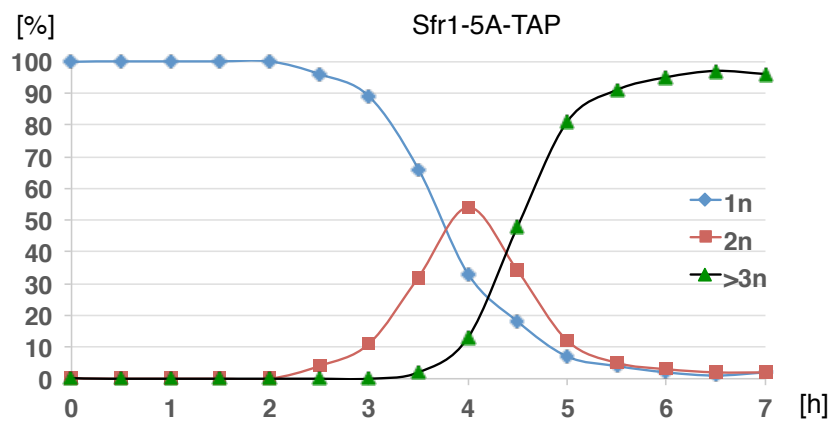


Figure S1

A



B

Sfr1-5A-TAP		unique peptides	coverage	phosphorylated residues
	Sfr1-5A (299 amino acids)	24	82%	S52 (or S48), T73, S135, S147 (or T146), T152, S175, S253
	Swi5 (85 amino acids)	7	60%	

Figure S1. Sfr1 phosphorylation sites identified by mass spectrometry.

(A) Haploid *pat1-114* cells expressing Sfr1-5A-TAP were arrested by nitrogen starvation and released into meiosis at 34°C by inactivation of Pat1. Small aliquots of the cell culture were harvested at the indicated time points (hours). Fixed cells were stained with DAPI and nuclei were counted in 100 cells per time point. Shown are the fractions of cells that contained one nucleus (1n), two nuclei (2n) or more than two nuclei (>3n) at the indicated time points.

(B) The cells were harvested around 3 hours after meiosis induction and Sfr1-5A-TAP was isolated by tandem affinity purification. Purified proteins were analyzed by mass spectrometry. Phosphorylation sites identified on Sfr1 by mass spectrometry are shown. For the full list of identified proteins see Table S1.

