

Article

Novel Lipidized Derivatives of the Bioflavonoid Hesperidin: Dermatological, Cosmetic and Chemopreventive Applications

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Abstract: Hesperidin is one of the most important natural flavonoids, known for its antioxidant, anti-inflammatory, anti-mutagenic, and anti-hypertensive properties. Despite its various biological activities, hesperidin is rarely used in the dermo-cosmetic field because of its poor solubility in both water and oil phases that makes difficult formulation, distribution and bioavailability through the skin layers. Moreover, hesperidin is still underestimated in skin care products, and literature data on its stability into a topical formulation are not yet available. In this paper we report the synthesis of five different derivatives of hesperidin and their evaluation in terms of antioxidant, antifungal, antiproliferative, and apoptotic effects on human leukemic K562 cells. Preliminary antiproliferative effects were considered since hyper-proliferation is involved in several cutaneous problems particularly in the case of photo-exposition and environmental pollution. **Esp4** and **Esp5** were found to be more active in inhibiting K562 cell growth than parent hesperidin. **Esp3** exhibited different biological properties, i.e., antioxidant activity in the absence of antiproliferative effects.

Keywords: hesperidin; antioxidant; pro-apoptotic; antifungal; dermo-cosmetic

1. Introduction

Among phenolic compounds, flavonoids represent one of the most important classes [1]. These molecules are commonly found in different plant materials such as fruit, herbs, vegetables, and cereals [2]. Flavonoids display many different biological activities. Antioxidant, photo-protective, photo-aging, anti-inflammatory, metal-chelating, anti-allergic, and anti-bacterial are the most important flavonoid-mediated activities on the skin [3–6]. Hesperidin is a flavanone glycoside abundant in citrus fruit peel [7], it is formed by the aglycone also called “hesperetin” to which a 6-*O*- α -L-rhamnosyl-D-glucose in position seven is bound. Since its discovery in 1872 by Lebreton, hesperidin became the object of continuous research focusing on both extraction methods and biological/biomedical activities [8].

Hesperidin has many biological activities, displaying antioxidant, prostaglandin-synthesis inhibitory, anti-inflammatory, and anti-mutagenic effects [9–12]. The recent interest in the use of flavonoids as anti-cancer agents has highlighted the effects of hesperidin in prevention and treatment of various human diseases [13–15]. Among the many beneficial effects on health, it has been reported that hesperidin is able to inhibit skin tumorigenesis [16].

With respect to structure activity relationships, the main biological activities of hesperidin, such as the antioxidant power, are closely associated with the chemical structure of the aglycone [17].

Although appealing for biomedical applications, flavonoids have general problems in terms of bioavailability, stability, and formulation [18]. Literature data about the pharmacokinetics of dietary flavonoids are widely discussed, but studies about the skin absorption of all these natural molecules are not yet available [19]. Glycosylated flavonoids are subjected to fibroblast glucosidases activity, leading to the release of pharmacologically active aglycone moiety [20]. Moreover, glycosylated drugs have low cellular toxicity as compared to their aglycones [21].

Skin absorption is a multifactorial and complex process considering that skin is the first barrier against biological toxins, and is designed to reduce water loss [22]. A topical effective molecule must cross the outer skin layers (first the stratum corneum and then the epidermis or using alternative routes such as glands and interstitial passages) and subsequently might stay within the papillary dermis to have its local effect. Conversely, achieving a systemic circulation by a topical molecule is considered a side effect [23]. The Log P value of the molecule is one of the most important factors to allow skin absorption. The Log P value indicates the rough estimate capacity to permeate directly across skin layers. Since the stratum corneum is constituted up of ceramides, cholesterol, and fatty acids, an hydrophilic molecule cannot efficiently penetrate the skin [24,25]. Hesperidin and many natural flavonoids do not have enough lipophilic molecules to cross the skin [26]. On the other hand, hesperidin is also poorly soluble in water [27]. These solubility problems reduce its skin topical use due to the difficulty in formulation, and consequently poor availability for skin [28,29]. Therefore, despite its various biological activity, hesperidin is still underestimated in skin care products, and literature data on its stability into a topical formulation are not yet available.

In the present study, continuing our investigation on phenylpropanoid structures [30–32], we synthesized lipophilic ester derivatives of hesperidin in order to improve its stability in topical formulations and in finished cosmetic products.

Moreover, following our recent approach, aimed to investigate newly synthesized molecules, possibly endowed with multifunctional properties, preliminary antioxidant, antimicrobial, anti-proliferative, and apoptosis induction tests were performed because all of this might allow a preventive role for such a class of molecules, that might be useful for early ageing and oxidative damage prevention, especially when considering daily cosmetic use.

2. Materials and Methods

2.1. General

All reagents and solvents were supplied by Sigma-Aldrich, Milan, Italy and CARLO ERBA Reagents, Milan, Italy. ^1H NMR spectra were recorded on VXR-200 Varian spectrometer and Mercury Plus-400 in d_6 -DMSO. Chemical shift values are expressed in ppm using tetramethylsilane as the reference standard (TMS). UV spectra were recorded on a UV-VIS spectrophotometer (Shimadzu UV-2600, Shimadzu Italia, Milano, Italia). HPLC analysis was performed using an Agilent 1100 Series HPLC System equipped with a G1315A DAD, autosampler and with a Phenomenex Synergi Hydro-RP C18 80 Å column (4.6×150 mm, $4 \mu\text{m}$).

