

Supplementary material

# A new high-throughput screening method to detect antimicrobial volatiles from metagenomic clone libraries

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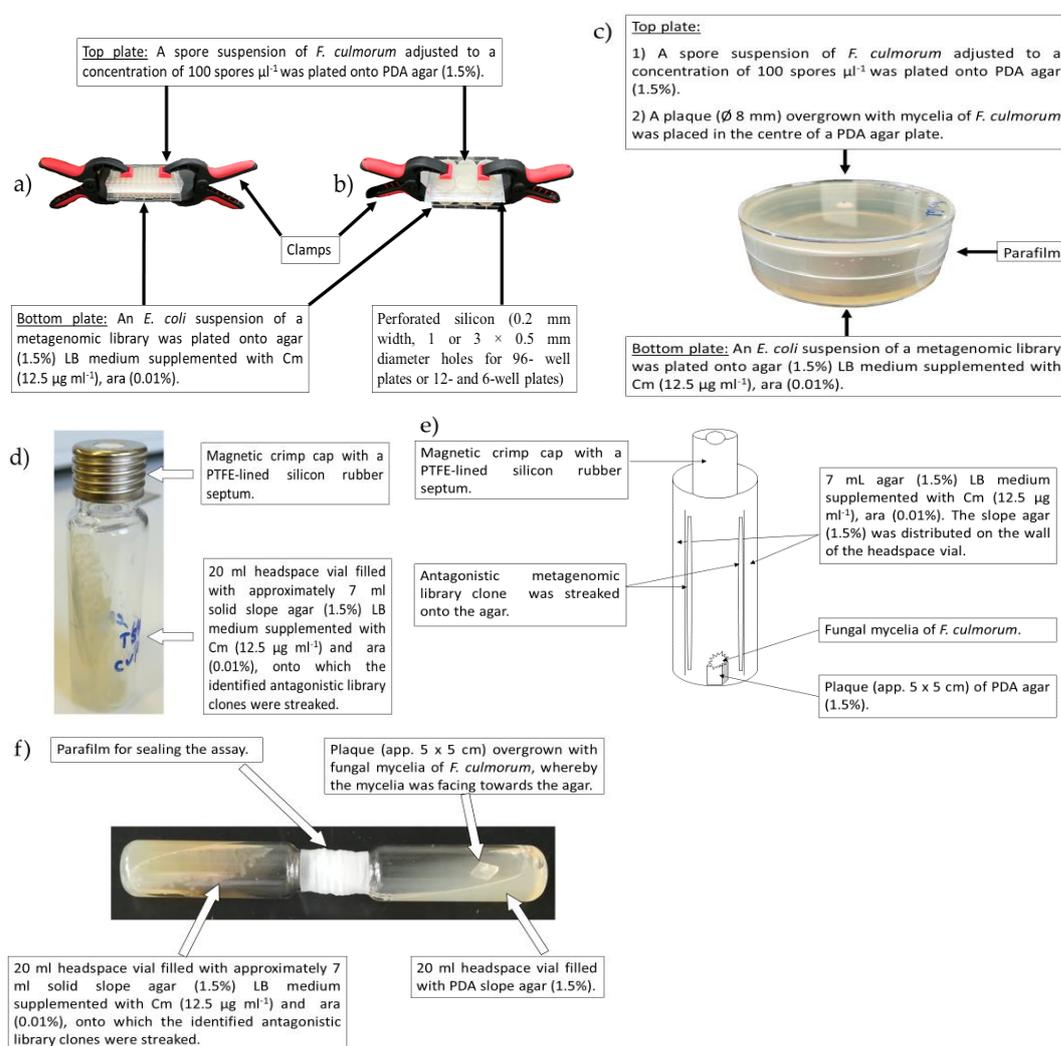
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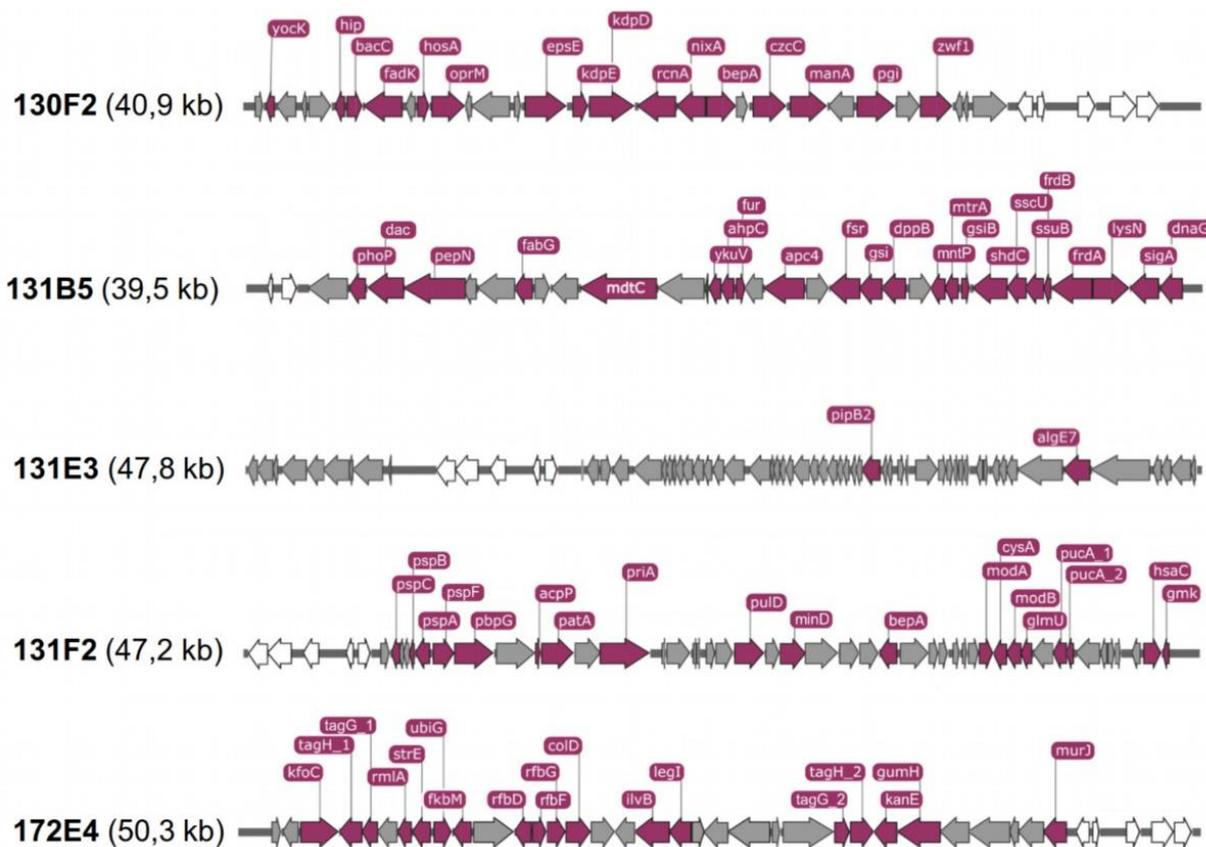
## FIGURES