Figure S2

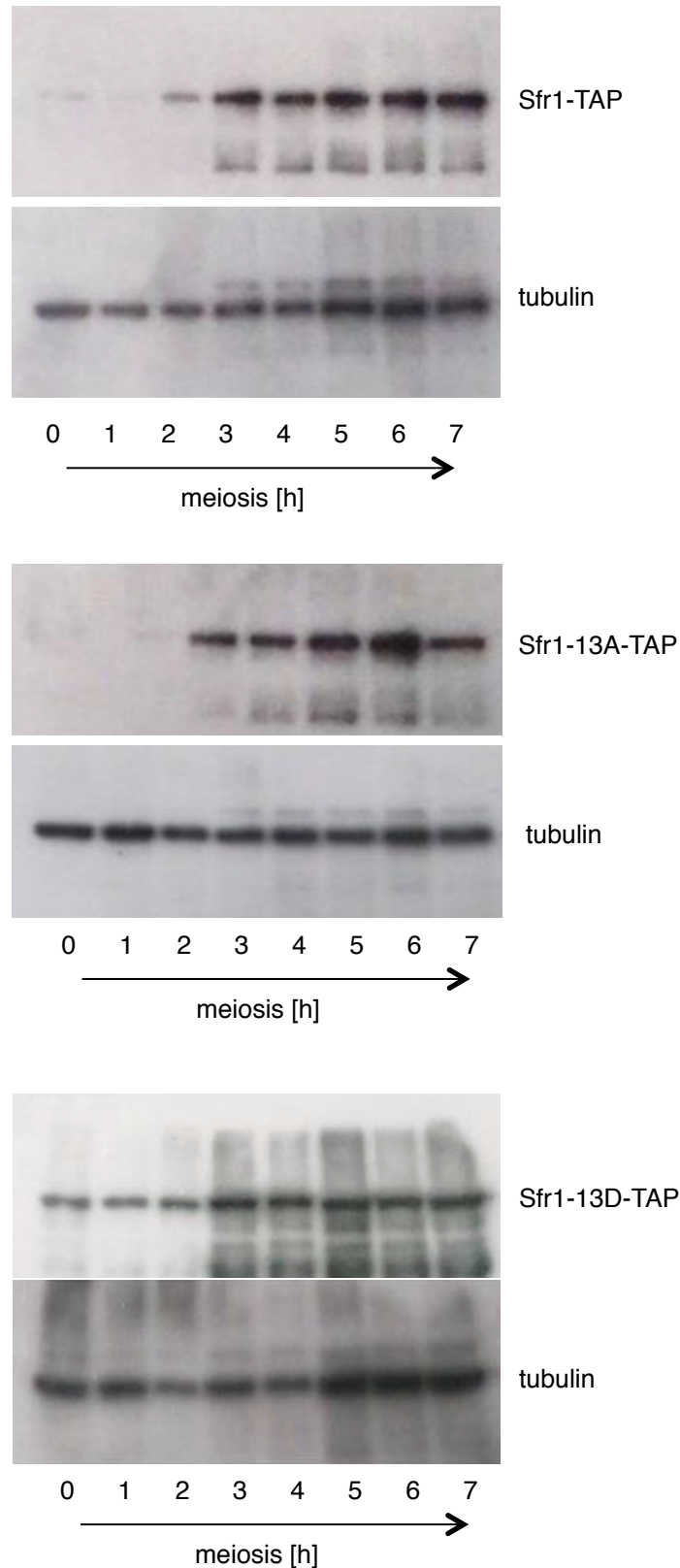


Figure S2. Expression of Sfr1-TAP, Sfr1-13A-TAP and Sfr1-13D-TAP proteins during meiosis.

pat1-114 cells expressing Sfr1-TAP, Sfr1-13A-TAP and Sfr1-13D-TAP were arrested by nitrogen starvation and released into meiosis at 34°C by inactivation of Pat1. Cells were harvested at the indicated time points (hours). Proteins extracted from meiotic cells were analyzed by gel electrophoresis and Western blotting using anti-tubulin antibodies. The TAP epitope was detected using PAP antibodies (rabbit antiperoxidase antibody linked to peroxidase).

Figure S3

A

Swi5

Sc (Sae3)	-----	0
Sp	-----	0
Mm	-----	0
Hs	MQRRGQRDLWRHNKSCARNRCPRPPRERGGAGFPWVRAQLSVRQFTLRVRVPGPVHLRGR	60
Sc (Sae3)	-----	0
Sp	-----	0
Mm	-----	0
Hs	SPTPALDPLAPLNPLIRGPRTPGLRRWIQSLALLLPNCSSSRIPTVPRPHSGLWVQSDFF	120
Sc (Sae3)	-----MNYLETQLNKKQKQIQEYE--SMNGNLIK	28
Sp	-----MEKSQLESRVHLEQQKEQLLESSLQDA	27
Mm	-----MIDENNDVSEALSSDIKKLKEKHDMLDKEISQL	34
Hs	LGFLSRTEPRLTRSCGAFRSRPLPKSGQADGTSEESLHLDIQKLKEKRDMLDKEISQF	180
	: * : : : . : : .	
Sc (Sae3)	FEQLSKEKKNDETPKKISSTYIKELKEYNELRDAGLRLAQIIADEKQCKIKDVFEEIGYS	88
Sp	LAKLKNRD-----AKQTVQKHIDLLHTYNEIRDIALGMIGKVAEHEKCTSVELFDRFGVN	82
Mm	---IAEGY-----RVIELEKHISLLHEYNDIKDVSQMLLGKLAIVRGVTTKELYPDFDLN	86
Hs	---VSEGY-----SVDELEDHITQLHEYNDIKDVGQMLMGKLAIVRGVTTKELYPEFGLD	232
	: : . : * : * : : : . : : * . . : : : . .	
Sc (Sae3)	MKD 91	
Sp	GSE 85	
Mm	LND 89	
Hs	MND 235	
	. :	

