

## **Supporting Information**

### **New peptaibiotics and a cyclodepsipeptide from *Ijuhyia vitellina*: Isolation, identification, cytotoxic and nematicidal activities.**

**Ashaimaa Y. Moussa<sup>1,2</sup>, Christopher Lambert<sup>3</sup>, Theresia E.B. Stradal<sup>3</sup>, Samad Ashrafi<sup>4</sup>, Wolfgang Maier<sup>4</sup>, Marc Stadler<sup>1</sup>, Soleiman Helaly<sup>1,5\*</sup>**

<sup>1</sup> Department of Microbial Drugs, Helmholtz Centre for Infection Research, Inhoffenstraße 7, 38124 Braunschweig, Germany.

<sup>2</sup> Department of Pharmacognosy, Faculty of Pharmacy, Ain shams University, Abbassia, 11566, Cairo, Egypt.

<sup>3</sup> Department of Cell Biology, Helmholtz Centre for Infection Research (HZI), Inhoffenstraße 7, 38124, Braunschweig, Germany.

<sup>4</sup> Institute for Epidemiology and Pathogen Diagnostics, Julius Kühn-Institut (JKI)–Federal Research Centre for Cultivated Plants, Braunschweig, Germany.

<sup>5</sup> Department of Chemistry, Faculty of Science, Aswan University, Aswan, Egypt.

\*Correspondence: soleiman.helaly@helmholtz-hzi.de.

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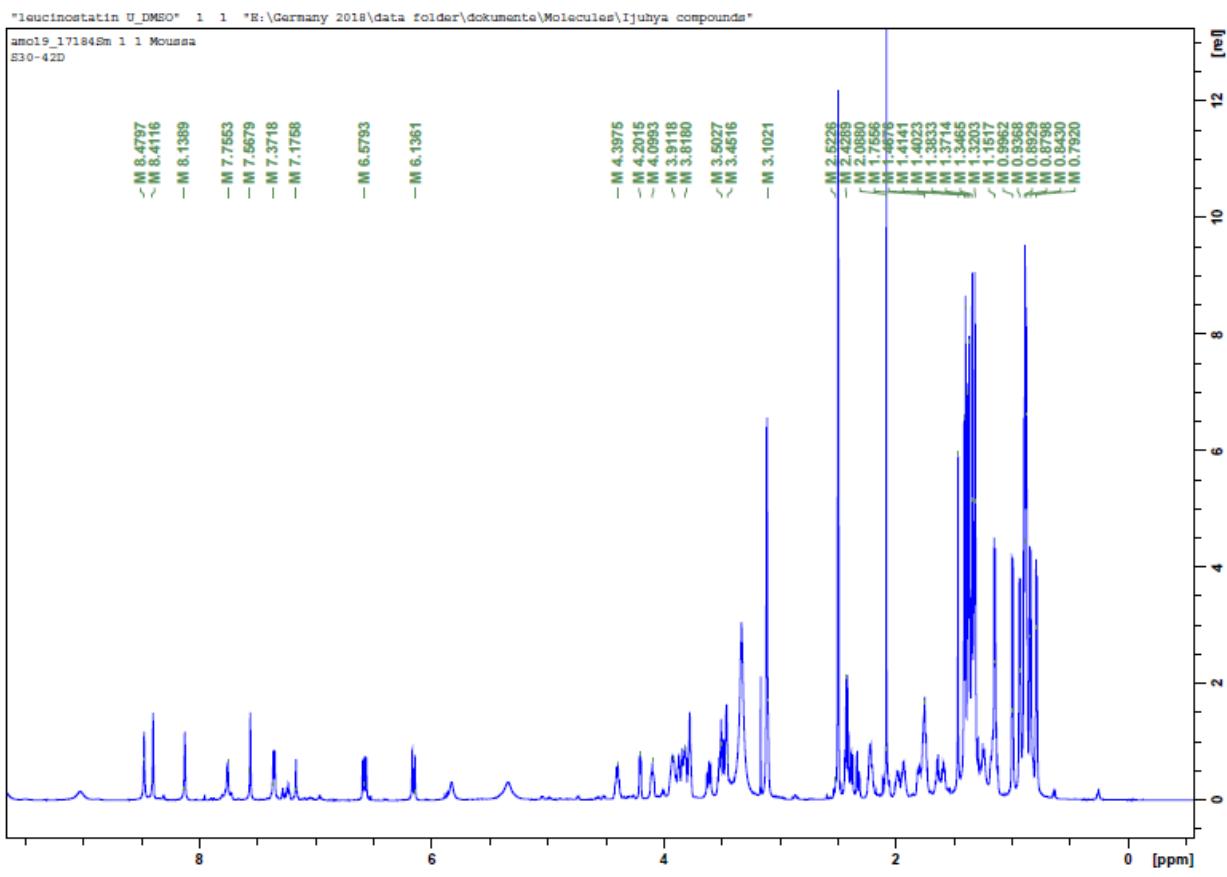
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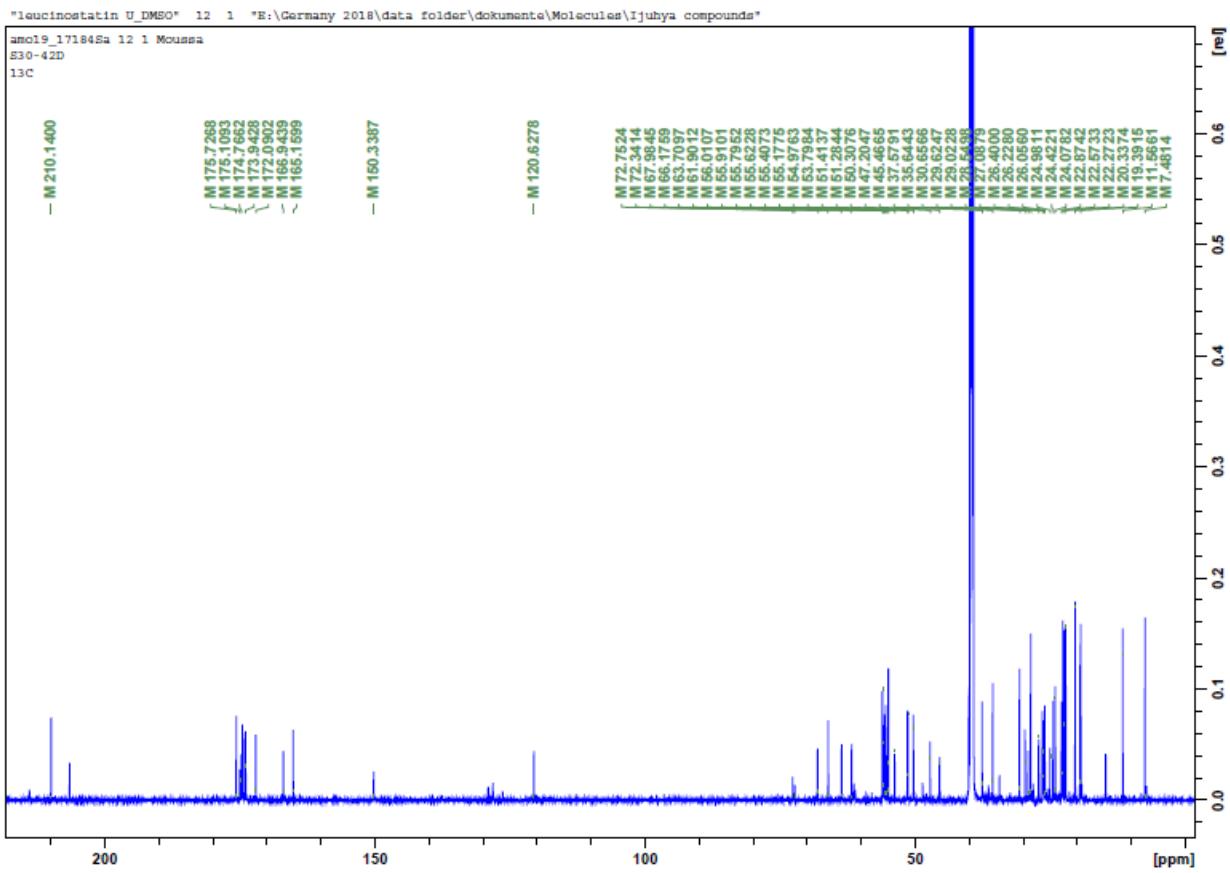
**S1 Table.**  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR data of **1** & **2** and the characteristic chemical shift values.

residue	position	leucinostatin U (1)		leucinostatin Q (2)	
		$\delta\text{C}$ , type	$\delta\text{H}$ ( J in Hz)	$\delta\text{C}$ , type	$\delta\text{H}$ ( J in Hz)
<b>ΔMHA</b>	1	165.15, C 120.27,		165.45,C	
	2	CH	6.14, (15.04) 6.58 (7.7,	120.65,CH	6.25
	3	150.02,CH	15.0)	151.24,CH	6.66
	4	38.10,CH	2.22	37.82,CH	2.26
	5	29.23,CH2	1.36	28.86,CH2	1.38
	6	12.10,CH3	0.88	11.88,CH3	0.87
	7	19.86,CH3	0.99(6.3)	18.85,CH3	1.04
<b>Pro</b>	1	174.76,C		173.6,C	
	2	62.28,CH	4.2(4.12, 8.9)	61.61,CH	4.26
	3	30.22,CH2	2.22 1.8	29.84,CH2	2.19 1.82
	4	24.80,CH2	1.94	24.80,CH2	1.94
	5	47.75,CH2	3.91 3.52	47.53,CH2	3.81 3.6
	6				
	7				
<b>AHMOD</b>	1	175.05, C		173.5,C	
	2	54.38,CH 34.82,	4.09	52.84,CH	4.17
	3	CH2	2.07 1.16	36.01,CH2	1.88 1.36
	4	25.47,CH	1.76	26.18,CH	1.68
	5	46.19,CH2	1.25 1.17	45.58,CH2	1.29 1.24
	6	64.23,CH	3.94	64.86,CH	3.97
	7	50.98,CH2	2.39(3.4,14.3) 2.33(8.7,14.3)	50.57,CH2	2.42
	8	210.41,C		210.12,C	
	9	36.14,CH2	2.42	36.26,CH2	2.44
	10	7.89,CH3	0.89(7.4)	7.79,CH3	0.9
	11	22.91,CH3	0.89	23.32,CH3	0.9
	NH		9.68		6.97
<b>Hyleu</b>	1	172.10,C		172.16	
	2	61.77,CH	3.81	58.64,CH	4.11
	3	73.24,CH	3.76	75.05,CH	3.64
	4	29.67,CH	1.59	30.34,CH	1.58
	5	15.13,CH3	0.88	17.48,CH3	0.91
	6	19.88,CH3	0.79	19.70,CH3	0.83
	NH		9.03		7.74
<b>Aib1</b>	1	174.56,C		174.6	

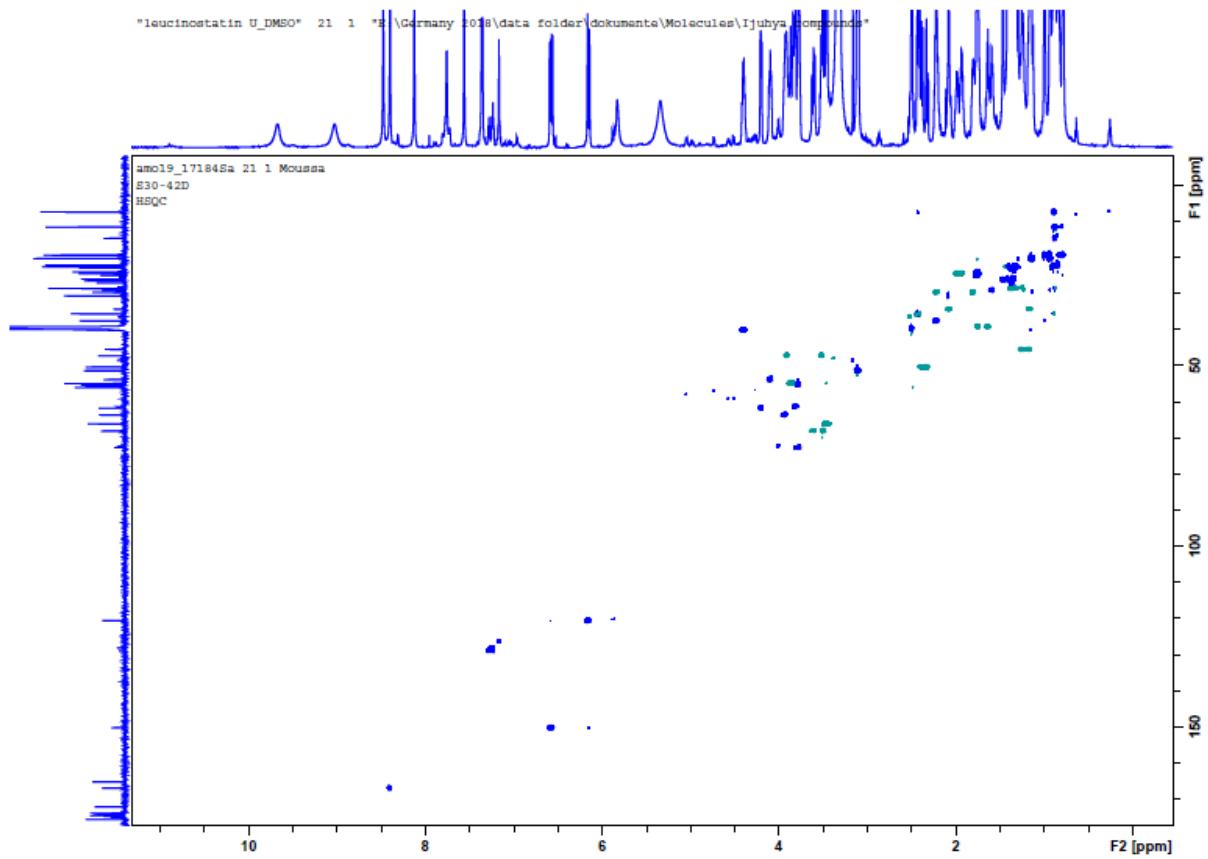
	2	55.7,C		55.80,C	
	3	22.30,CH3	1.34	23.39,CH3	1.38
	4	26.10,CH3	1.41	25.14,CH3	1.34
	NH		8.47		8.16
<b>Aib2</b>	1	175.64,C		173.24,C	
	2	56.0,C		55.94,C	
	3	22.6,CH3	1.32	25.63,CH3	1.31
	4	25.83,CH3	1.46	23.32,CH3	1.38
	NH		7.56		7.43
<b>Leu</b>	1	174.43,C		173.16,C	
	2	55.82,CH	3.78	53.76,CH	3.83
	3	39.80,CH2	1.75	39.41,CH2	1.77
			1.64		1.58
	4	24.54,CH	1.76	24.55,CH	1.76
	5	22.57,CH3	0.84(6.3)	21.75,CH3	0.84
	6	22.91,CH3	0.89	23.32,CH3	0.9
	NH		7.75		6.92
<b>Aib3</b>	1	173.97,C		175.2,C	
	2	56.04,C		56.09,C	
	3	22.3,CH3	1.36	22.90,CH3	1.38
	4	26.54,CH3	1.4	25.66,CH3	1.35
	NH		8.12		7.57
<b>Aib4</b>	1	173.90,C		173.31,C	
	2	55.73,C		55.97,C	
	3	26.79,CH3	1.47	25.76,CH3	1.34
	4	22.90,CH3	1.38	23.44,CH3	1.37
	NH		7.16		7.57
<b>C-terminal</b>	1	40.64,CH	4.4(m)	43.15,CH	3.81
	2	68.65,CH2	3.61(9.8)	63.31,CH2	2.31
			3.5(7.6)		
	3	66.59,CH2	3.46 (5.3)	60.21,CH2	2.45
	4	55.48,CH2	3.86 (5.3)	59.22,CH2	3.44
	5	51.91,CH3	3.11(5.9)	19.27,CH3	1.02
	6	51.91,CH3	3.11(5.9)		
	7	20.86,CH3	1.15(6.4)		
	NH		7.36		7.75
	N-CH3			43.41,CH3	2.2



