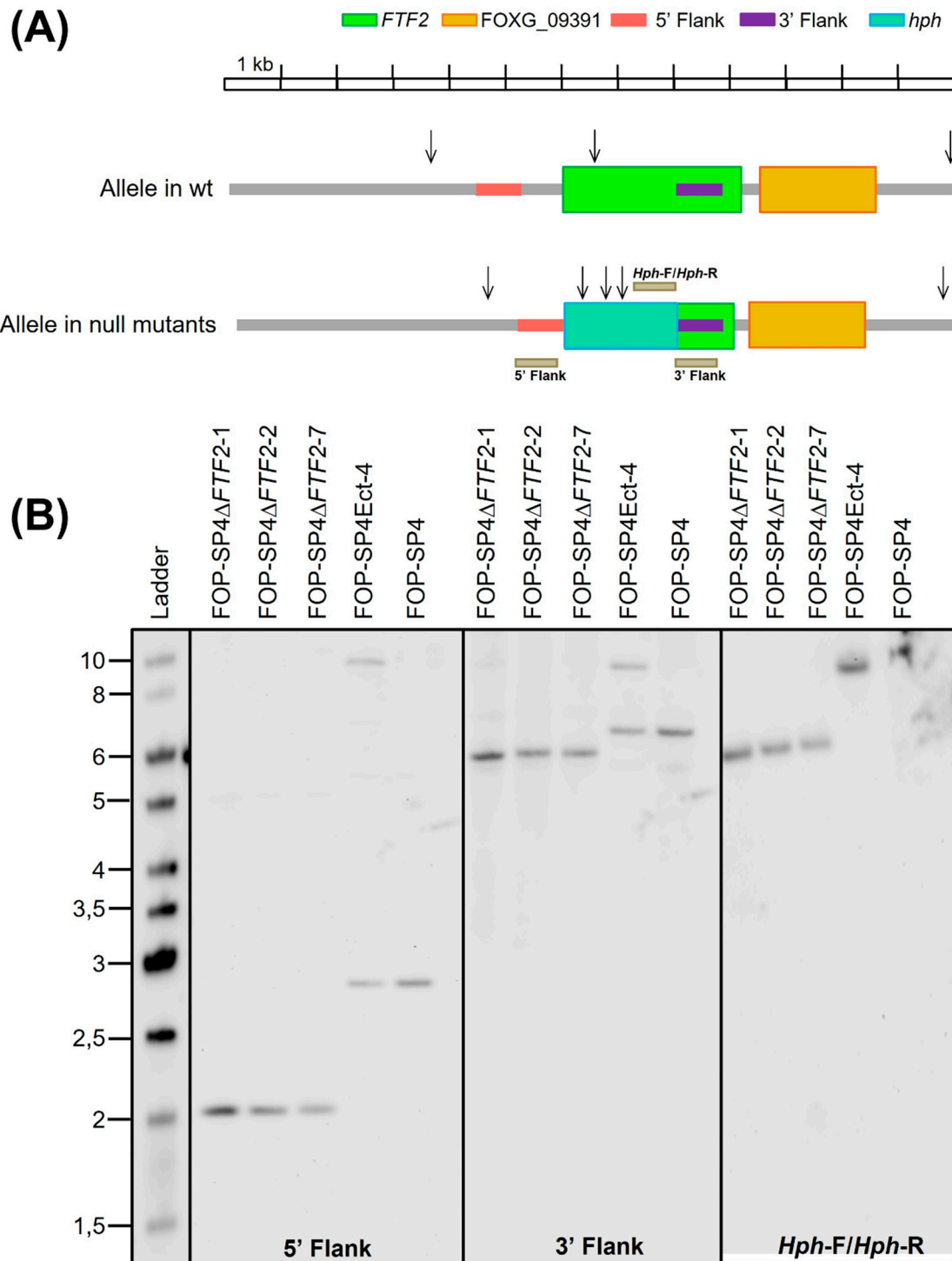
