



Article

Clinicopathologic Characteristics of Epstein–Barr Virus-Associated Gastric Cancer Over the Past Decade in Japan

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Abstract: Epstein–Barr virus (EBV) is a ubiquitous human herpes virus, but related with several types of malignancies. Among EBV-related malignancies, EBV-associated gastric carcinoma (EBVaGC) has the largest patient’s number. We screened for EBV infection in 1067 GC lesions of 1132 patients who underwent surgical resection from 2007 to 2017 in Japan and examined clinicopathological features of EBVaGC. EBV infection was detected by *in situ* hybridization with EBV-encoded small RNA 1 (EBER-1 ISH). EBV was infected in 80 GC lesions (7.1%). Mean age was significantly lower in patients with EBVaGC than with EBV-negative GC. EBVaGC was more frequent in men than in women. EBVaGC was found twice as frequent in the upper or middle stomach as in the lower stomach. Early EBVaGC was more frequent, and submucosally invaded cases were dominant. The presence of lymphatic vessel invasion was less in EBVaGC, but frequency of lymph node metastasis was similar. Carcinoma with lymphoid stroma (CLS) was found in 3.8% (43/1132) of all lesions with 60.5% of EBV positivity. The synchronous or metachronous multiple GC was frequent in EBVaGC. We clarified clinicopathologic characteristics of EBVaGC over the past decade in Japan. EBV infection should be examined in gastric cancer cases showing these characteristics.

Keywords: Epstein–Barr virus (EBV); gastric carcinoma; carcinoma with lymphoid stroma; lymph node metastasis; immune checkpoint inhibitor

1. Introduction

Epstein–Barr virus (EBV) is a ubiquitous human herpes virus discovered from a Burkitt lymphoma cell. EBV is associated with a variety of tumors derived from B cells, such as Burkitt lymphoma, post-transplant lymphoproliferative disease, and Hodgkin’s disease. EBV is also detected from epithelial tumors, including nasopharyngeal carcinoma and gastric carcinoma (GC). Although not frequent, the virus is sometimes associated with peripheral T-cell lymphomas and nasal natural killer T-cell lymphomas [1,2].

Gastric cancer is the fifth frequently diagnosed cancer and the third leading cause of cancer deaths worldwide [3]. EBV-associated GC (EBVaGC) is about 10% of all GC worldwide [4,5]. EBVaGC is not endemic like EBV-related Burkitt lymphoma and nasopharyngeal carcinoma. Among EBV-related malignancies, EBVaGC is the most common. However, EBVaGC has not received clinical attention because there is no specific treatment for this malignancy.

The Cancer Genome Atlas (TCGA) 2014 divided gastric cancer into four types according to characterization based on molecular biology: EBVaGC, microsatellite unstable tumors, genomically stable tumors, and chromosomally unstable tumors. EBVaGC characteristically shows recurrent PIK3CA mutations, extreme DNA hypermethylation, and overexpression of each of JAK2, PD-L1, and PD-L2 [6]. Therefore, demethylating agents, PI3K or JAK2 inhibitors, and immune checkpoint inhibitors may be effective in the treatment of EBVaGC [7]. Selection of therapy based on the mechanism of tumor development is required, and it is important to diagnose EBVaGC prior to treatment.

EBVaGC shows that infrequent lymph node metastasis and EBV positivity in gastric cancers confers a better survival advantage than EBV-negative cancers [8,9]. Minimally invasive endoscopic treatment has been applied to early gastric cancer. Because endoscopic treatment is a local treatment inside the stomach, gastric cancer without lymph node metastasis is the principal indication [10]. The number of reports that mention infrequent lymph node metastasis in EBVaGC is increasing, especially in early-stage cancer [11,12]. Early-stage EBVaGC can be an indicator for more extensive endoscopic treatment than other types of gastric cancer.

Because important features of EBVaGC related to treatment strategies have been reported, it is necessary to diagnose EBVaGC before treatment. Thus, we examined EBV infection in a series of about 1000 gastric cancers over the past decades in Japan to clarify the clinicopathologic features of EBVaGC.

2. Materials and Methods

2.1. Patients

The subjects were 1067 GC lesions of 1132 patients with gastric cancer who underwent surgical resection at Yamaguchi University Hospital and Hofu Gastroenterology Center from 2007 to 2017. This retrospective study was approved by the institutional review board of Yamaguchi University Hospital (approval number: H26-119-3) (Date: 24 December 2014).