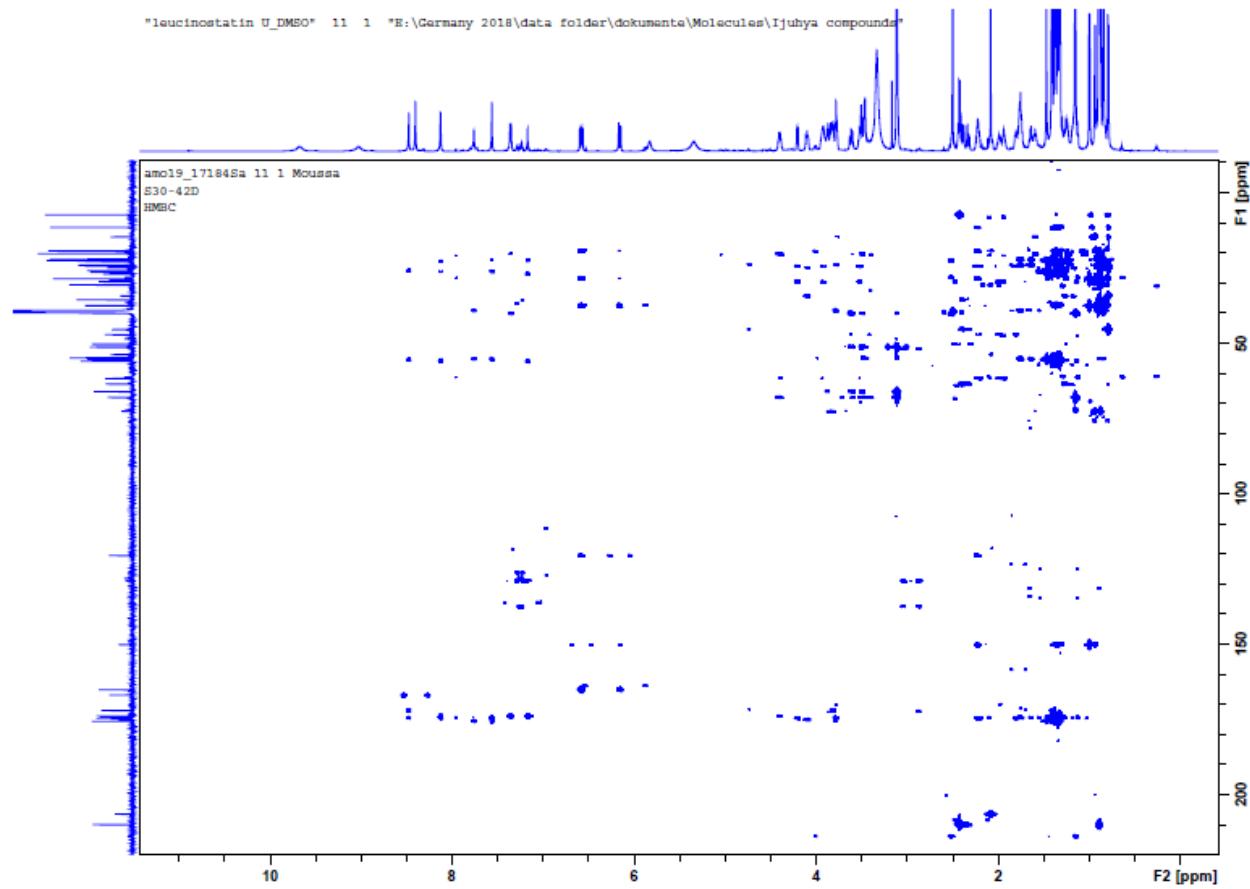
**S1 figure:**  $^1\text{H}$ NMR spectra of **leucinostatin U 1.**



**S2. Figure:**  $^{13}\text{C}$ NMR spectra of leucinostatin U.1.



S3 Figure: HSQC spectra of leucinostatin U 1.

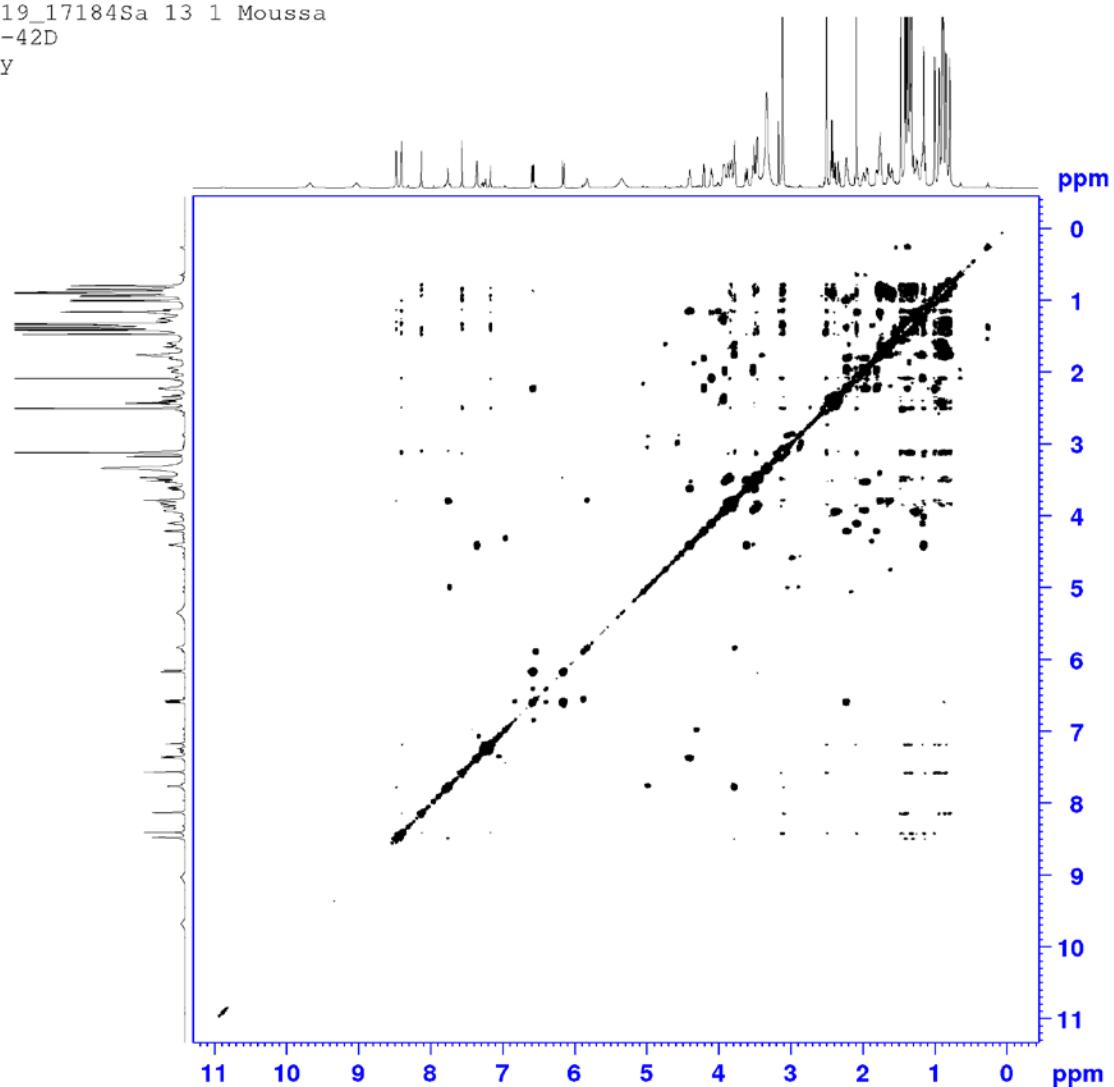


S4 Figure: HMBC spectra of leucinostatin U 1

amo19\_17184Sa 13 1 Moussa

S30-42D

Cosy

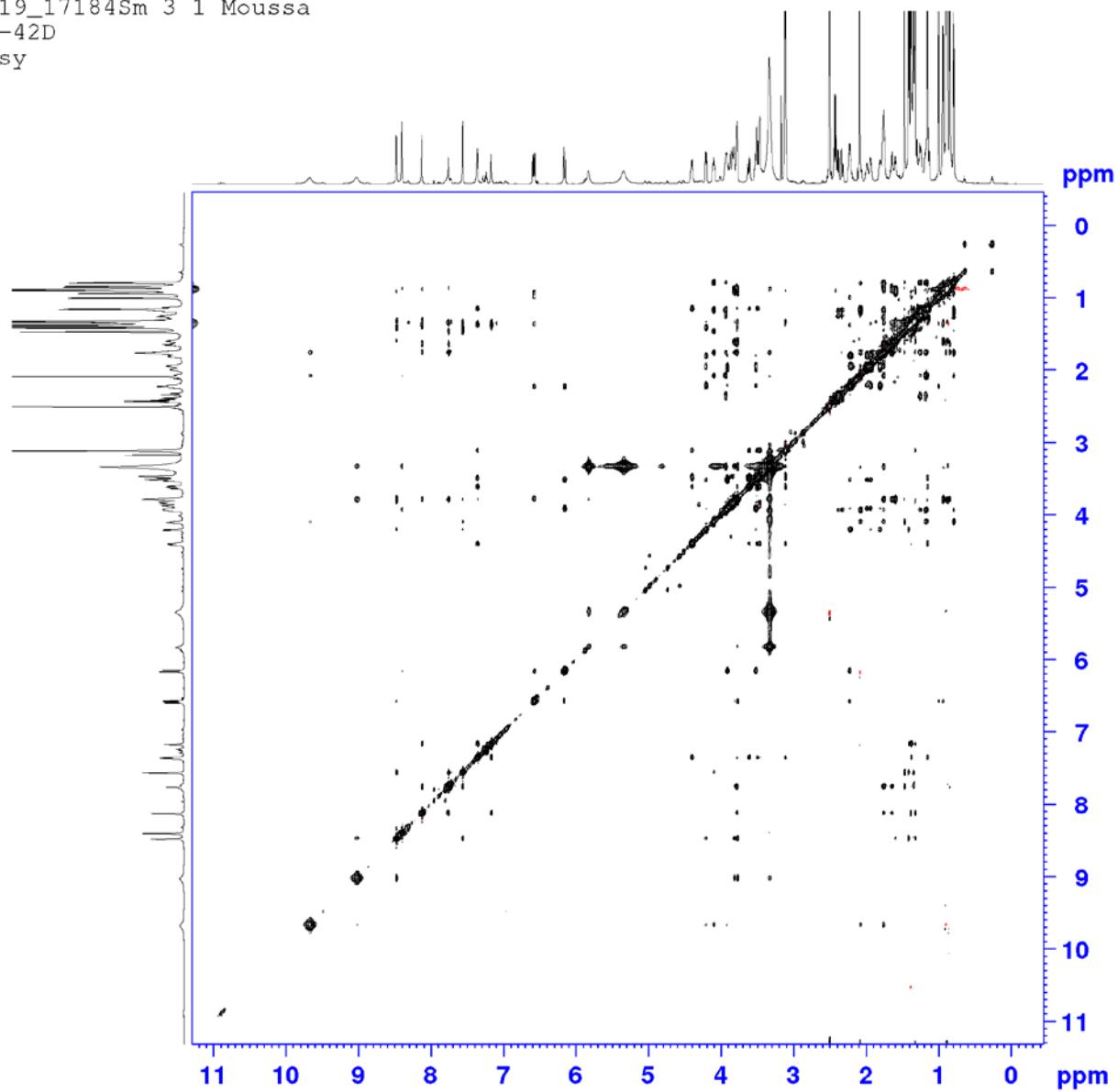


S5 Figure: COSY spectra of leucinostatin U 1.

>19\_17184Sm 3 1 Moussa

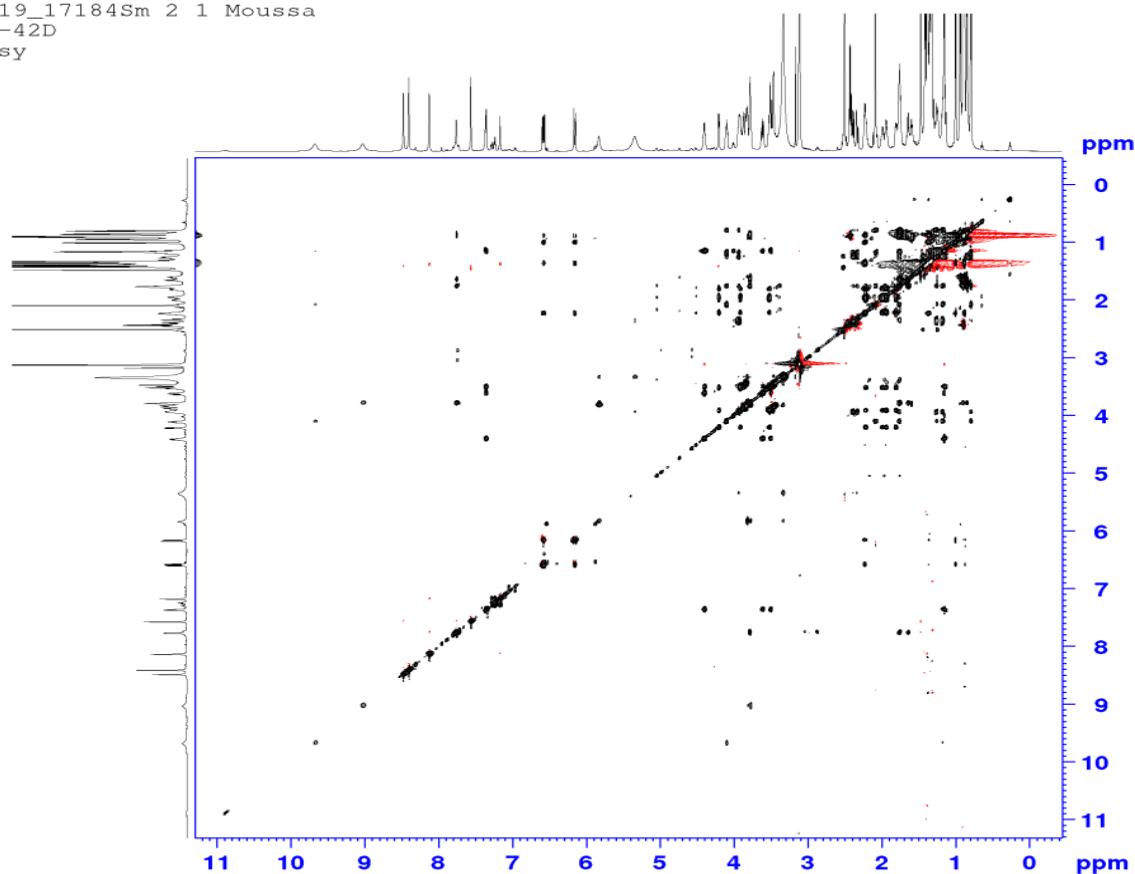
)-42D

=SY

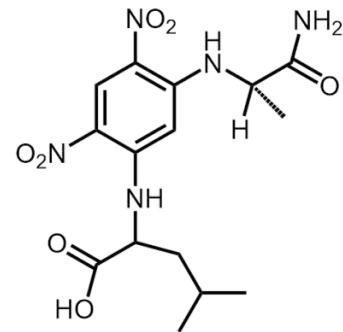
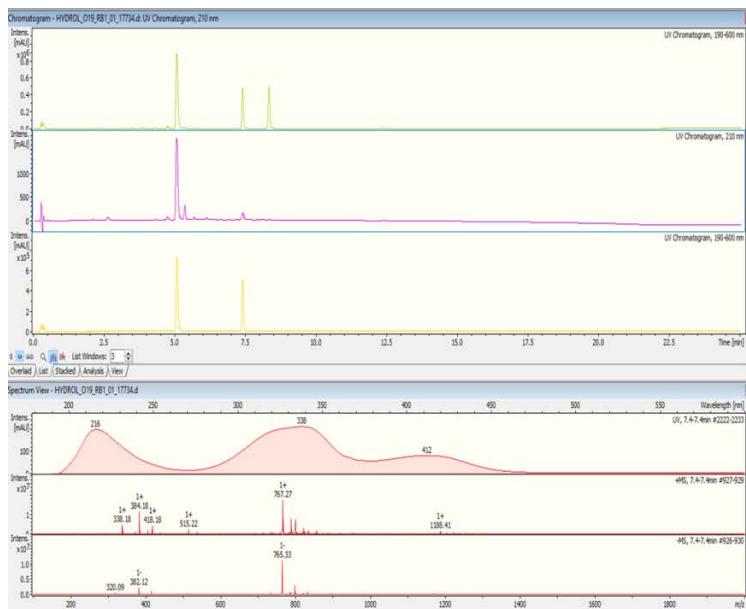


S6 Figure: NOESY spectra of leucinostatin U 1

amo19\_17184Sm 2 1 Moussa  
S30-42D  
Tocsy

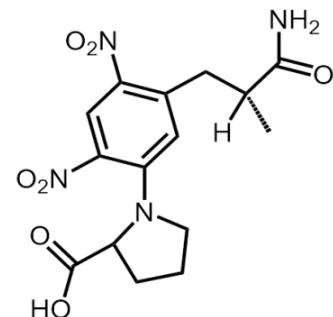
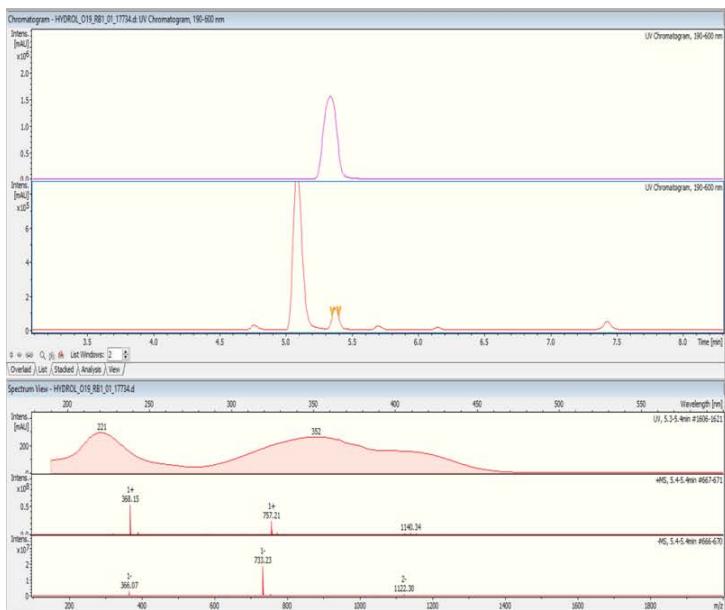


S7 Figure: TOCSY spectra of leucinostatin U 1.