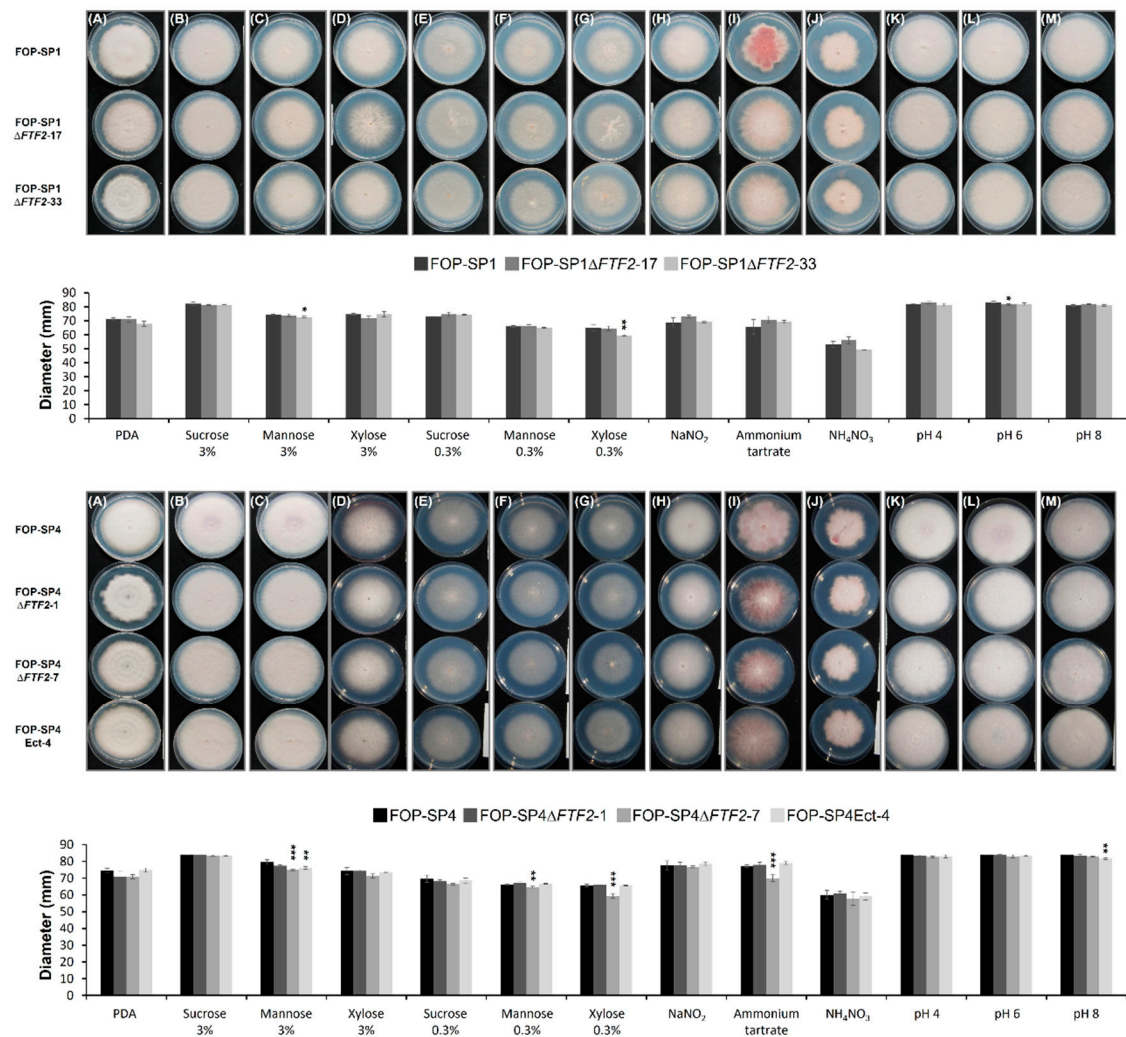


