

Supplementary materials

New Lipidyl-Cyclodextrins Obtained by Ring Opening of Methyl Oleate Epoxide using Ball Milling

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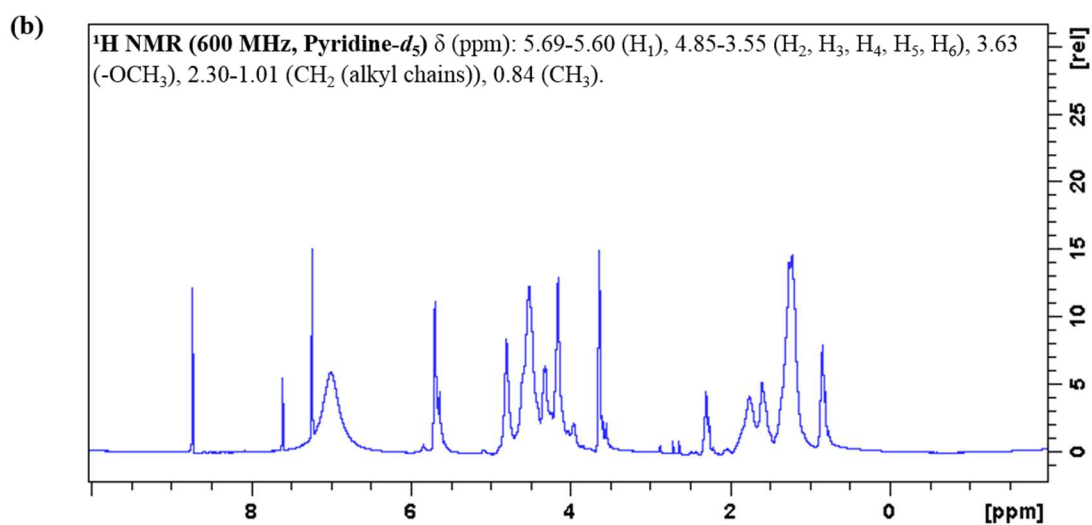
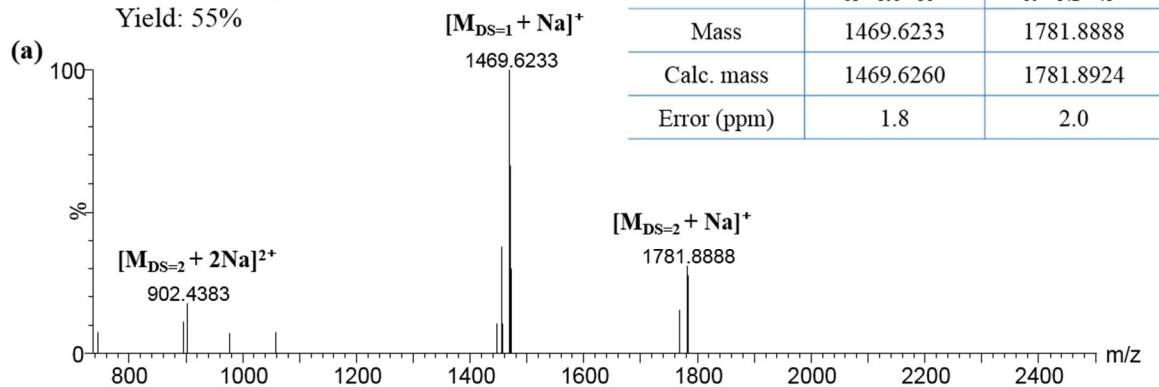
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β -CD(C₉)₂OOME, **1**

$\overline{DS}$: 1.4

MWa: 1577.3 g/mol

Yield: 55%



(c) ¹³C NMR (151 MHz, Pyridine-*d*₅) δ (ppm): 174.4 (C=O), 104.3 (C₁), 85.2-83.0 (C₄), 75.3-72.7 (C₂, C₃, C₅), 62.4-61.6 (C₆), 51.7 (-OCH₃), 34.5-23.0 (CH₂ (alkyl chains)), 14.6 (CH₃).