Chemical Formula: C<sub>15</sub>H<sub>21</sub>N<sub>5</sub>O<sub>7</sub>  
Exact Mass: 383,14  
Molecular Weight: 383,36

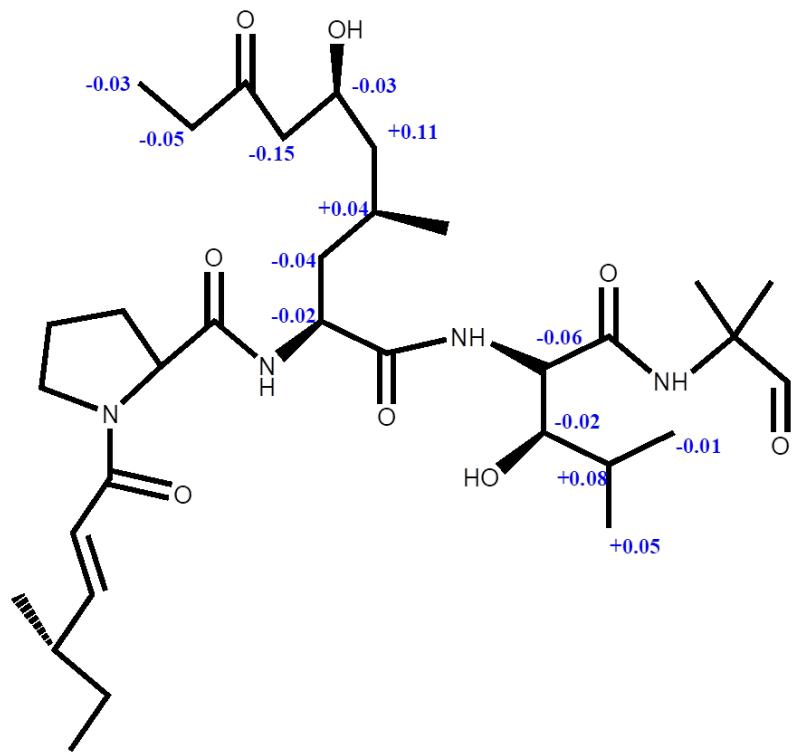
#### FDLA- D-leucine and DL-leucine



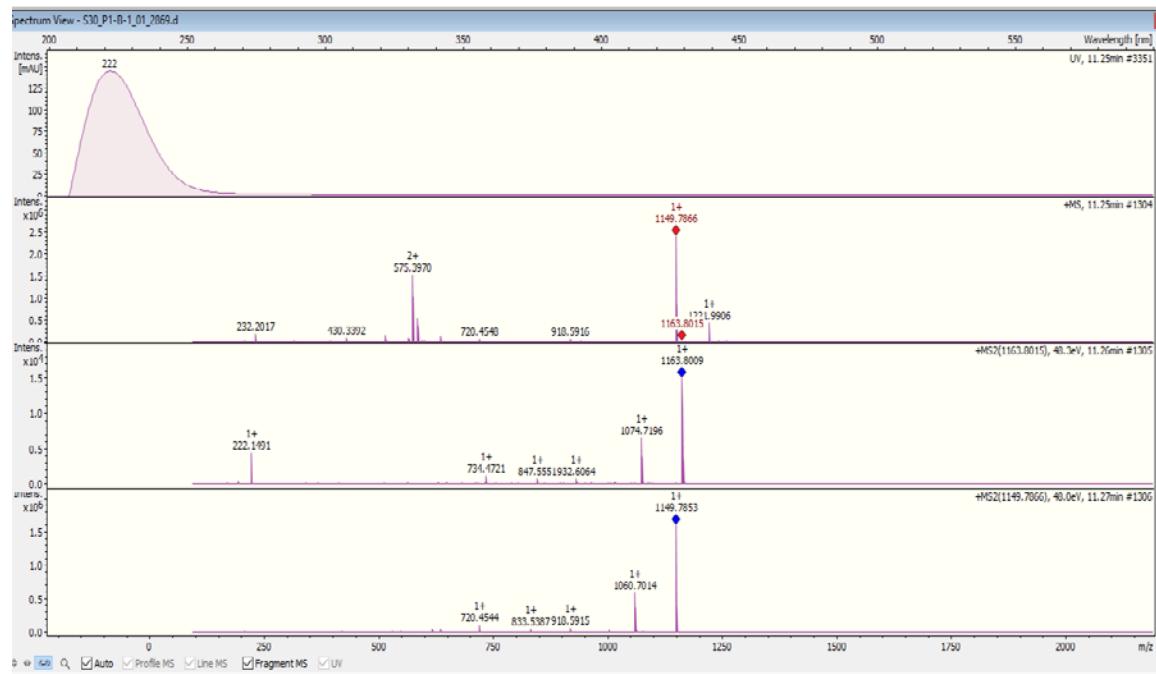
Chemical Formula:  
C<sub>15</sub>H<sub>18</sub>N<sub>4</sub>O<sub>7</sub>  
Exact Mass: 366,12  
Molecular Weight: 366,33

#### FDLA-L- Proline and D-proline

S8 Figure. HR-ESI-MS of Marfay's reaction products of leucinostatin **U 1**.



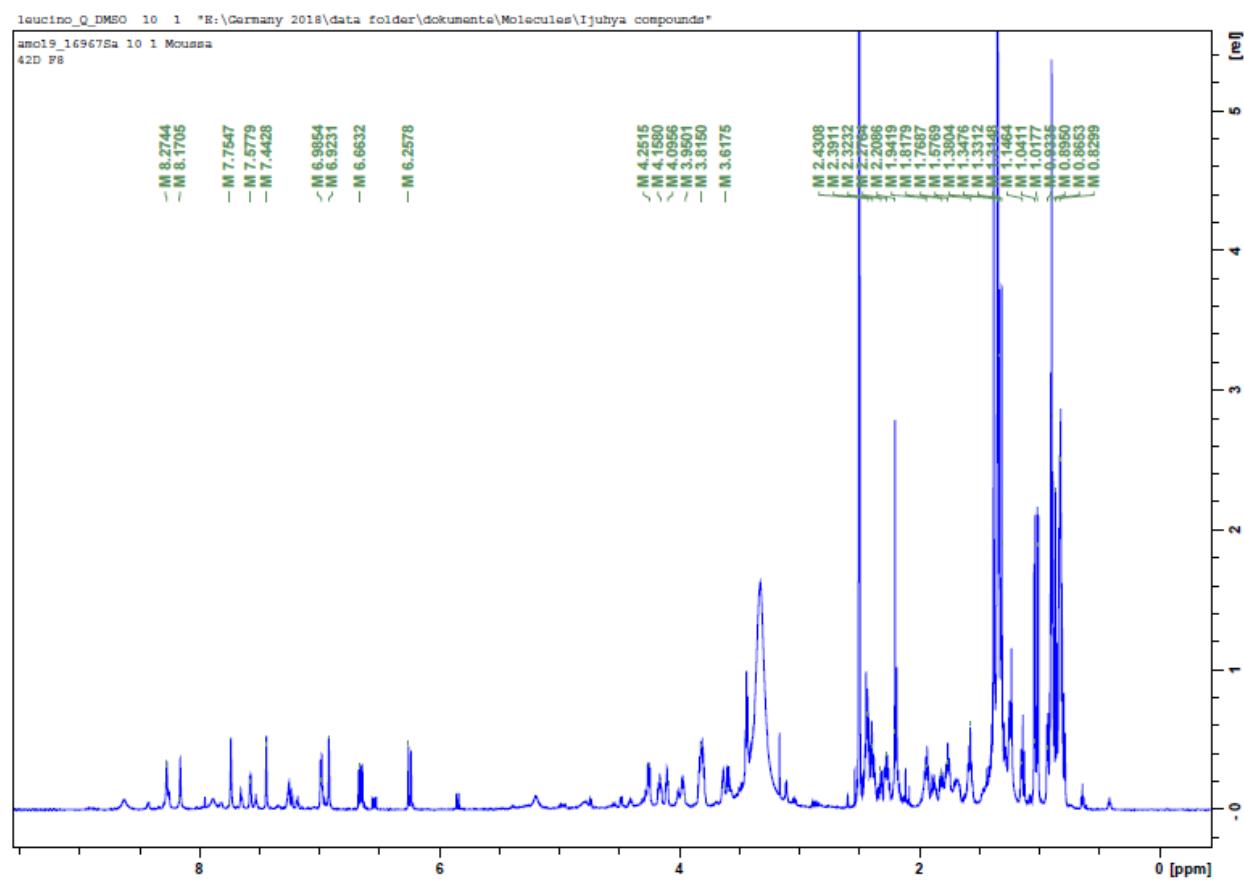
**S9 Figure.**  $\Delta\delta$ SR chemical shifts of MTPA ( $\alpha$ -methoxy- $\alpha$ -trifluoromethylphenylacetic acid) (Mosher) derivatives of the proline, AHMOD and Hy-Leu of **1**.



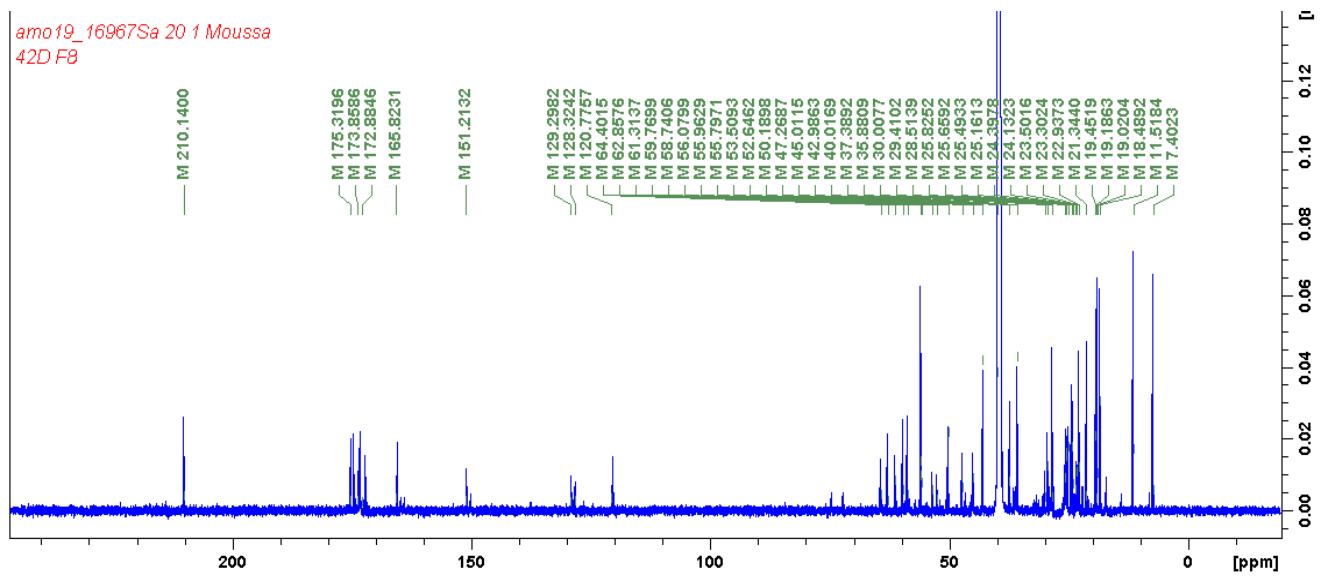
## S10 Figure. HR-ESI-MS-MS fragmentation pattern of leucinostatin U 1.

**S2.Table.** 1H NMR data of **leucinostatin U 1** and the characteristic chemical shift value differences  $\Delta\delta_{SR}$  of the corresponding Mosher esters, recorded at 700 MHz (75% CH3CN/D2O).

position	$\delta^H$ leucinostatin U (1)	$\delta^H$ of leucinostatin U (1) reacted with R-MTPA-Cl	$\delta^H$ of leucinostatin U (1) reacted with S-MTPA-Cl	$\Delta S - \Delta R$ (ppm)	$\Delta S - \Delta R$ (Hz)
<b>AHMOD</b>					
2	4.67	4.73,CH	4.71,CH	-0.02	-14
3	2.57, 1.29	1.52, 2.51CH2	1.92, 2.76,CH2	-0.04	-28
4	1.74	1.72	1.68, CH2	0.04	28
10	1	0.98	0.97	0.01	7
5	1.55, 1.19	1.62, 1.30	1.73	0.11	77
6	4.58	4.68	4.51,CH2	-0.03	-21
7	2.98	2.56, 2.72	3.22, 2.99, CH2	-0.15	-105
8	2.57, 1.29	2.64, 2.53	2.59,2.45,CH2	-0.05	-35
9	1.06	0.99	1.02	-0.03	-21
<b>Hyleu</b>					
2	4.39	4.60,CH	4.55,CH	-0.06	-42
3	4.52	4.45,CH	4.47,CH	-0.02	-14
4	1.88	1.86,CH	1.94,CH	0.08	56
5	0.85	0.88,CH3	0.93,CH3	0.05	35
6	1.42	1.34,CH3	1.35,CH3	-0.01	-1