2.2. Chemistry

2.2.1. Synthesis of Epta-Tert-Butyl Hesperidin (Esp1)

To a solution of hesperidin (0.64 mmol) and DMAP (23.04 mmol) in CH_2Cl_2 (40 mL), tert-butyl-acetyl chloride (17.28 mmol) was added. The reaction mixture was stirred for 14 h at room temperature and then washed with H_2O , NaHCO_3 and Brine. The combined organic phases were dried and evaporated under reduced pressure. The crude residue was purified by column chromatography (ethyl acetate) to obtain Esp1 as orange oil (yield 65.25%). ^1H -NMR (DMSO) δ ppm: 0.96 to 1.03 (m, 66H); 2.06 to 2.12 (m, 12H); 2.43 (m, 2H); 2.50 (m, 2H); 2.70 (m, 1H); 3.30 (m, 2H); 3.59 to 3.61 (m, 2H); 3.80 (s, 3H); 4.30 (t, 1H); 5.10 (m, 4H); 5.43 (d, 1H); 5.50–5.60 (m, 1H); 5.80 (d, 1H); 6.25 to

6.65 (m, 2H); 7.15 to 7.25 (d, 2H), 7.40 (m, 1H); 12.00 (s, 1H). ESI-MS $[M + H]^+$: found for $C_{70}H_{104}O_{22}$, 1298.56. HPLC purity 99.2%.

2.2.2. Synthesis of Epta-Cyclohexylacetyl Hesperidin (Esp2)

To a solution of hesperidin (0.64 mmol) and DMAP (23.04 mmol) in CH_2Cl_2 (40 mL), cyclohexylacetyl chloride (17.28 mmol) was added. After stirring for 14 h at room temperature, the mixture was washed with water, $NaHCO_3$, and Brine and the organic phase was then dried and concentrated under reduced pressure. The residue was purified by column chromatography (ethyl acetate) to obtain Esp2 (final yield 79.28%) as orange oil. 1H -NMR (DMSO) δ ppm: 1.20 (d, 3H, CH_3); 1.35 to 1.50 (m, 70H); 2.06 to 2.12, (m, 19H); 2.35 (d, 2H); 3.43 (m, 2H); 3.50 (m, 2H); 3.79 (s, 3H); 4.37 to 4.50 (m, 2H); 5.30 (t, 2H); 5.51 to 5.62 (m, 2H); 5.82 (m, 1H); 5.90 (m, 3H); 6.23 to 6.65 (m, 2H); 7.12 (d, 1H), 7.25 (d, 1H), 7.40 (m, 2H); 12.00 (s, 1H). ESI-MS $[M + H]^+$: found for $C_{84}H_{118}O_{22}$, 1480.85. HPLC purity 99.6%.

2.2.3. Synthesis of Epta-Benzoyl Hesperidin (Esp3)

To a solution of hesperidin (0.64 mmol) and DMAP (23.04 mmol) in CH_2Cl_2 (40 mL), benzoyl chloride (17.28 mmol) was added. After stirring at room temperature for 14 h, the reaction mixture was washed with H_2O , $NaHCO_3$ and Brine. The organic phase was subsequently dried and evaporated under reduced pressure to give a crude residue purified by column chromatography (ethyl acetate). Esp3 was obtained by crystallization from methanol as a light yellow powder (final yield 56.87%). 1H -NMR (DMSO) δ ppm: 1.10 (t, 3H); 3.20 to 3.40 (m, 2H); 3.52 to 3.61 (m, 2H); 3.80 (s, 3H); 4.28 to 4.37 (m, 2H); 5.28 (t, 2H); 5.50 to 5.61 (m, 2H); 5.85 to 5.96 (m, 4H); 6.18 (t, 2H); 6.39 (d, 1H); 6.89 (d, 1H); 7.32 (m, 2H); 7.56 to 7.71 (m, 21H); 8.02 (m, 12H); 8.14 (8d, 2H); 12.00 (s, 1H, OH aromatico). ESI-MS $[M + H]^+$: found for $C_{77}H_{62}O_{22}$, 1340.30. HPLC purity 98.7%.

2.2.4. Synthesis of Epta-Naphtoyl Hesperidin (Esp4)

To a solution of hesperidin (0.64 mmol) and DMAP (23.04 mmol) in CH_2Cl_2 (40 mL), naphthoyl chloride (17.28 mmol) was added. The reaction was stirring for 14 h at room temperature and then washed with H_2O , $NaHCO_3$ and Brine. The combined organic phase was dried and evaporated under reduced pressure. The crude residue was purified by column chromatography (petroleum ether/ethyl acetate: 50/50) to obtain Esp4 was obtained as white powder (final yield 77.23%). 1H -NMR (DMSO) δ ppm: 1.14 (d, 3H); 2.70 (m, 1H); 3.30 (m, 2H); 3.59–3.61 (m, 1H); 3.81 (s, 3H); 4.29–4.35 (m, 2H); 5.30 (t, 2H); 5.39–5.42 (m, 2H); 5.89–6.02 (m, 4H); 6.18 (t, 2H); 6.39 (d, 1H); 6.91 (d, 1H); 7.15–7.25 (m, 2H); 7.37–7.52 (m, 35H); 8.01–8.12 (m, 14H); 12.00 (s, 1H). ESI-MS $[M + H]^+$: found for $C_{105}H_{76}O_{22}$, 1689.71. HPLC purity 99.2%.

