

Supplementary Material

Table S1. The Cre-lox system plasmids and knockout plasmid constructs

Plasmids	Relevant features	References
pNZ5319	Cm ^r , Ery ^r , containing <i>lox66-P32-cm-lox71</i> fragment	(Lambert et al., 2007)
pNZ5348	Ery ^r , Cre-recombinase expression	(Lambert et al., 2007)
pNZ5319-Δ <i>menA</i>	Cm ^r , Ery ^r ; pNZ5319 derivative containing homologous regions upstream and downstream of NZ9000 <i>menA</i>	This study
pNZ5319-Δ <i>menB</i>	Cm ^r , Ery ^r ; pNZ5319 derivative containing homologous regions upstream and downstream of NZ9000 <i>menB</i>	This study
pNZ5319-Δ <i>menE</i>	Cm ^r , Ery ^r ; pNZ5319 derivative containing homologous regions upstream and downstream of NZ9000 <i>menE</i>	This study
pNZ5319-Δ <i>menG</i>	Cm ^r , Ery ^r ; pNZ5319 derivative containing homologous regions upstream and downstream of NZ9000 <i>menG</i>	This study

Table S2. Sequence of oligonucleotide primers

Primer	Nucleotides
up A F <i>Xba</i> I	CGTCTCGAGATTGTCCTAATGGCTAAAA
up A R <i>Swa</i> I	TGCATTAAATTCCTTTAATGTGATTATCAA
down A F <i>Sac</i> I	TCCGAGCTCATAAAAACTCCAATTAATTAA
down A R <i>Bgl</i> II	CGAAGATCTTTATTAAAGCTTATAACTG
up B F <i>Xba</i> I	GATCTCGAGTCGGCTCCGTTGAAAAA
up B R <i>Swa</i> I	GAAATTAAATGATTCTCCTTAAGATGTGAGGG
down B F <i>Ban</i> II	TCCGRGCYCATAAAGCGTCATTTGGCGC
down B R <i>Bgl</i> II	CGAAGATCTATGTTATTTCAGAATTGACCACTTC
up E F <i>Xba</i> I	CCGCTCGAGACTGGGTTGCCCTTA
up E R <i>Swa</i> I	CGCATCATTAAATAGCGCCAAAATGACG
down E F <i>Ban</i> II	TCCGAGCTCAAGGGCCTTCCGGAT
down E R <i>Bgl</i> II	CGAAGATCTACCGGCCATAGCATTCC
up G F <i>Xba</i> I	GATCTCGAGCCTGACGTCGAGTC
up G R <i>Swa</i> I	GTCATTAAATACTGGTCACAAGGTCA
down G F <i>Sac</i> I	GCCGAGCTCAGTTGAACGCTTGGATAG
down G R <i>Bam</i> HI	TACGGATCCTCGAGCACGCTTCTTTAT
lox F	AAATCTACCGTCGATAATGTATGC
lox R	CTCATGCCGGCTGTACCG
menA F	TTTCTGCTGGCCTTCGTAC

men A R	GACAATGGTTTGGGCCTCCTG
men B F	TCGCAATGCTTTCGTCAA
men B R	TCCCGCAAATTGTTGCAGTC
men E F	ATGAAATGGTAAAAAACAGGCGG
men E R	TCATGCTTGAGCTCTTT
men G F	AACGAAGAACGTGTGCAAGA
men G R	AAGCGTCTTGGCATCTGGAA
Ery F	CGATACCGTTACGAAAT
Ery R	CTTGCTCATAAGTAACGG

*Underlined sequences indicate the restriction sites.