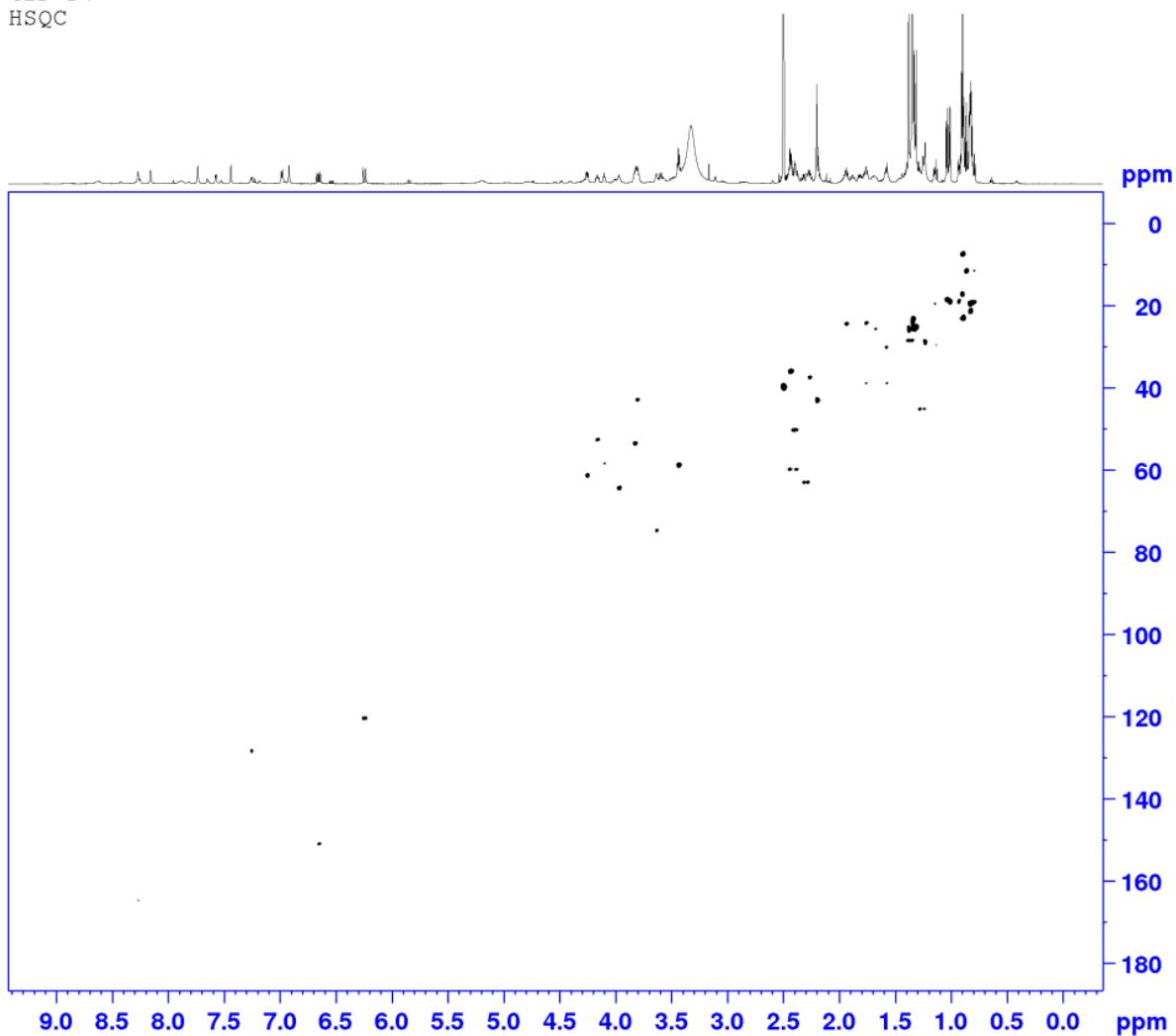
S11 figure:  $^1\text{H}$ NMR spectra of leucinostatin Q 2



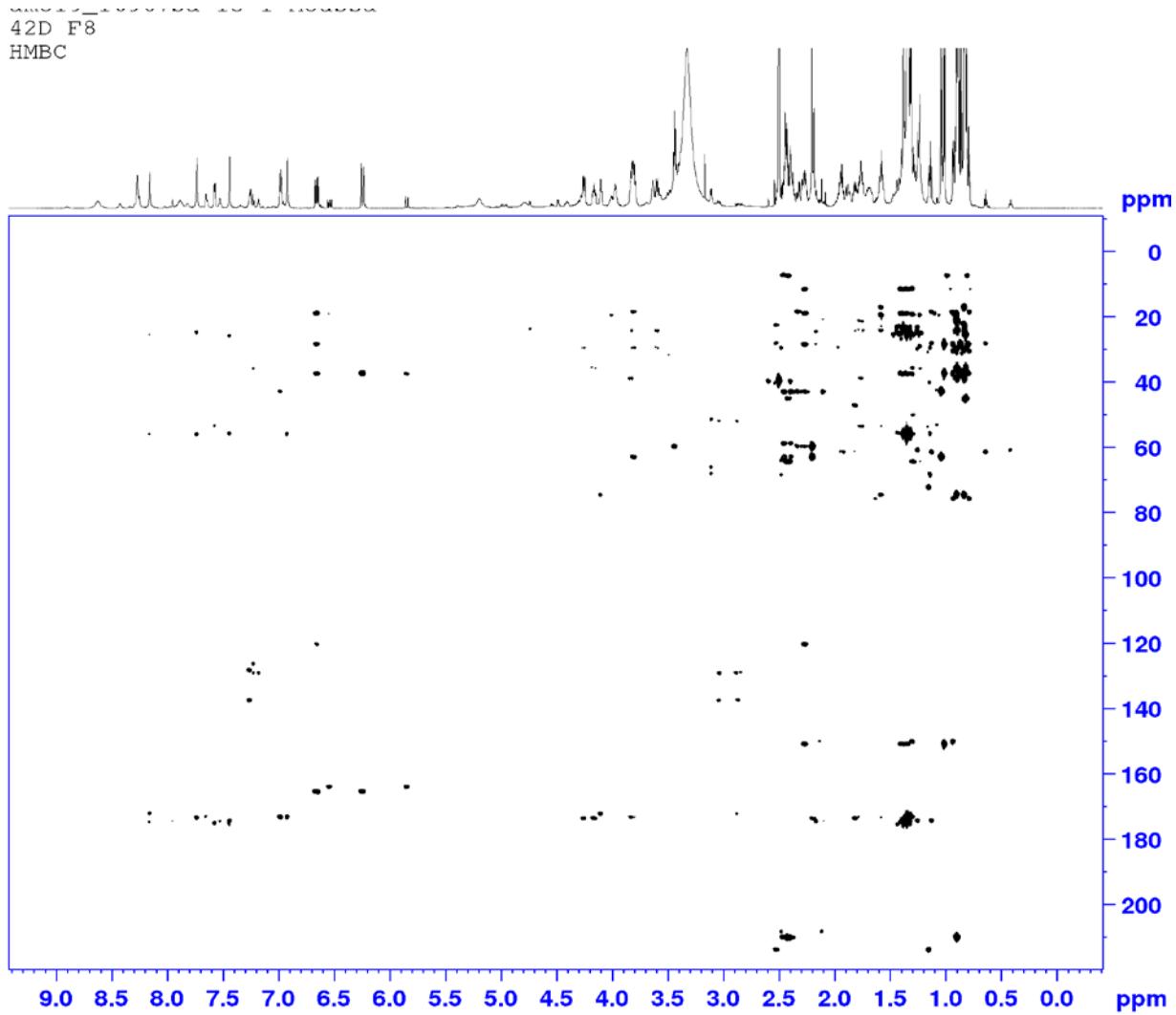
S12. Figure:  $^{13}\text{C}$ NMR spectra of leucinostatin Q 2..



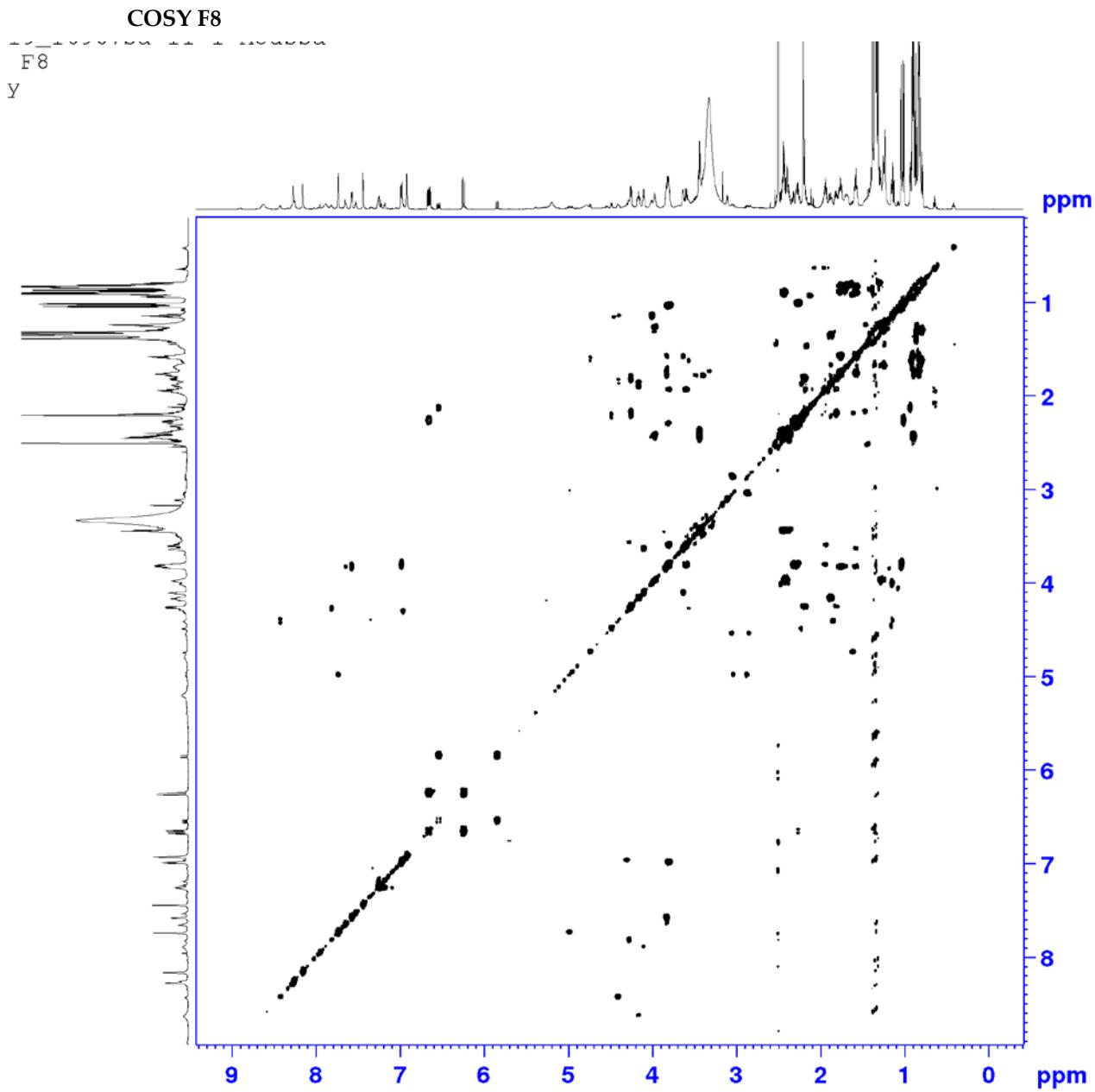
42D F8  
HSQC



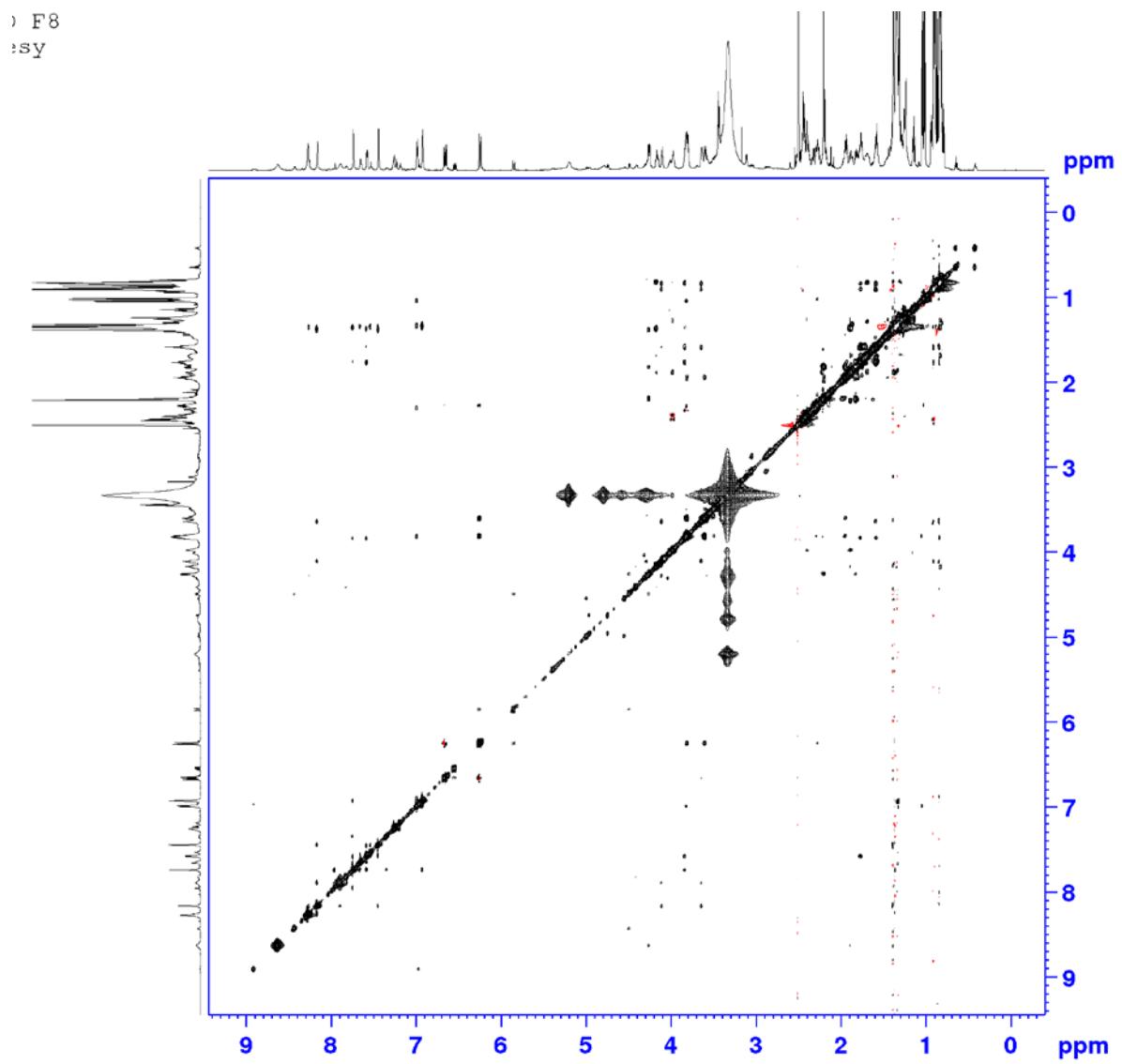
**S13 Figure:** HSQC spectra of leucinostatin Q 2



