

## Supplementary Materials

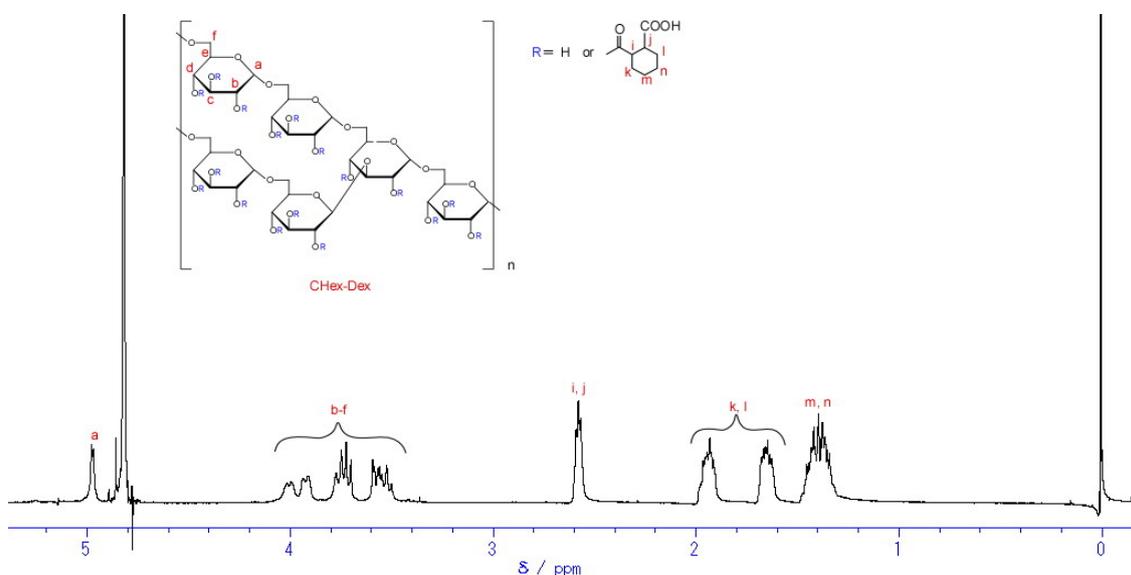
### Development of pH-sensitive dextran derivatives with strong adjuvant function and their application to antigen delivery

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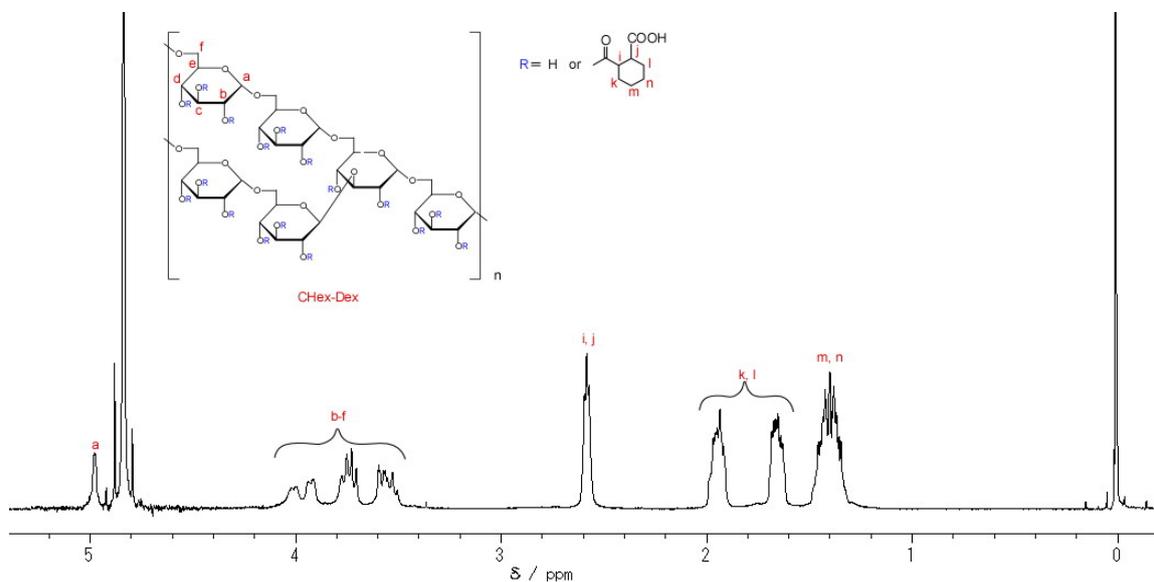
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**\*Corresponding author: Eiji Yuba**