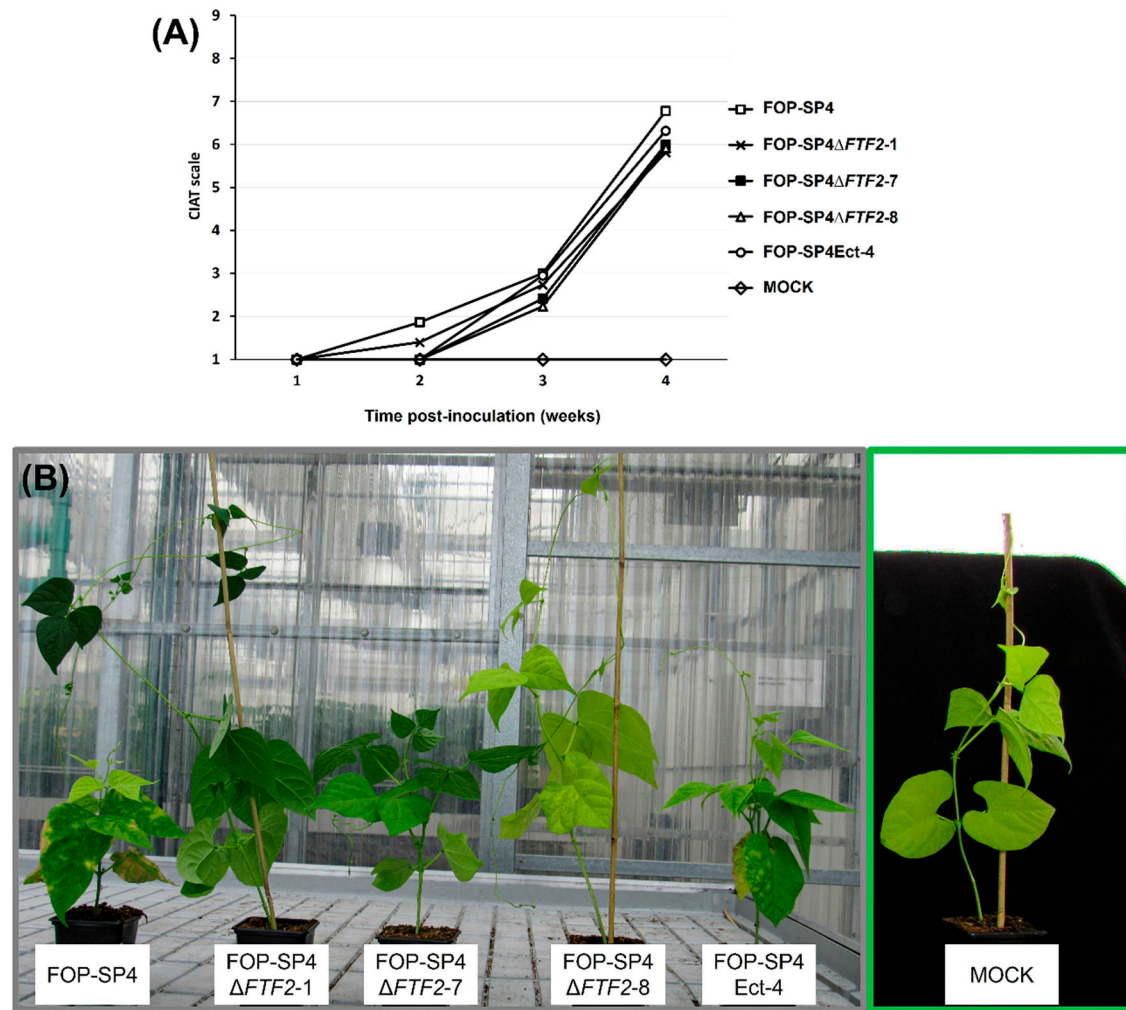
Supporting information



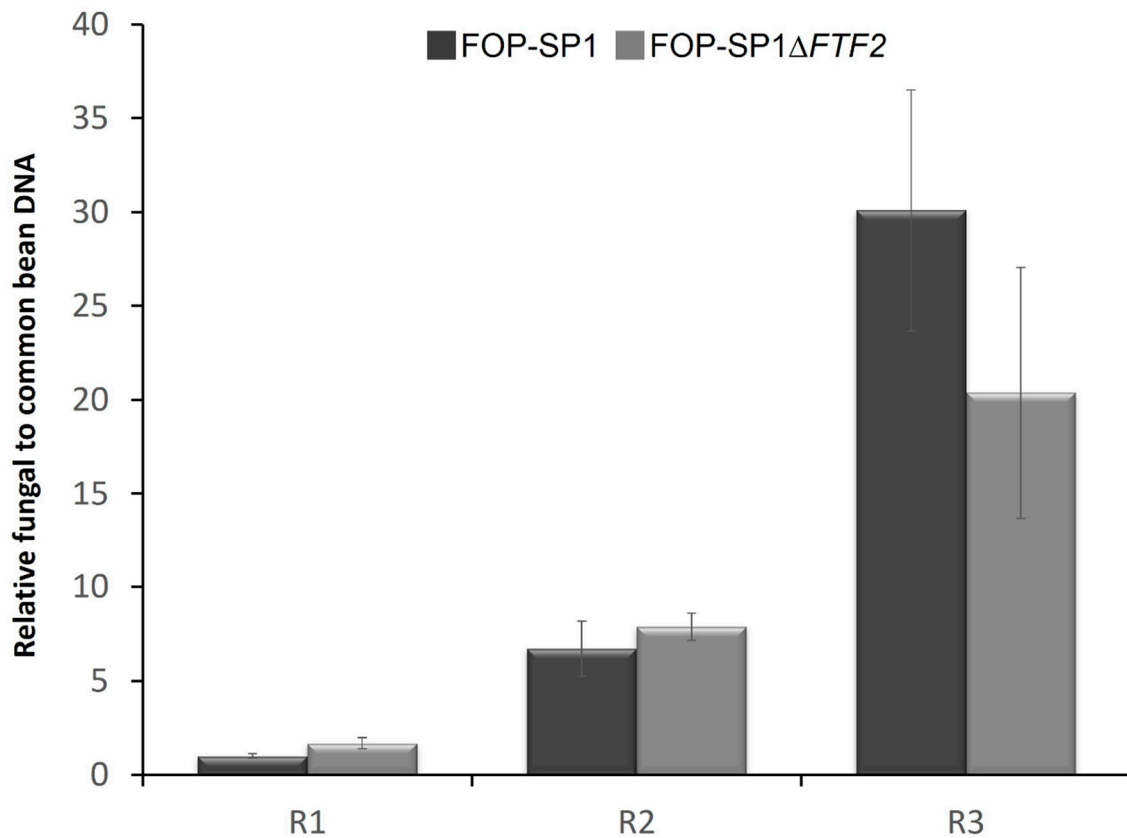
Supplementary Figure S1. (A) Schematic representation of the recognition sites of *Bam*HI (black arrows) in the wild-type allele of *FTF2* and the allele in the null mutants. Brown boxes show the location of the probes used in the Southern blot analysis. (B) Southern blot hybridization analysis of *DFTF2* mutants in FOP-SP4 strain. *Bam*HI-digested DNA from mutants FOP-SP4Δ*FTF2*-1, FOP-SP4Δ*FTF2*-2, FOP-SP4Δ*FTF2*-7 and FOP-SP4Ect-4 and from wild-type strain FOP-SP4 was hybridized with a probe corresponding to the 5' flanking region (probe 5' Flank), a probe corresponding to the 3' flanking region (probe 3' Flank) and a probe derived from the *hph* ORF (probe *Hph-F/Hph-R*). 'Ladder' identifies the molecular marker 1 kb DNA ladder (Biotools), sizes are shown in kb.