Table S3. Primers used for the functional verification of menaquinone biosynthesis genes study

Primers	Nucleotides
LpA1 F	GGAGTACTTGAAACCAAAAGTGTCTTGAA
LpA1 R	CTAGAGCTCCTAGTTAAAAATCAGTCCTAACGCC
LpA2 F	GGAGTACTTGTTGACGAACGAACCAAC
LpA2 R	CTAGAGCTCTAAAAGAAAATACCCAGGATCAGTC
LpA3 F	GGAGTACTATGTTAAAAAATGGCTAACTTG
LpA3 R	CTAGAGCTCTAACCAAGTCCCTAAAACAAA
LpG1 F	GGAGTACTATGACAGCTTAGAAAGAAGTTG
LpG1 R	TACGAGCTCTCAATCACGACGAGCCA
LpG2 F	GGAGTACTATGGCAAATCGTTATTAC
LpG2 R	CTAGAGCTCTTACTTGGCCTCCTAG
LbA F	GGAGTACTATGAGTTATCAACTTTCCCAG
LbA R	CTAGAGCTCCTAGTGGCAATCAGGG
LbG F	GGAGTACTATGACCGCTGACGAACAA
LbG R	CTAGAGCTCTCAGGGCTTCATTCCCCA
LbB F	GGAGTACTATGACTTCAGTTAAATGGGAATC
LbB R	CTAGAGCTCCTATGGAACTTAGGAAATTG
LbE F	GGAGTACTATGAAAGTGGATAATTGGATTTAA
LbE R	CTAGAGCTCTATAACAACGTTCAGCTTGAATC
LLA F	GGAGTACTATGAATTAAAACATTGCT

LLA R	GGCGAGCTCTTAAAATCTAACAAACTAATAAAGA
LLB F	GGAGTACTATGTCAAAATTAACTGGTTGC
LLB R	GGCGAGCTCTATGGAAATTTGGAAATTGGTC
LLE F	GGAGTACTATGAAATGGTTAAAAACAGGCG
LLE R	AACGGYRCCTCATGCTTGAGCTTTCTTAA
LLG F	GGAGTACTATGACTAAAGTAAACGAAGAACGT
LLG R	GGCGAGCTCTTACTTTACCAATACGAATATTGATT
pnz8150 F	GCATAATAACGGCTTGAT
pnz8150 R	CAGCAATATCAGTAATTGCTTTATC

*Underlined sequences indicate the restriction sites.

Table S4. Plasmids and menaquinone expression plasmid constructs

Plasmids	Relevant features	References
pNZ8150	Cm ^r , nisA promoter, expression vector	(Mierau and Kleerebezem, 2005)
pNZ8150- <i>lpmenA1</i>	Cm ^r ; nisA promoter, <i>lpmenA1</i> gene cloned to the MCS	This study
pNZ8150- <i>lpmenA2</i>	Cm ^r ; nisA promoter, <i>lpmenA2</i> gene cloned to the MCS	This study
pNZ8150- <i>lpmenA3</i>	Cm ^r ; nisA promoter, <i>lpmenA3</i> gene cloned to the MCS	This study
pNZ8150- <i>lpmenG1</i>	Cm ^r ; nisA promoter, <i>lpmenG1</i> gene cloned to the MCS	This study
pNZ8150- <i>lpmenG2</i>	Cm ^r ; nisA promoter, <i>lpmenG2</i> gene cloned to the MCS	This study
pNZ8150- <i>lbmenA</i>	Cm ^r ; nisA promoter, <i>lbmenA</i> gene cloned to the MCS	This study
pNZ8150- <i>lbmenB</i>	Cm ^r ; nisA promoter, <i>lbmenB</i> gene cloned to the MCS	This study
pNZ8150- <i>lbmenE</i>	Cm ^r ; nisA promoter, <i>lbmenE</i> gene cloned to the MCS	This study
pNZ8150- <i>lbmenG</i>	Cm ^r ; nisA promoter, <i>lbmenG</i> gene cloned to the MCS	This study
pNZ8150- <i>llmenA</i>	Cm ^r ; nisA promoter, <i>llmenA</i> gene cloned to the MCS	This study
pNZ8150- <i>llmenB</i>	Cm ^r ; nisA promoter, <i>llmenB</i> gene cloned to the MCS	This study
pNZ8150- <i>llmenE</i>	Cm ^r ; nisA promoter, <i>llmenE</i> gene cloned to the MCS	This study
pNZ8150- <i>llmenG</i>	Cm ^r ; nisA promoter, <i>llmenG</i> gene cloned to the MCS	This study