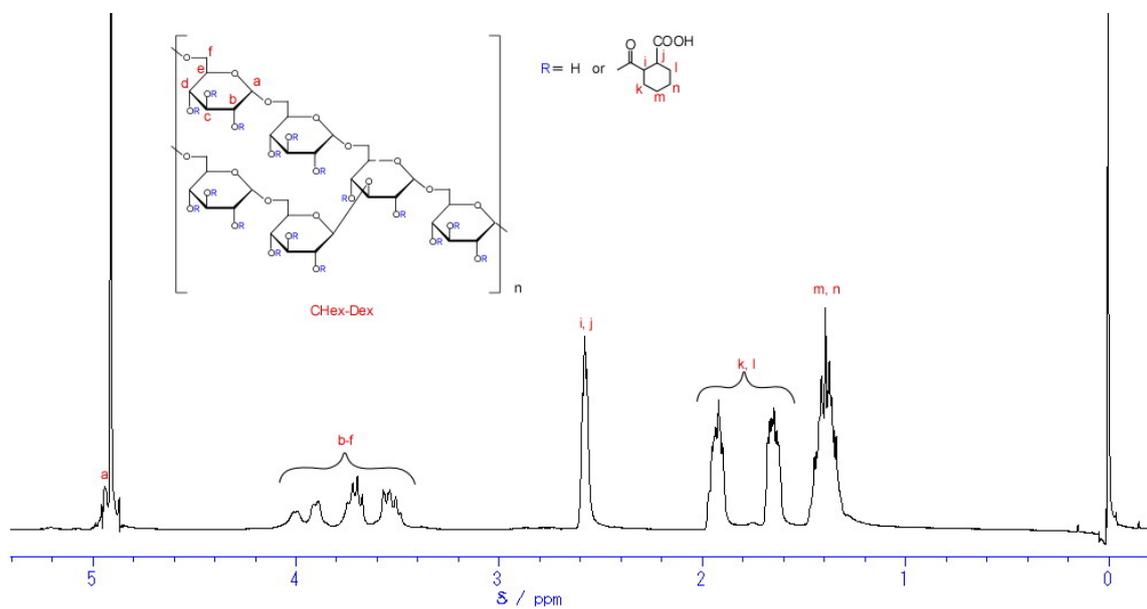
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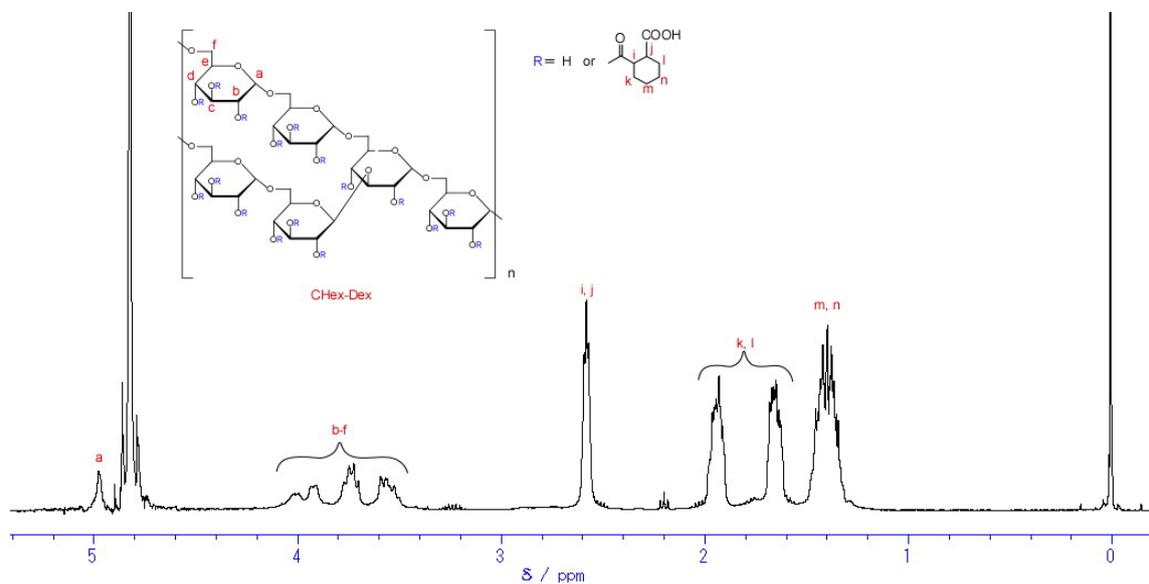
**Figure S1.**  $^1\text{H}$  NMR chart of CHex40-Dex (400 MHz,  $\text{D}_2\text{O} + \text{NaOD}$ ).



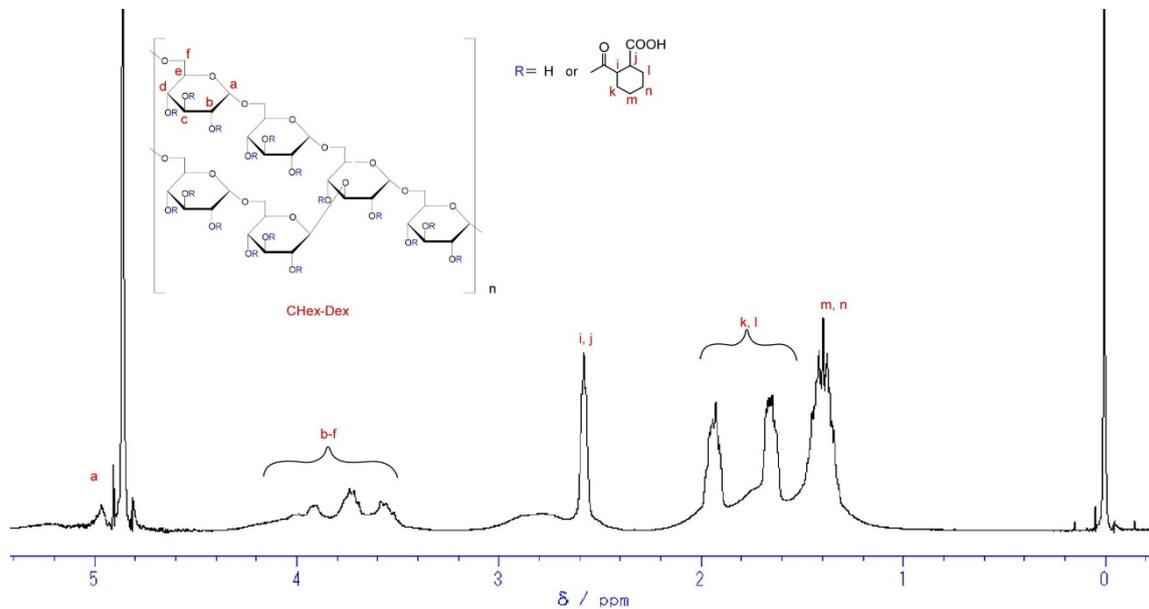
**Figure S2.** <sup>1</sup>H NMR chart of CHex57-Dex (400 MHz, D<sub>2</sub>O+NaOD).