2.2. EBV-Encoded Small RNA 1 (EBER-1) *in Situ* Hybridization

The presence of EBV was determined using *in situ* hybridization (ISH) with EBV-encoded small RNA 1 (EBER-1), which is known to be present in large amounts in EBV-infected cells. EBER-1 was detected with a biotin- or digoxigenin-labeled 30-base oligomer using previously described procedures [13]. Paraffin-embedded 4-mm sections were deparaffinized, rehydrated, predigested with pronase, prehybridized, and then hybridized overnight at 37 °C. After washing with 0.5× SSC, hybridization was detected according to the manufacturer's instructions. EBVaGC was diagnosed by EBER-1 ISH, where most of tumor cell nuclei were positively stained.

2.3. Evaluation of Clinicopathologic Findings

The locations of the GCs were divided into three areas: upper, middle, and lower stomach. We had 28 GC lesions developed in the remnant stomach after partial gastrectomy. Histologic types were divided into differentiated and undifferentiated types. Carcinoma with lymphoid stroma (CLS), which is considered to be a pathological feature of EBVaGC, was classified separately. Lesions with cancerous invasion limited to the mucosa or submucosa were classified as early, and lesions with deeper invasion as advanced. The macroscopic types of the GCs were classified according to the Japanese Classification of Gastric Carcinoma [14].

2.4. Statistics

Data were analyzed using the Mann–Whitney U test. A p value of 0.05 was considered significant.

3. Results

A large series of 1132 GC lesions in 1067 consecutive patients was studied. EBV was detected in 80 GC lesions (7.1%) by EBER-1 ISH. EBER-1 signals were observed uniformly in the nuclei of tumor cells in these lesions. EBV was not present in infiltrating lymphocytes of the stroma. Epithelial cells in the adjacent mucosa were EBER-1 negative (Figures 1 and 2).

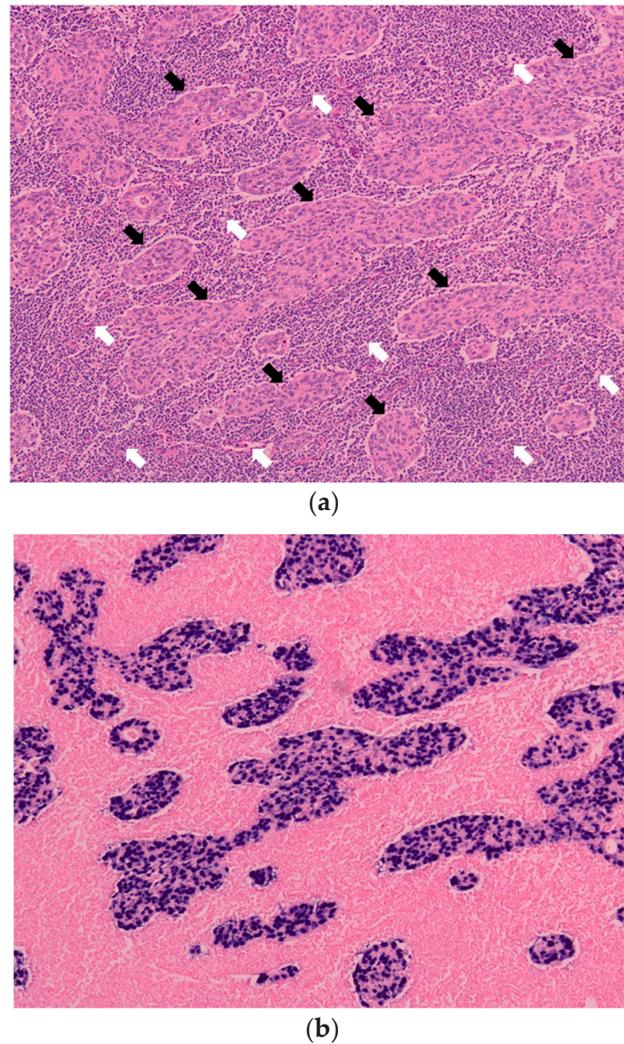
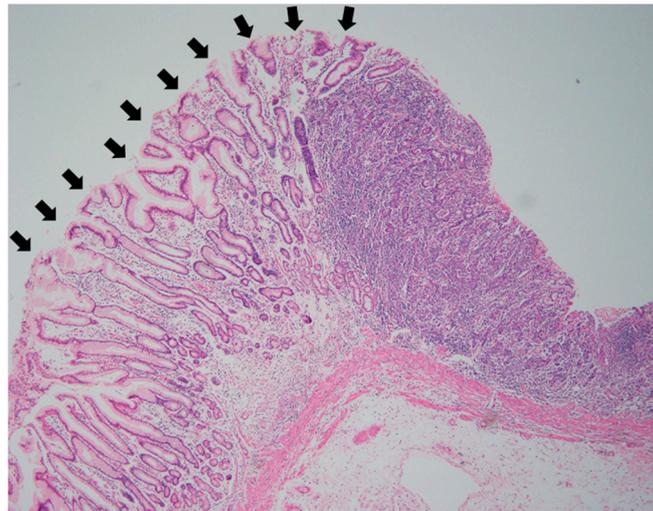
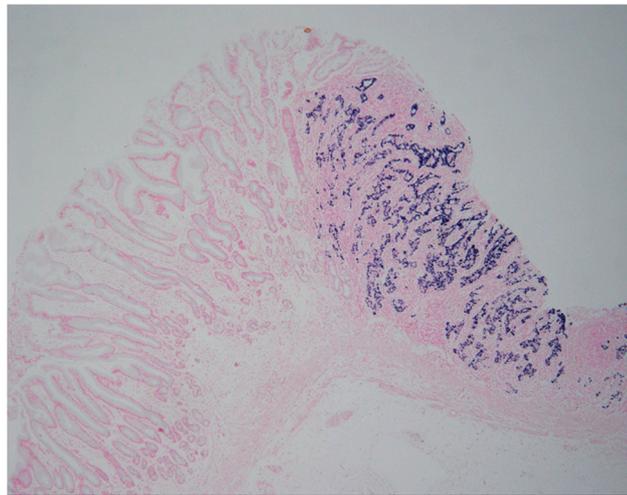