Table S5. *Lactococcus lactis* strains with menaquinone expression plasmid

Strains	Relevant features	References
<i>Lactococcus lactis</i> (II)		
<i>lpmenA1</i>	NZ9000- $\Delta menA$ carrying pNZ8150- <i>lpmenA1</i>	This study
<i>lpmenA2</i>	NZ9000- $\Delta menA$ carrying pNZ8150- <i>lpmenA2</i>	This study
<i>lpmenA3</i>	NZ9000- $\Delta menA$ carrying pNZ8150- <i>lpmenA3</i>	This study
<i>lpmenG1</i>	NZ9000- $\Delta menG$ carrying pNZ8150- <i>lpmenG1</i>	This study
<i>lpmenG2</i>	NZ9000- $\Delta menG$ carrying pNZ8150- <i>lpmenG2</i>	This study
<i>lbmenA</i>	NZ9000- $\Delta menA$ carrying pNZ8150- <i>lbmenA</i>	This study
<i>lbmenB</i>	NZ9000- $\Delta menB$ carrying pNZ8150- <i>lbmenB</i>	This study
<i>lbmenE</i>	NZ9000- $\Delta menE$ carrying pNZ8150- <i>lbmenE</i>	This study
<i>lbmenG</i>	NZ9000- $\Delta menG$ carrying pNZ8150- <i>lbmenG</i>	This study
<i>llmenA</i>	NZ9000- $\Delta menA$ carrying pNZ8150- <i>llmenA</i>	This study
<i>llmenB</i>	NZ9000- $\Delta menB$ carrying pNZ8150- <i>llmenB</i>	This study
<i>llmenE</i>	NZ9000- $\Delta menE$ carrying pNZ8150- <i>llmenE</i>	This study
<i>llmenG</i>	NZ9000- $\Delta menG$ carrying pNZ8150- <i>llmenG</i>	This study

Table S6. Primers used for the reconstitution of menaquinone biosynthesis pathway in *Lactipl. plantarum* and *Lent. buchneri* strains

Primers	Nucleotides
Psip 409 BsaIKO F	CACGTTACTAAAGGAAATGGAGACCGGGGT
Psip409 BsaIKO R	CGGTCGCCATTCCCTTAGAACGTGTAACTTCCAAT
Psip409 F BsaI	AAGGGTCT <u>CATGCGT</u> CTAGACTCGAGGAATT
Psip409 R BsaI	CATGGTCTCC <u>GATCGC</u> TAAAATCTCCTTGTAAATA

Golden Gate Assemble for *Lactipl. plantarum* WCFS1 menaquinone expression vector construction

Psip409 backbone F BsaI	AAGGGTCT <u>CATGCGT</u> CTAGACTCGAGGAATT
Psip409 backbone R BsaI	CAT <u>GGTCTCCCGCC</u> GCTAAAATCTCCTTGTAAATA
GO menF Lp F	TAT <u>GGTCTCA</u> GCGG ATTACAAGGAGATTTAGCCATGAAATATATAAAAAACGATTAAATTAA
GO menF Lp R	CGCGGTCTCAT CTTCATAAGGCTTCTAAA
GO menD Lp F	TAT <u>GGTCTCA</u> AAGA TATTACA <u>AGGAG</u> ATTTAGCCATGACCAATGAATATTAGCTCC
GO menD Lp R	CGCGGTCTCC <u>CTGA</u> TCAATTTCATAAGCAGTATTTTTTAT
GO menH Lp F	TAT <u>GGTCTCA</u> TCAG ATTACA <u>AGGAG</u> ATTTAGCCATGAAAATTGATAAAAAAATAATGACGA
GO menH Lp R	CGCGGTCTCG <u>TAAG</u> CCTAAGCCAAAATTCCCTC
PsppQ LP F	TAT <u>GGTCTCG</u> CTTA GGAGATCTACCGGTTAATTGAAA
PsppQ LP R	CGCGGTCTCG <u>GCT</u> CCGGCTAAAATCTCCTTGTAAATAGT