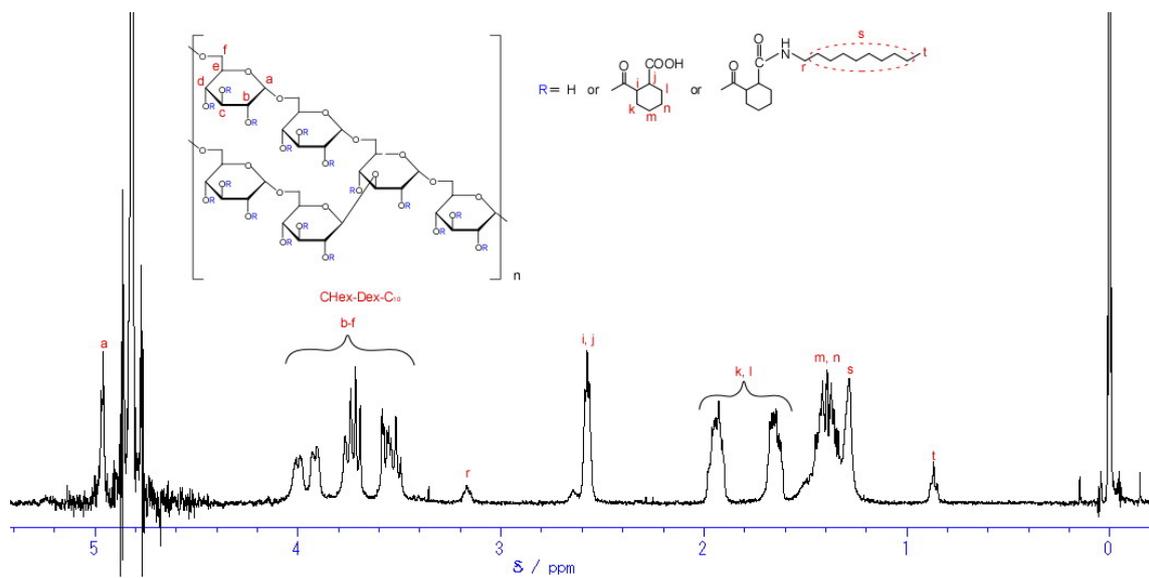
**Figure S3.** <sup>1</sup>H NMR chart of CHex73-Dex (400 MHz, D<sub>2</sub>O+NaOD).



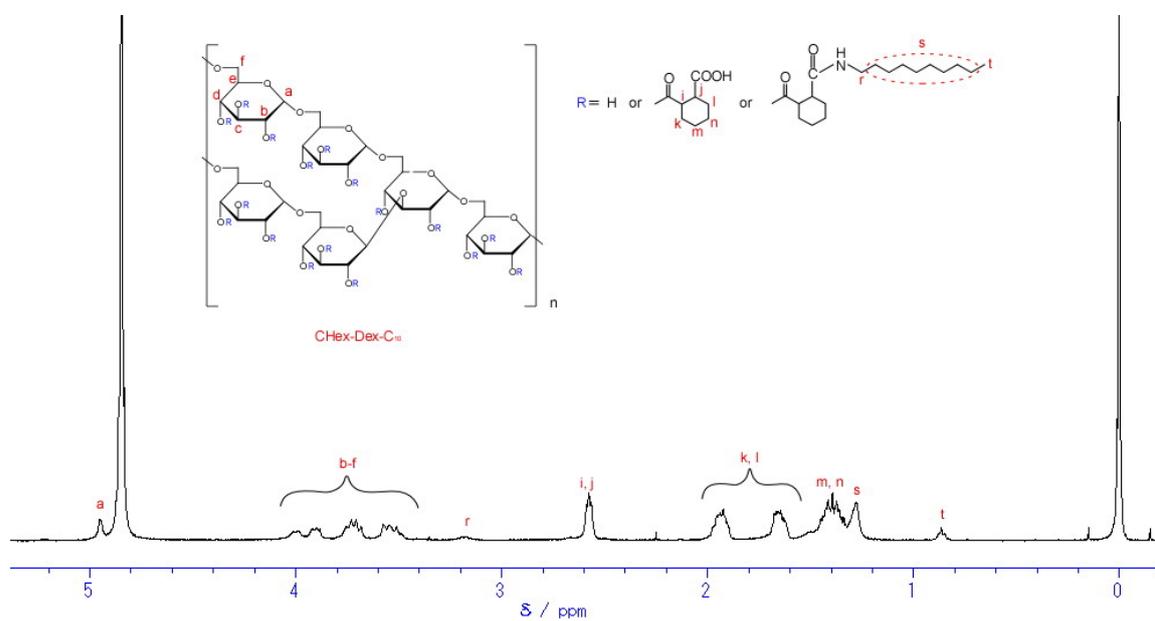
**Figure S4.**  $^1\text{H}$  NMR chart of CHex86-Dex (400 MHz,  $\text{D}_2\text{O}+\text{NaOD}$ ).