Figure 1. Histologic characteristics of Epstein-Barr virus-associated gastric carcinoma (EBVaGC). (a) Histology of carcinoma with lymphoid stroma (CLS). Undifferentiated adenocarcinomas (black arrows) are surrounded by infiltrating lymphocytes (white arrows). (H&E staining); (b) EBV-encoded small ribonucleic acid 1 in situ hybridization (EBER-1 ISH). Signals of EBER-1 are observed in nuclei of the carcinoma cells indicated by black arrows in (a). Infiltrating lymphocytes in the stroma indicated by white arrows in (a) are EBER-1 negative. Magnification (a) and (b)($\times 100$).



(a)



(b)

Figure 2. Epstein–Barr virus-associated gastric carcinoma (EBVaGC). (a) Moderately differentiated adenocarcinoma is composed of irregular tubules and moderate lymphocytic infiltration. Black arrows indicate non-cancerous epithelium; (b) Dark-blue signals of EBV encoded small RNA 1 (EBER-1) are observed in cancerous area. Epithelial cells in the non-cancerous mucosa indicated by black arrows in (a) are EBER-1 negative. Magnification (a) and (b) ($\times 40$).

The mean age of the patients with EBVaGC was significantly lower than that of patients with EBV-negative gastric cancers. EBVaGC was mostly detected in male patients, and it arose twice as frequent in the upper or middle stomach as in the lower stomach. The rate of early cancer was significantly high for EBVaGC, and submucosally invaded cancers were predominant. The degree of tumor differentiation and macroscopic type were similar. EBVaGC showed a significantly infrequent lymphatic vessel invasion and the frequency of lymph node metastasis was similar. The frequency of synchronous or metachronous multiple GC was higher in EBVaGC than in the EBV-negative cancers (Table 1).