GO menB Lp F	TAT <u>GGTCTCG</u> GAGC ATTACAAGGAGATTAGCCATGTCAAAATTAACTGGTTG
GO menB Lp R	CGCGGTCTCG AAC <u>CTT</u> ATGGAAATTGGAAATTGG
GO menE Lp F	TAT <u>GGTCTC</u> AGGT TTTAC <u>AGGAGA</u> TTTAGCCATGAAATGGTAAAAAACAGGC
GO menE Lp R	CGCGGTCTC T <u>GTAGTC</u> ATGCTTGAGCTTT
GO menC LP F	TAT <u>GGTCTC</u> ACTAC TTAC <u>AGGAGA</u> TTTAGCCATGAAAATTGAAAAATCACAATGT
GO menC Lp R	CGCGGTCTC <u>CGCAT</u> TTCAAGGAGGTCAAGC

*Underlined sequences indicate the restriction sites

Ribosome binding site are highlighted in grey

Colored nucleotides represent the overhangs for Golden Gate Assembly and the same color showed the complementary bases

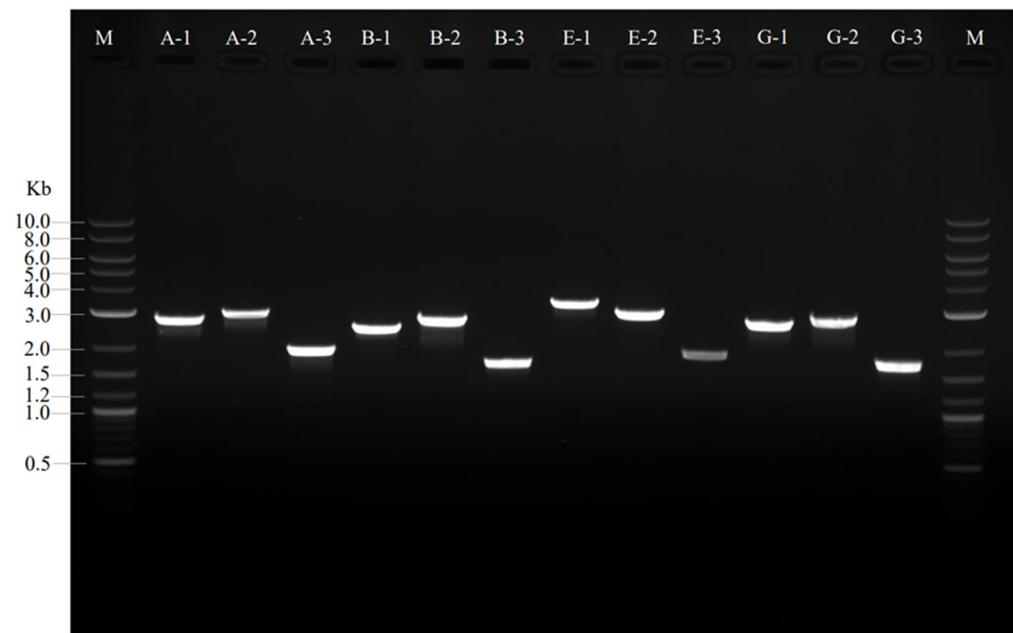


Figure S1. Diagnostic PCR of *L. lactis* NZ9000 *menA* (A-1), its gene replacement mutant *menA:: lox66-P32-cm-lox71* (A-2), Δ *menA* (A-3); *menB* (B-1), *menB:: lox66-P32-cm-lox71* (A-2), Δ *menB* (B-3); *menE* (E-1), *menE:: lox66-P32-cm-lox71* (E-2), Δ *menE* (E-3); *menG* (G-1), *menG:: lox66-P32-cm-lox71* (A-2), Δ *menG* (G-3); M is a 2 log DNA ladder