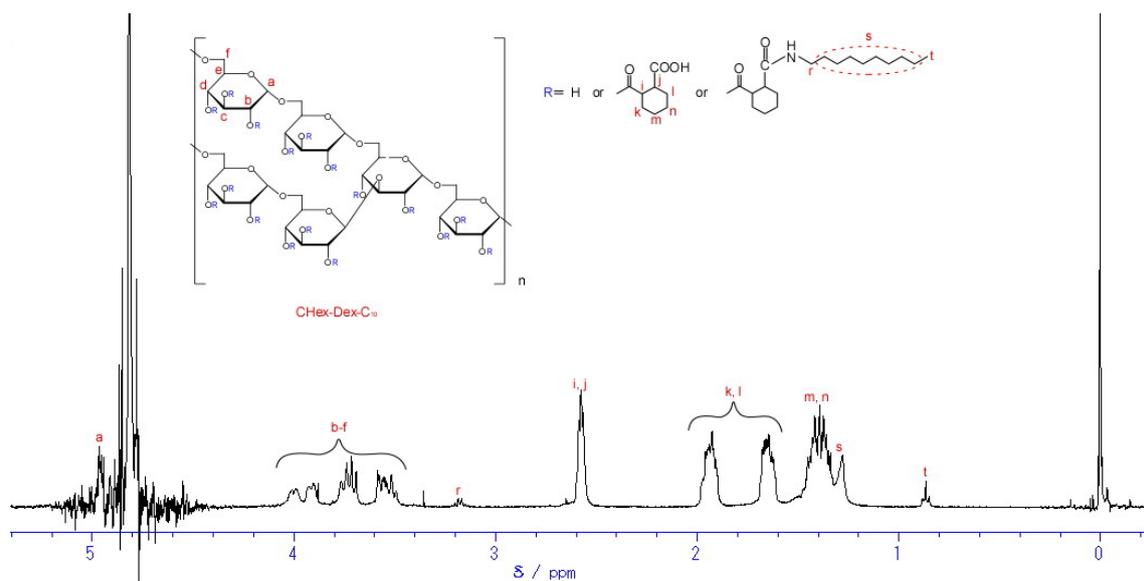
**Figure S5.**  $^1\text{H}$  NMR chart of CHex98-Dex (400 MHz,  $\text{D}_2\text{O}+\text{NaOD}$ ).



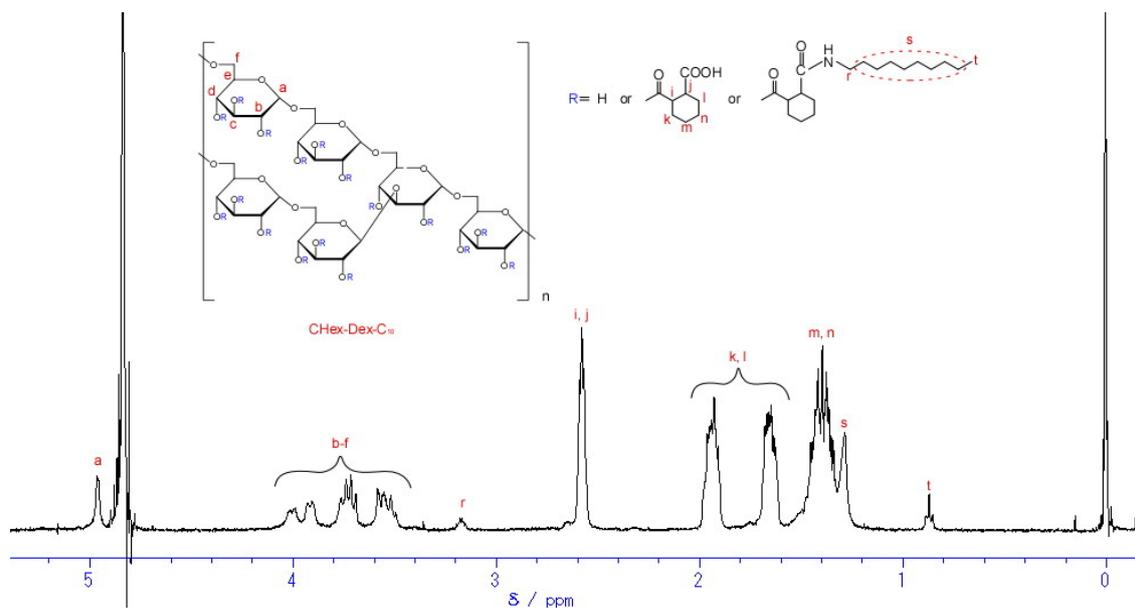
**Figure S6.**  $^1\text{H}$  NMR chart of CHex28-Dex-C<sub>10</sub> (400 MHz, D<sub>2</sub>O+NaOD).