S14 Figure: HMBC spectra of leucinostatin Q 2

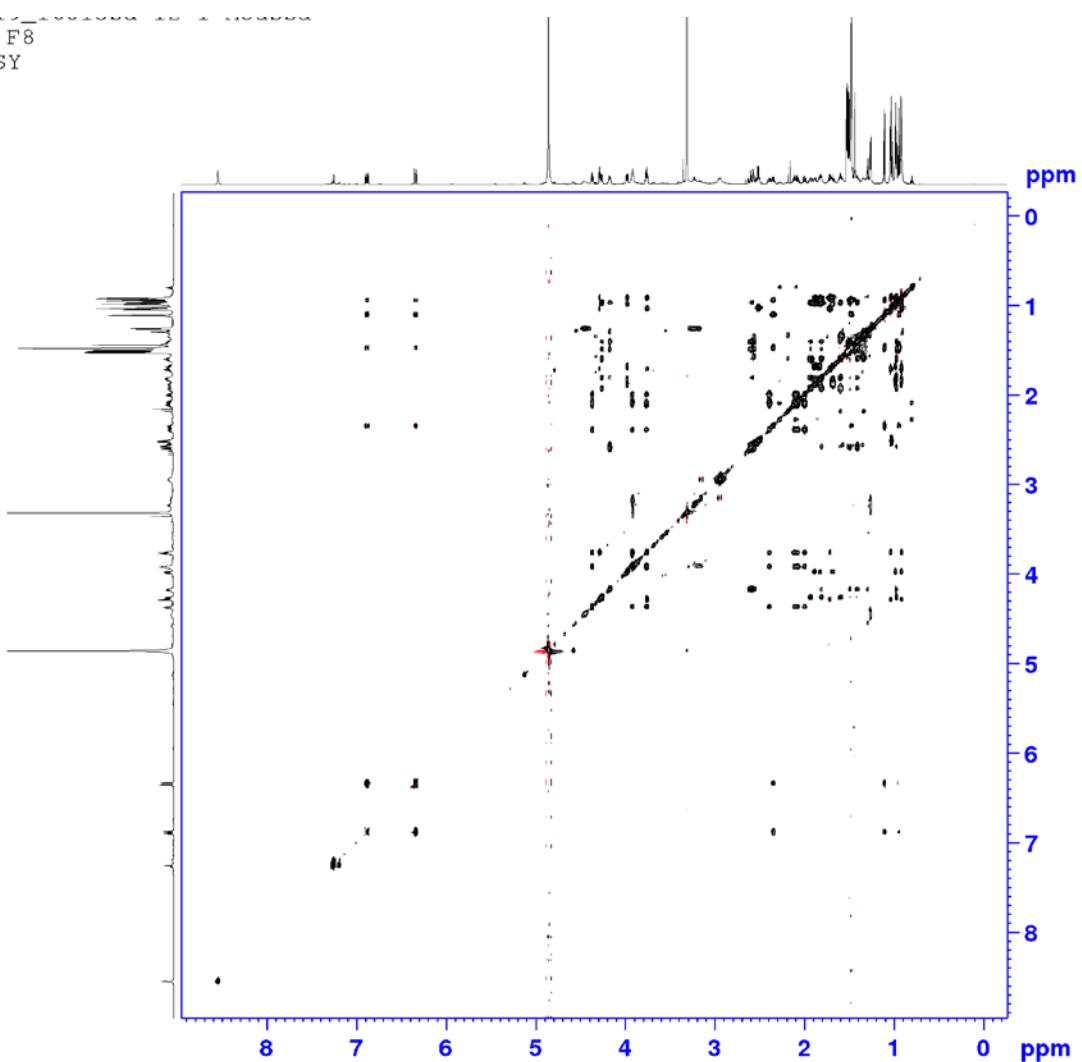


S15 Figure: COSY spectra of leucinostatin Q 2



S16 Figure: NOESY spectra of leucinostatin Q 2

42D F8  
TOCSY



**S17 Figure:** TOCSY spectra of leucinostatin Q 2

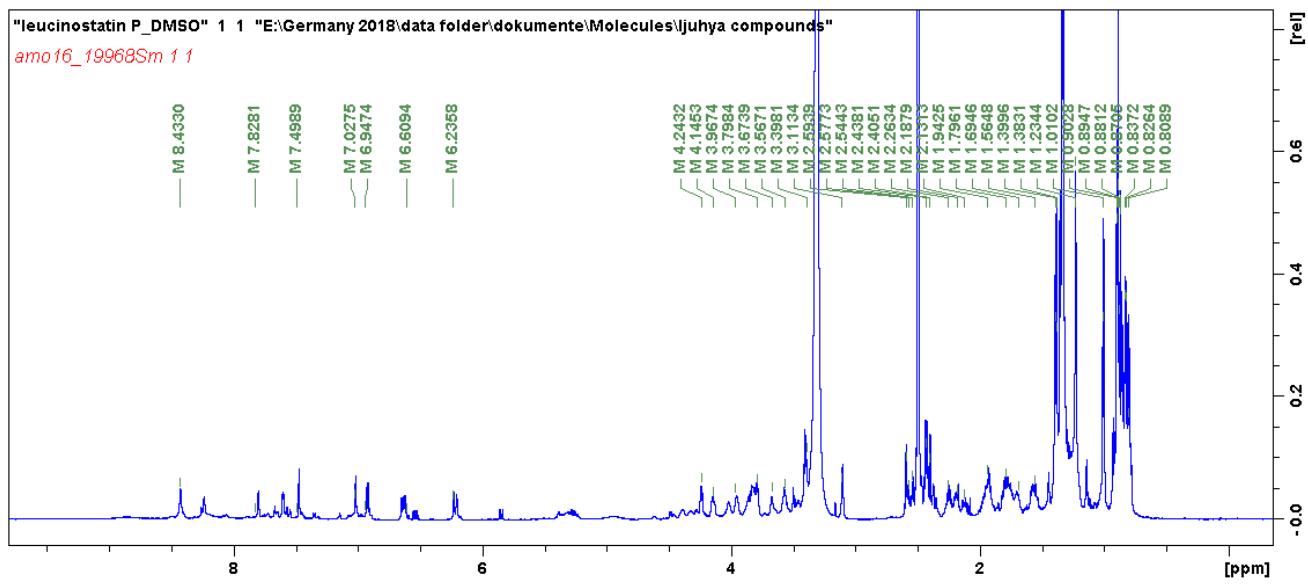


**S18 Figure:** HR-ESI-MS-MS fragmentation pattern of leucinostatin Q 2.

**S3 Table.**  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR data of leucinostatin P 3 and its characteristic chemical shift values.

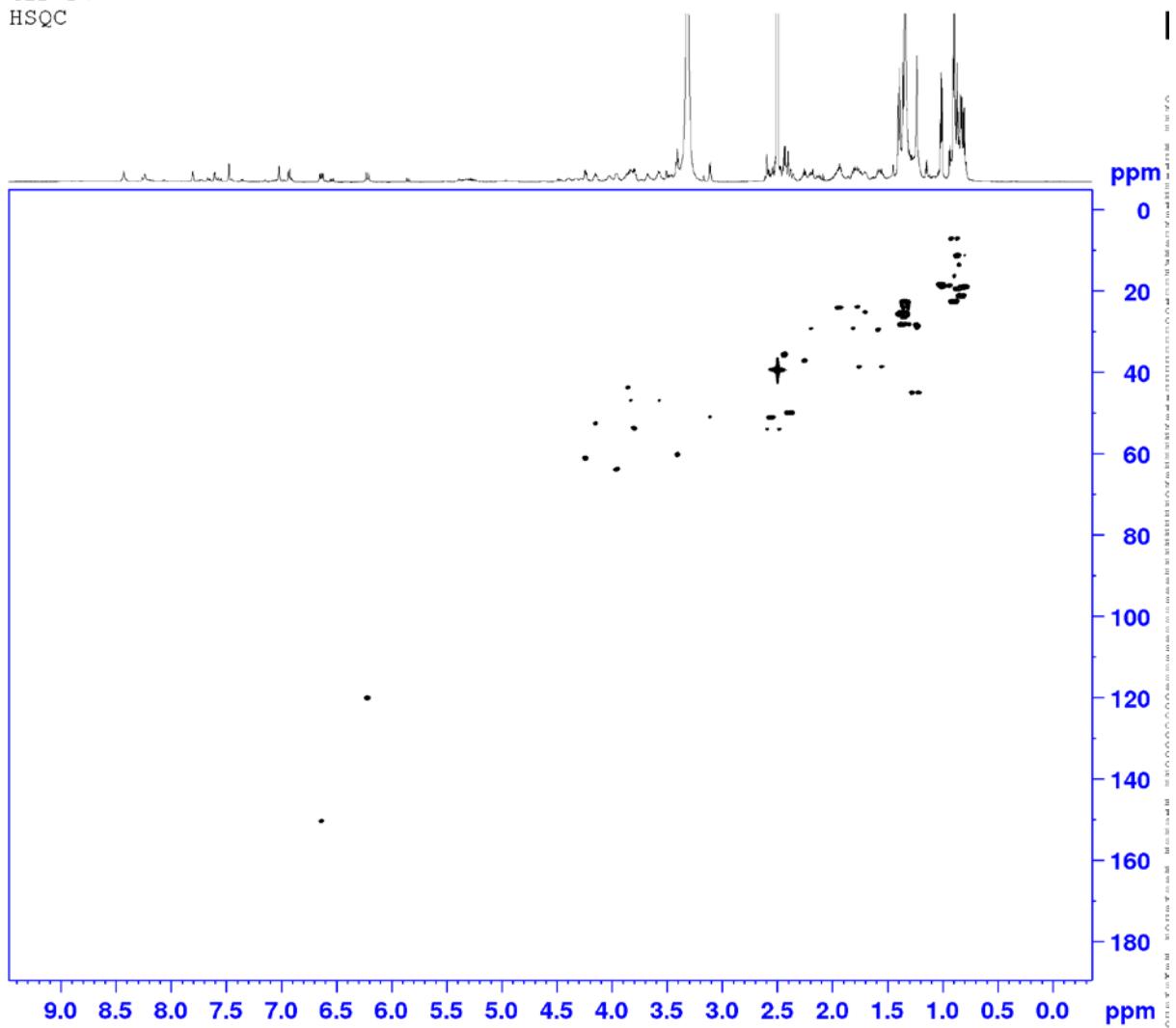
			$\delta\text{H}$ ( J in Hz)			$\delta\text{C}$ , type		$\delta\text{H}$ ( J in Hz)
residue	position			residue	position			
<b>ΔMHA</b>	<b>1</b>	165.28,C		<b>Aib2</b>	<b>1</b>	173.23,C		
	<b>2</b>	120.19,CH	6.22		<b>2</b>	55.8,C		
	<b>3</b>	150.59,CH	6.64		<b>3</b>	25.18,CH3	1.39	
	<b>4</b>	37.17,CH	2.25		<b>4</b>	23.06,CH3	1.33	
	<b>5</b>	28.22,CH2	1.36		<b>NH</b>			7.47
	<b>6</b>	11.29,CH3	0.87		<b>Leu</b>	<b>1</b>	173.78,C	
	<b>7</b>	18.62,CH3	1.01		<b>2</b>	53.88,CH	3.79	
<b>Pro</b>	<b>1</b>	173.16,C			<b>3</b>	38.74,CH2	1.55	
	<b>2</b>	61.28,CH	4.24					1.75
	<b>3</b>	29.19,CH2	1.81		<b>4</b>	23.79,CH	1.77	
			2.19		<b>5</b>	21.27,CH3	0.85	
	<b>4</b>	24.10,CH2	1.95		<b>6</b>	22.58,CH3	0.91	
	<b>5</b>	46.90,CH2	3.82		<b>NH</b>			7.02
			3.56		<b>Aib3</b>	<b>1</b>	175.23,C	
<b>AHMOD</b>	<b>1</b>	173.63,C			<b>2</b>	24.30,CH3	1.34	
	<b>2</b>	52.65,CH	4.15		<b>3</b>	25.01,CH3	1.33	
	<b>3</b>				<b>4</b>	56.5,C		
					<b>NH</b>			7.6
	<b>4</b>	25.11,CH	1.7	<b>Aib4</b>	<b>1</b>	171.4,C		
	<b>5</b>	44.90,CH2	1.21		<b>2</b>	55.7,C		
			1.28		<b>3</b>	23.85,CH3	1.35	
	<b>6</b>	64.02,CH	3.96		<b>4</b>	24.10,CH3	1.32	
	<b>7</b>	51.12,CH2	2.57		<b>NH</b>			7.6
					<b>C-terminal</b>	<b>1</b>	43.62,CH	3.85
			2.54					

	<b>8</b>	209.95,C		<b>2</b>	54.13,CH2	2.5
	<b>9</b>	35.66,CH2	2.43			2.48
	<b>10</b>	7.20,CH3	0.9	<b>3</b>	51.12,CH2	2.57
	<b>11</b>	22.58,CH3	0.91			2.54
	<b>NH</b>		6.93	<b>4</b>	60.18,CH2	3.4
<b>Hyleu</b>	<b>1</b>	172.16,C		<b>NH</b>		8.25
	<b>2</b>	63.85,CH	4.01			
	<b>3</b>	73.88,CH	3.67			
	<b>4</b>	29.57,CH	1.58			
	<b>5</b>	13.50,CH3	0.85			
	<b>6</b>	18.98,CH3	0.81			
	<b>NH</b>		7.8			
<b>Aib1</b>	<b>1</b>	174.68,C				
	<b>2</b>	56.03,C				
	<b>3</b>	23.57,CH3	1.33			
	<b>4</b>	26.01,CH3	1.36			
	<b>NH</b>					
			8.43			



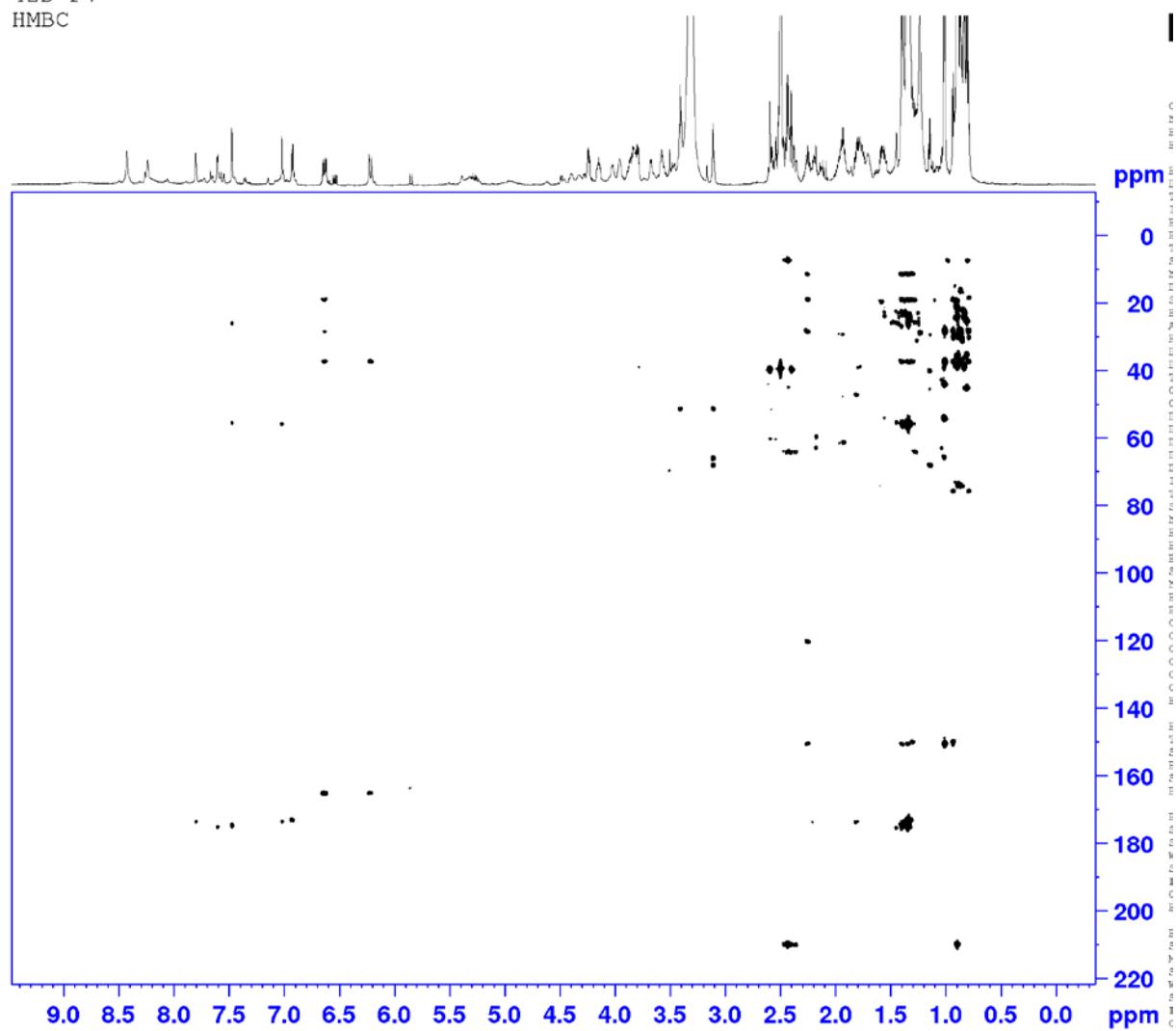
**S19 figure:**  $^1\text{H}$ NMR spectra of leucinostatin P 3