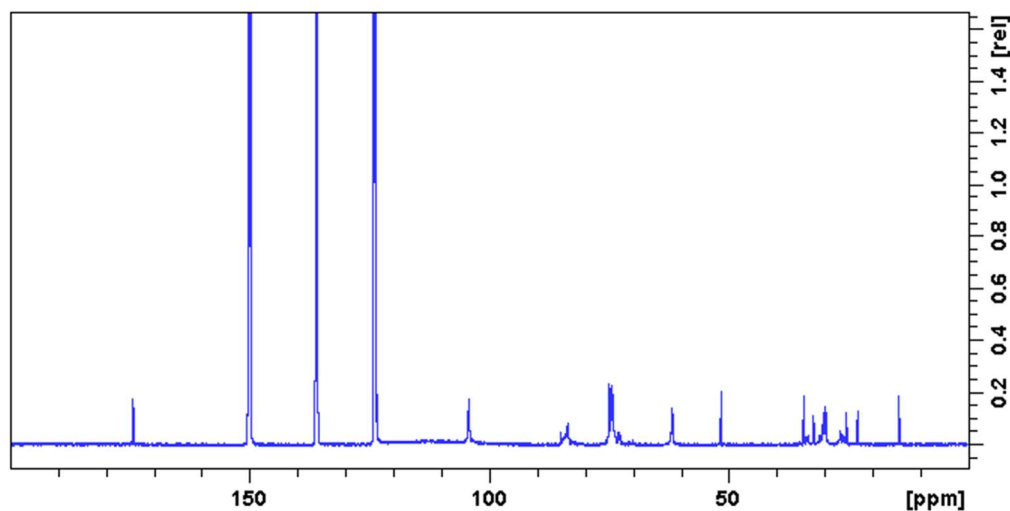


Figure S1: ESI⁺-HRMS (a), ¹H NMR (b) and ¹³C NMR spectra (c) of β -CD(C₉)₂OOME **1**.

α -CD(C₉)₂OOMe, **2**

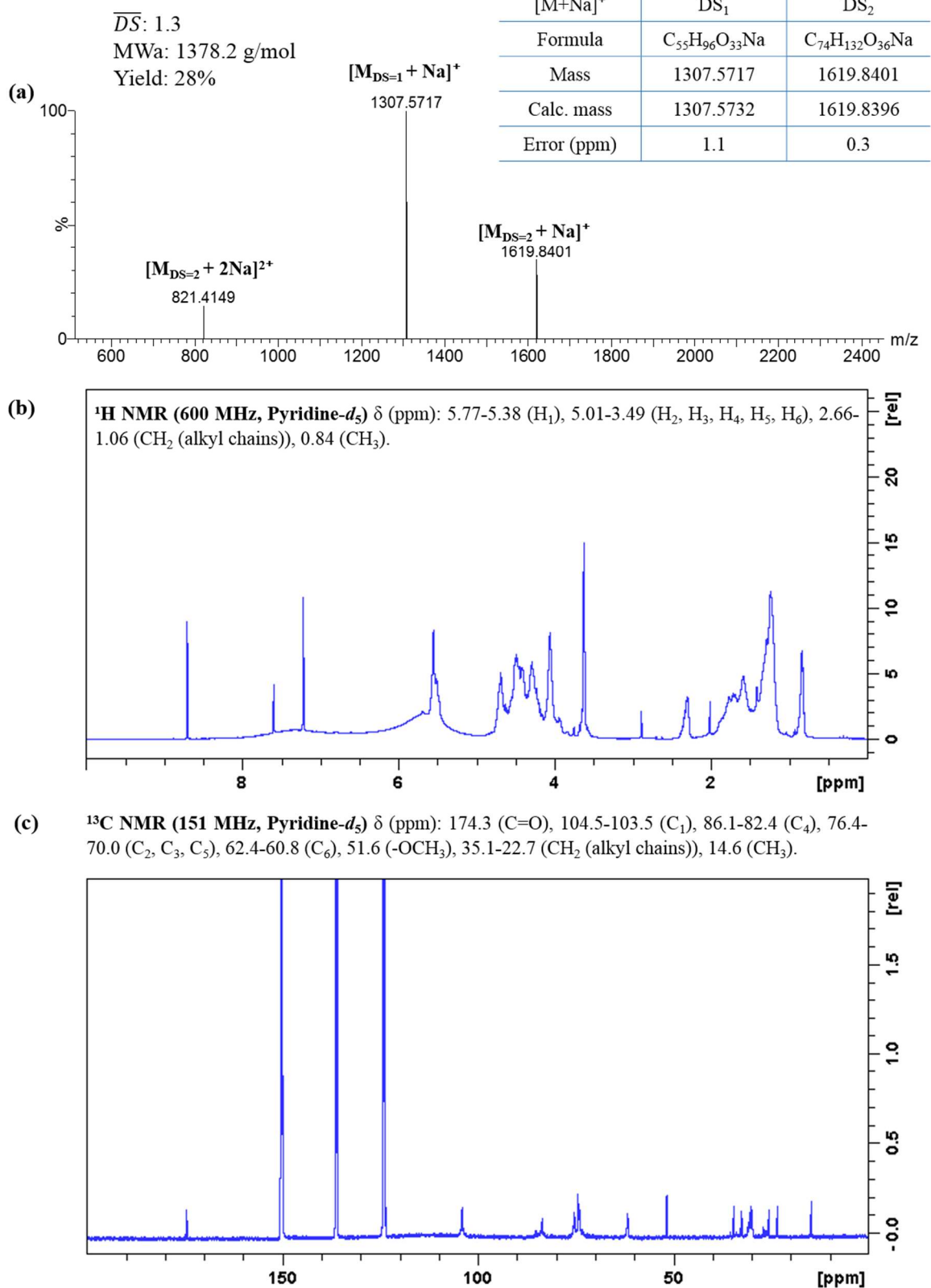


Figure S2: ESI⁺-HRMS (a), ¹H NMR (b) and ¹³C NMR spectra (c) of α -CD(C₉)₂OOMe **2**.