Table 1. Clinicopathologic features of Epstein–Barr virus (EBV)-associated gastric cancer.

| Feature | Total | EBV Positive | EBV Negative | p Value |
|-----------------------|--------------|--------------|--------------|---------|
| Sex | | | | |
| Male | 735 (68.8%) | 61 (88.4%) | 674 (67.5%) | 0.022 |
| Female | 332 (31.2%) | 8 (11.6%) | 324 (32.5%) | |
| Age (yrs, mean) | | 65.6 | 68.9 | <0.001 |
| Location | | | | |
| U | 228 (20.1%) | 30 (37.5%) | 198 (18.8%) | <0.001 |
| M | 520 (45.9%) | 40 (50.0%) | 480 (45.6%) | |
| L | 356 (31.4%) | 6 (7.5%) | 350 (33.3%) | |
| Remnant stomach | 28 (2.5%) | 4 (5.0%) | 24 (2.3%) | |
| Stage | | | | |
| Early | 603 (58.6%) | 52 (65.0%) | 551 (52.4%) | 0.029 |
| Advanced | 529 (41.4%) | 28 (35.0%) | 501 (47.5%) | |
| Depth of invasion | | | | |
| m | 318 (28.1%) | 19 (23.7%) | 299 (28.4%) | <0.001 |
| sm | 285 (25.2%) | 33 (41.3%) | 252 (24.0%) | |
| mp | 126 (11.1%) | 8 (10.0%) | 118 (11.2%) | |
| ss | 203 (17.9%) | 10 (12.5%) | 193 (18.3%) | |
| sei | 200 (17.7%) | 10 (12.5%) | 190 (18.1%) | |
| Macroscopic type | | | | |
| Protruded | 192 (17.0%) | 11 (13.8%) | 181 (17.2%) | 0.428 |
| Depressed | 940 (83.0%) | 69 (86.2%) | 871 (82.8%) | |
| Pathologic type | | | | |
| Differentiated | 610 (53.9%) | 37 (46.3%) | 573 (54.5%) | 0.155 |
| Undifferentiated | 522 (46.1%) | 43 (53.7%) | 479 (45.5%) | |
| Lymphatic invasion | | | | |
| Presence | 536 (47.3%) | 25 (31.3%) | 511 (48.6%) | 0.003 |
| Absence | 596 (52.7%) | 55 (68.7%) | 541 (51.4%) | |
| Venous invasion | | | | |
| Presence | 533 (47.1%) | 25 (31.3%) | 408 (38.8%) | 0.182 |
| Absence | 599 (52.9%) | 55 (68.7%) | 644 (61.2%) | |
| Lymph node metastasis | | | | |
| Presence | 445 (41.6%) | 25 (37.3%) | 420 (42.1%) | 0.169 |
| Absence | 622 (58.4%) | 44 (63.7%) | 578 (57.9%) | |
| CLS | | | | |
| Presence | 43 (3.8%) | 26 (32.5%) | 17 (1.6%) | <0.001 |
| Absence | 1089 (96.2%) | 54 (67.5%) | 1035 (98.4%) | |
| Multiple cancers | | | | |
| Presence | 60 (5.6%) | 10 (14.5%) | 50 (5.0%) | <0.001 |
| Absence | 1007 (94.4%) | 59 (85.5%) | 948 (95.0%) | |

EBV: Epstein-Barr virus; CLS: carcinoma with lymphoid stroma; m: mucosa; sm: submucosa; mp: muscularis propria; ss: subserosa; sei: serosal exposed cancer infiltrating to the adjacent organ.

Carcinoma with lymphoid stroma (CLS) was classified separately. CLS was found in 3.8% (43/1132) of all lesions, and 60.5% (26/43) of CLS was positive for EBV Figure 3 and Table 1.