2.2.5. Synthesis of Octa-Cinnamoyl Hesperidin (Esp5)

To a solution of hesperidin (0.64 mmol) and DMAP (23.04 mmol) in CH_2Cl_2 (40 mL), cinnamoyl chloride (17.28 mmol) was added. The reaction was stirring for 14 h at room temperature and then washed with H_2O , $NaHCO_3$ and Brine. The combined organic phase was dried and evaporated under reduced pressure. The crude residue was purified by column chromatography (petroleum ether/ethyl acetate: 52/48). Esp5 was obtained as light yellow powder (final yield 74.07%). 1H -NMR (DMSO) δ ppm: 1.20 (t, 3H); 3.10 (m, 1H); 3.35(m, 2H); 3.58 (m, 1H); 3.82 (s, 3H); 4.28 to 4.36 (m, 2H); 5.32 (t, 2H); 5.48 to 5.52 (m, 2H); 5.86 to 6.01 (m, 4H); 6.28 to 6.32 (d, 8H); 6.41 to 6.46 (d, 2H); 6.78 to 6.82 (d, 1H); 7.02 (m, 2H); 7.18 (d, 1H); 7.50 to 7.56 (m, 16H); 7.61 to 7.64 (m, 8H); 7.70 to 7.78 (d, 8H); 7.95 to 8.02 (m, 16H). ESI-MS $[M + H]^+$: found for $C_{100}H_{82}O_{23}$, 1652.70. HPLC purity 98.8%.

2.3. Determination of LogP

The determination of the partition ratio (LogP), was calculated as the logarithm of the concentration of substance that was in octanol and water. A test was carried out in a mixture of octanol and water (pH 12) in 1:1 ratio at room temperature. The biphasic system was stirred for 30 min and then centrifuged for 10 min (4000 rpm). An exact volume of each phase was analyzed with a spectrophotometer (λ_{max} fixed for each single compound). LogP is the result of the following expression:

$$\text{LogP} = \log_{10} [\text{mg}/20 \text{ mL}]_{\text{octanol}} / [\text{mg}/20 \text{ mL}]_{\text{H}_2\text{O}}$$

2.4. Antioxidant Assays

2.4.1. PCL Test (Photochemiluminescence)

PCL assay was performed using the Popov and Lewin method [33]. This test permitted us to measure the antioxidant capacity of a compound with a Photochem[®] apparatus (Analytik Jena, Leipzig, Germany) against superoxide anion radicals generated from Luminol, a photo-sensitizer, when exposed to UV light (Double Bore[®] phosphor lamp, output 351 nm, 3 m Watt/cm²). The antioxidant activity of a liposoluble substance could be observed using the ACL (Antioxidant Capacity of Liposoluble substance) kit provided by the manufacturer [34]. Standard Trolox solution and Luminol reagent were prepared according to the protocol. An antioxidant molecule generated a kinetic light emission curve, monitored for 180 s and expressed as micromoles of Trolox per gram of compound. The areas under the curves were calculated using the PCLsoft control and analysis software.

2.4.2. DPPH Test (Diphenylpicrylhydrazyl)

DPPH assay was carried out using method proposed by Wang et al. [35]. At 0.75 mL of each molecule tested, 1.5 mL of DPPH methanolic solution was added. Each sample was tested in absorbance using a spectrophotometer UV-VIS at 517 nm. The radical-scavenging activity was expressed as inhibition ratio of initial concentration of DPPH radical. For the best interpretation of the results, each compound was tested at a concentration able to cause 50% of the radical inhibition. DPPH values were expressed as μmol equivalents of Trolox for each gram of product.

2.4.3. FRAP Assay (Ferric Reducing Antioxidant Power)

FRAP assay was made according to the modified protocol described by Guihua et al. [36]. Each sample was dissolved with an appropriate solvent (water and/or methanol) to evaluate its ability in reducing ferric. Three solution were freshly prepared and then mixed to obtain the reagent for analysis: Acetate buffer pH 3.6, 2,4,6-tripyridyl-s-triazine (TPTZ) in HCl, and ferric chloride 20 mM, in the ratio 10:1:1, respectively. 0,11 mL of each sample solution or solvent (blank) was added to 1,9 mL of the reagent. The standard used for the calibration curve was Trolox. The absorbance was measured at 593 nm after 10 min of incubation in the dark at 37 °C.

2.5. Antifungal Activity

2.5.1. Microorganisms

The dermatophytes used were *Arthroderma cajetani* Ajello, CBS 495.70 strain; *Epidermophyton floccosum* (Hartz) Langerone Milochevitch, CBS 358.93 strain; *Trichophyton violaceum* Malmsten, CBS 459.61 strain; *Trichophyton tonsurans* Malmsten, CBS 483.76 strain, *Trichophyton mentagrophytes* (Robin) Blanchard, CBS 160.66 strain; *Microsporium canis* Bodin CBS 4727 strain; *Nannizzia gypsea* (Bodin) Guiart et Grigoraki CBS 286.63 strain purchased by the Centraal Bureau voor Schimmelcultures (CBS), Baarn, Netherlands; *Trichophyton rubrum* (Castellani) Sabouraud IHME 4321; *Microsporium gypseum* (Bodin) Guiarte Grigorakis IHME 3999 from Institute of Hygiene and Epidemiology-Mycolgy (IHME,

Brussels, Belgium). Cultures were maintained at 4 °C as agar slants, on Sabouraud Dextrose Agar (SDA, Sigma-Aldrich SRL, Milan, Italy).