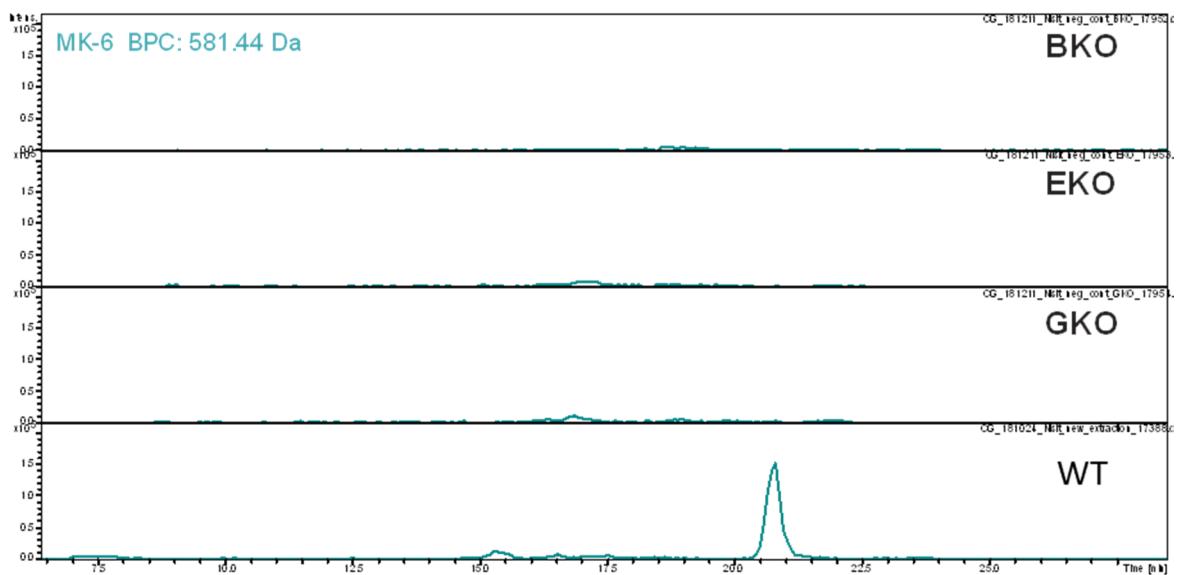


Figure S2. Base peak chromatogram of menaquinone extracts from *L. lactis* NZ9000 (WT) and its menaquinone deficient strains $\Delta menB$ (BKO), $\Delta menE$ (EKO), $\Delta menG$ (GKO) for MK-6.