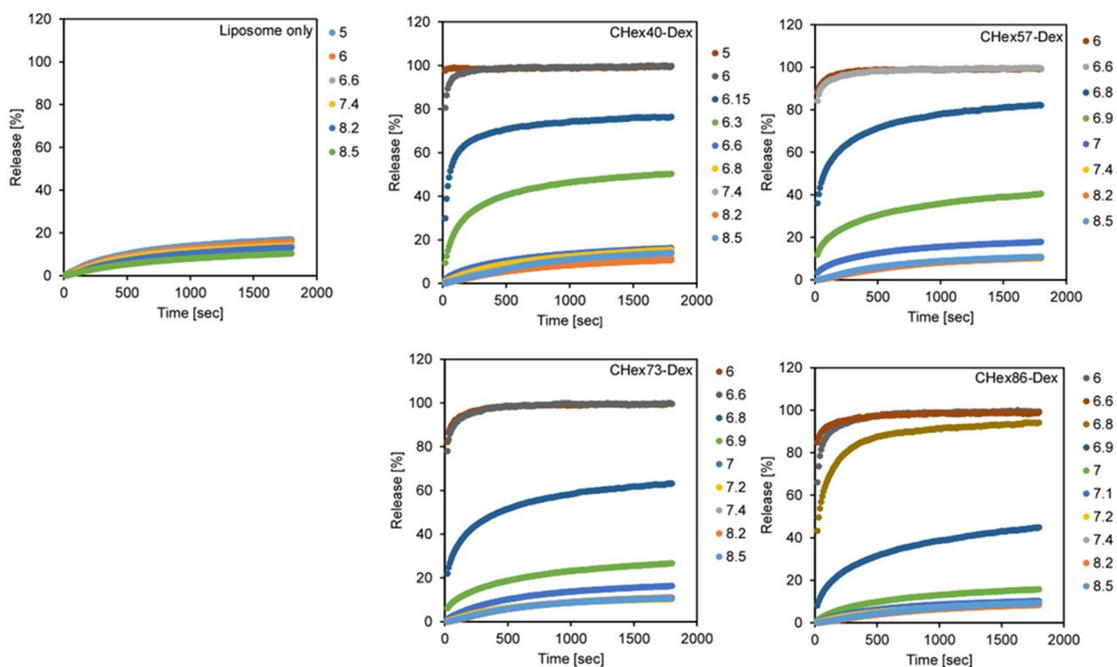
**Figure S7.**  $^1\text{H}$  NMR chart of CHex42-Dex-C<sub>10</sub> (400 MHz, D<sub>2</sub>O+NaOD).



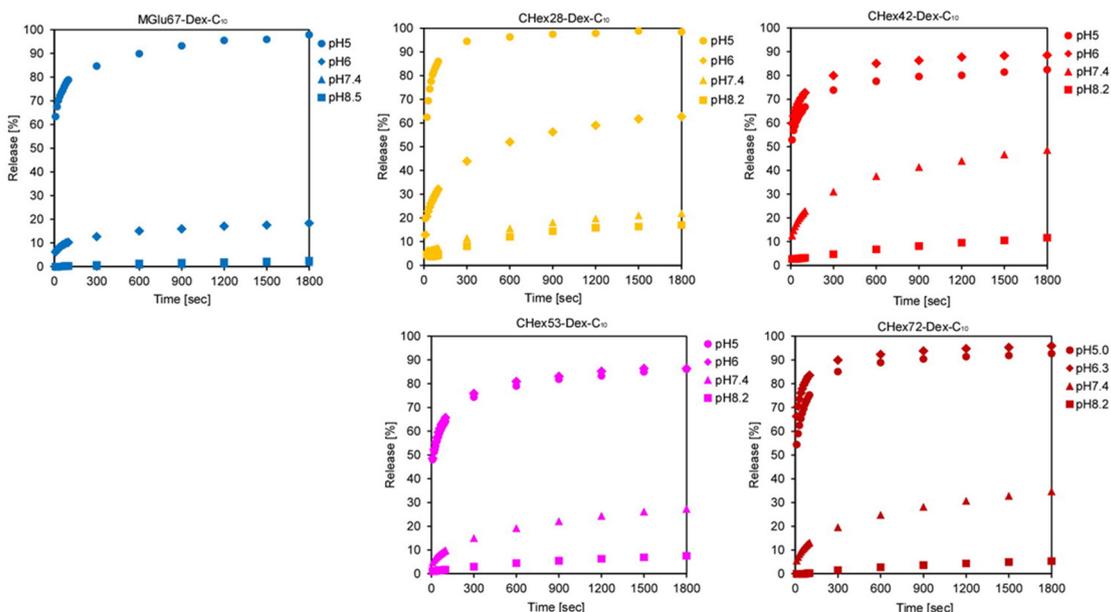
**Figure S8.**  $^1\text{H}$  NMR chart of CHex53-Dex- $\text{C}_{10}$  (400 MHz,  $\text{D}_2\text{O}+\text{NaOD}$ ).