2.5.2. Antifungal Inhibition Growth Test

Antifungal activity was determined as follows. Each test substance was dissolved in dimethylsulfoxide (DMSO) and aseptically mixed with sterile medium (SDA) at 45 °C in order to obtain two different concentrations, 20 and 100 µg/mL. The DMSO concentration in the final solution was adjusted to 0.1%. The same concentration of DMSO (0.1% *v/v*) was used to prepare the controls. For experiments, cultures were obtained by transplanting mycelium disks (10 mm diameters) from a single mother culture in the stationary phase. Before starting, mycelia were incubated on SDA on thin sheets of cellophane at 26 ± 1 °C until the logarithmic growth phase. Subsequently, the cultures were transferred to Petri plates with media containing 20 or 100 µg/mL of the single substance and incubated under growth conditions.

The fungal growth was evaluated daily by measuring colony diameters (in millimeters) for seven days from the treatment onset [37]. The percentage of inhibition is calculated according to the following equation:

$$\text{Percentage of relative inhibition (\%)} = [(d_{\text{ex}} - d_{\text{ex}'})/d_{\text{ex}}] \times 100\% \quad (1)$$

where *dex* is the diameter of the mycelium in the control; and *dex'* is the diameter of the mycelium measured in the experiment plates at different concentrations.

2.6. Preparation and Stability Test of Cosmetic Formulations

In the present study, hesperidin and its five synthetic derivatives were formulated using the same base formulation (at a concentration of 0.3%) and subjected to accelerated aging in an oven at 40 °C for 120 days. At each time point, the concentration of the active ingredient was monitored by HPLC analysis.

The formulation used is as follows:

INCI: Aqua, Glyceryl stearate citrate (8%), Caprylic/Capric Triglyceride (6%), Coco-caprylate (7%), glycerin (5%), cetearyl alcohol (2%), Xanthan gum (0.2%), Phenoxyethanol and Ethylhexylglycerin (1%).

For each molecule analyzed, a standard O/W (oil/water) emulsion was prepared. Once the ingredients were carefully weighted, three different phases were prepared. Phase A (aqueous phase) and phase B (oil phase) were heated separately up to a temperature between 60 and 70 °C, and then the oil phase was added to the aqueous phase, under mechanical stirring. During the subsequent cooling phase of the emulsion, maintained in continuous agitation, the compound investigated (0.3%) was added as a third step, to preserve it from possible thermal degradation. Finally, the emulsion was cooled and kept in the fridge until the next day for analysis. The final pH value of each emulsion was adjusted to 5.5.

For the analytical method, a different mobile phase was chosen for each formulation (Table 1).

Table 1. Analytical parameters of HPLC analysis of hesperidin and its derivatives.

Compound	Analytical Conditions				
	% Solvent A	% Solvent B	Flow Rate (mL/min)	λ (nm)	Column Temperature (°C)
Hesperidin	77	23	1.5	270	27
Esp1	2	98	1.5	270	27
Esp2	77	23	1.2	280	27
Esp3, Esp5	5	95	1.2	280	27
Esp4	55	45	1.2	280	27

2.7. Cell Cultures and Assays

2.7.1. K562 Cell Culture

K562, human erythroleukemic cell line, was the first human immortalized myelogenous leukemia line isolated and characterized by Lozzio CB and Lozzio BB, from a patient with chronic myelogenous leukemia (CML) in blast crisis [38]. K562 cells were cultured in a humidified atmosphere of 5% CO₂, in RPMI-1640 medium (Lonza, Verviers, Belgium) supplemented with 10% fetal bovine serum (FBS; Biowest, Nuaille, France), 50 units/mL penicillin (Lonza, Verviers, Belgium) and 50 µg/mL streptomycin (Lonza, Verviers, Belgium) [39].

2.7.2. Anti-Proliferative Activities

The K562 cells were counted with the BECKMAN COULTER® Z2 (Beckman, Pasadena, CA, USA) to evaluate the possible anti-proliferative effect caused by the derivatives after 72 and 96 h from the treatment, when cells are in the log phase of growth. Preliminary tests were done to identify the IC₅₀ on K562 cells: 25,000 cells/mL were seeded in 24-well plates and treated with hesperidin and synthetic analogues at different concentrations.

2.7.3. Pro-Apoptotic Effects

Annexin V and Dead Cell assays on K562 cell line, untreated and treated with increasing doses of the derivatives for three days, were performed with the Muse cell analyzer (Millipore, Billerica, MA, USA), according to the instructions supplied by the manufacturer. This procedure utilized Annexin V to detect PS (PhosphatidylSerine) on the external membrane of apoptotic cells. A dead cell marker was also used as an indicator of cell membrane structural integrity. Cells were first washed with sterile PBS 1X, second trypsinized and resuspended in the original medium, and finally diluted (1:2) with the one step addition of the Muse Annexin V and Dead Cell reagent. After incubation of 20 min at room temperature, cell samples were analyzed, using Triton X 0.01%, as positive control [40]. Data from prepared samples are acquired and recorded utilizing the Annexin V and Dead Cell Software Module (Millipore, Billerica, MA, USA). Four populations of cells were distinguished using this assay: Live, early apoptotic, late apoptotic, and dead cells.

2.8. Statistical Evaluations

Relative standard deviations and statistical significance (Student's *t* test; $p \leq 0.05$) were given where appropriate for all data collected. One-way ANOVA and LSD post hoc Tukey's honest significant difference test were used for comparing the bioactive effects of different samples. All computations were made using the statistical software STATISTICA 6.0 (StatSoft Italia srl).