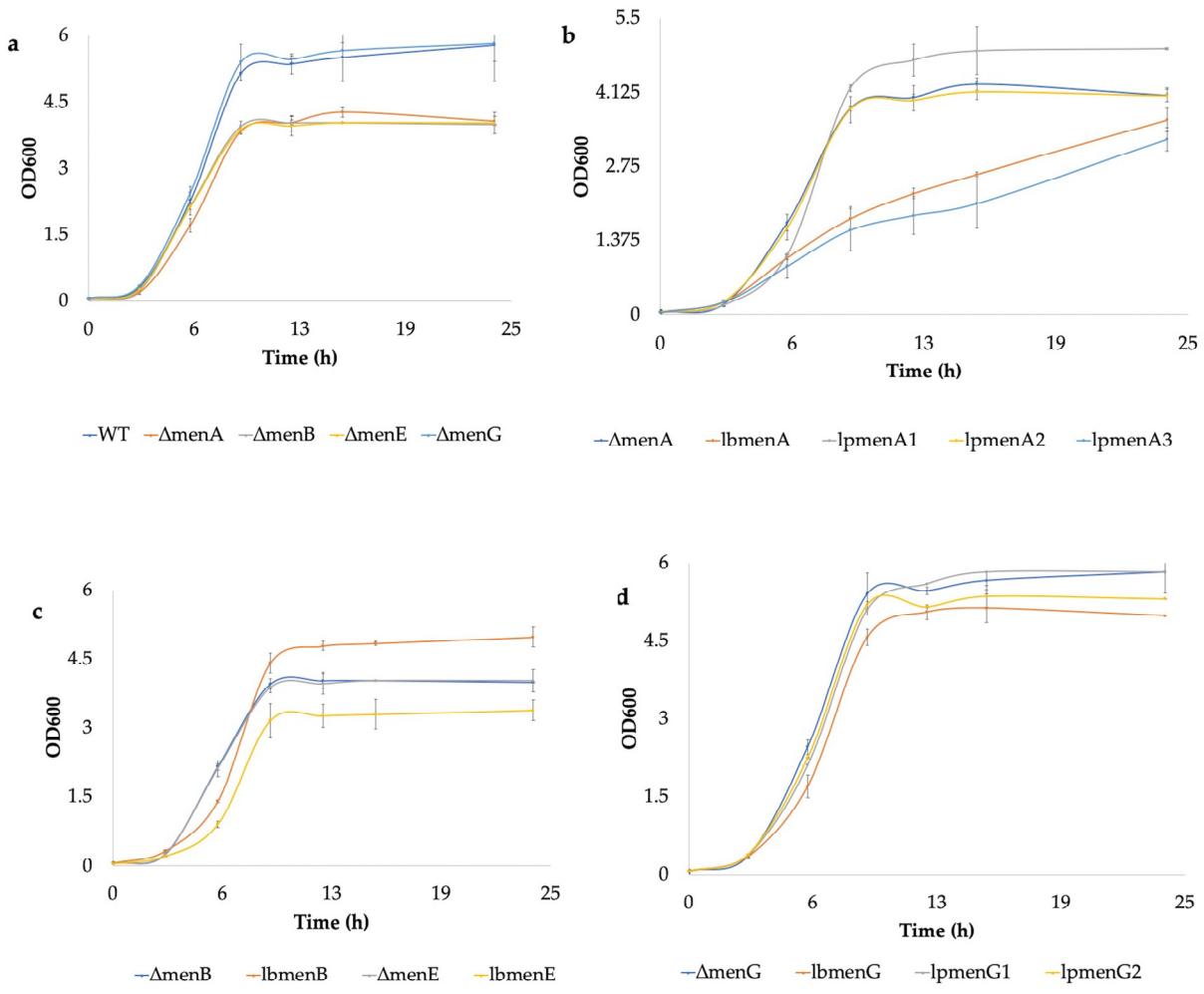


Figure S3. Growth profile analysis of (a) *L. lactis* NZ9000 (WT) and the knockout strains (ΔmenA , ΔmenB , ΔmenE , ΔmenG), (b) *L. lactis* ΔmenA and its engineered derivatives carrying *lbmenA*, *lpmenA1*, *lpmenA2* and *lpmenA3*), (c) *L. lactis* ΔmenB , *L. lactis* ΔmenE and the engineered derivatives (carrying *lbmenB*, *lbmenE*) and (d) *L. lactis* ΔmenG and its engineered derivatives carrying *lbmenG*, *lpmenG1* and *lpmenG2*. 300 mL of GM17 medium supplemented with heme (2 $\mu\text{g}/\text{mL}$) were used, 1 ng/mL of nisin was added as an inducer at $\text{OD}_{600} = 0.4$.