**Figure S1: Illustration of the different assays.** Set-up of the high-throughput Two Clamp VOCs Assay (htTCVA) (a), the Two Clamp VOCs Assay (TCVA) (b) and the Petri Dish VOCs Assay (PDVA) (c) for the high throughput screening and the set-up of single cultivation (d), one-vial co-cultivation (e) and separated co-cultivation (f) prior to the SPME GC-MS measurements.

### De novo sequencing and gene annotation

Metagenomic DNA inserts of positive tested clones were sequenced by the Göttingen Genomics Laboratory (Göttingen, Germany) as previously described [1]. In short, using the Illumina MiSeq technology (Illumina, California, USA) 300 bp reads were generated, quality filtered with Trimmomatic v0.36 [2], assembled with SPAdes v3.10.0 [3] and genes annotated with Prokka [4].

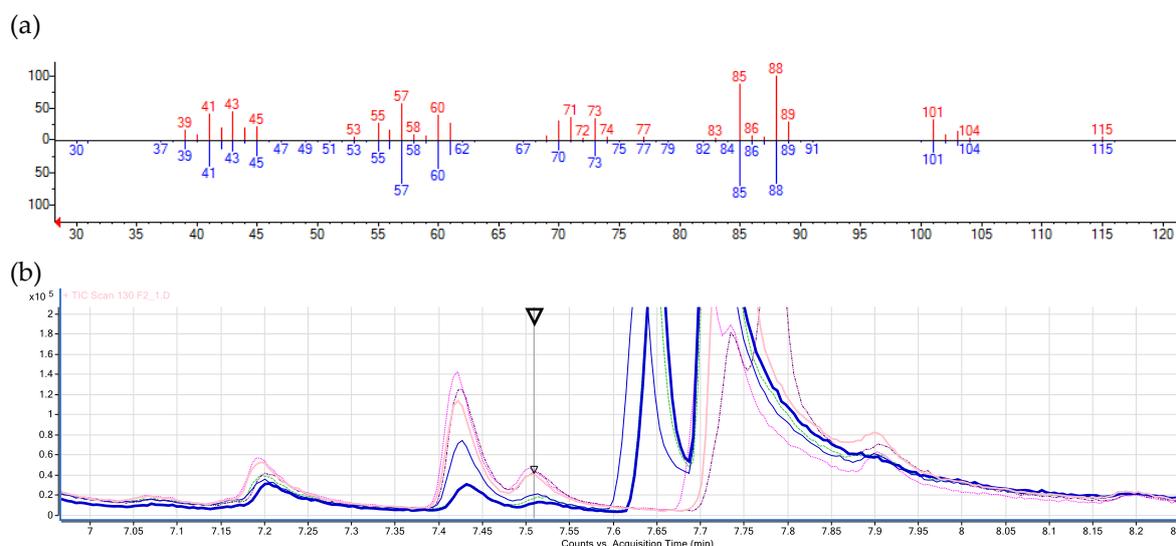


**Figure S2: Insert maps of the fosmid clones.** Sequences of fosmid clones displaying annotated genes (pink), unknown genes (grey) and genes from the fosmid backbone (white).

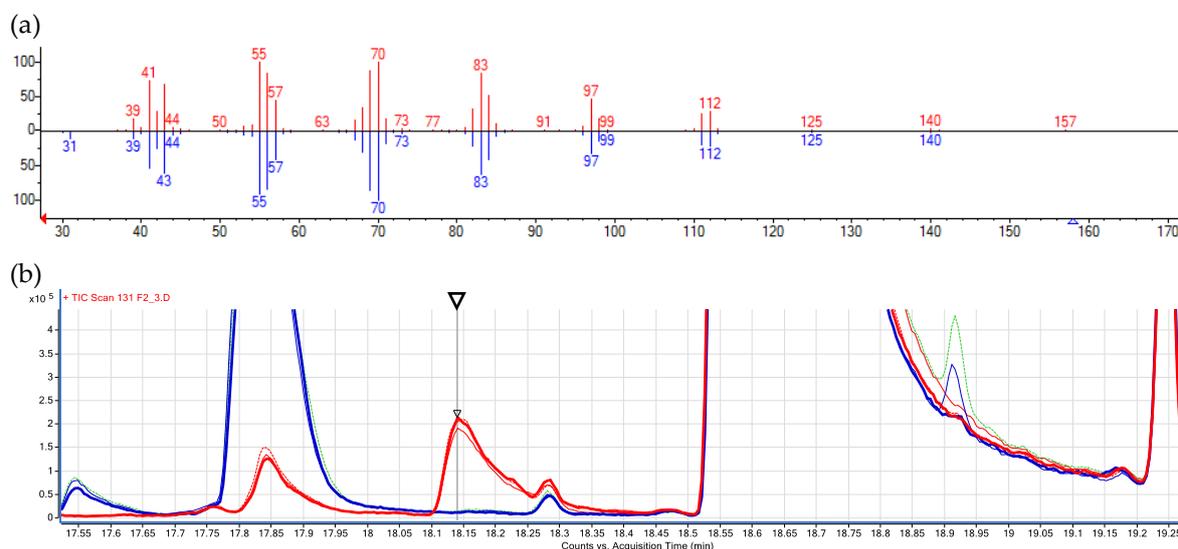


**Figure S3. Volatile inhibition of *F. culmorum*.** Growth inhibition of 7.5% by the metagenomic library clone *E. coli* EPI300 pCC2FOS 131 E3 (center and right) as compared to the empty vector control *E. coli* EPI300 pCC2FOS (left).