γ -CD(C₉)₂OOMe, **3**

$\overline{DS}$: 1.3

MW_a: 1712.5 g/mol

Yield: 33%

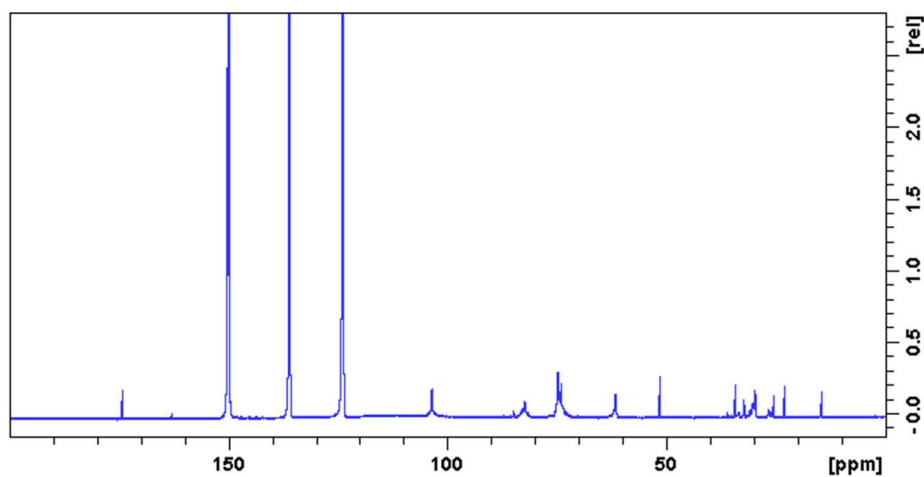
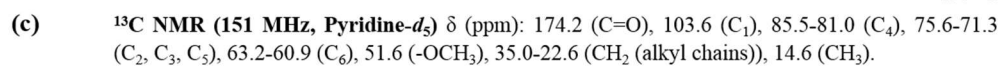
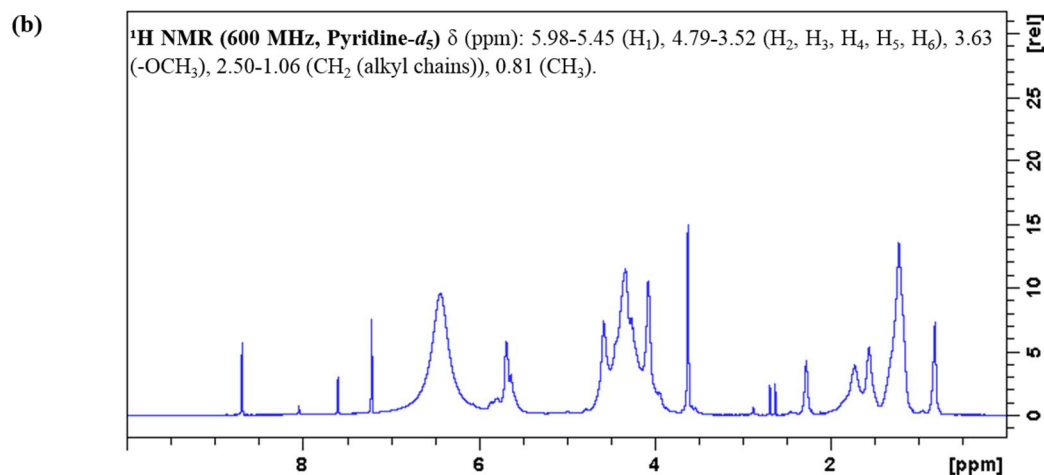
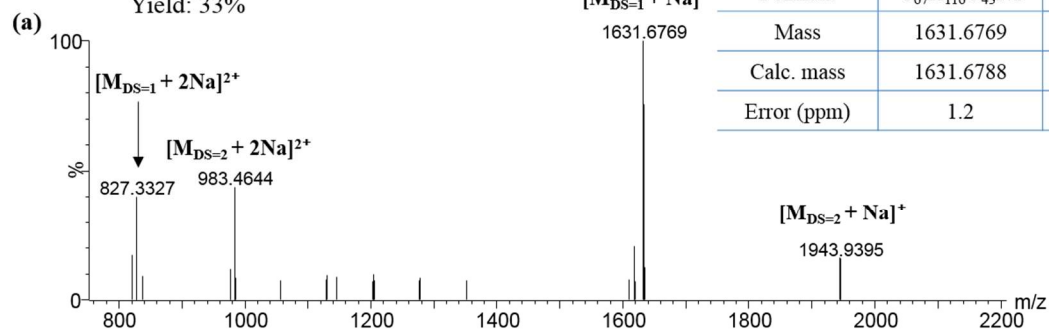


Figure S3: ESI⁺-HRMS (a), ¹H NMR (b) and ¹³C NMR spectra (c) of γ -CD(C₉)₂OOMe **3**.