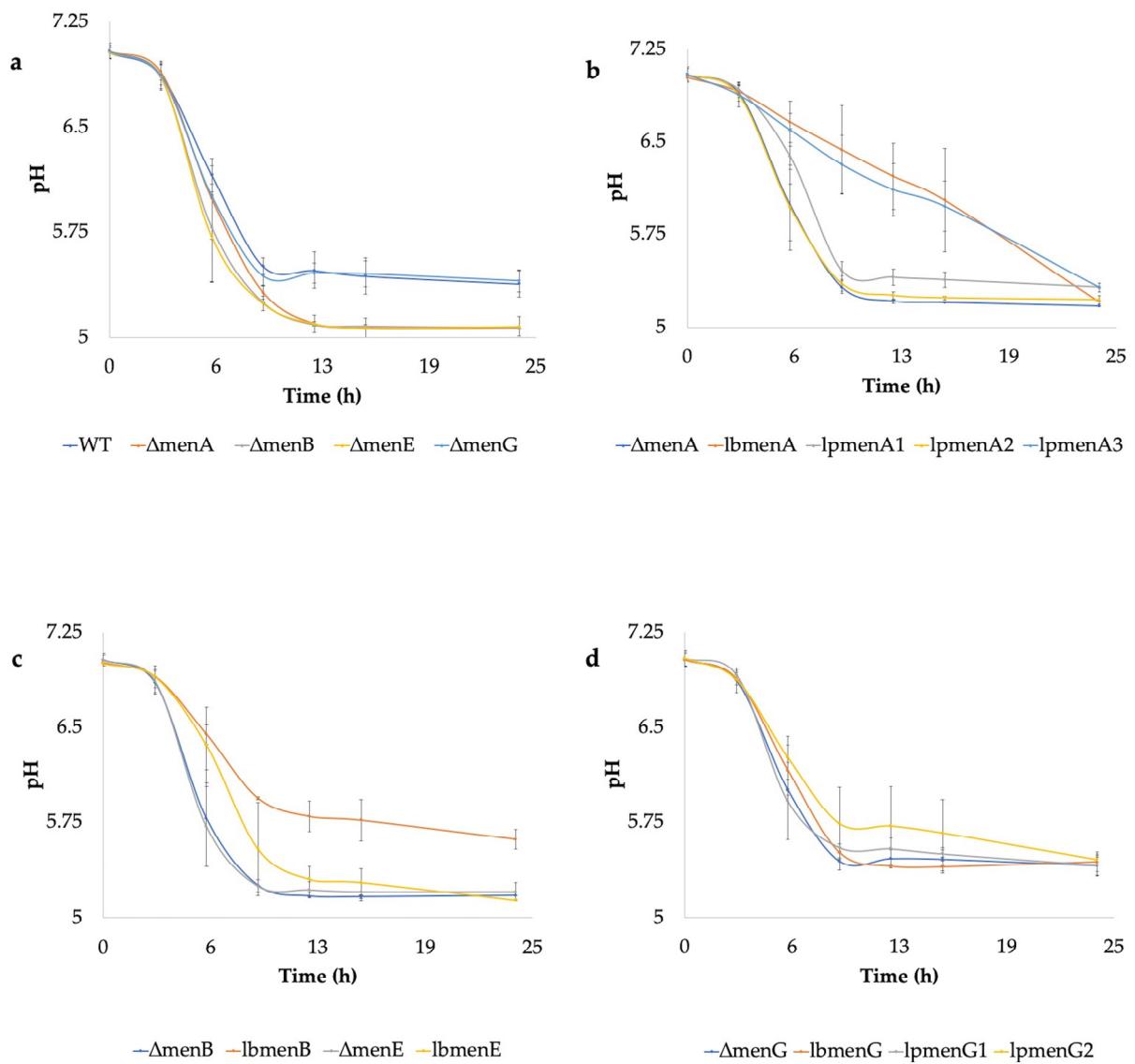


Figure S4. pH profile of (a) *L. lactis* NZ9000 (WT) and its menaquinone deficient strains (Δ menA, Δ menB, Δ menE, Δ menG), (b) *L. lactis* Δ menA and its engineered strains (lbmenA, lpmenA1, lpmenA2, lpmenA3), (c) *L. lactis* Δ menB, *L. lactis* Δ menE and its engineered strains (lbmenB, lbmenE) and (d) *L. lactis* Δ menG and its engineered strains (lbmenG, lpmenG1, lpmenG2). 300 mL of GM17 medium supplemented with heme (2 μ g/mL) was used, 1 ng/mL of nisin was added as inducer at OD₆₀₀ = 0.4.