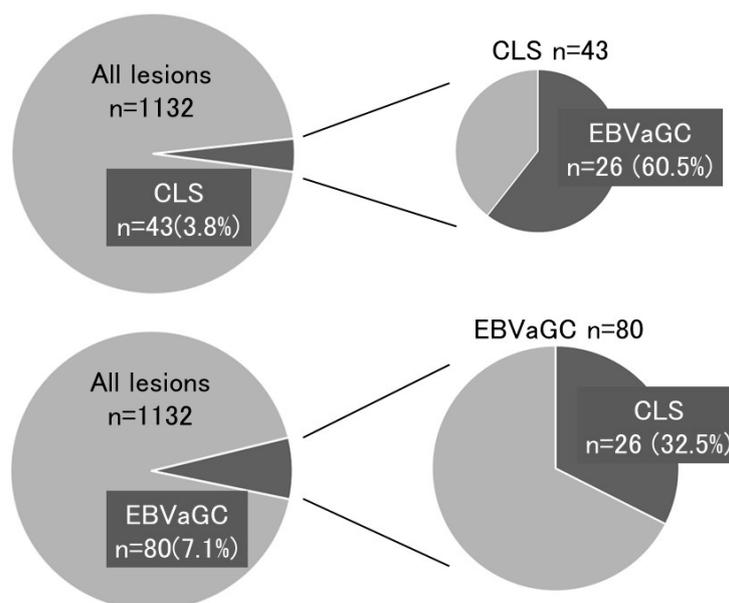


Figure 3. Relationship between Epstein–Barr virus-associated gastric carcinoma (EBVaGC) and carcinoma with lymphoid stroma (CLS).

4. Discussion

EBV was detected in 80 GC lesions (7.1%). The results shown in this study were almost identical to the previous meta-analytical examinations [5]. In our cohort, early stage EBVaGC was more frequent, and submucosally invaded cancers were dominant. Synchronous or metachronous multiple GC were frequently found. Because frequent recurrence of EBVaGC is an important issue, careful follow-up of the remnant stomach after surgical or endoscopic resection of EBVaGC is necessary [15,16].

The characteristic pathological feature of EBVaGC is lymphocyte infiltration (CLS) [17–20]. Therefore, if this histopathologic feature is observed in the clinical setting, the patient is examined for EBV infection. Patients are also examined for EBV infection when endoscopic and endosonographic features reflecting the histology of CLS are found. As EBVaGC is comprised of a poorly differentiated tumor mass that accompanies infiltrating lymphocytes in a submucosal or deeper layer, EBVaGC exhibits a submucosal tumor-like morphology on endoscopic observation or a well-defined hypoechoic mass in the submucosa on endosonographic examination [21,22]. Watanabe et al. showed that CLS was found in 4% of gastric carcinomas removed surgically [23]. The present cohort showed frequencies of 3.8% for CLS and 7.1% for EBVaGC (Figure 3), indicating that EBVaGC is made up of two histological lesions, CLS, and ordinary gastric cancer at a similar frequency. Thus, if EBVaGC is searched simply according to the characteristics of CLS, half of EBVaGC cases will be undetected.

EBVaGC is classified into one of the molecular subtypes in TCGA, and overexpression of PD-L1 has been shown in this cancer [24–28]. Responders to immune checkpoint inhibitors show high PD-L1 expression in tumor cells, high lymphocyte infiltration in tumor stroma, and high IFN- γ content in tumors [29–32]. EBVaGC exhibits all of these characteristics of responders. In a prospective phase II clinical trial of pembrolizumab, dramatic responses were observed in patients with EBVaGC and microsatellite instability-high (MSI-H) tumors [33]. It is desirable to check for EBV infection prior to chemotherapy because EBVaGC can become a good indication to use immune checkpoint inhibitors. Unresectable recurrent gastric cancer that will be treated by chemotherapy need to be investigated for EBV infection.

In the present study, the rate of early gastric cancer was high in EBVaGC, with 33 cases of submucosal EBVaGC. Endoscopic local treatment has recently been expanded to treat submucosally invaded gastric cancer and can be applied to patients without lymph node metastasis. The rate of lymph node metastasis of submucosal gastric cancer is usually 20%, whereas that of the current

cases of EBV-positive submucosal gastric cancer was 12.1%. Regarding to the submucosal cancer without vascular invasion, only 1 case (3.0%) was found to show lymph node metastasis. According to Tokunaga et al., there was no lymph node metastasis in early EBVaGC [11]. Osumi et al. analyzed 847 consecutive patients with submucosal GC without vascular invasion in a multicenter observational study and showed that EBVaGC accounted for 11.3% (96/847) of cases, and lymph node metastasis was less frequent in EBVaGC versus non-EBVaGC (1 of 96, 1.0% vs. 71 of 751, 9.5%) [34]. For the treatment of submucosally invaded gastric cancer with EBV and without lymphovascular invasion, endoscopic local treatment might be applied instead of surgical resection.