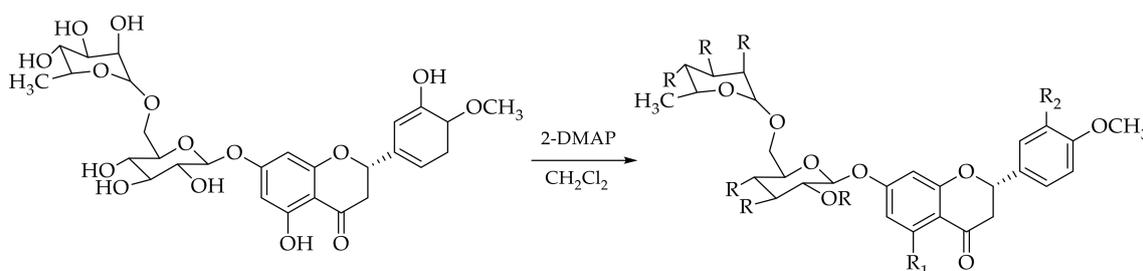
3. Results and Discussion

3.1. Chemistry

In order to extend potential applications of hesperidin, five derivatives were synthesized (Table 2). Different substituent groups were used to explore lipophilicity changes: We chose cyclic substituent and not with increasing steric hindrance. Obtained derivatives had none or one free hydroxyl phenolic group in order to evaluate the differences in terms of biological activity and stability. Esterification reactions were carried out using different substituents with 4-dimethylaminopyridine (DMAP) in CH₂Cl₂ (Scheme 1) [32]. All derivatives were obtained by the same synthetic procedure, but they were purified with different methods.

Table 2. Synthetic derivatives of hesperidin.

Compounds	R	R ₁	R ₂
Esp1	tert-butyl-acetyl	-H	tert-butyl-acetyl
Esp2	cyclo-hexyl-acetyl	-H	cyclo-hexyl-acetyl
Esp3	benzoyl	-H	benzoyl
Esp4	naphthoyl	-H	naphthoyl
Esp5	cinnamoyl	cinnamoyl	cinnamoyl

**Scheme 1.** Synthetic procedure of **Esp1–Esp5**.

3.2. LogP Values

The partition ratio (LogP), was calculated in order to evaluate changes in the lipophilic properties of the molecules induced by derivatizations. LogP values (Table 3) confirmed the increased lipophilicity of synthetic derivatives than hesperidin that had a negative LogP value.

Table 3. LogP values of hesperidin and its synthetic derivatives.

Compounds	LogP ^[*]
Hesperidin	−0.87
Esp1	1.38
Esp2	0.40
Esp3	2.04
Esp4	1.08
Esp5	1.04

[*] LogP = log₁₀ [mg/20 mL]octanol/[mg/20 mL]H₂O.

3.3. Antifungal Activity

Recent studies have shown that a particular flavonoid's antimicrobial activity is correlated with its lipophilicity. Indeed, a quantitative structure-activity relationship (QSAR) study has shown that the activity of these compounds can be related to the hydrophobicity of the molecules themselves (CLogP) and to the presence of substituents in particular positions. The investigation of our derivatives on antifungal activity is based on this scientific evidence [41]. Taking into consideration the possible application as a multifunctional ingredient for dermo-cosmetic applications, antifungal activity of hesperidin and its derivatives were tested on six dermatophytes by the diffusion method in Sabouraud Dextrose Agar (SDA), in comparison with the well-known antifungal agent, Fluconazole. Antifungal activity of the natural molecule and its derivatives (data not shown) was not significant. The inhibition values were less than 20% for each compound on all dermatophytes, at the concentration of 20 and 100 µg/mL. In several cases the compounds have shown hormone-like effects with a fungal growth higher than that of the control. IC₅₀ values of Fluconazole on the same dermatophytes were between 3.5 to 37.2 µg/mL, underlining the poor power of hesperidin and its derivatives on these microorganisms.

3.4. Antioxidant Activity

The oxidative process is an important part for UV and pollution skin damage. However, it is responsible for the great instability of hesperidin in the dermo-cosmetic formulation due to accelerated degradation. To evaluate the antioxidant activity of semisynthetic compounds and hesperidin, three different *in vitro* tests were used. We chose to test the natural compound and its derivatives against different radicals (DPPH, FRAP and Photochemoluminescence assays) for a broader spectrum of antioxidant activity (Table 4).

Table 4. Antioxidant assays. Each value was the result of three different experiments (Mean \pm SE).

Compounds	PCL $\mu\text{molTE/g}$	FRAP $\mu\text{molTE/g}$	DPPH $\mu\text{molTE/g}$
Hesperidin	1068.63 \pm 1.12	1255.32 \pm 4.04	216.61 \pm 0.87
Esp1	2.16 \pm 0.09	44.37 \pm 0.96	No activity
Esp2	1.14 \pm 0.50	20.15 \pm 0.58	No activity
Esp3	22.58 \pm 2.92	14.18 \pm 0.64	6.03 \pm 0.31
Esp4	6.06 \pm 1.70	19.61 \pm 0.84	No activity
Esp5	8.12 \pm 1.83	18.19 \pm 0.77	No activity

As expected, hesperidin confirmed to be a potent antioxidant molecule, demonstrating a consistent activity against all of the three different radicals. On the other hand, being the new molecule precursor acylated to at least at one of the phenolic groups, they showed lower or no antioxidant activity in these *in vitro* assays, as expected. **Esp1** demonstrated a two-fold increase in antioxidant activity (44.37 $\mu\text{mol TE/g}$) than the values of the other derivatives in FRAP test. Saturated alkyl chain (**Esp1** and **Esp2**) exhibited similar antioxidant values in DPPH and PCL tests, like the aromatic ones. **Esp5** was the only derivative which had both hydroxyl groups esterified. However, no significant difference in antioxidant activity with respect to the mono derivatives was found, demonstrating that the activity of the natural compound resides in the OH phenolic group on the aromatic ring in position two on bicycle croman-4-one, which was protected in all the obtained derivatives. In this respect, it is worth to note that the structure-activity relationship on antioxidant property of hesperidin, has not been deeply investigated so far [42].