42D-F7  
HSQC



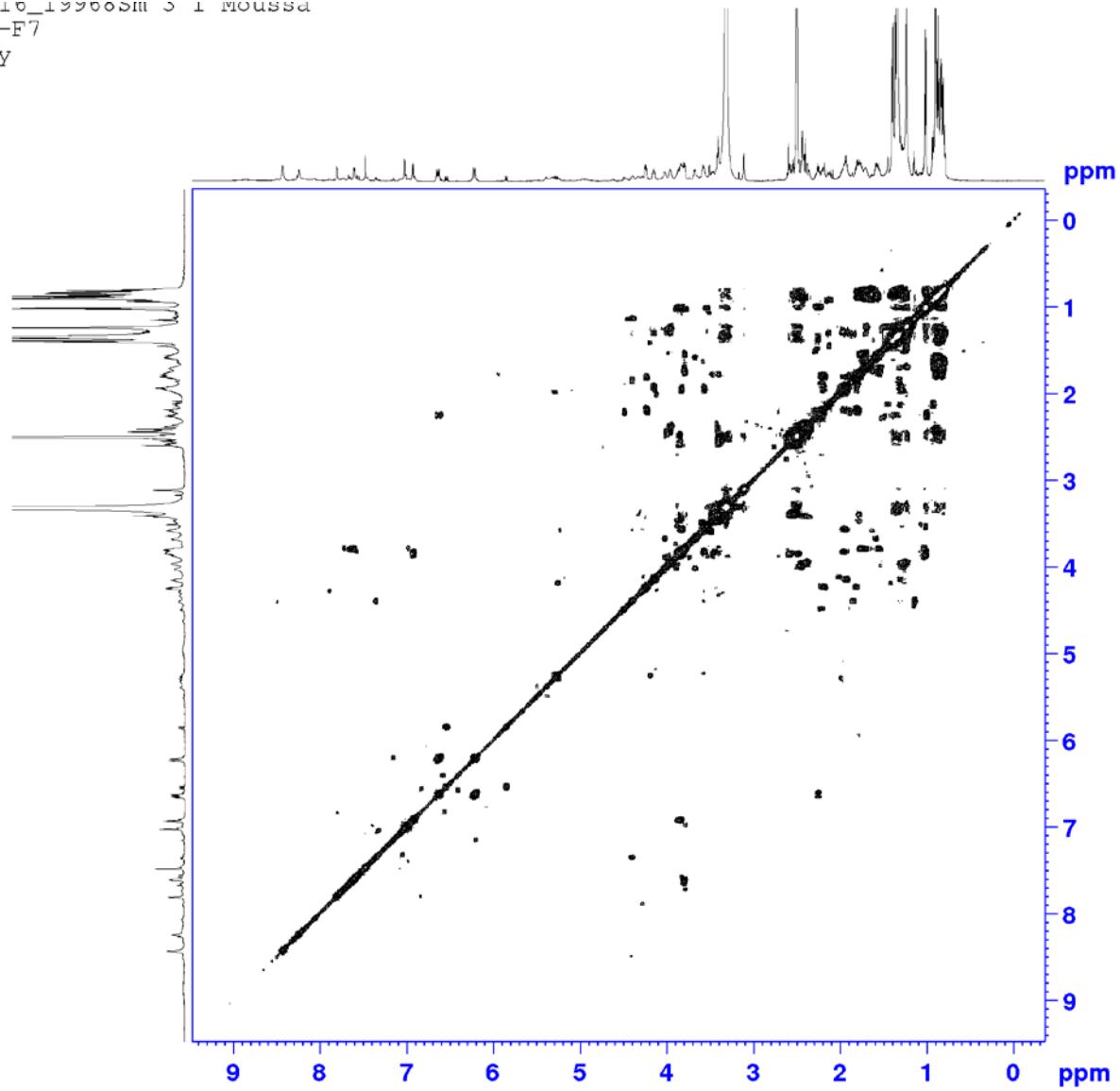
S20 Figure: HSQC spectra of leucinostatin P 3

42D-F7  
HMBC



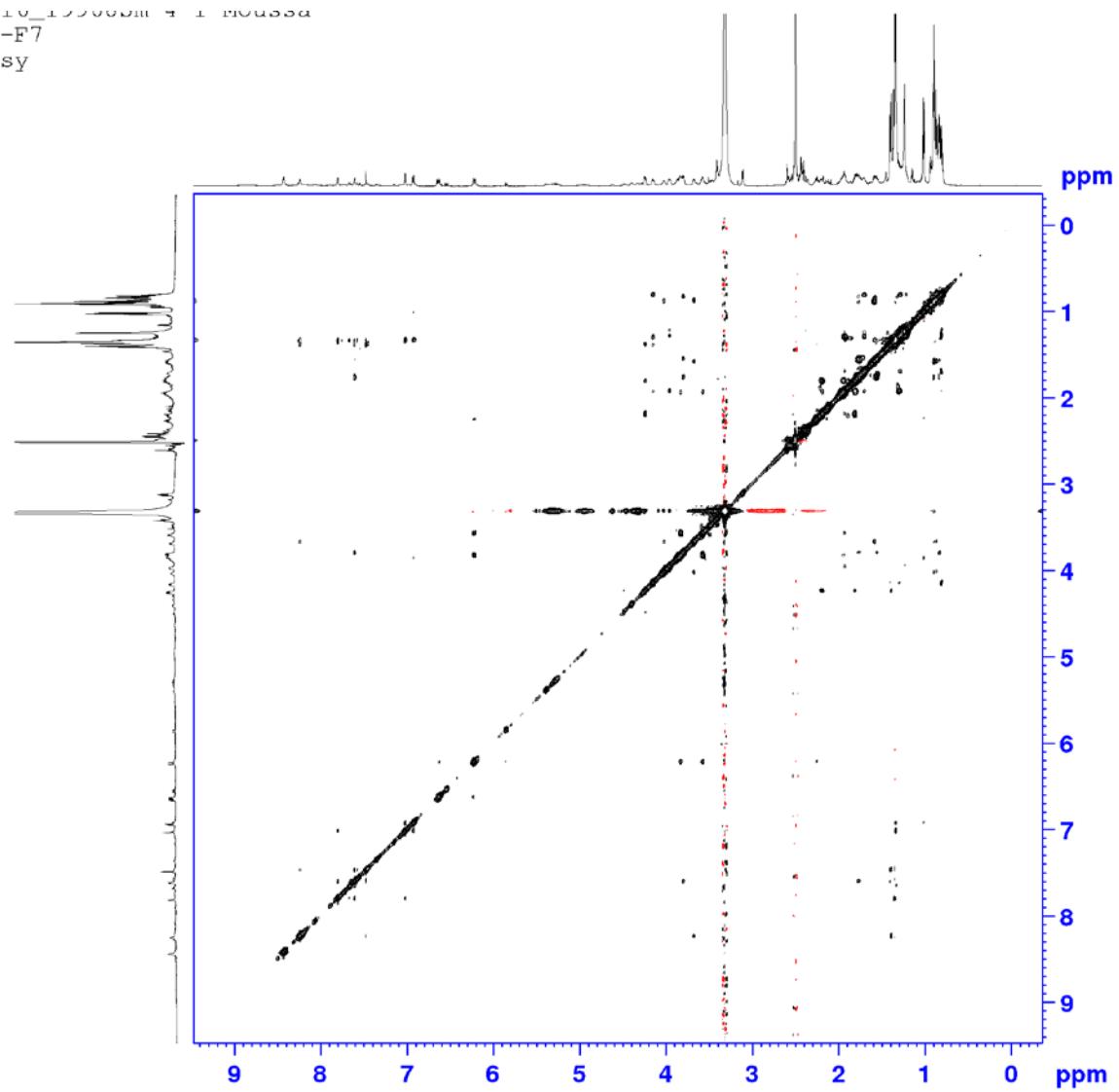
S21 Figure: HMBC spectra of leucinostatin P 3

42D-F7  
Cosy

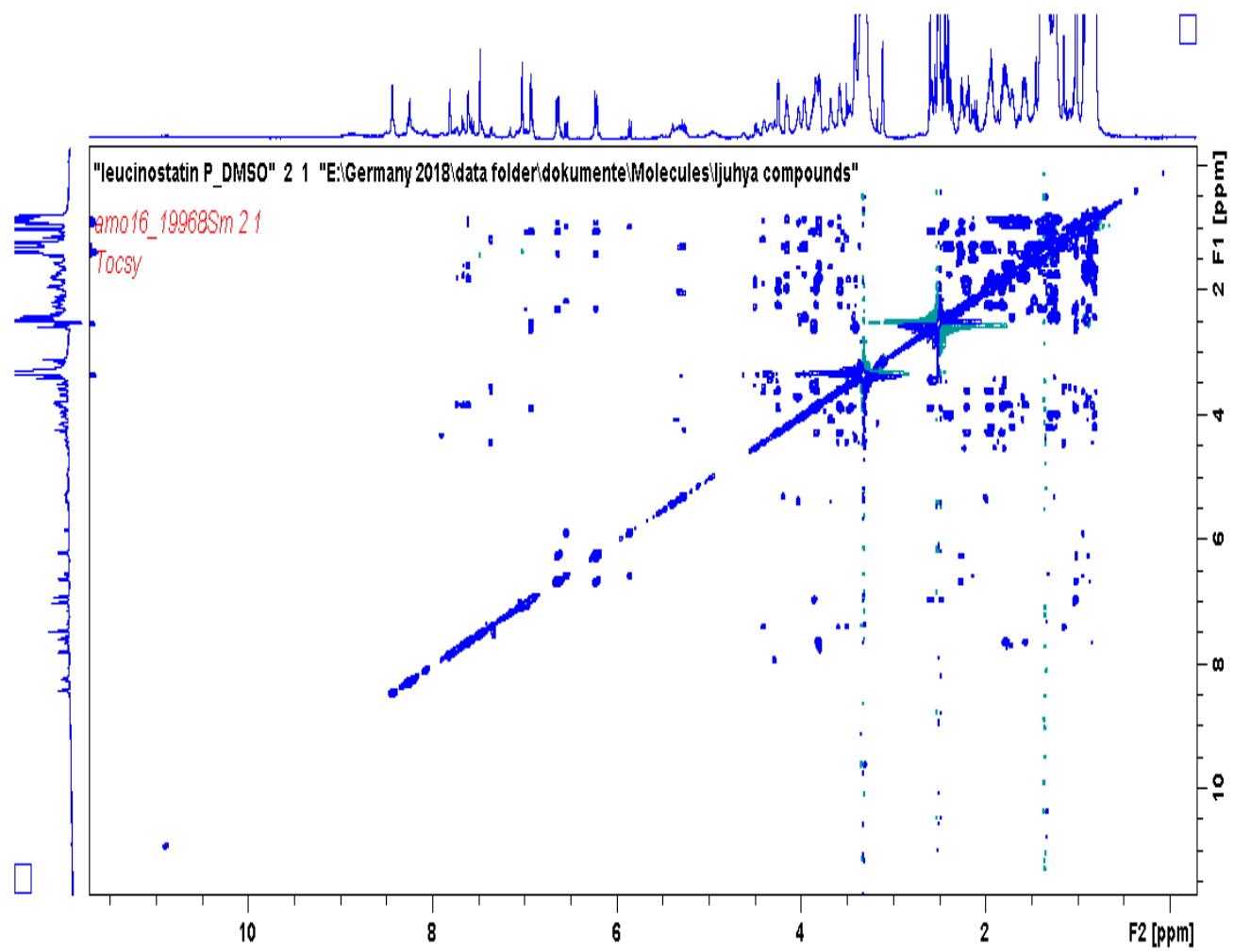


S22 Figure: COSY spectra of leucinostatin P 3.

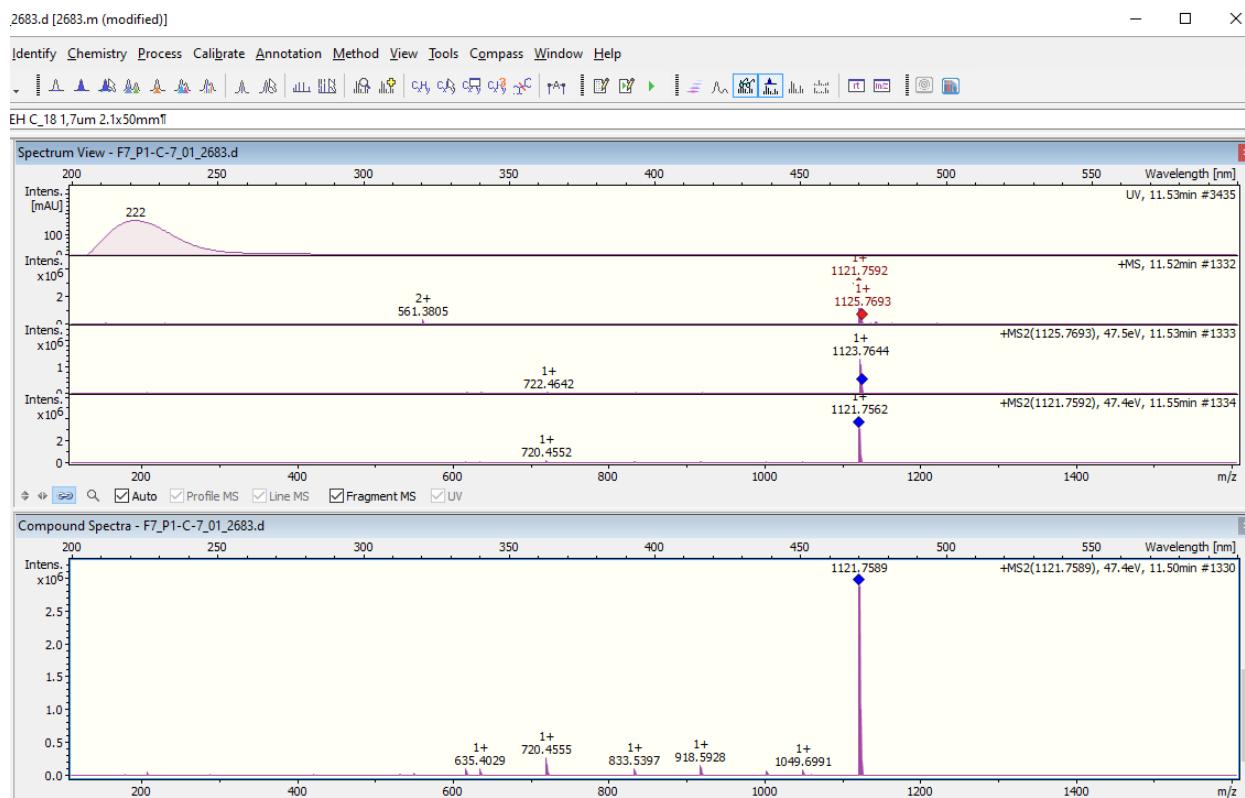
42D-F7  
Noesy



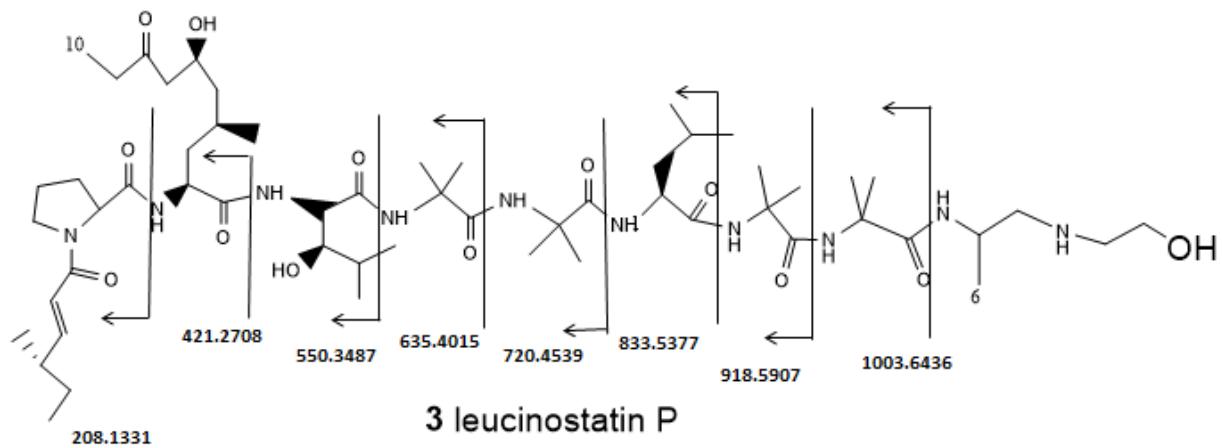
S23 Figure: NOESY spectra of leucinostatin P 3



S24 Figure: TOCSY spectra of leucinostatin P 3.