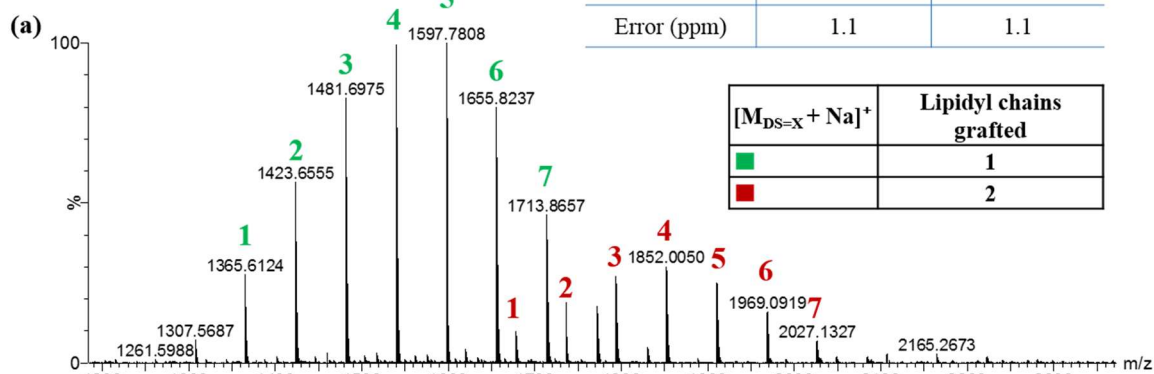
HP- α -CD(C₉)₂OOMe, **4**

$\overline{DS}$: 1.2

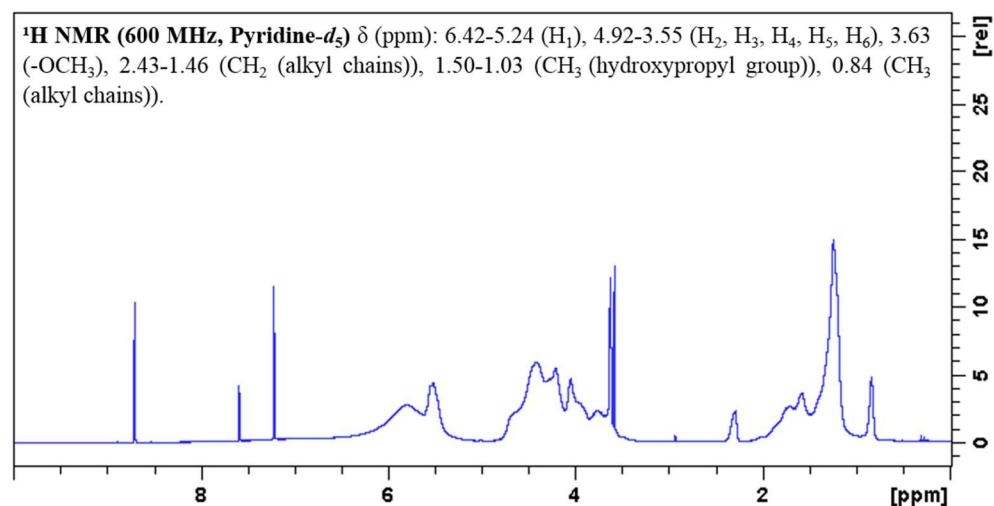
MW_a: 1571.7 g/mol

Yield: 33%

[M+Na] ⁺	DS ₁	DS ₂
Formula	C ₇₀ H ₁₂₆ O ₃₈ Na	C ₈₆ H ₁₅₆ O ₄₀ Na
Mass	1597.7808	1852.0050
Calc. mass	1597.7825	1852.0071
Error (ppm)	1.1	1.1



(b) ¹H NMR (600 MHz, Pyridine-*d*₅) δ (ppm): 6.42-5.24 (H₁), 4.92-3.55 (H₂, H₃, H₄, H₅, H₆), 3.63 (-OCH₃), 2.43-1.46 (CH₂ (alkyl chains)), 1.50-1.03 (CH₃ (hydroxypropyl group)), 0.84 (CH₃ (alkyl chains)).



(c) ¹³C NMR (151 MHz, Pyridine-*d*₅) δ (ppm): 174.3 (C=O), 105.0-100.5 (C₁), 86.1-77.4 (C₄), 76.0-66.0 (C₂, C₃, C₅), 63.0-60.7 (C₆), 51.6 (-OCH₃), 34.8-22.9 (CH₂ (alkyl chains)), 21.0-19.9 (CH₃ (hydroxypropyl group)), 14.6 (CH₃).