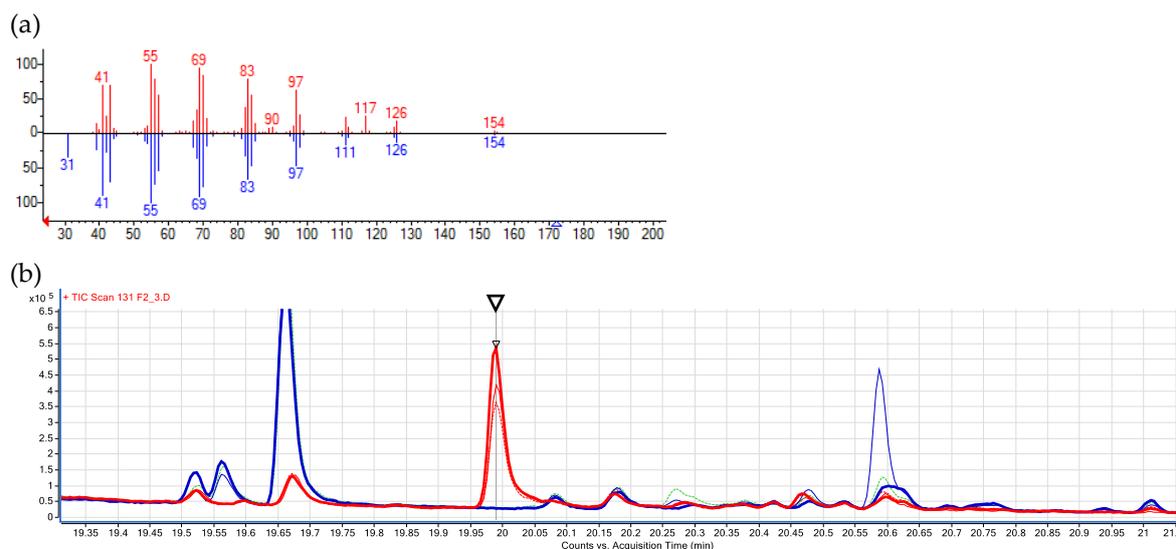
## VOCs profiling through SPME GC-MS



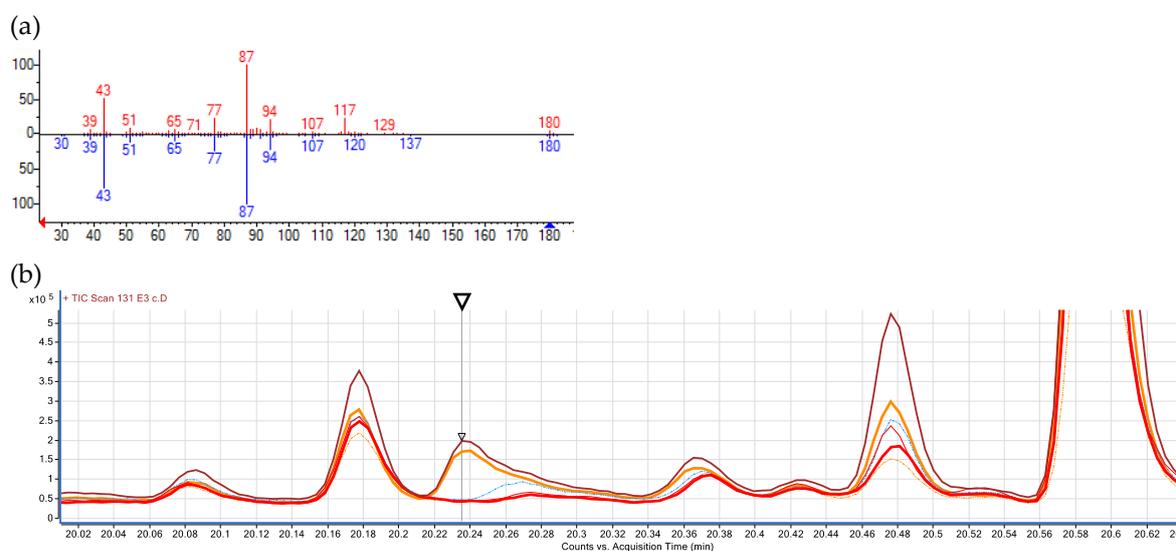
**FIGURE S4. Detection of valeric acid.** (a) Comparison of the recorded mass spectrum (red) and the spectrum of the NIST reference (blue) for valeric acid. (b) SPME GC-MS measurement displaying scans of the metagenomic clone (pink) as compared to the empty vector strain *E. coli* EPI300 pCC2FOS (blue and green) with sample size n=3. The arrow at retention time 7.509 minutes marks the significantly enriched peak, later identified as valeric acid.