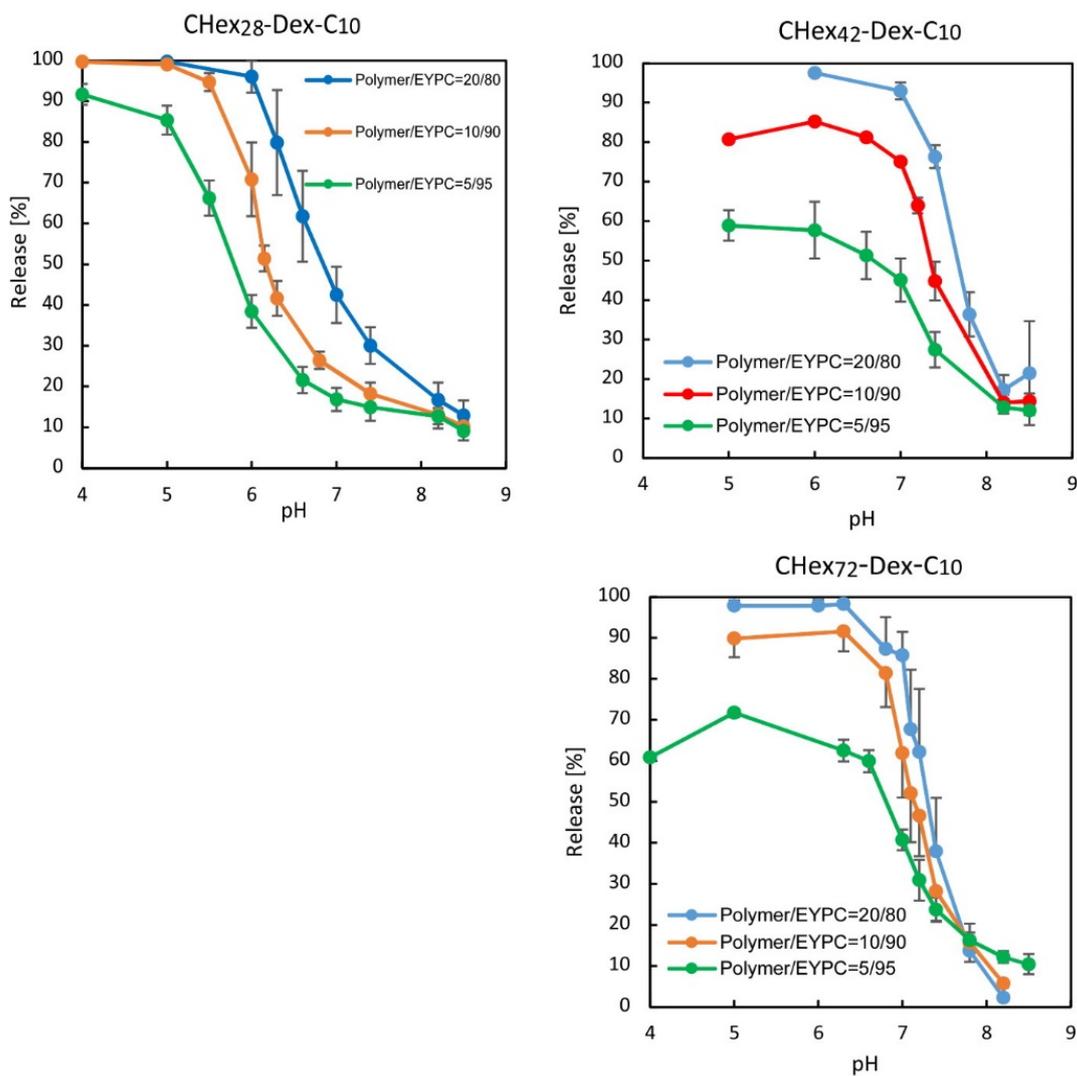
**Figure S9.**  $^1\text{H}$  NMR chart of CHex72-Dex- $\text{C}_{10}$  (400 MHz,  $\text{D}_2\text{O}+\text{NaOD}$ ).



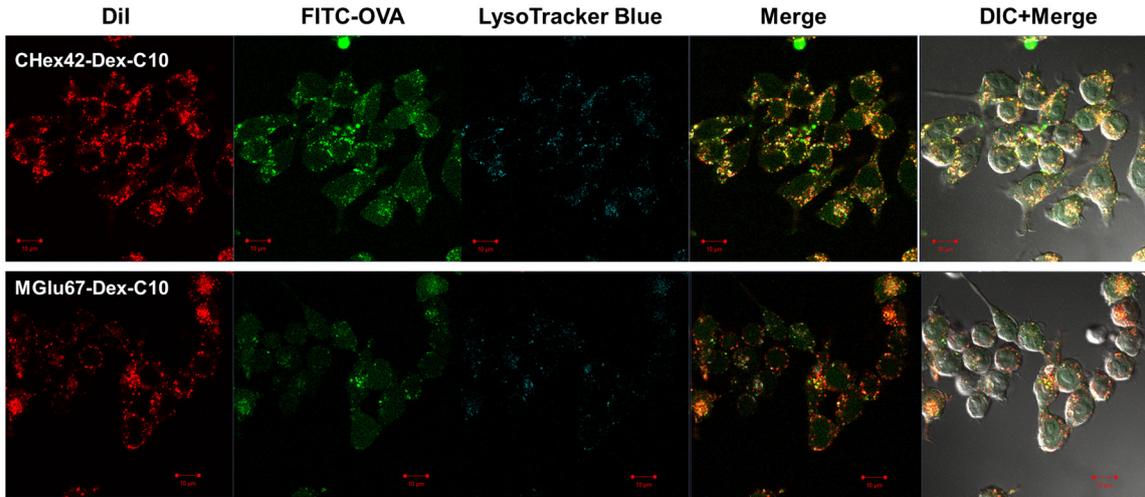
**Figure S10.** Time courses of pyranine release from EYPC liposomes at various pH after addition of various CHex-Dex. Lipid concentration was  $2.0 \times 10^{-5}$  M. The ratio by weight of lipid to polymer is 9 to 1. Measurements were performed in PBS solution at 37 °C.