Sfr1

Sc (Mei5)	-----	0
Sp	MSQTINSELNENATSQCKEDLKVSLSESDDLRSQGLGIENPPKCNN---GNHSNGLGF	57
Mm	---MAEEGNQFTSKMENSS---DSASTSPDAPQPSNPSPPTSPAAPQTSNPPS	51
Hs	---MAEGEKNQDFTFKME-----	15
Sc (Mei5)	-----	0
Sp	IEQSETVHPENKA-----LTPDLRDT-----KIHTSLPITT-----	89
Mm	PPTSPAAPQPRENPPSPPTSPAAPQPRENPPSPPTSPAAPQPRENPPSPPTSPAAPQPRE	111
Hs	-----SPSDSAVVLPTPQASA	32
Sc (Mei5)	-----	0
Sp	-PFSKKRAREAKNILLKPFKSPLRQIASPQVADTNLKPSLAVTNLNSDETNTSSEPVTSP	148
Mm	NPPSPHSNSSGKQPLSGTPKERLKKARSSSHSFCSVVKRMKVENDENN--ETLSEPGESS	169
Hs	NPSSPYTNSSRKQPM SATLRERLRKTRFSNSSYNVVKRLKVESEEND--QTFSEKPASS	90
Sc (Mei5)	-----MHNQEEWLDKDKTL--VNEEENTCINHSTYTKKDTNNYRVGKSGIKD-LK	46
Sp	LR-----TPNSIKRQKRL-----F---KSPISNCLNPKSDP-----EITQLLS	184
Mm	KEENC SKAQESLKNKDEPGEKS-----SEEKNTCESKSDTGSSNALPKES-E-NAIIR	222
Hs	TEENCLEFQESFKHIDSEFEENTNLKNTLKNLVNCEQSQSDSGSCSALQNEF-VSEKLPK	149
	: : . . . * . .	
Sc (Mei5)	KPTNQKEIAIKNRELTKQLTLLRQENHLQACKILSENKIIENRKSIEKWRTICEMELS	106
Sp	R-----R-----LKLEKEVRNLQEQLITAETARKVEAKNEDKDLQTLIQKWNAAQQAEE	234
Mm	EKLKQEK-----IRLIRQVEEKEDLLRRLKLVKMYRIKNDVTELENLIKWRKCGQRLLC	277
Hs	QRLNAEK-----AKLVKQVQEKEDLLRRLKLVKMYRSKNDLSQLQLLIKWRSCSQQLLLY	204
	. . . * : : . : . : * . : . * : * : . :	
Sc (Mei5)	FILNSTLIKINRMGGYKDFLEKEMEAKRR-----L-EYQIDNGMEDQICEIKESD	156
Sp	VLFPKMAERIRLAGGVTSFRIE EGENKGQIQEVRTFTMSFLNQFGVPVHLMSFDE--	292
Mm	ELQSIMSED-----EDELTLTELIDFYGIDDNLHYNR--	311
Hs	ELQSAVSE-----ENKKLSLTQLIDHYGLDDKLLHYNR--	237
	: . : * : : . .	
Sc (Mei5)	DFRQLSEVEKQEWESQMNEQLKELEKKKIAELEKLNKVLHDSEGKDFGMAELCTRLKLDY	216
Sp	-----ENGDWKS-----	299
Mm	-----SEEEFTGV-----	319
Hs	-----SEEEFIDV-----	245
	. : : : .	
Sc (Mei5)	SLIFPQ 222	
Sp	----- 299	
Mm	----- 319	
Hs	----- 245	