Supplementary Figure S2. Saprophytic growth on solid media of FOP-SP1ΔFTF2 and FOP-SP4ΔFTF2 mutant strains. For each media, colony diameter was determined 6 days post-inoculation. Bars represent the media \pm standard deviation of 3 independent biological experiments. Differences between mutant and control strains were test with an ANOVA analysis followed by a Dunnett's test and are indicated as * ($P < 0.05$), ** ($P < 0.01$) and *** ($P < 0.001$). Colony morphology and pigmentation were tested 6 dpi on PDA (A), on minimal media amended with 3% sucrose (B), 3% mannose (C), 3% xylose (D), 0.3% sucrose (E), 0.3% mannose (F) or 0.3% xylose (G), on minimal media amended with 3% sucrose and NaNO₂ (H), ammonium tartrate (I) or NH₄NO₃ (J), and on minimal media amended with NaNO₃ and 3% sucrose at pH 4 (K), pH 6 (L) or pH 8 (M).



Supplementary Figure S3. Fusarium wilt induced in common bean plants by *Fusarium oxysporum* f. sp. *phaseoli* *DFTF2* mutants derived from the weakly virulent wild type FOP-SP4. (A) Severity of Fusarium wilt disease symptoms measured by the CIAT (International Center for Tropical Agriculture) scale (the disease index is assigned according to the percentage of chlorotic or necrotic leaves). (B) Common bean plants 4 weeks after inoculation with three different *FTF2* null mutants derived from FOP-SP4: FOP-SP4 Δ FTF2-1, FOP-SP4 Δ FTF2-7 and FOP-SP4 Δ FTF2-8 and ectopic transformant obtained with the gene replacement construction (FOP-SP4Ect-4).



Supplementary Figure S4. Quantification of fungal biomass in *Phaseolus vulgaris* plants colonized by FOP-SP1 Δ FTF2 strains. Fungal DNA relative to that of common bean was measured by assaying the fungal *SGE1* gene and the plant *PR1* gene by RT-qPCR using DNA extracted from the root system at 1, 2 and 3 dpi (R1, R2 and R3). All measurements were referred to the value obtained for FOP-SP1 colonization at R1 (arbitrary value of 1.0).

Table S1. Oligonucleotides used in this work.