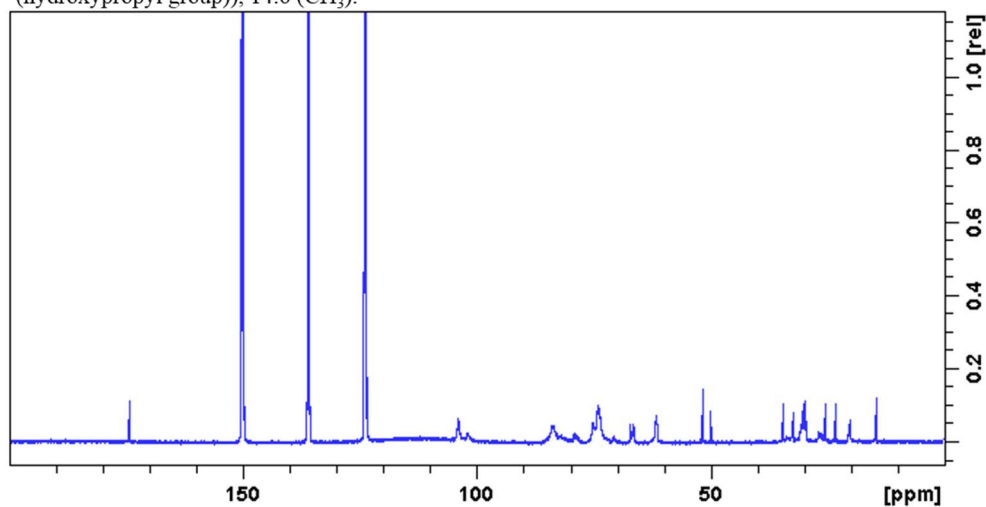


Figure S4: ESI⁺-HRMS (a), ¹H NMR (b) and ¹³C NMR spectra (c) of HP- α -CD(C₉)₂OOMe **4**.

HP- β -CD(C₉)₂OOME, 5

$\overline{DS}$: 1.3

MWa: 1892.5 g/mol

Yield: 35%

[M+Na] ⁺	DS ₁	DS ₂	DS ₃
Formula	C ₈₂ H ₁₄₈ O ₄₅ Na	C ₁₀₁ H ₁₈₄ O ₄₈ Na	C ₁₂₀ H ₂₂₀ O ₅₁ Na
Mass	1875.9177	2188.1821	2500.4465
Calc. mass	1875.9190	2188.1855	2500.4519
Error (ppm)	0.7	1.6	2.2

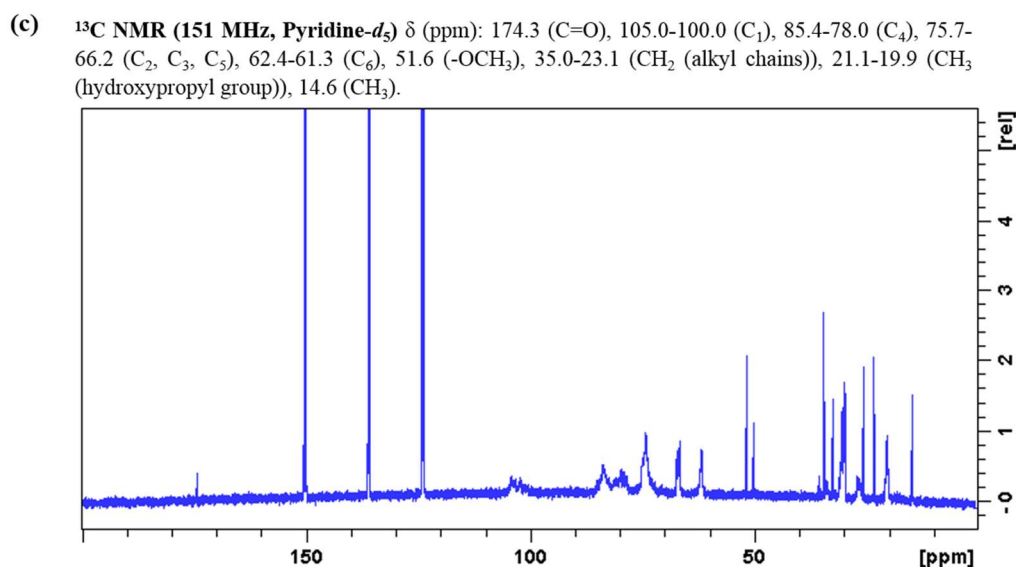
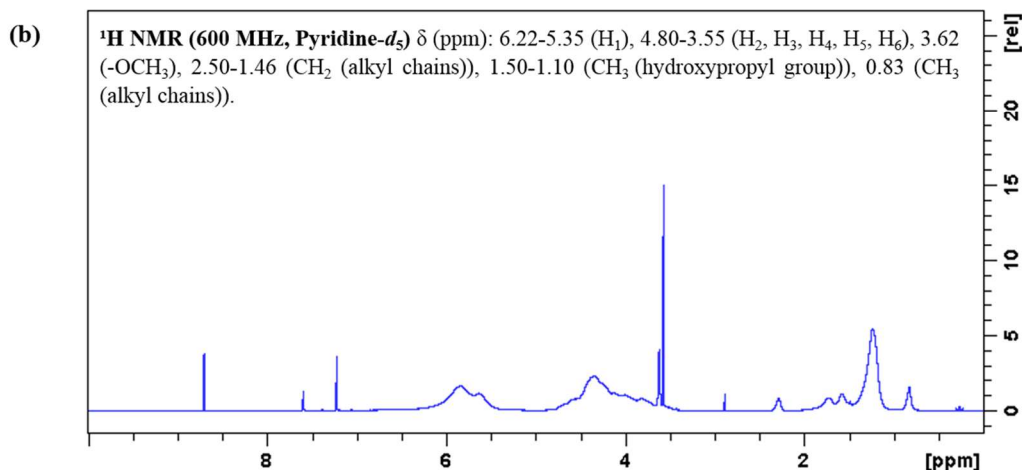
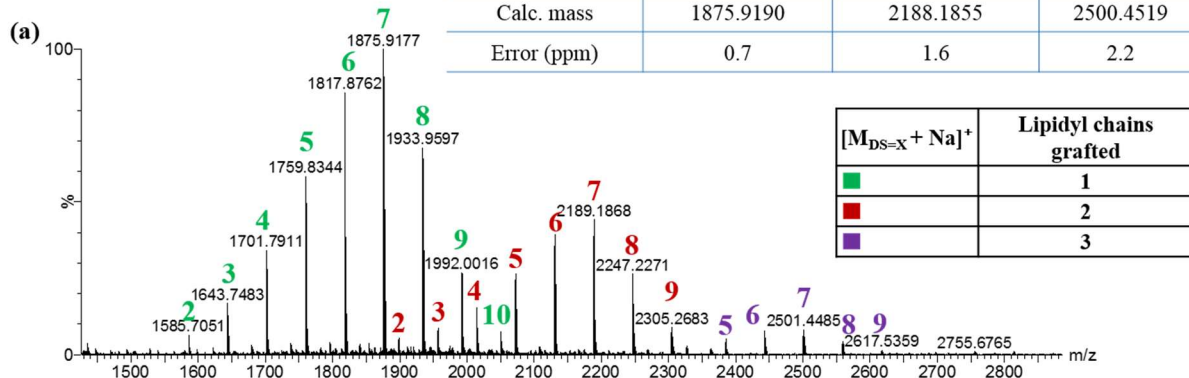