B

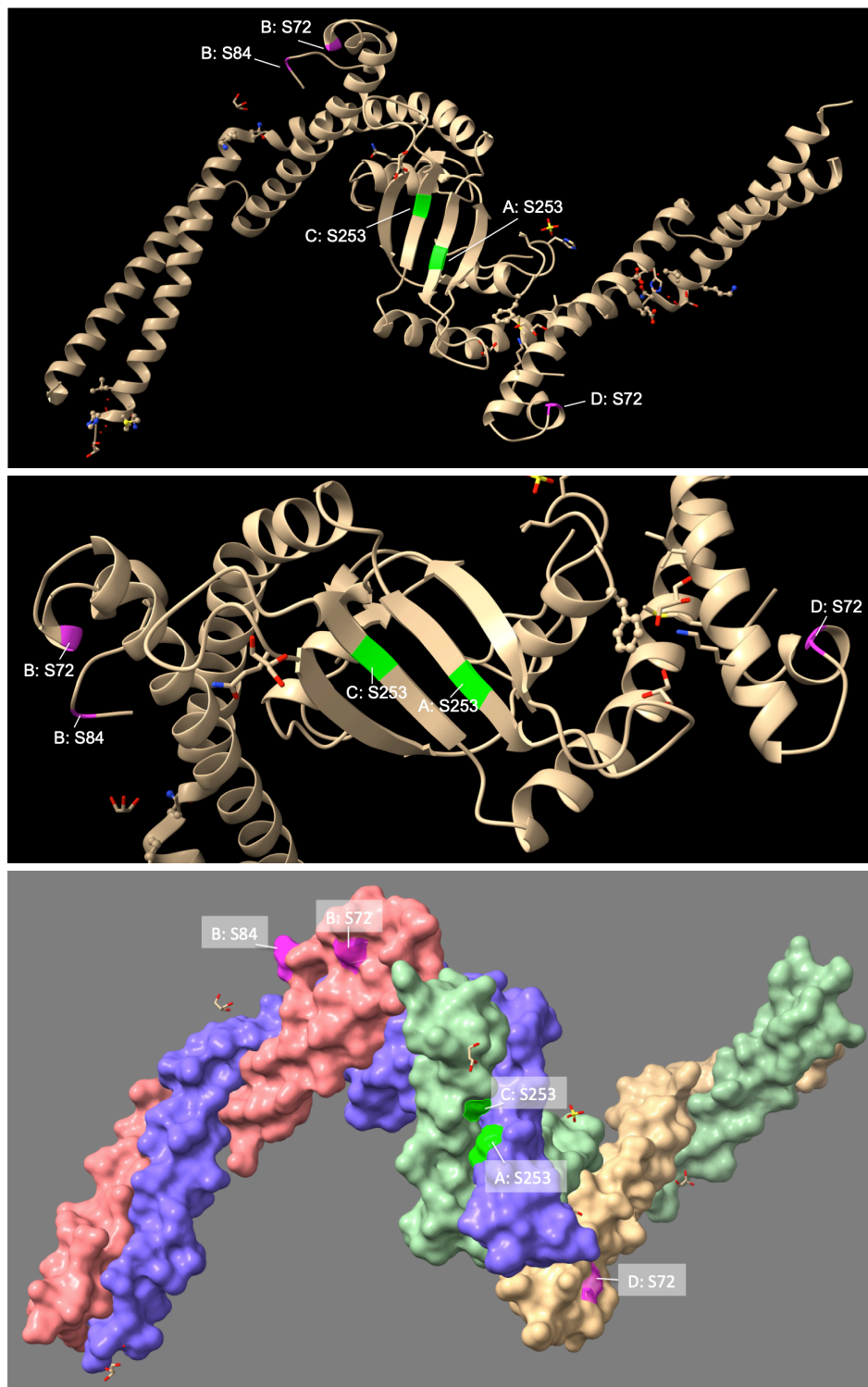


Figure S3. Visualization of Swi5 and Sfr1 phosphorylation sites.

(A) Swi5 and Sfr1 protein sequences from *Saccharomyces cerevisiae* (Sc), *Schizosaccharomyces pombe* (Sp), *Mus musculus* (Mm) and *Homo sapiens* (Hs) were aligned using Clustal Omega (Sievers et al., 2017; <https://pubmed.ncbi.nlm.nih.gov/21988835>). An asterisk (*) indicates positions of fully conserved residues, a colon (:) indicates conservation between residues of strongly similar properties and a period (.) indicates conservation between residues of weakly similar properties. Swi5 and Sfr1 phosphorylation sites are highlighted in red.