3.5. Stability Studies

The stability of the newly synthesized compounds was evaluated in dermo-cosmetic formulations containing 0.3% (*w/w*) of each molecule. Finished formulations were subjected to accelerated aging in an oven at 40 °C and monitored by HPLC for 120 days.

It is noteworthy (Figure 1) that the increase in lipophilicity greatly improved stability of hesperidin, thus making it possible to use the in finished formulations (less than 18% decrease after 120 days at 40 °C). Indeed, hesperidin, which is susceptible to degradation, induced important changes also in the appearance of the formulation when introduced in significant amount in the finished products (color range went from yellow to brown after 30 days at 40 °C). This latter occurrence explains why it is impossible to reach effective concentrations of free hesperidin in finished dermo-cosmetic formulations. On the contrary, each formulation, with its derivatives, remained uncolored until the end of its respective stability study.

3.6. Antiproliferative Activity on Human Leukemic K562 Cells

Hesperidin and its synthetic derivatives were tested to evaluate their potential antiproliferative effects on human chronic myelogenous leukemia K562 cell line, and demonstrated as a very useful experimental model system for the screening of potential antitumor drug [43]. This was done for two reasons: (a) To identify derivatives exhibiting antioxidant activity, with low effects on cell proliferation of eukaryotic cells and (b) to identify derivatives exhibiting both biological activities. Increasing

amounts of hesperidin and its derivatives (from 1 μM to 500 μM) were added to the cell cultures in order to analyze their effects on cell growth, evaluating the IC_{50} values after three and four days of cell culture. The results (Table 5) showed that the IC_{50} values of hesperidin and **Esp2** are very similar (IC_{50} values: $389.96 \pm 66.33 \mu\text{M}$ and $376.13 \pm 45.53 \mu\text{M}$, respectively), on the contrary we found that **Esp4** and **Esp5** are more active in inhibiting K562 cell growth (IC_{50} values: $11.12 \pm 3.68 \mu\text{M}$ and $21.68 \pm 4.36 \mu\text{M}$, respectively). The data of **Esp1** and **Esp3** were not determined in detail since **Esp1** and **Esp3** were unstable during the test with clear evidence of precipitation (ppt) at concentrations exceeding 50 μM (**Esp1**) and 400 μM (**Esp3**).

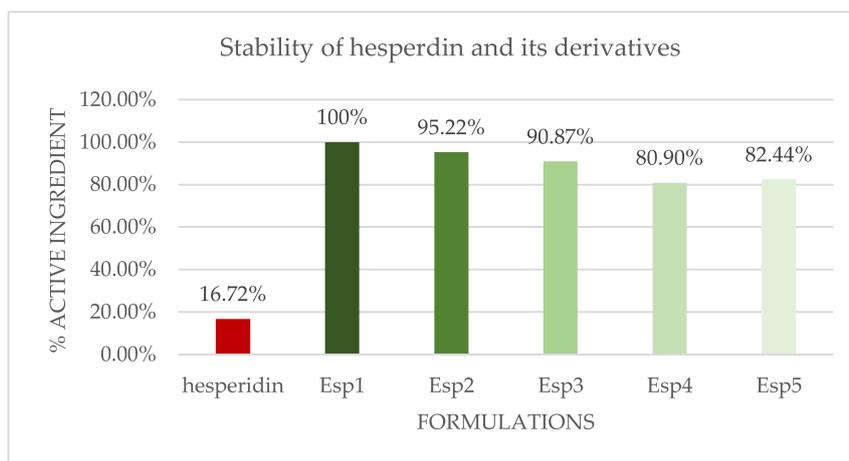


Figure 1. T120 Stability data of formulations with hesperidin and its derivatives (results are percentage decrease compared to T0).

Table 5. Effects of hesperidin and its derivatives on cell proliferation of K562 cells (the data are the mean of three independent experiments \pm SD).

Compound	IC_{50} (μM) on K562 Cell Line
Hesperidin	389.96 ± 35.49
Esp1	>50 (ppt)
Esp2	376.13 ± 32.29
Esp3	>400 (ppt)
Esp4	11.12 ± 1.97
Esp5	21.68 ± 2.82

3.7. Pro-Apoptotic Effect of Hesperidin and Its Analogues Esp1-5 on the K562 Cell Line

Since anti-proliferative effects might be associated with the activation of apoptotic pathways, apoptosis was analyzed on the K562 cell line after treatment with hesperidin and its analogues for three days. Two different concentrations were used close to the relative IC_{50} values. All these concentrations were selected starting from the anti-proliferative effects shown in Table 5, except for **Esp1** and **Eps3**, which were added at concentrations never exceeding the maximum concentrations used for these hesperidin analogues in the cell proliferation assays reported in Table 5. This was conducted in consideration of their already discussed precipitation effects in the aqueous medium, requiring the use of concentration values lower than their solubility limit. Table 6 shows the obtained results on the pro-apoptotic effects of these compounds, using the assay system, which enables the identification of early and late apoptosis stage, after comparison to untreated control cells (C-). The data shown in Table 6 originated from the analysis of the corresponding Muse Analyzer plots, after 72 h of treatment (representative results shown in Figure 2).