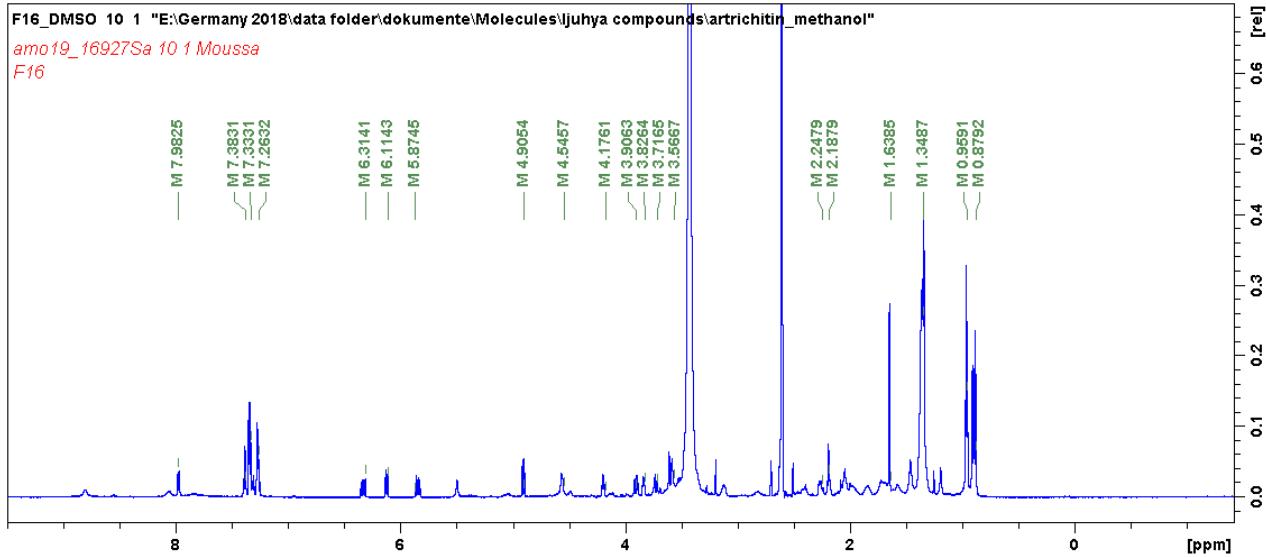
**S25 Figure:** HR-ESI-MSMS fragmentation pattern of leucinostatin P 3.

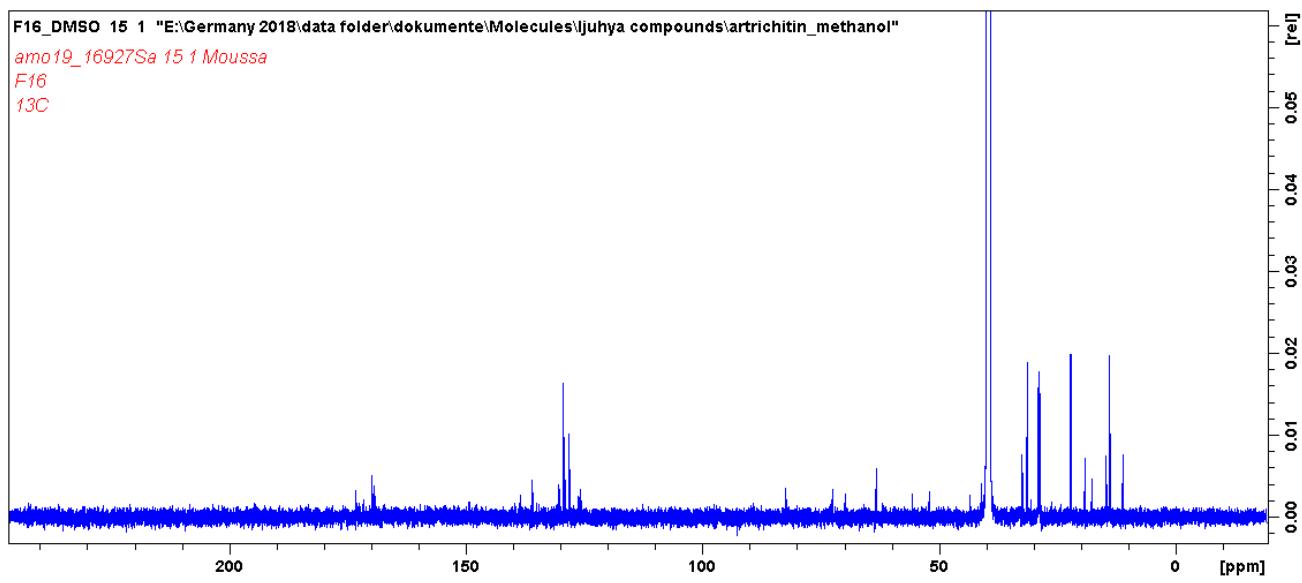


**S26 Figure:** HR-ESI-MS-MS fragments of leucinostatin P 3.

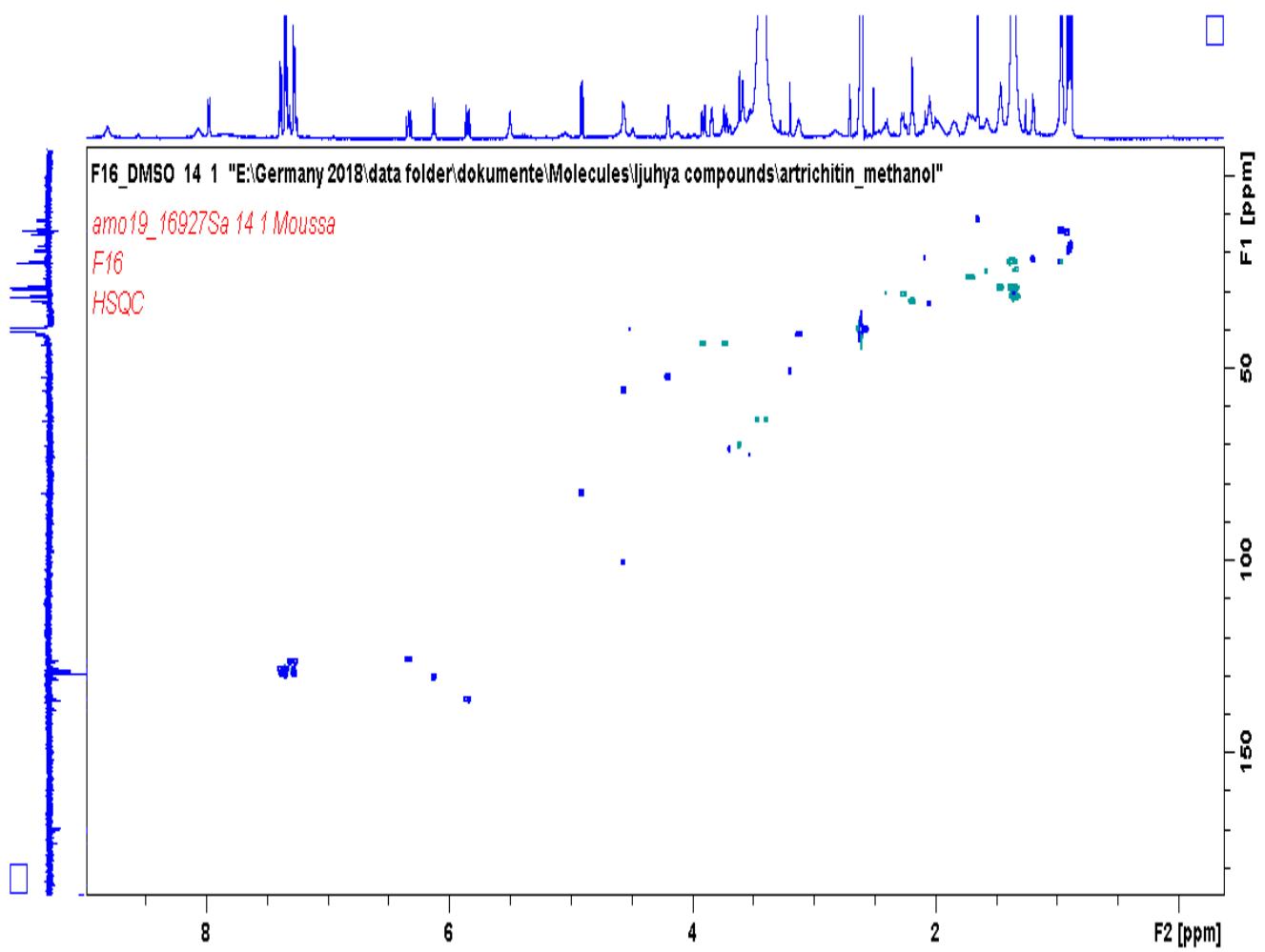
**S4 Table.**  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR data of arthrichitin D 4 and its characteristic chemical shift values

		Arthrichitin D			
residue	position	$\delta\text{C}$ , type	$\delta\text{H}$ ( J in Hz)	$\delta\text{H}$ ( J in Hz)	$\delta\text{C}$ , type
L-Glu	1	169.92,C		Dimethyl-oxy-hexadecanoyl	
	2	52.50,CH	4.19	27	173.19,C
	3	25.20,CH2	1.82	28	40.70,CH
			2.04	29	82.30,CH
	4	30.50,CH2	2.25	30	130.20,C
			2.4	31	129.9,CH
	5	173.02,C		32	125.70,CH
L-Gly	NH		7.35	33	136.50,CH
	6	169.30,C	4.05	34	32.10,CH2
	7	43.40,CH2	3.73	35	31.50,CH2
			3.9	36	28.9,CH2
	NH		8.06	37	24.40,CH2
L-Phe				38-	
	8	171.50,C		40	22.10,CH2
	9	55.01,CH	4.13	42	14.74,CH3
	10	36.60,CH2	3.43	43	14.04,CH3
				44	11.25,CH3
	11	138.55,C			
	12	129.40,CH	7.34		
	13	128.34,CH	7.34		
	14	126.25,CH	7.26		
	15	128.22,CH	7.38		
	16	129.30,CH	7.27		
L-Gln	NH		8.81		
	17	171.60,C			
	18	56.60,CH	3.83		
	19	26.00,CH2	1.71		
	20	32.37,CH2	2.18		
	21	170.13,C			
L-Val	NH		7.97		
	22	170.0,C			
	23	55.6,CH	4.56		
	24	33.27,CH	2.04		
	25	19.17,CH3	0.89		
	26	17.79,CH3	0.88		
	NH		7.98		