Figure S5: ESI⁺-HRMS (a), ¹H NMR (b) and ¹³C NMR spectra (c) of HP- β -CD(C₉)₂OOME 5.

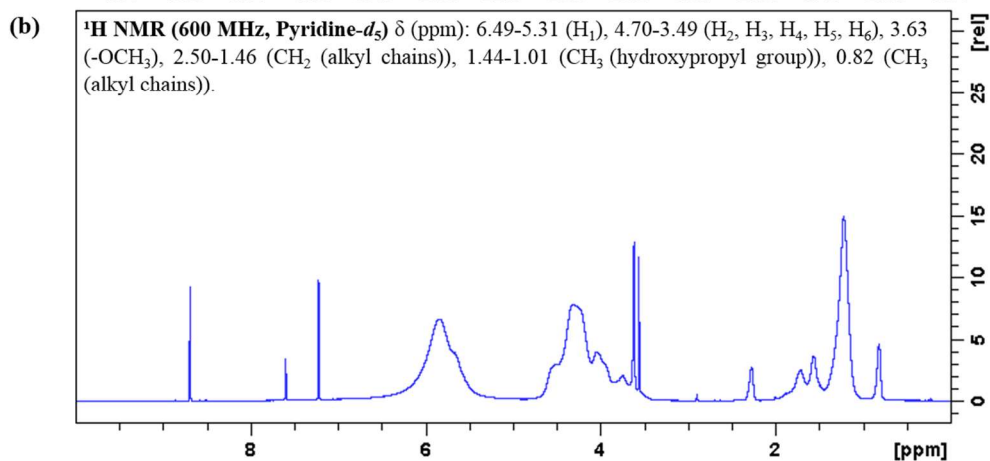
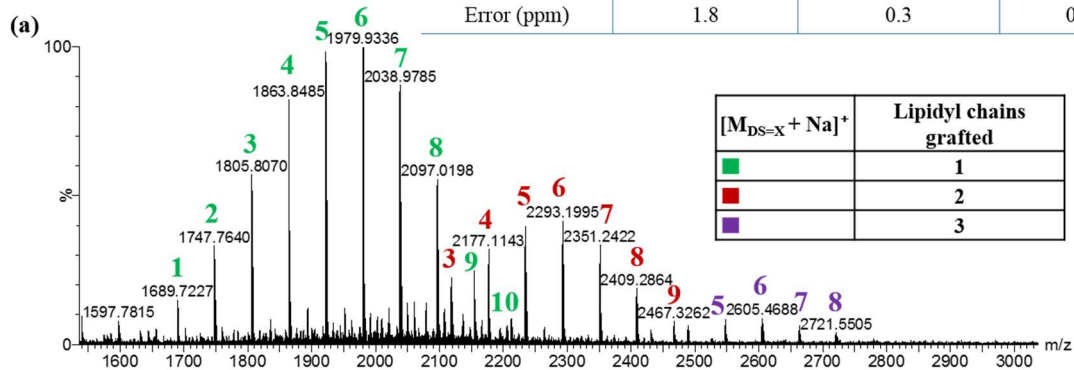
HP- γ -CD(C₉)₂OOMe, **6**