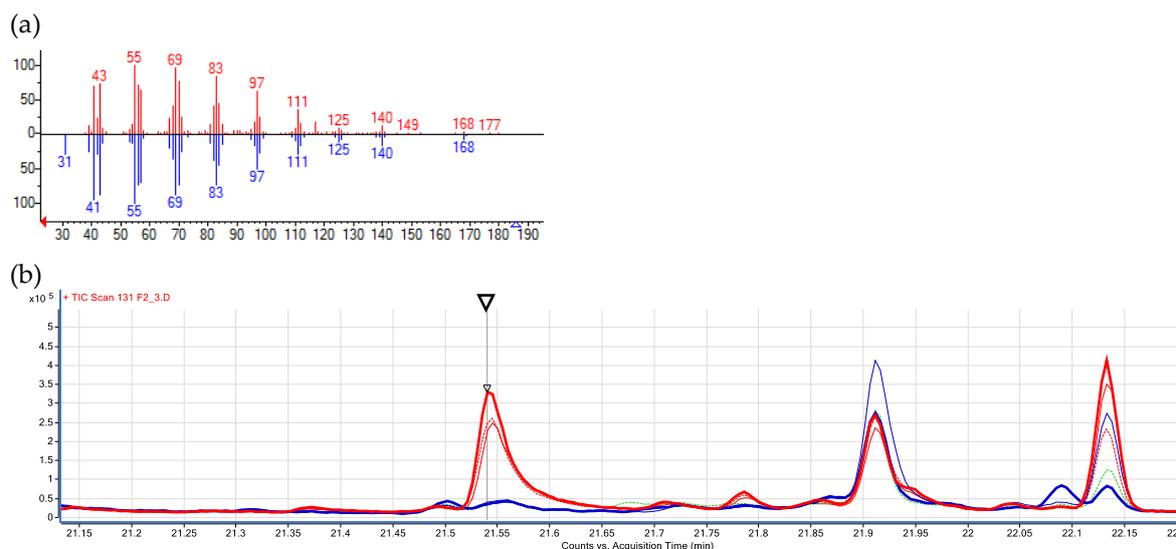
**FIGURE S5. Detection of 1-decanol.** (a) Comparison of the recorded mass spectrum (red) and the spectrum of the NIST reference (blue) for 1-decanol. (b) SPME GC-MS measurement displaying scans of the metagenomic clone (red) as compared to the empty vector strain *E. coli* EPI300 pCC2FOS (blue and green) with sample size n=3. The arrow at retention time 18.570 minutes marks the unique peak later identified as 1-decanol.



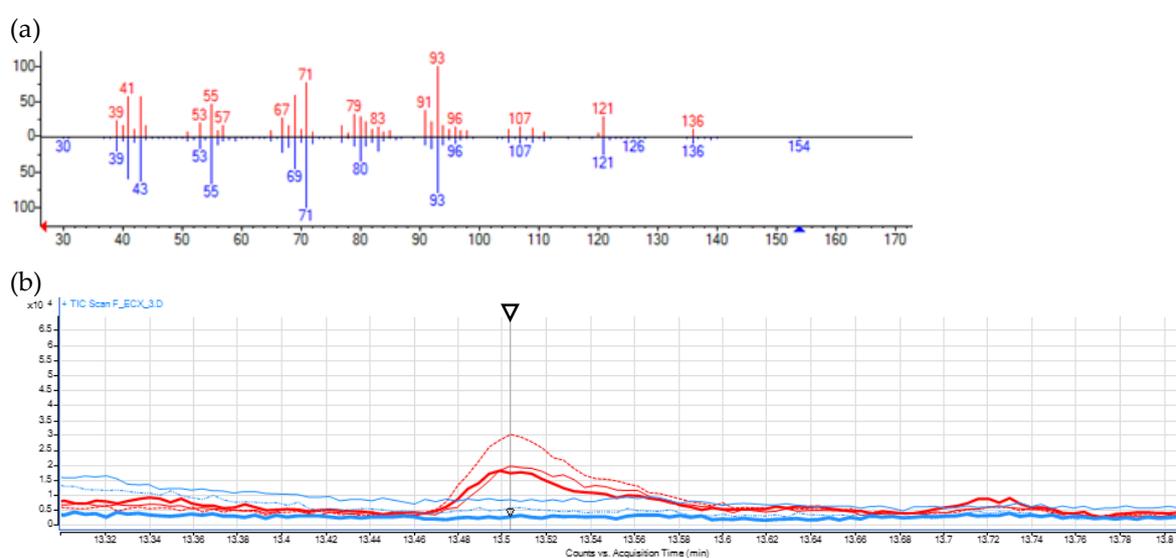
**FIGURE S6. Detection of 1-undecanol.** (a) Comparison of the recorded mass spectrum (red) and the spectrum of the NIST reference (blue) for 1-undecanol. (b) SPME GC-MS measurement displaying scans of the metagenomic clone (red) as compared to the empty vector strain *E. coli* EPI300 pCC2FOS (blue and green) with sample size n=3. The arrow at retention time 19.990 minutes marks the unique peak later identified as 1-undecanol.