Among the tested compounds, the most interesting response was found when hesperidin and **Esp2** were employed. Both of them were found to exert a very important and relevant activity, inducing

apoptosis (when total apoptotic cells were considered) in more than 30% and 50% of the total cell population, respectively. The most interesting data obtained highlighted the shift caused by **Esp2** of almost all the population to early apoptotic cells (Figure 2A) (Annexin-V positive, PI negative). Moreover, **Esp5** seems to induce a slight pro-apoptotic effect (11.60%). Among the anti-proliferative hesperidin analogues (see Table 4), a clear increment of late apoptotic cells was found with **Esp5**. For all three active derivatives, it is evident that a dose-dependent apoptosis increased, though it possible to observe some differences: Hesperidin and **Esp5** are able to induce late apoptosis in the K562 cells in 72 h, while **Esp2**, that possess the more evident pro-apoptotic effect, stimulates early apoptosis (Table 5 and Figure 2A).

Table 6. Pro-apoptotic activity (%) of hesperidin and semi-synthetic analogues on K562 cells.

Compound	Live Cells (%)	Early Apoptotic Cells (%)	Late Apoptotic Cells (%)	Total Apoptotic Cells (%)
C- (untreated cells)	94.35	1.65	3.15	4.80
Hesperidin 300 μ M	74.05	3.50	21.65	25.15
Hesperidin 400 μ M	68.20	4.20	26.85	31.05
Esp1 20 μ M	94.00	1.65	3.50	5.15
Esp1 50 μ M	95.20	1.65	2.55	4.20
Esp2 300 μ M	96.15	1.20	2.25	3.45
Esp2 400 μ M	40.20	53.20	5.50	58.70
Esp3 200 μ M	90.70	2.60	6.05	4.20
Esp3 400 μ M	95.45	1.25	2.95	8.65
Esp4 5 μ M	96.45	1.05	1.70	2.75
Esp4 10 μ M	93.15	2.40	3.85	6.25
Esp5 10 μ M	92.35	1.65	5.05	6.70
Esp5 20 μ M	87.40	2.00	9.60	11.60

This rather unexpected finding was confirmed in five different independent experiments aimed at comparing hesperidin and **Esp2**. The data obtained are reported in Figure 2B and give clear evidence that, while 400 μ M hesperidin induced late apoptosis, 400 μ M **Esp2** was able to induce only early apoptosis, characterized by the alteration of the charge ratio of the membrane causing Phosphatidylserine (PS) to be exposed to the cell surface. In the case of **Esp2** treatment (unlike the treatment with hesperidin) this is not associated with disruption of the cellular membrane integrity. In fact, most of the hesperidin-treated cells (34.89 ± 12.17 in five independent experiments) were found to be Annexin-V positive and propidium iodide (PI) negative, while the majority of **Esp2** treated cells (44.52 ± 8.59 in five independent experiments) were found to be Annexin-V positive, PI negative. This effect is highly significant ($p < 0.001$). The induction of early apoptosis (but not of late apoptosis) was confirmed by forcing the system employing **Esp2** at 700 μ M. In this case, the % of early apoptotic cells increased, in five independent experiments, to 83.68 ± 5.16 (data not shown), the % of late apoptotic cells being 8.96 ± 1.51 .

The data shown in Figure 2 suggest that hesperidin and **Esp2** are potent inducers of apoptosis, but exhibit a sharply different mechanism of action, leading to activation of the early apoptosis pathway (**Esp2**) or to late apoptosis (hesperidin). Further experiments are necessary to understand the molecular basis of this differential effect.

