
Decitabine	approved	Belinostat	approved	Birabresib	CT	AMG 193	CT
Aza-TdCyd	CT	Valproic Acid	approved	CPI-610	CT	JNJ-64619178	CT
		Romidepsin	approved	FT-1101	CT	PF-06939999	CT
		Vorinostat	approved	ZEN-3694	CT	PRT811	CT
		Pracinostat	approved as orphan drug	BMS-986158	CT	IDE397	CT
		Entinostat	CT	OTX-015	CT	PRT543	CT
		Abexinostat	CT	ABBV-075	CT		
		CUDC-101	CT	GS-5829	CT		
		Givinostat	CT	PLX-51107	CT		
		Mocetinostat	CT	TEN-010	CT		

The names of the epidrugs approved or under clinical trials (CT) are given, although some of them were approved for diseases other than cancer. Further details can be obtained from [16,17]. The progress of the clinical trials can be checked at <https://www.clinicaltrials.gov/>

Table S5. Drugs targeting mRNA splicing, approved or under clinical trials.

Splicing modulators		Drugs targeting enzymes involved in splicing		
Name	status	name	status	target
Risdiplam	approved	SM08502	CT	CLKs
Branaplam	approved as orphan drug			
Aclarubicin	CT			

The names of the drugs approved or under clinical trials (CT) are given. CLK stands for CDC2-like kinases. Further details can be obtained from [18–20]. The progress of the clinical trials can be checked at <https://www.clinicaltrials.gov/>

ADDITIONAL REFERENCES

1. Nishiyama, A.; Nakanishi, M. Navigating the DNA methylation landscape of cancer. *Trends Genet.* **2021**, *37*, 1012–1027.
2. Ginder, G.D.; Williams, D.C. Readers of DNA methylation, the MBD family as potential therapeutic targets. *Pharmacol. Ther.* **2018**, *184*, 98–111.
3. Wu, X.; Zhang, Y. TET-mediated active DNA demethylation: mechanism, function and beyond. *Nat. Rev. Genet.* **2017**, *18*, 517–534.
4. Roth, S.Y.; Denu, J.M.; Allis, C.D. Histone acetyltransferases. *Annu. Rev. Biochem* **2001**, *81*–120.
5. Marmorstein, R.; Roth, S.Y. Histone acetyltransferases: Function, structure, and catalysis. *Curr. Opin. Genet. Dev.* **2001**, *11*, 155–161.
6. Sapountzi, V.; Côté, J. MYST-family histone acetyltransferases: Beyond chromatin. *Cell. Mol. Life Sci.* **2011**, *68*, 1147–1156.
7. Filippakopoulos, P.; Qi, J.; Picaud, S.; Shen, Y.; Smith, W.B.; Fedorov, O.; Morse, E.M.; Keates, T.; Hickman, T.T.; Felletar, I.; et al. Selective inhibition of BET bromodomains. *Nature* **2010**, *468*, 1067–1073.
8. Yun, M.; Wu, J.; Workman, J.L.; Li, B. Readers of histone modifications. *Cell Res.* **2011**, *21*, 564–578.
9. Marmorstein, R.; Zhou, M.M. Writers and readers of histone acetylation: Structure, mechanism, and inhibition. *Cold Spring Harb. Perspect. Biol.* **2014**, *6*, a018762.
10. Jain, A.K.; Barton, M.C. Bromodomain histone readers and cancer. *J. Mol. Biol.* **2017**, *429*, 2003–2010.
11. Biswas, S.; Rao, C.M. Epigenetic tools (The Writers, The Readers and The Erasers) and their implications in cancer therapy. *Eur. J. Pharmacol.* **2018**, *837*, 8–24.
12. Seto, E.; Yoshida, M. Erasers of histone acetylation: The histone deacetylase enzymes. *Cold Spring Harb. Perspect. Biol.* **2014**, *6*, a018713.
13. Hyun, K.; Jeon, J.; Park, K.; Kim, J. Writing, erasing and reading histone lysine methylations. *Exp. Mol. Med.* **2017**, *49*, e324.
14. Park, S.Y.; Kim, J.S. A short guide to histone deacetylases including recent progress on class II enzymes. *Exp. Mol. Med.* **2020**, *52*, 204–212.
15. Yang, Q.; Yang, Y.; Zhou, N.; Tang, K.; Lau, W.B.; Lau, B.; Wang, W.; Xu, L.; Yang, Z.; Huang, S.; et al. Epigenetics in ovarian cancer: Premise, properties, and perspectives. *Mol. Cancer* **2018**, *17*, 109.
16. Miranda Furtado, C.L.; Dos Santos Luciano, M.C.; Silva Santos, R. Da; Furtado, G.P.; Moraes, M.O.; Pessoa, C. Epidrugs: targeting epigenetic marks in cancer treatment. *Epigenetics* **2019**, *14*, 1164–1176.
17. Lu, Y.; Chan, Y.T.; Tan, H.Y.; Li, S.; Wang, N.; Feng, Y. Epigenetic regulation in human cancer: The potential role of epi-drug in cancer therapy. *Mol. Cancer* **2020**, *19*, 79.

18. Martinez-Montiel, N.; Rosas-Murrieta, N.H.; Ruiz, M.A.; Monjaraz-Guzman, E.; Martinez-Contreras, R. Alternative splicing as a target for cancer treatment. *Int. J. Mol. Sci.* **2018**, *19*, 545.
19. Black, A.J.; Gamarra, J.R.; Giudice, J. More than a messenger: Alternative splicing as a therapeutic target. *Biochim. Biophys. Acta - Gene Regul. Mech.* **2019**, *1862*, 194395.
20. Tang, Z.; Zhao, J.; Pearson, Z.J.; Boskovic, Z. V; Wang, J. RNA-targeting splicing modifiers: Drug development and screening assays. *Molecules* **2021**, *26*, 2263.