Endoscopic examination is mainly used for diagnosing GC. However, treatment strategies are decided by pathologic diagnosis of biopsy samples. The macroscopic findings in the present study were too diverse to find specific features. It is not easy to predict EBVaGC based on clinical characteristics. Only half of EBVaGC can be diagnosed, if there were the pathological findings such as CLS. Because it is not practical to perform EBER-1 ISH for all GC patients, we should carefully focus on cases where treatment strategies may change depending on the EBV infection. Patients with unresectable recurrent gastric cancer and off-label lesions after endoscopic treatment should be examined for EBV infection.

5. Conclusions

We clarified the clinicopathological features of EBVaGC in a series of about 1000 gastric cancers over the past decades in Japan. EBV infection should be examined for gastric cancer showing certain characteristics.

Author Contributions: A.Y., J.N. and H.Y. conceived and designed the manuscript. K.S., T.S., T.T., F.T., Y.K., M.Y., and Y.S. collected and assembled the data. O.M., H.Y., T.Y., and I.S. critically revised and approved the final article.

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Conflicts of Interest: The authors declare no financial or commercial conflict of interest.

References

1. Rickinson, A.B.; Kieff, E. Epstein-Barr virus. In *Fields Virology*, 5th ed.; Fields, B.N., Knipe, D.M., Howley, P.M., Eds.; Lippincott-Williams & Wilkins: Philadelphia, PA, USA, 2007; Volume 2, pp. 2655–2700.
2. Zur Hausen, H.; Schulte-Holthausen, H.; Klein, G.; Henle, W.; Henle, G.; Clifford, P.; Santesson, L. EBV DNA in biopsies of Burkitt tumours and anaplastic carcinomas of the nasopharynx. *Nature* **1970**, *228*, 1056–1058. [[CrossRef](#)]
3. Bray, F.; Ferlay, J.; Soerjomataram, I.; Siegel, R.L.; Torre, L.A.; Jemal, A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J. Clin.* **2018**, *68*, 394–424. [[CrossRef](#)]
4. Takada, K. Epstein-Barr virus and gastric carcinoma. *Mol. Pathol.* **2000**, *53*, 255–261. [[CrossRef](#)]
5. Murphy, G.; Pfeiffer, R.; Camargo, M.C.; Rabkin, C.S. Meta-analysis shows that prevalence of Epstein-Barr virus-positive gastric cancer differs based on sex and anatomic location. *Gastroenterology* **2009**, *137*, 824–833. [[CrossRef](#)]
6. Cancer Genome Atlas Research Network. Comprehensive molecular characterization of gastric adenocarcinoma. *Nature* **2014**, *513*, 202–209. [[CrossRef](#)]
7. Nishikawa, J.; Iizasa, H.; Yoshiyama, H.; Shimokuri, K.; Kobayashi, Y.; Sasaki, S.; Nakamura, M.; Yanai, H.; Sakai, K.; Suehiro, Y.; et al. Clinical importance of Epstein-Barr virus-associated gastric cancer. *Cancers* **2018**, *10*, 167. [[CrossRef](#)]
8. Van Beek, J.; zur Hausen, A.; Klein Kranenbarg, E.; van de Velde, C.J.; Middeldorp, J.M.; van den Brule, A.J.; Meijer, C.J.; Bloemena, E. EBV-positive gastric adenocarcinomas: A distinct clinicopathologic entity with a low frequency of lymph node involvement. *J. Clin. Oncol.* **2004**, *22*, 664–670. [[CrossRef](#)]
9. Camargo, M.C.; Kim, W.H.; Chiaravalli, A.M.; Kim, K.M.; Corvalan, A.H.; Matsuo, K.; Yu, J.; Sung, J.J.; Herrera-Goepfert, R.; Meneses-Gonzalez, F.; et al. Improved survival of gastric cancer with tumour Epstein-Barr virus positivity: An international pooled analysis. *Gut* **2014**, *63*, 236–243. [[CrossRef](#)]