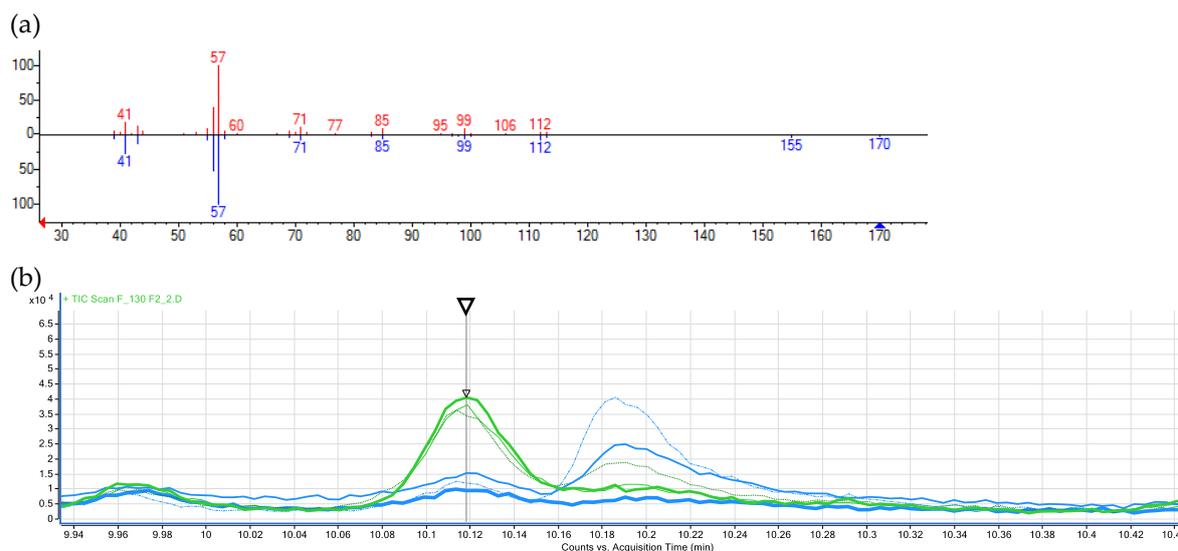
**FIGURE S7. Detection of 2-phenoxyethyl acetate.** (a) Comparison of the recorded mass spectrum (red) and the spectrum of the NIST reference (blue) for 2-phenoxyethyl acetate. (b) SPME GC-MS measurement displaying scans of the metagenomic clone (wine red to orange) as compared to the empty vector strain *E. coli* EPI300 pCC2FOS (blue and red) with sample size n=3. The arrow at retention time 20.235 minutes marks the unique peak later identified as 2-phenoxyethyl acetate.



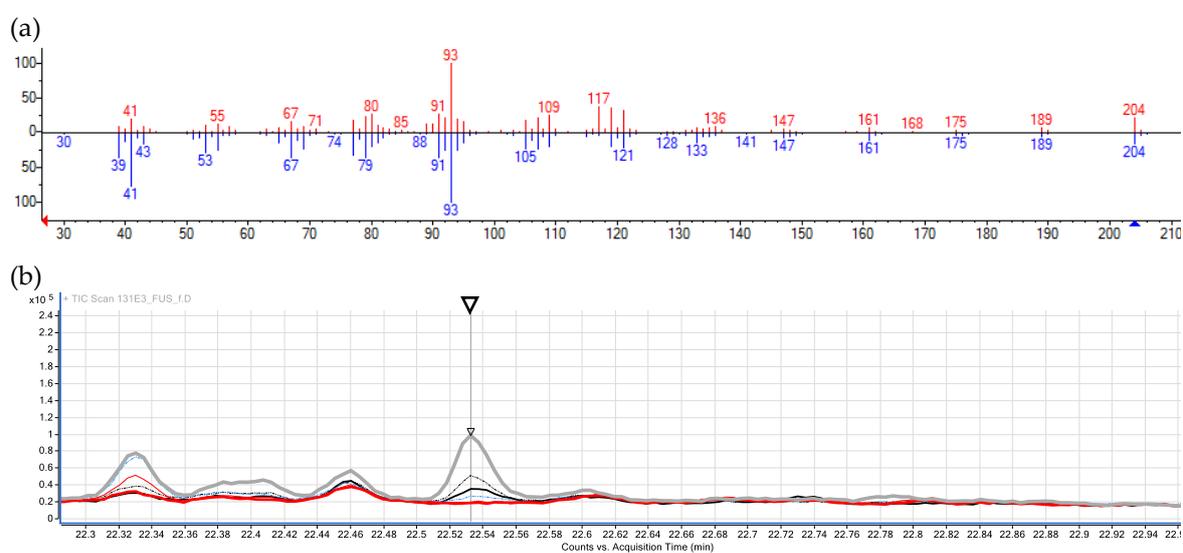
**FIGURE S8. Detection of 1-dodecanol.** (a) Comparison of the recorded mass spectrum (red) and the spectrum of the NIST reference (blue) for 1-dodecanol. (b) SPME GC-MS measurement displaying scans of the metagenomic clone (red) as compared to the empty vector strain *E. coli* EPI300 pCC2FOS (blue) with sample size  $n=3$ . The arrow at retention time 21.541 minutes marks the unique peak later identified as 1-dodecanol.