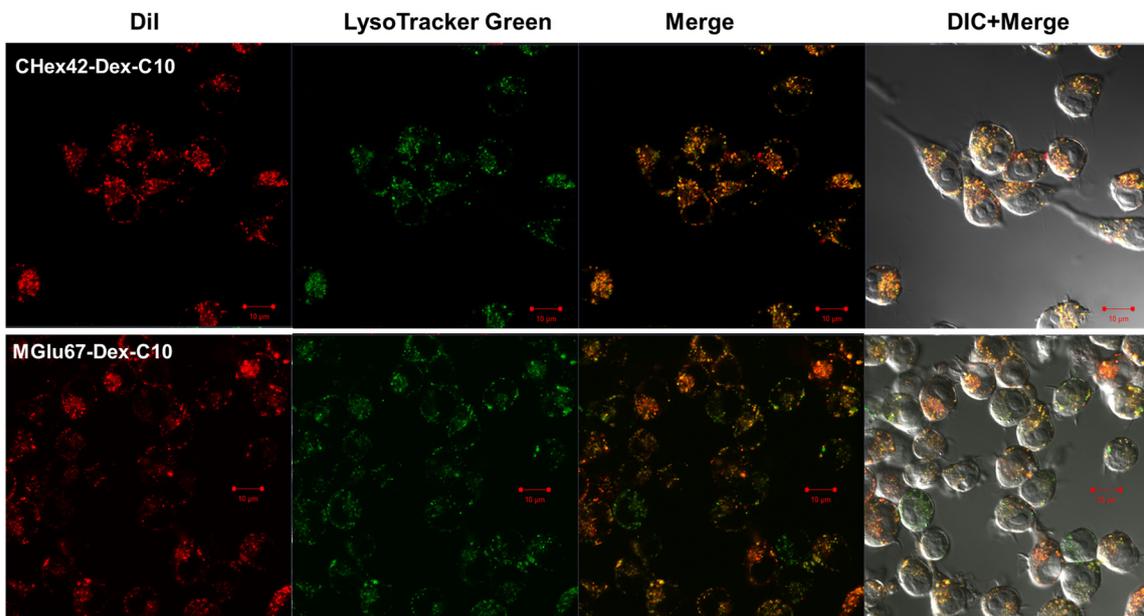
**Figure S11.** Time courses of pyranine release from EYPC liposomes modified with or without 10 wt% CHex-Dex-C<sub>10</sub> or 30 wt% MGLu67-Dex-C<sub>10</sub> at 37 °C. Lipid concentrations were  $2.0 \times 10^{-5}$  M.



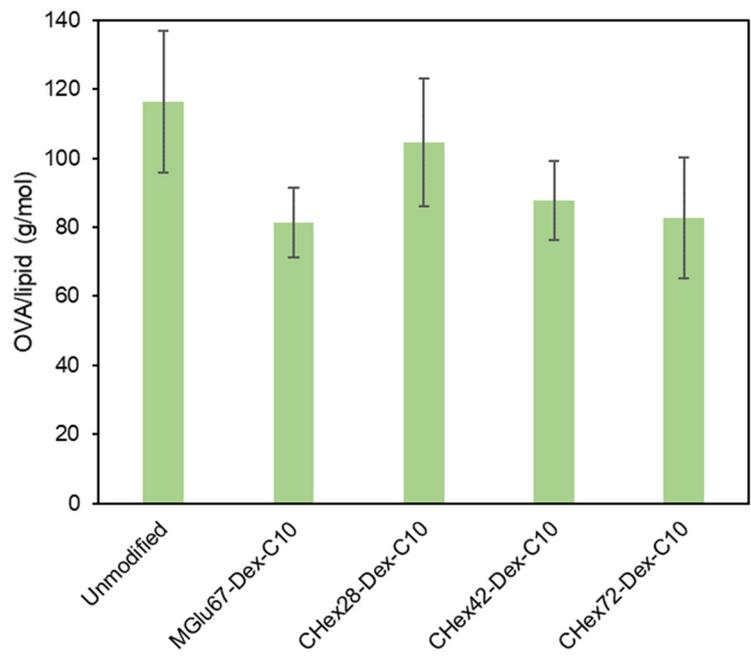
**Figure S12. Effect of polymer/lipid ratio on pH-sensitivity of CHex-Dex-C<sub>10</sub>-modified liposomes.** Pyranine release from EYPC liposomes modified with various amounts of CHex-Dex-C<sub>10</sub> at 37 °C after 30 min-incubation was evaluated. Lipid concentrations were  $2.0 \times 10^{-5}$  M.