(B) Crystal structure of fission yeast Swi5 and its complex with the Sfr1C (C-terminal domain, residues 181-299) was retrieved from the RCSB Protein Data Bank (PDB ID: 3VIQ) (Kuwabara et al., 2012; <https://pubmed.ncbi.nlm.nih.gov/22405003>) and visualized using ChimeraX software (Pettersen et al., 2021; <https://pubmed.ncbi.nlm.nih.gov/32881101>). Presented are the two Swi5-Sfr1C heterodimers in the asymmetric unit. Swi5 and Sfr1 phosphorylation sites located on individual protein chains (denoted A, B, C and D) are indicated in magenta and green, respectively. Upper and middle figures: ribbon diagram of Swi5-Sfr1C, lower figure: surface representation of Swi5-Sfr1C.

Figure S4

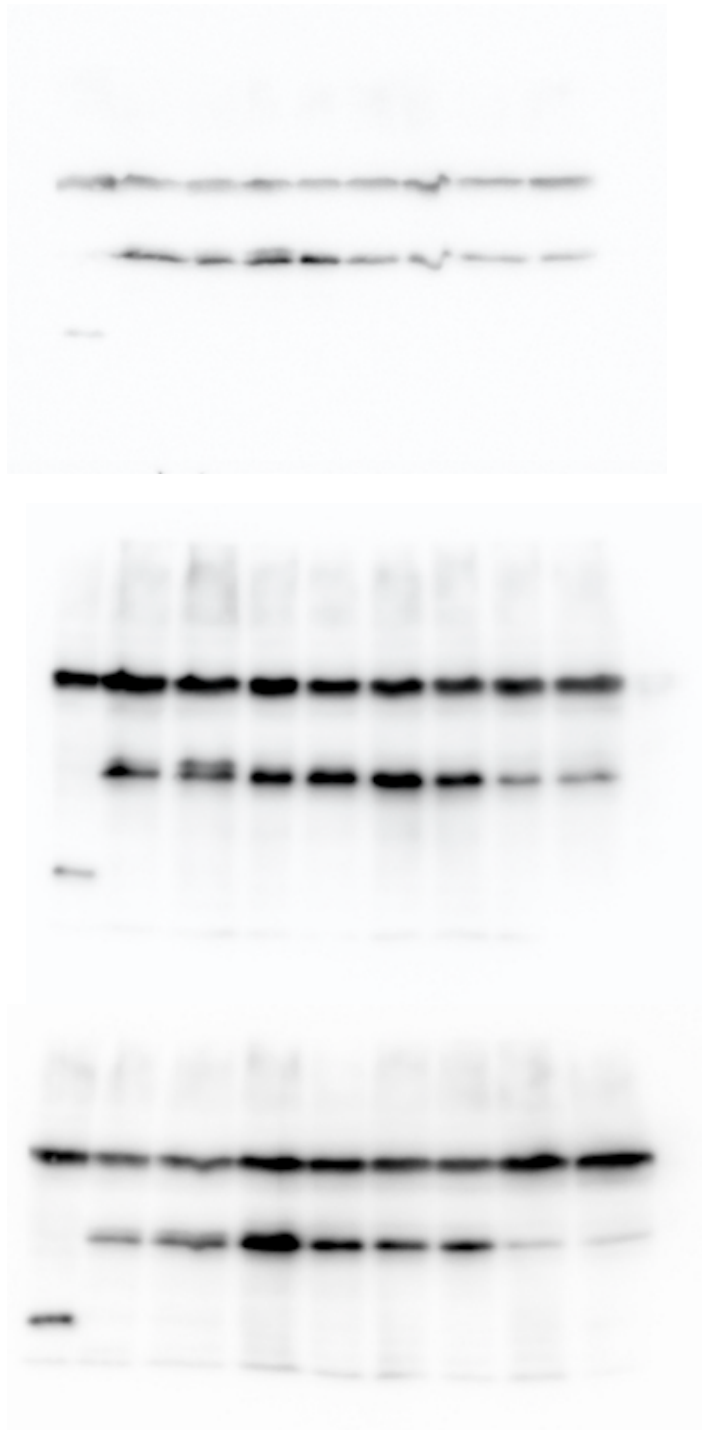


Figure S4. Full original images of Western blots shown in Figure 3.