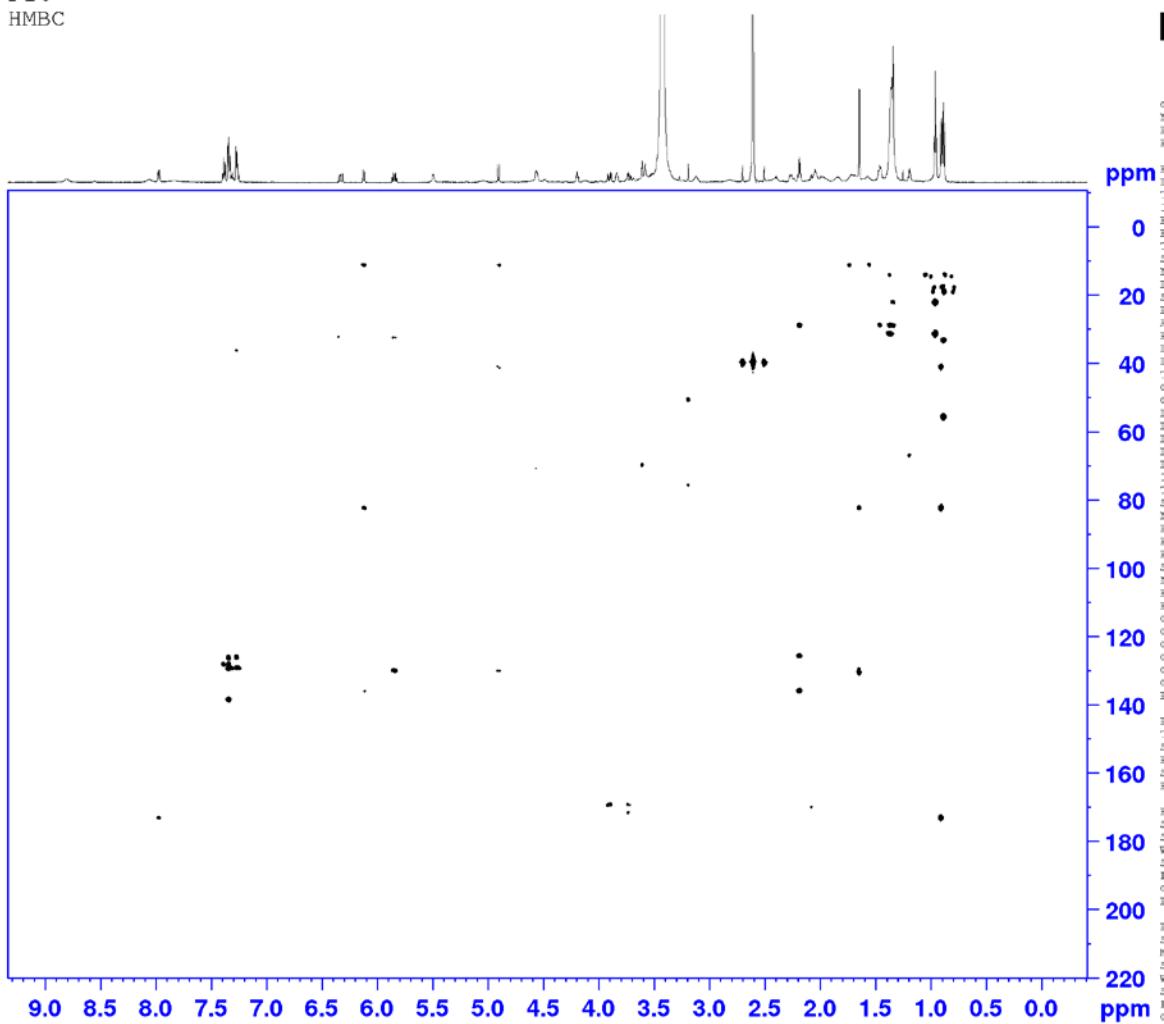


S28 Figure: <sup>13</sup>CNMR spectra of arthrichitin D 4

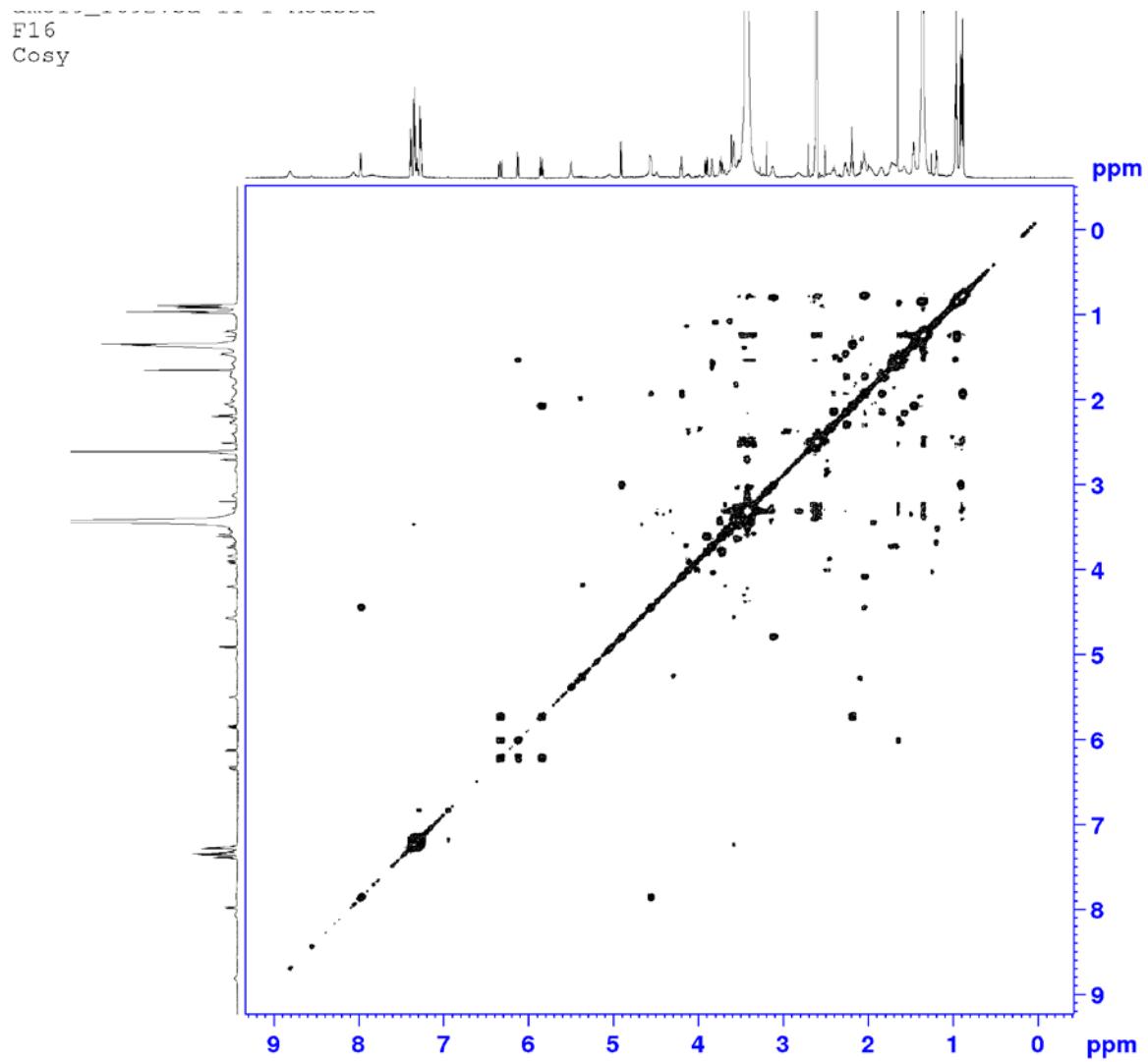


S29 Figure: HSQC spectra of arthrichitin D 4

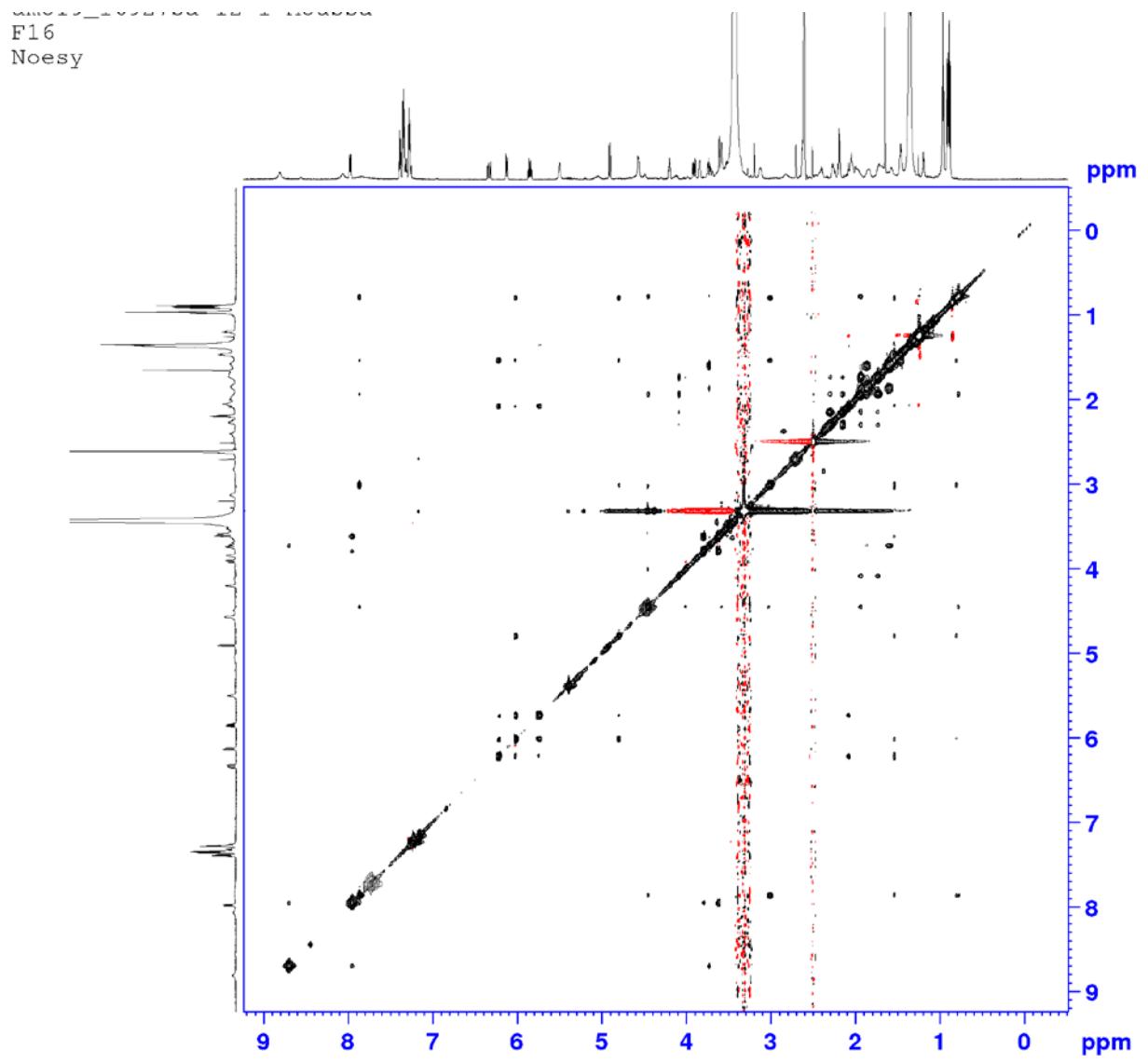
F16  
HMBC



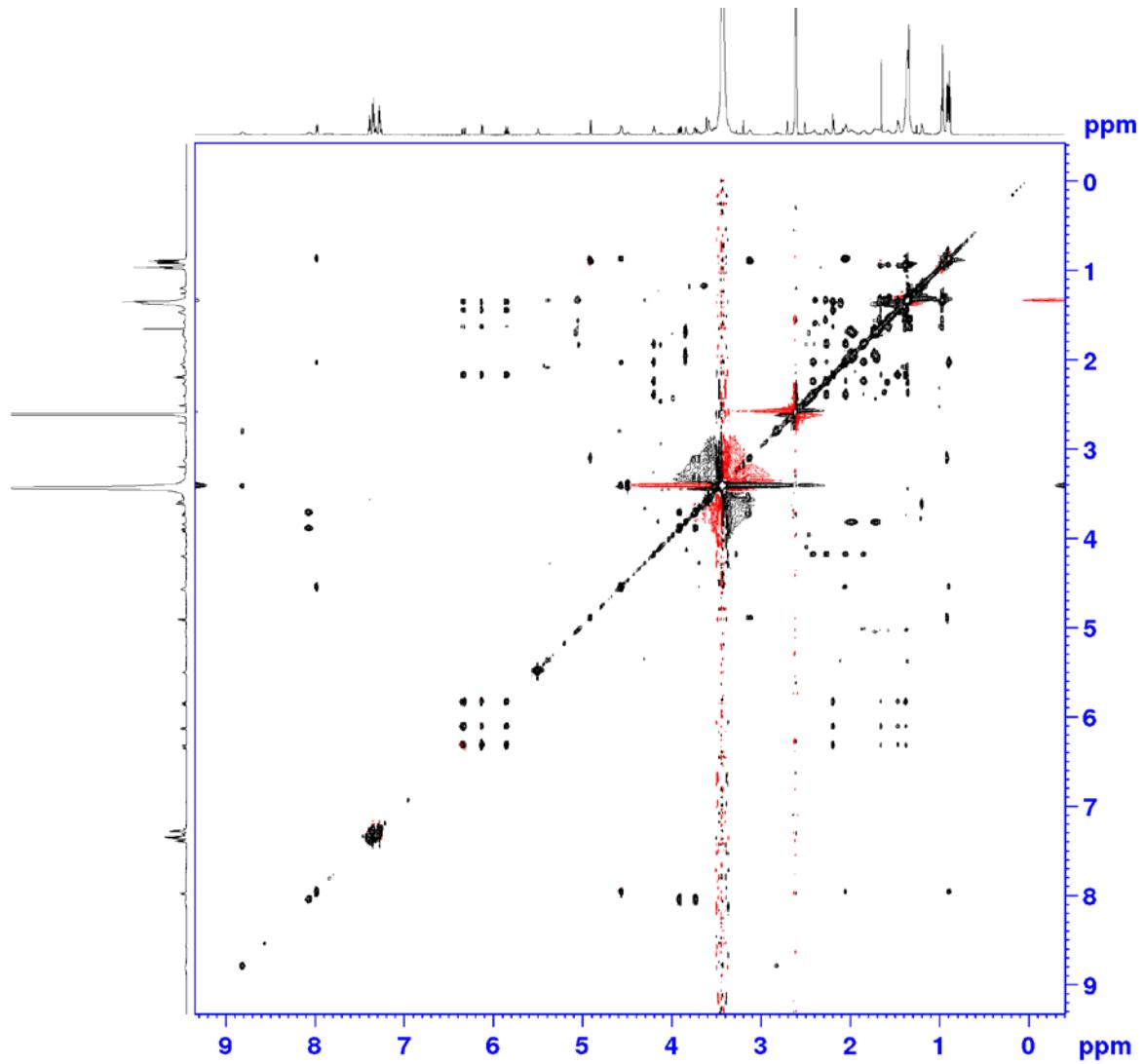
S30 Figure: HMBC spectra of arthrichitin D 4..



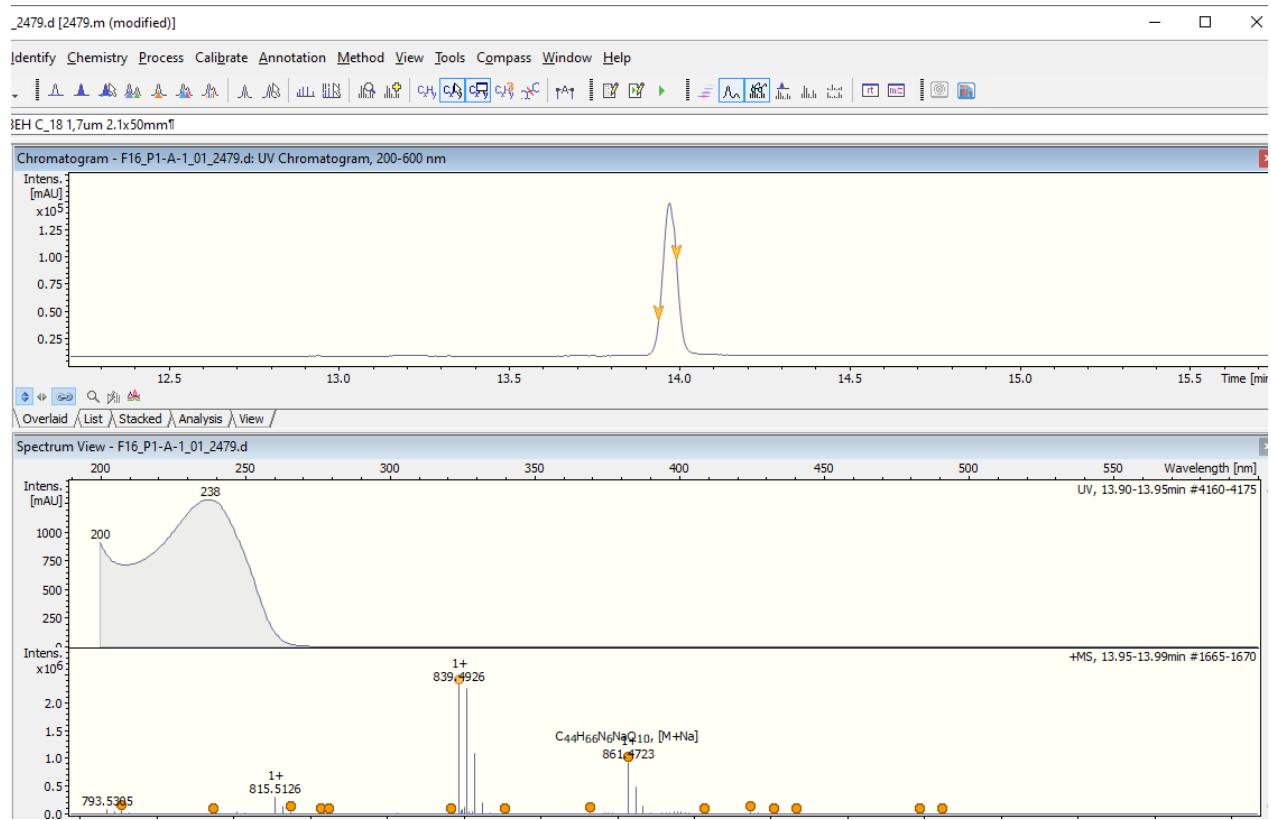
S31 Figure: COSY spectra of arthrichitin D 4



S32 Figure: NOESY spectra of arthrichitin D 4



S33 Figure: ROESY spectra of arthrichitin D 4



**S34. Figure: HR-ESI-MS of arthrichitin D 4.**

**S12 Text:**  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR data of **chaetoglobosin B 5** and its characteristic chemical shift values.

### Chaetoglobosin B 5

Yellow powder, UV absorption  $\lambda_{\max}$  200, 222, 281 nm , HR-ESI-MS m/z 1057.5321 [2M+H] $^+$  (calcd for C<sub>32</sub>H<sub>36</sub>N<sub>2</sub>O<sub>5</sub>);  $^1\text{H}$  NMR (700 MHz, methanol-d4)  $\delta$ H 3.53 (H-3, dd, J=8.4, 5.2 Hz), 3.84 (H-7, d, J= 9.4 Hz), 2.07 (H-8, dd, J= 9.4, 12.3 Hz), 2.87 (H-10a, dd, J=5.3, 14.5 Hz), 2.57 (H-10b, dd, J=5.3, 14.5 Hz), 1.28 (H-11, s), 1.65 (H-12, s), 6.05 (H-13, dd, J=9.4, 15.9 Hz), 5.15 (H-14, ddd, J=3.5, 10.2, 15.5 Hz), 2.32(H-15a, m), 2.01 (H-15b,m), 2.58 (H-16, m), 5.57(H-17, d, J=9.6 Hz), 5.11 (H-19,s), 6.68 (H-21, d, J=16.2 Hz), 7.46 (H-22,d J=5.6 Hz), 7.02 (H-2', brs), 7.48 (H-4', m), 7.01 (H-5', t, J=7.1 Hz), 7.07(H-6', t, J= 7.3 Hz), 7.32 (H-7', d, J= 8.3Hz ), 1.01(Me-16, d, J=6.8 Hz), 1.42 (Me-18, s).

**S13 Text:**  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR data of **19-O-acetyl chaetoglobosin B (6)** and its characteristic chemical shift values.

### 19-O-acetyl chaetoglobosin B 6

Yellow powder, UV absorption  $\lambda_{\max}$  199, 223, 280 nm, HR-ESI-MS m/z 571.2801 [M+H] $^+$ , HR-ESI-MS m/z (calcd for C<sub>34</sub>H<sub>38</sub>N<sub>2</sub>O<sub>6</sub>);  $^1\text{H}$  NMR (700 MHz, methanol-d4)  $\delta$ H 3.60 (H-3, dd, J=3.2,8.5 Hz), 3.26 (H-4, m), 3.89 (H-7, d, J= 9.4 Hz), 2.10 (H-8, m), 2.89 (H-10a,dd, J=14.3, 4.3 Hz), 2.60 (H-10b,dd, J=14.3, 4.3 Hz), 1.32 (H-11, s), 1.65 (H-12, s), 6.17 (H-13, dd, J=15.3, 9.5 Hz), 5.19 (H-14, ddd, J= 3.5, 9.5, 15.4 Hz), 2.01 (H-15a, m), 2.32 (H-15b, m), 2.61 (H-16, m), 5.77 (H-17, d, J=9.4 Hz), 5.92 (H-19, s), 6.80 (H-21, m), 7.66 (H-22, m), 7.15 (H-2', brs), 7.53 (H-4', d, J=7.9 Hz), 7.01 (H-5', 7.09 (H-6', t), 7.38 (H-7', d, J= 8.15 Hz), 2.11 ( 19-O-acetyl); $^{13}\text{C}$  NMR (175 MHz, methanol-d4)  $\delta$ H $^{13}\text{C}$  59.0 (C-3), 48.4 (C-4), 127.1 (C-5), 134.0 (C-6), 69.2 (C-7), 52.3 ( C-8), 62.2 (C-9), 33.2 (C-10), 17.7 (C-11), 14.6 (C-12), 129.1 (C-13), 135.5 (C-14), 41.9 (C-15), 33.0 (C-16), 143.3 (C-17), 127.9 (C-18), 84.2 (C-19), 194.9 (C-20), 135.0 (C-21), 136.8 (C-22), 199.7 (C-23), 172.5 ( C-24), 124.5 (C-2'), 111.7 (C-3'), 119.2(C-4'), 119.6 (C-5'), 122.3 (C-6'), 112.3 (C-7'), 169.4 ( 19-O-acetyl C=O), 20.2 ( 19-O-acetyl -Me), 137.1 (C-1a'), 128.0 (C-3a').