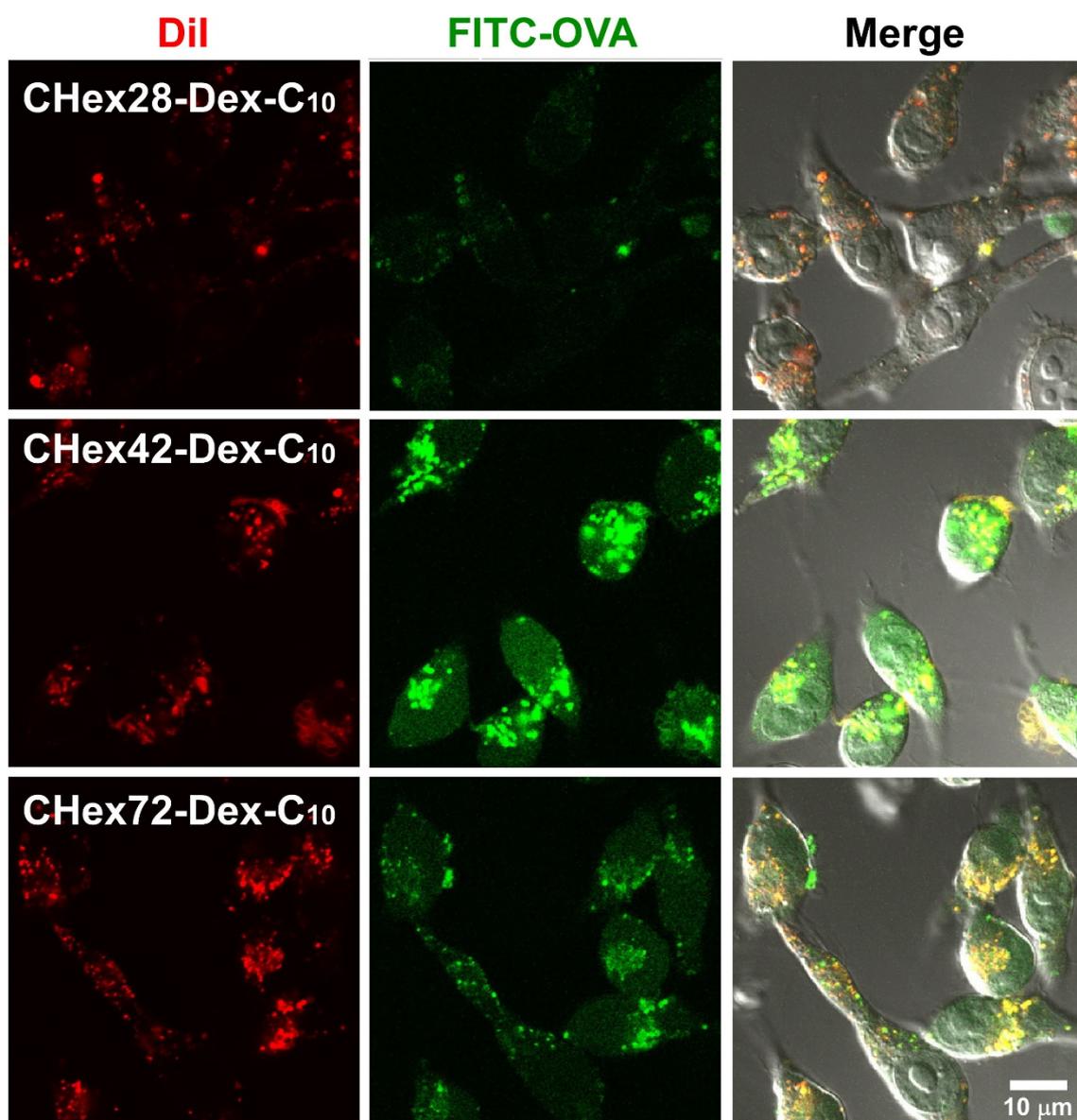
**Figure S13.** Confocal laser scanning microscopy (CLSM) images of DC2.4 cells treated with DiI-labeled and FITC-OVA-loaded EYPC liposomes modified with CHex42-Dex-C<sub>10</sub> or MGLu67-Dex-C<sub>10</sub> for 2 h at 37 °C in serum-free medium. Scale bar represents 10 µm. Lipid concentration was  $5.0 \times 10^{-4}$  M. Intracellular acidic compartments were stained using LysoTracker Blue.



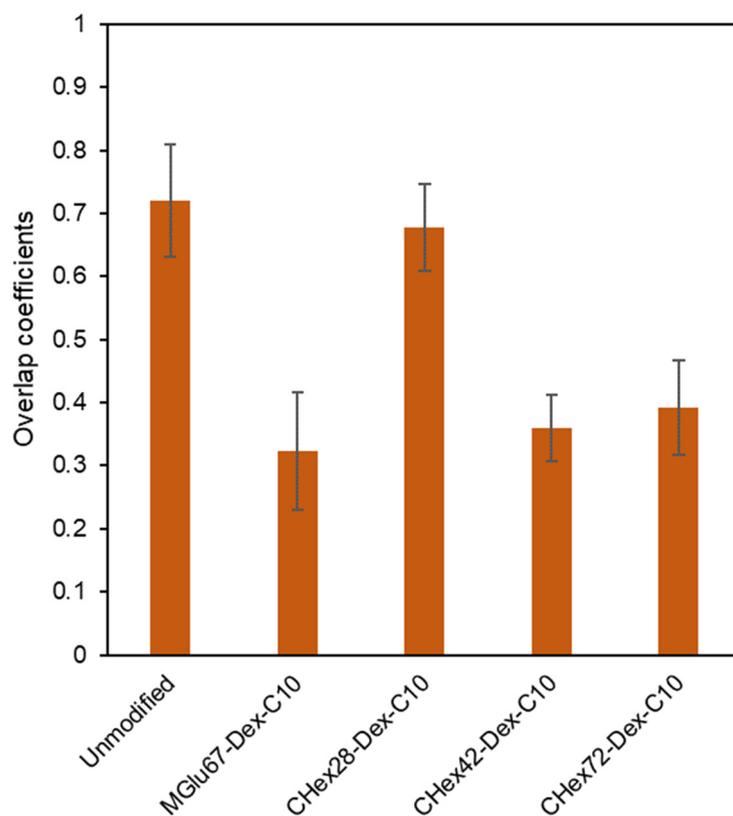
**Figure S14.** Most DiI fluorescence derived from liposomes co-localized with endo/lysosomes. CLSM images of DC2.4 cells treated with DiI-labeled EYPC liposomes modified with CHex42-Dex-C<sub>10</sub> or MGLu67-Dex-C<sub>10</sub> for 2 h at 37 °C in serum-free medium. Scale bar represents 10 µm. Lipid concentration was  $5.0 \times 10^{-4}$  M. Intracellular acidic compartments were stained using LysoTracker Green.



**Figure S15.** OVA amounts per lipid in various liposomes.



**Figure S16.** CLSM images of DC2.4 cells treated with DiI-labeled and FITC-OVA-loaded EYPC liposomes modified with CHex-Dex-C<sub>10</sub> for 4 h at 37 °C in serum-free medium. Scale bar represents 10 μm. Lipid concentration was  $5.0 \times 10^{-4}$  M.



**Figure S17.** Colocalization for FITC fluorescence derived from FITC-OVA with DiI fluorescence. Overlap Coefficient of FITC fluorescence with DiI fluorescence was calculated from CLSM images.