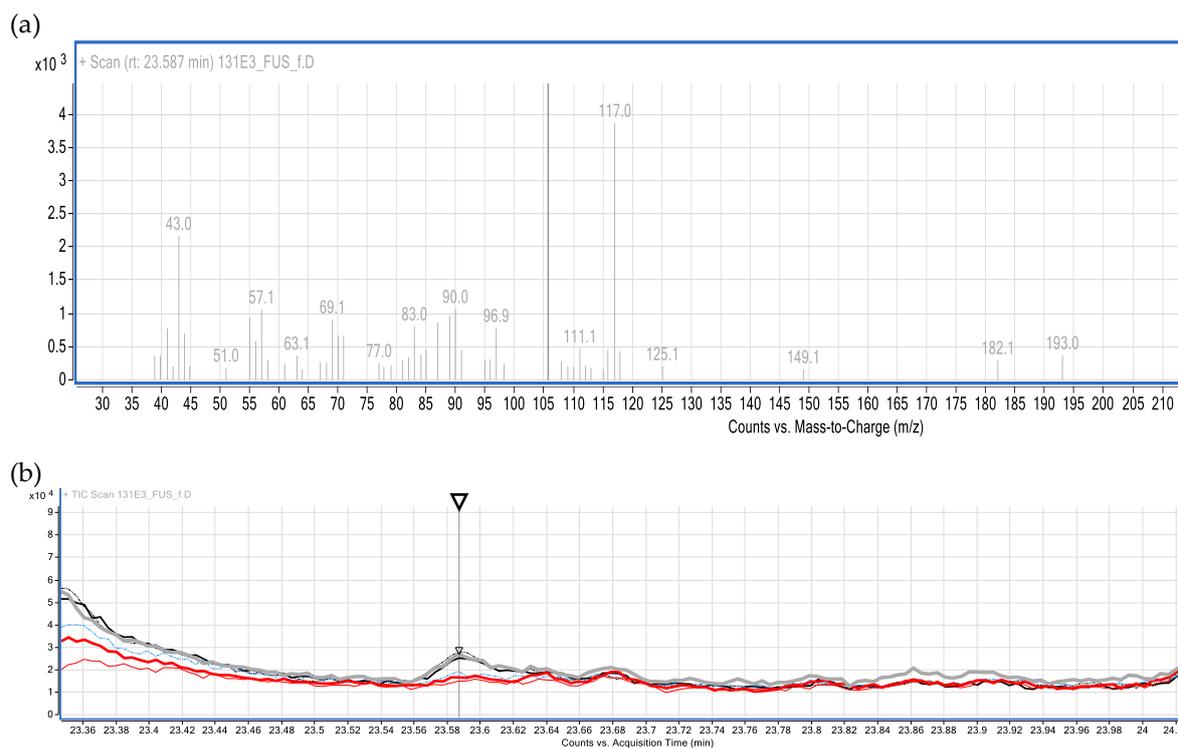
**FIGURE S9. Detection of linalool.** (a) Comparison of the recorded mass spectrum (red) and the spectrum of the NIST reference (blue) for linalool. (b) SPME GC-MS measurement displaying scans of the metagenomic clone (red) as compared to the empty vector strain *E. coli* EPI300 pCC2FOS (blue) with sample size  $n=3$ . The arrow at retention time 13.504 minutes marks the unique peak later identified as linalool.



**FIGURE S10. Detection of 2,2,4,6,6-pentamethylheptane.** (a) Comparison of the recorded mass spectrum (red) and the spectrum of the NIST reference (blue) for 2,2,4,6,6-pentamethylheptane. (b) SPME GC-MS measurement displaying scans of the metagenomic clone (green) as compared to the empty vector strain *E. coli* EPI300 pCC2FOS (cyan) with sample size  $n=3$ . The arrow at retention time 10.119 minutes marks the unique peak later identified as 2,2,4,6,6-pentamethylheptane.