10. Tada, M.; Murakami, A.; Karita, M.; Yanai, H.; Okita, K. Endoscopic resection of early gastric cancer. *Endoscopy* **1993**, *25*, 445–450. [[CrossRef](#)]
11. Tokunaga, M.; Land, C.E. Epstein-Barr virus involvement in gastric cancer: Biomarker for lymph node metastasis. *Cancer Epidemiol. Biomarkers Prev.* **1998**, *7*, 449–450.
12. Park, J.H.; Kim, E.K.; Kim, Y.H.; Kim, J.H.; Bae, Y.S.; Lee, Y.C.; Cheong, J.H.; Noh, S.H.; Kim, H. Epstein-Barr virus positivity, not mismatch repair-deficiency, is a favorable risk factor for lymph node metastasis in submucosa-invasive early gastric cancer. *Gastric Cancer.* **2016**, *19*, 1041–1051. [[CrossRef](#)]
13. Shibata, D.; Weiss, L.M. Epstein-Barr virus-associated gastric adenocarcinoma. *Am. J. Pathol.* **1992**, *140*, 769–774.
14. Japanese Gastric Cancer Association. Japanese classification of gastric carcinoma: 3rd English edition. *Gastric Cancer* **2011**, *14*, 101–112. [[CrossRef](#)]
15. Arikawa, J.; Tokunaga, M.; Tashiro, Y.; Tanaka, S.; Sato, E.; Haraguchi, K.; Yamamoto, A.; Toyohira, O.; Tsuchimochi, A. Epstein-Barr virus-positive multiple early gastric cancers and dysplastic lesions: A case report. *Pathol. Int.* **1997**, *47*, 730–734. [[CrossRef](#)]
16. Kaizaki, Y.; Hosokawa, O.; Sakurai, S.; Fukayama, M. Epstein-Barr virus-associated gastric carcinoma in the remnant stomach: De novo and metachronous gastric remnant carcinoma. *J. Gastroenterol.* **2005**, *40*, 570–577. [[CrossRef](#)]
17. Shibata, D.; Tokunaga, M.; Uemura, Y.; Sato, E.; Tanaka, S.; Weiss, L.M. Association of Epstein-Barr virus with undifferentiated gastric carcinomas with intense lymphoid infiltration. Lymphoepithelioma-like carcinoma. *Am. J. Pathol.* **1991**, *139*, 469–474.
18. Oda, K.; Tamaru, J.; Takenouchi, T.; Mikata, A.; Nunomura, M.; Saitoh, N.; Sarashina, H.; Nakajima, N. Association of Epstein-Barr virus with gastric carcinoma with lymphoid stroma. *Am. J. Pathol.* **1993**, *143*, 1063–1071.
19. Nakamura, S.; Ueki, T.; Yao, T.; Ueyama, T.; Tsuneyoshi, M. Epstein-Barr virus in gastric carcinoma with lymphoid stroma. Special reference to its detection by the polymerase chain reaction and in situ hybridization in 99 tumors, including a morphologic analysis. *Cancer* **1994**, *73*, 2239–2249. [[CrossRef](#)]
20. Matsunou, H.; Konishi, F.; Hori, H.; Ikeda, T.; Sasaki, K.; Hirose, Y.; Yamamichi, N. Characteristics of Epstein-Barr virus-associated gastric carcinoma with lymphoid stroma in Japan. *Cancer* **1996**, *77*, 1998–2004. [[CrossRef](#)]
21. Yanai, H.; Nishikawa, J.; Mizugaki, Y.; Shimizu, N.; Takada, K.; Matsusaki, K.; Toda, T.; Matsumoto, Y.; Tada, M.; Okita, K. Endoscopic and pathologic features of Epstein-Barr virus-associated gastric carcinoma. *Gastrointest. Endosc.* **1997**, *45*, 236–242. [[CrossRef](#)]
22. Nishikawa, J.; Yanai, H.; Mizugaki, Y.; Takada, K.; Tada, M.; Okita, K. Case report: Hypoechoic submucosal nodules: A sign of Epstein-Barr virus-associated early gastric cancer. *J. Gastroenterol. Hepatol.* **1998**, *13*, 585–590. [[CrossRef](#)]
23. Watanabe, H.; Enjoji, M.; Imai, T. Gastric carcinoma with lymphoid stroma. Its morphologic characteristics and prognostic correlations. *Cancer* **1976**, *38*, 232–243. [[CrossRef](#)]
24. Kawazoe, A.; Kuwata, T.; Kuboki, Y.; Shitara, K.; Nagatsuma, A.K.; Aizawa, M.; Yoshino, T.; Doi, T.; Ohtsu, A.; Ochiai, A. Clinicopathological features of programmed death ligand 1 expression with tumor-infiltrating lymphocyte, mismatch repair, and Epstein-Barr virus status in a large cohort of gastric cancer patients. *Gastric Cancer.* **2017**, *20*, 407–415. [[CrossRef](#)]
25. Böger, C.; Behrens, H.M.; Mathiak, M.; Krüger, S.; Kalthoff, H.; Röcken, C. PD-L1 is an independent prognostic predictor in gastric cancer of Western patients. *Oncotarget.* **2016**, *7*, 24269–24283. [[CrossRef](#)]
26. Derks, S.; Liao, X.; Chiaravalli, A.M.; Xu, X.; Camargo, M.C.; Solcia, E.; Sessa, F.; Fleitas, T.; Freeman, G.J.; Rodig, S.J.; et al. Abundant PD-L1 expression in Epstein-Barr Virus-infected gastric cancers. *Oncotarget.* **2016**, *7*, 32925–32932. [[CrossRef](#)]
27. Ma, C.; Patel, K.; Singhi, A.D.; Ren, B.; Zhu, B.; Shaikh, F.; Sun, W. Programmed Death-Ligand 1 Expression Is Common in Gastric Cancer Associated with Epstein-Barr Virus or Microsatellite Instability. *Am. J. Surg. Pathol.* **2016**, *40*, 1496–1506. [[CrossRef](#)]
28. Saito, R.; Abe, H.; Kunita, A.; Yamashita, H.; Seto, Y.; Fukayama, M. Overexpression and gene amplification of PD-L1 in cancer cells and PD-L1+ immune cells in Epstein-Barr virus-associated gastric cancer: The prognostic implications. *Mod Pathol.* **2017**, *30*, 427–439. [[CrossRef](#)]