$\overline{DS}$: 1.2

MW_a: 2031.9 g/mol

Yield: 44%

[M+Na] ⁺	DS ₁	DS ₂	DS ₃
Formula	C ₈₅ H ₁₅₂ O ₄₉ Na	C ₁₀₄ H ₁₈₈ O ₅₂ Na	C ₁₂₃ H ₂₂₄ O ₅₅ Na
Mass	1979.9336	2292.1956	2604.4609
Calc. mass	1979.9300	2292.1964	2604.4629
Error (ppm)	1.8	0.3	0.8



(c) ¹³C NMR (151 MHz, Pyridine-*d*₅) spectrum showing chemical shift (ppm) versus relative intensity [rel]. The x-axis ranges from 0 to 150 ppm. The y-axis ranges from 0 to 0.8 [rel]. The spectrum shows peaks corresponding to the following chemical shifts (ppm): 174.3 (C=O), 104.6-98.9 (C₁), 86.0-78.4 (C₄), 75.8-65.6 (C₂, C₃, C₅), 63.0-60.2 (C₆), 51.7 (-OCH₃), 34.9-22.7 (CH₂ (alkyl chains)), 20.9-19.6 (CH₃ (hydroxypropyl group)), 14.6 (CH₃).

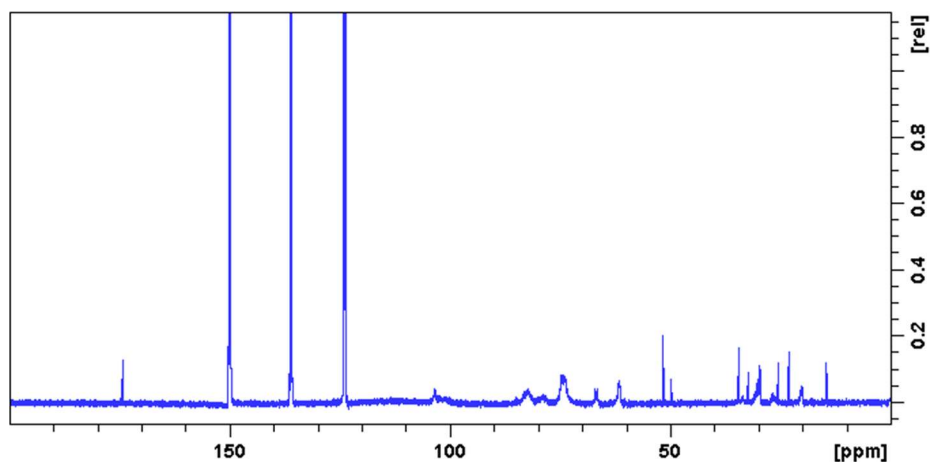


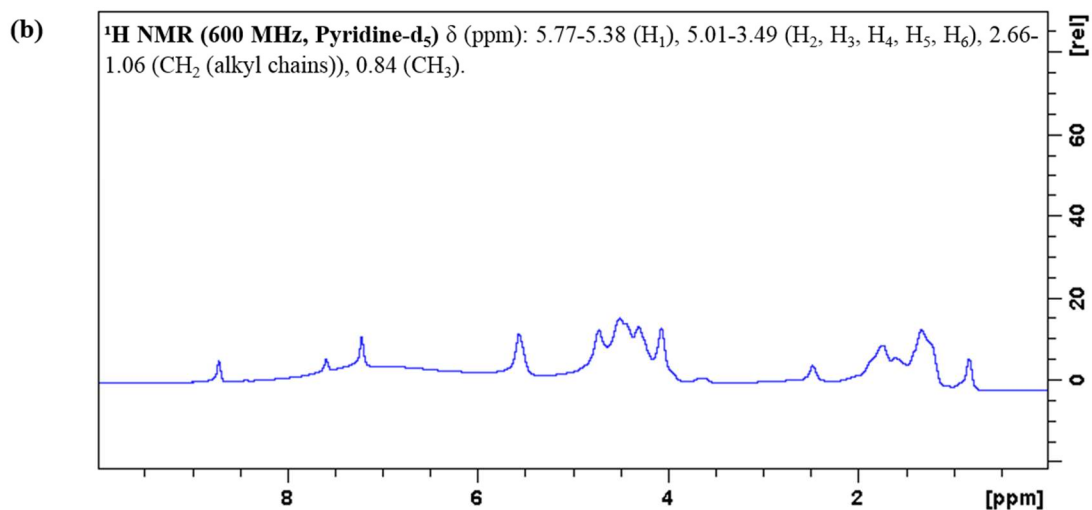
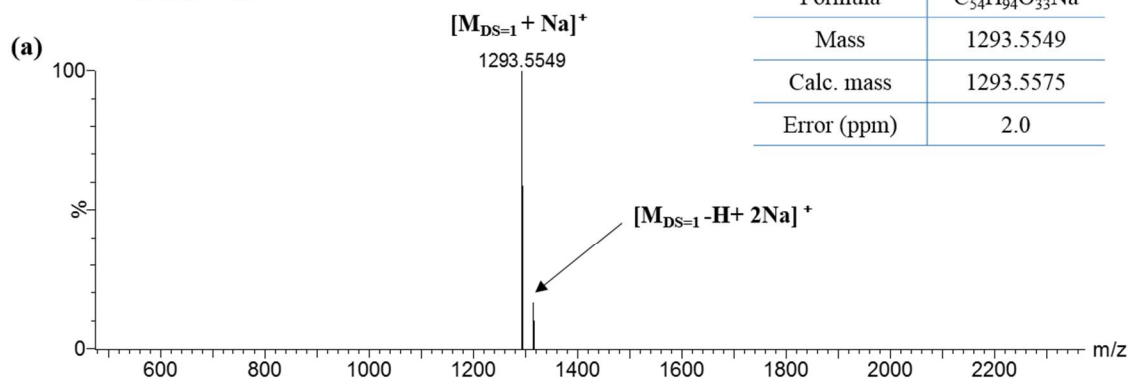
Figure S6: ESI⁺-HRMS (a), ¹H NMR (b) and ¹³C NMR spectra (c) of HP- γ -CD(C₉)₂OOMe **6**.