**FIGURE S11. Detection of  $\alpha$ -bisabolene.** (a) Comparison of the recorded mass spectrum (red) and the spectrum of the NIST reference (blue) for  $\alpha$ -bisabolene. (b) SPME GC-MS measurement displaying scans of the metagenomic clone (blue, grey and black) as compared to the empty vector strain *E. coli* EPI300 pCC2FOS (red) with sample size  $n=3$ . The arrow at retention time 22.533 minutes marks the unique peak later identified as  $\alpha$ -bisabolene.



**FIGURE S12. Detection of a not identified substance produced during interaction between *F. culmorum* and metagenomic library clone 131 E3.** (a) The mass spectrum of the measured substance is depicted. (b) SPME GC-MS measurement displaying scans of the metagenomic clone (blue, grey and black) as compared to the empty vector strain *E. coli* EPI300 pCC2FOS (red) with sample size  $n=3$ . The arrow at retention time 23.587 minutes marks the unique peak later identified as an unknown substance.

## TABLES

Table S1. Inhibition efficacy for different concentrations of the detected substances against *F. culmorum*.

Substance	Amount of substance ( $\mu\text{mol}$ )	Concentration (mM)	Inhibition Rate (%)	Standard deviation (%)
1-Undecanol	0.1	0.7	0	0
	1	6.7	0	0
	25	166.7	0	0
	50	333.3	0	0
	100	666.7	0	0
	250	1,666.7	0	0
	500	3,333.3	3.1	5.4
	716	4,773.3	48.2	5.6
1-Decanol	0.1	0.7	0	0
	1	6.7	1.8	2.5
	25	166.7	0.8	0.7
	50	333.3	0	0
	100	666.7	1.2	1.2
	250	1,666.7	1.6	1.8
	500	3,333.3	5.1	3.8
	770	5,133.3	51.8	4.6
1-Dodecanol	0.1	0.7	2.4	0
	1	6.7	0.4	0.7
	25	166.7	1.6	1.8
	50	333.3	2.0	3.4
	100	666.7	3.5	6.1
	250	1,666.7	10.2	3.6
	500	3,333.3	12.9	1.7
	657	4,380.0	14.3	9.3
2-Phenoxyethyl acetate	0.1	0.7	4.7	2.0
	1	6.7	0.4	0.7
	25	166.7	1.6	2.7
	50	333.3	0.4	0.7
	100	666.7	1.2	1.2
	250	1,666.7	2.4	2.0
	500	3,333.3	2.4	1.2
	933	6,220.0	25.0	7.1
$\alpha$ -Bisabolol	0.1	0.7	0	0
	1	6.7	0	0
	25	166.7	0	0
	50	333.3	0	0
	100	666.7	0.4	0.7
	250	1,666.7	0.6	0.8

	500	3,333.3	11.8	6.1
	503	3,353.3	11.5	2.3
<b>Linalool</b>	0.1	0.7	5.1	3.6
	1	6.7	17.6	5.4
	25	166.7	38.8	7.3
	50	333.3	69.0	6.9
	100	666.7	84.7	7.1
	250	1,666.7	100	0
	500	3,333.3	100	0
	812	5,413.3	100	0
	<b>2,2,4,6,6-Pentamethylheptane</b>	0.1	0.7	0.4
1		6.7	0.8	1.4
25		166.7	0	0
50		333.3	0	0
100		666.7	0	0
250		1,666.7	0	0
500		3,333.3	0	0
647		4,313.3	11.3	2.6
<b>Valeric acid</b>	0.1	0.7	10.2	7.7
	1	6.7	100	0
	25	166.7	100	0
	50	333.3	100	0
	100	666.7	100	0
	250	1,666.7	100	0
	500	3,333.3	100	0

## References

- [1] Obermeier, M.M.; Wicaksono, W.A.; Taffner, J.; Bergna, A.; Poehlein, A.; Cernava, T.; Lindstaedt, S.; Lovric, M.; Müller Bogotá, C.A.; Berg, G. Plant resistome profiling in evolutionary old bog vegetation provides new clues to understand emergence of multi-resistance. *ISME J* (under review).
- [2] Bolger, A.M.; Lohse, M.; Usadel, B. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* **2014**, *30*, 2114–20.
- [3] Bankevich, A.; Nurk, S.; Antipov, D.; Gurevich, A.A.; Dvorkin, M.; Kulikov, A.s.; Lesin,, V.M.; Nikolenko, S.I.; Pham, S.; Prjibelski, A.D.; Pyshkin, A.V.; Sirotkin, A.V.; Vyahhi, N.; Tesler, G.; Alekseyev, M.A.; Pevzner, P.A. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* **2012**, *19*, 455–77.
- [4] Seemann T. Prokka: rapid prokaryotic genome annotation. *Bioinformatics* **2014**, *30*, 2068–9.