Oligonucleotide	Accession code or plasmid	Sequence (5'-3')
InacFTF2-O1	FOXG_09390 ^a	GGTCTTAAUGACAGCGACAACCCAAGCCAAAC
InacFTF2-O2	FOXG_09390 ^a	GGCATTAAUCCCTGCCCTGCGAGAGAGACAAG
InacFTF2-A3	FOXG_09390 ^a	GGACTTAAUATGAAGCCAACAGCGGAAGCC
InacFTF2-A4	FOXG_09390 ^a	GGGTTTAAUATATCCTGACCACCACCCAACCC
Hph-F	pRF-HU ^d	GCGCTTCTGCGGGCGATTTG
Hph-R	pRF-HU ^d	CGGGTTCGGCCCATTCGGAC
EF1alpha-Fwd	FOXG_03515 ^a	CATCGGCCACGTCGACTCT
EF1alpha-Rev	FOXG_03515 ^a	AGAACCCAGGCGTACTTGAA
Int-qFTF2-Fwd	FOXG_09390 ^a	CGCTGTGCCATCTCCCTCTCA

Int-qFTF2-Rev	FOXG_09390 ^a	CGTGGGCGTGGGCGTGAT
SIX1-Fwd	AJ608702 ^b	GAGCCGCCTCAATCGCCTG
SIX1-Rev	AJ608702 ^b	GGCCAAGTTGCGCGATATGTG
SIX6-Fwd	ACY39286,1 ^b	GCTTTTGCGTGGCGAACCC
SIX6-Rev	ACY39286,1 ^b	TTTTCCTGCTGAGATTGCG
SGE1-Fwd	FOXG_10510 ^a	CAGCCGTATCCTTGGCAACTA
SGE1-Rev	FOXG_10510 ^a	TGGTTGACTTGCCGTTCTT
14731-Fwd	FOXG_14731 ^a	TCGTCATATCGCCAGGGGTG
14731-Rev	FOXG_14731 ^a	CAACGTATCGTGAGCCCCGA
02748-Fwd	FOXG_02748 ^a	GATCCTTGCCCCGACAGTCT
02748-Rev	FOXG_02748 ^a	GCGCAGATGGCTTGGAAGTT
02746-Fwd	FOXG_02746 ^a	TCTTCGGAAACCCCCAGTGC
02746-Rev	FOXG_02746 ^a	CCTTCTGGCCAGACTCGGC
14730-Fwd	FOXG_14730 ^a	CGTCATATCGCCAGGGGTGA
14730-Rev	FOXG_14730 ^a	TCAACGTATCGTGAGCCCCG
04430-Fwd	FOXG_04430 ^a	CCAAGACCGCAAGGGTAACATGCT
04430-Rev	FOXG_04430 ^a	TCGAACTTGAAGGGATAGGATGTCGC
Actin-Fwd	Phvul.011G064500 _c	GAAGTTCTCTTCCAACCATCC
Actin-Rev	Phvul.011G064500 _c	TTTCCTTGCTCATTCTGTCCG
ERF1-Fwd	Phvul.007G127800 _c	CCTGTTGGGCTCTGAAGAGGAAAC
ERF1-Rev	Phvul.007G127800 _c	AGGACCAAGGTCTTCAAACACGAC
ERF2-Fwd	XM_003549886 ^d	GGGAAAGTTCGCGGCGGAG
ERF2-Rev	XM_003549886 ^d	CGGAGTTAACCCTCAACGGAAAATTC
PR1-Fwd	Phvul.003G109100 _c	CAGGCACTACACTCAGGTTGTTTGA
PR1-Rev	Phvul.003G109100 _c	TTGCCAGGAGGAGCATAGTTGCAA

(a) *Locus* in the genome of the strain 4287 of *F. oxysporum* f. sp. *lycopersici*; (b) *GenBank* accession number; (c) *locus* in the genome of *P. vulgaris*; (d) *locus* in the genome of *Glicine max*.

Table S2. Quantification of macroconidia (%) produced by FOP-SP1 Δ FTF2 strains.

	FOP-SP1	FOP-SP1 Δ FTF2-17	FOP-SP1 Δ FTF2-33
PDA	0	21.63 \pm 15.2 ***	22.66 \pm 5 ***
Glucose	0	78.07 \pm 6.4 ***	91.74 \pm 8.9 ***
Sucrose	0	81.31 \pm 10.9 ***	84.72 \pm 9.6 ***
Xylose	0	72.8 \pm 8 ***	80.48 \pm 8.8 ***
Mannose	0	75.59 \pm 5.4 ***	67.28 \pm 6.4 ***
Glycerol	0	81.81 \pm 16.2 ***	84.63 \pm 18.7 ***

Numbers show the percentage of macroconidia over the total harvested conidia per condition. The data represent the media \pm standard deviation of three independent biological experiments; differences were tested using an ANOVA analysis followed by a Dunnett's test and are indicated by *** ($P < 0.001$).