29. Kang, Y.K.; Boku, N.; Satoh, T.; Ryu, M.H.; Chao, Y.; Kato, K.; Chung, H.C.; Chen, J.S.; Muro, K.; Kang, W.K.; et al. Nivolumab in patients with advanced gastric or gastro-oesophageal junction cancer refractory to, or intolerant of, at least two previous chemotherapy regimens (ONO-4538-12, ATTRACTION-2): A randomised, double-blind, placebo-controlled, phase 3 trial. *Lancet* **2017**, *390*, 2461–2471. [[CrossRef](#)]
30. Lawrence, M.S.; Stojanov, P.; Polak, P.; Kryukov, G.V.; Cibulskis, K.; Sivachenko, A.; Carter, S.L.; Stewart, C.; Mermel, C.H.; Roberts, S.A.; et al. Mutational heterogeneity in cancer and the search for new cancer-associated genes. *Nature* **2013**, *499*, 214–218. [[CrossRef](#)]
31. Panda, A.; Mehnert, J.M.; Hirshfield, K.M.; Riedlinger, G.; Damare, S.; Saunders, T.; Kane, M.; Sokol, L.; Stein, M.N.; Poplin, E.; et al. Immune activation and benefit from avelumab in EBV-positive gastric cancer. *J. Natl. Cancer Inst.* **2018**, *110*, 316–320. [[CrossRef](#)]
32. Ayers, M.; Lunceford, J.; Nebozhyn, M.; Murphy, E.; Loboda, A.; Kaufman, D.R.; Albright, A.; Cheng, J.D.; Kang, S.P.; Shankaran, V.; et al. IFN- γ -related mRNA profile predicts clinical response to PD-1 blockade. *J. Clin. Investig.* **2017**, *127*, 2930–2940. [[CrossRef](#)]
33. Kim, S.T.; Cristescu, R.; Bass, A.J.; Kim, K.M.; Odegaard, J.I.; Kim, K.; Liu, X.Q.; Sher, X.; Jung, H.; Lee, M.; et al. Comprehensive molecular characterization of clinical responses to PD-1 inhibition in metastatic gastric cancer. *Nat. Med.* **2018**, *24*, 1449–1458. [[CrossRef](#)]
34. Osumi, H.; Kawachi, H.; Murai, K.; Kusafuka, K.; Inoue, S.; Kitamura, M.; Yoshio, T.; Kakusima, N.; Ishihara, R.; Ono, H.; et al. Risk stratification for lymph node metastasis using Epstein-Barr virus status in submucosal invasive (pT1) gastric cancer without lymphovascular invasion: A multicenter observational study. *Gastric Cancer* **2019**. [[CrossRef](#)]



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