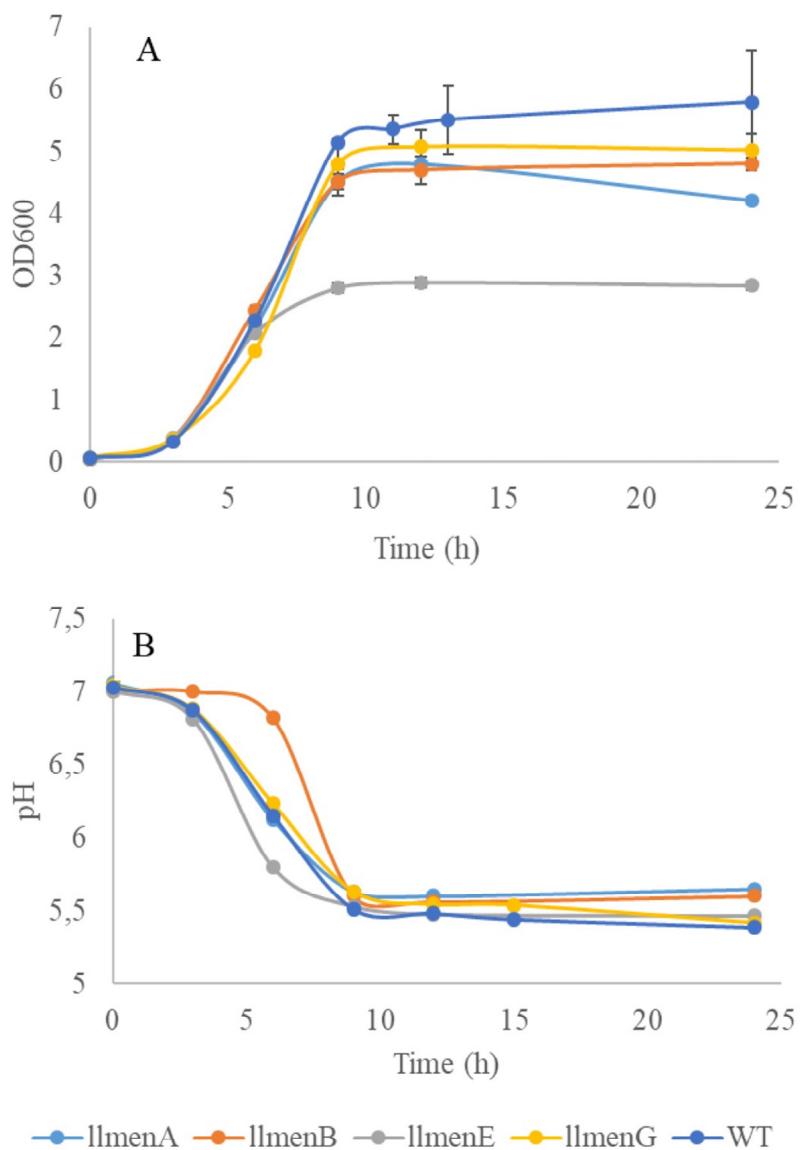


Figure S5. Growth profile (A) and pH analysis (B) of *L. lactis* NZ9000 (WT) and engineered deficient strains complemented by homologous genes (*llmenA*, *llmenB*, *llmenE*, *llmenG*). 300 mL of GM17 medium supplemented with heme (2 μ g/mL) were used, 1 ng/mL of nisin was added as an inducer at OD₆₀₀ = 0.4.