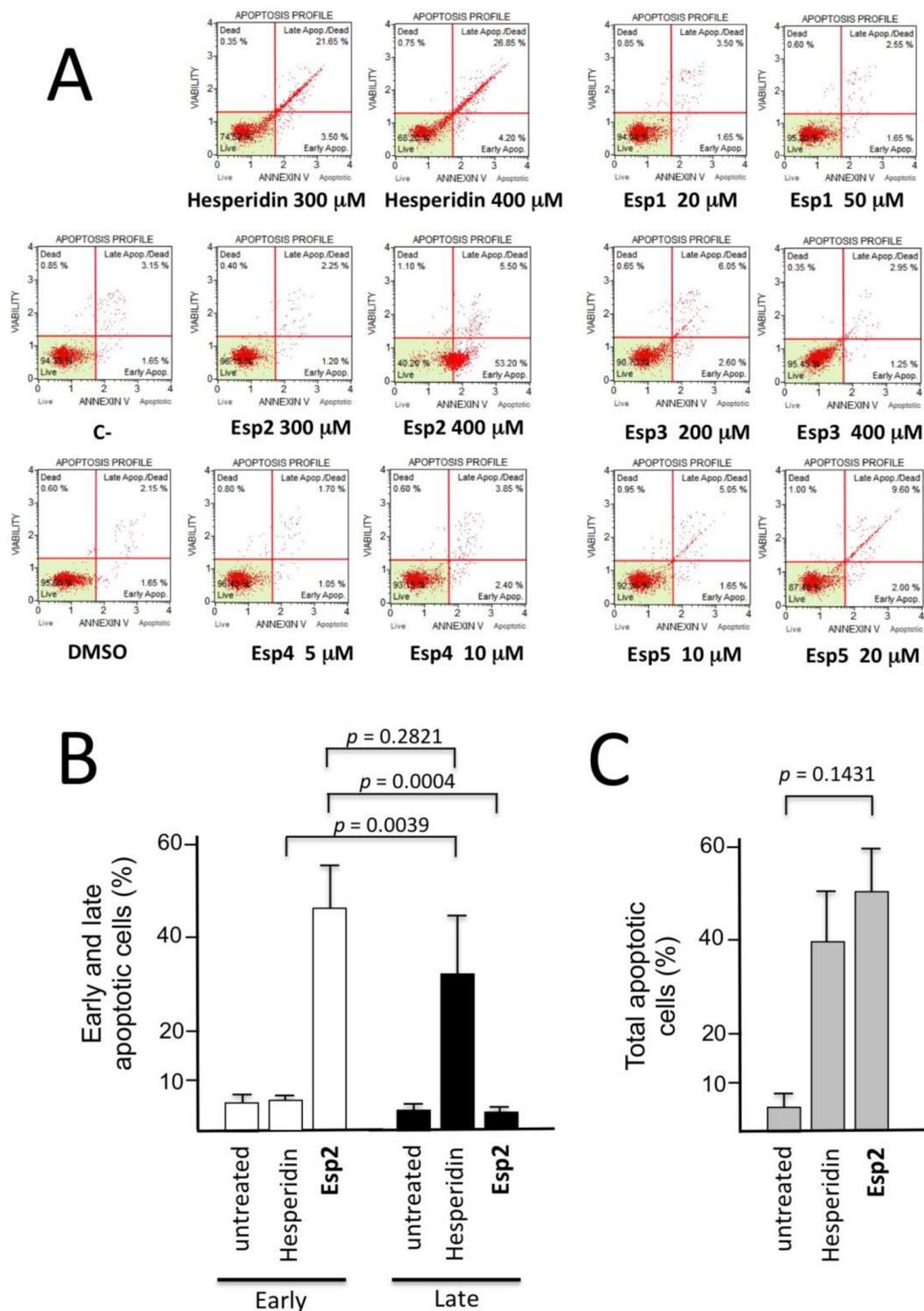


Figure 2. Annexin-V assays were performed on K562 cells treated for 72 h with the indicated concentrations of hesperidin and semi-synthetic analogues. (A) Representative examples showing that the best effects (total apoptosis >30%) on K562 cells treated with the hesperidin and its derivative **Esp2** are here reported in comparison with untreated cells (negative control, C-). Also **Esp5** demonstrated to induce late apoptosis (11.60%). (B,C) Summary of the results obtained from five independent experiments. K562 cells were treated for 72 h with 400 μ M hesperidin and **Esp2**, as indicated and the apoptosis assay was performed as described in panel A. The results represent the average \pm S.D., of five independent experiments.

4. Conclusions

The present preliminary study was designed in order to investigate the effects of lipidization of hesperidin, as a strategy to improve stability and the chance of inclusion in skin topical formulations of hesperidin derivatives. Five ester derivatives were prepared characterized by different substituting groups, ranging from a simple alkyl chain up to aromatic structures increasingly conjugated. Our results allow for us to hypothesize that synthetic derivatives can behave as pro-drugs of hesperidin, although further studies of bioavailability and determination of antioxidant activity in cells and tissues are necessary.

All synthetic derivatives proved to be much more stable in dermo-cosmetic formulations than the formulation containing hesperidin itself as active ingredient, confirming its applicability in products for dermo-cosmetic use. One obvious control was to verify the possible influence on activity, thus antiproliferative and pro-apoptotic effects of the hesperidin derivatives on eukaryotic cells were investigated. We therefore performed preliminary tests using erythroleukemic K562 cells as the experimental model system, since this cell line has been extensively used for these analyses. The results obtained indicated that **Esp3** exhibited very low pro-apoptotic and anti-proliferative effects (Table 5 and Figure 2), while maintaining strong anti-oxidant activity (Table 4). Some derivatives exhibited a multifunctional activity that can complement, in a preventive way, the dermo-cosmetic application. Indeed, during the K562-based preliminary screening, they were interesting candidates for a multifunctional approach, exhibiting, in addition to anti-oxidant activity, cell growth inhibitory and pro-apoptotic activity. This has the potential to have a significant impact, also when considering the known skin tumorigenesis inhibition of hesperidin [44], which drives attention towards this, and related compounds, as a possible preventive agent in the treatment of some human pathologies, such as UV induced neoplastic diseases [45], squamous cell carcinoma (SCC) [46], malignant melanomas [47], non-melanoma skin tumors [48], and psoriasis-associated skin cancers [49]. In this context, hesperidin derivatives should be tested on the several “in vitro” experimental model systems available for these pathologies [50,51], as well as on “in vivo” mouse models for skin cancers [52,53].

As far as the effects on apoptosis, hesperidin and the analogue **Esp2** behave differently, since hesperidin induces activation of late apoptosis, while **Esp2** induces only early apoptosis. This should be considered in dermo-cosmetics, given that **Esp2** is expected to differentially influence gene expression when compared to hesperidin. In respect to anti-tumor treatment, while hesperidin might be proposed as such, **Esp2** might be used in combination with other anti-tumor drugs as proposed for other inducers of early stag apoptosis, such as fluoxetine [54] and carnorsol [55]. Combination therapy might be of interest since is based on the use of sub-optimal (and less toxic) concentrations of each agent.

Finally, the structure-activity relationship on antioxidant property of hesperidin, has not been deeply investigated so far and, due to the results here presented, further studies (in vitro and in vivo) are planned in order to deepen the knowledge of this interesting natural molecule, in particular its behavior towards normal and carcinogenic skin cells.

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