α -CD(C₉)₂OOH, 7

MWa: 1271.0 g/mol

Yield: 78%

[M+Na] ⁺	DS ₁
Formula	C ₅₄ H ₉₄ O ₃₃ Na
Mass	1293.5549
Calc. mass	1293.5575
Error (ppm)	2.0



(c) ¹³C NMR (151 MHz, Pyridine-d₅) δ (ppm): 176.7 (C=O), 104.7-103.1 (C₁), 85.7-81.9 (C₄), 76.3-69.4 (C₂, C₃, C₅), 62.7-60.8 (C₆), 35.9-22.9 (CH₂ (alkyl chains)), 14.6 (CH₃).

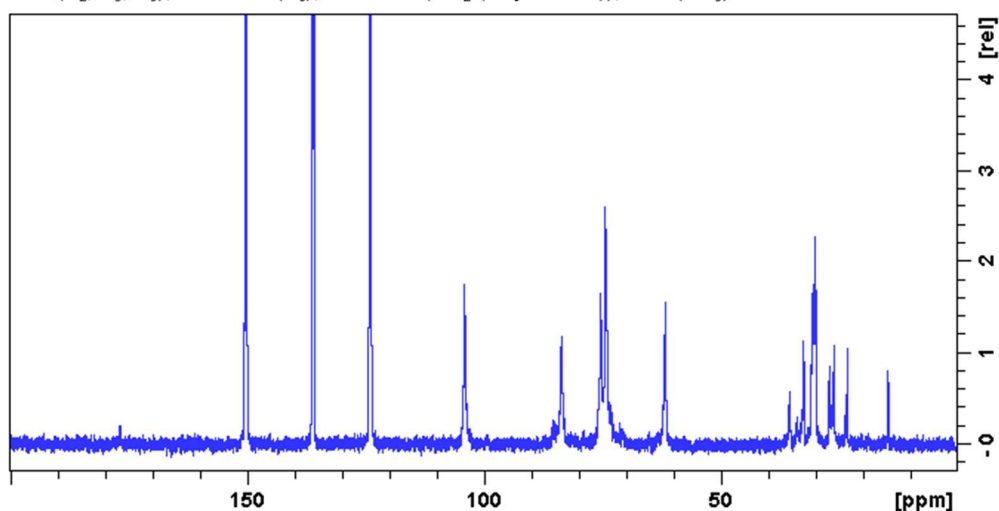


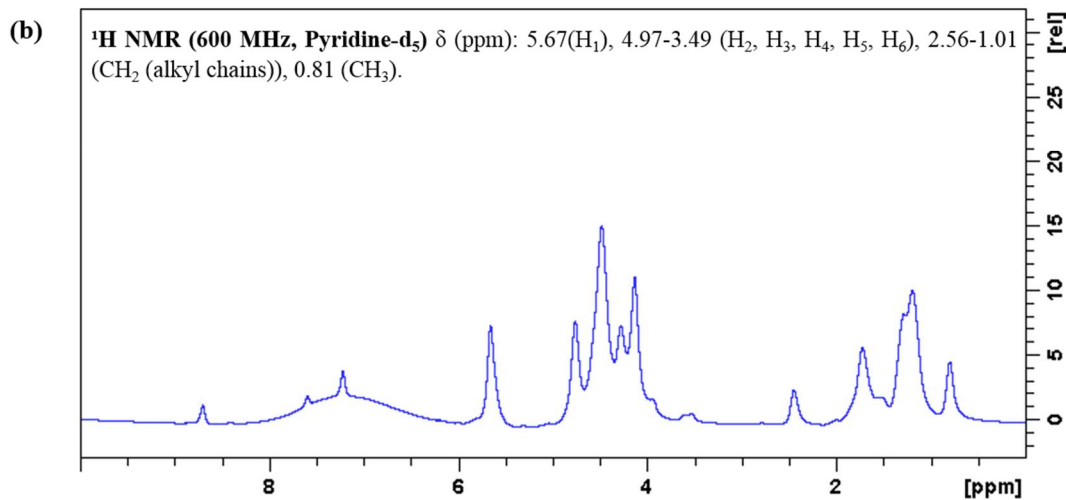
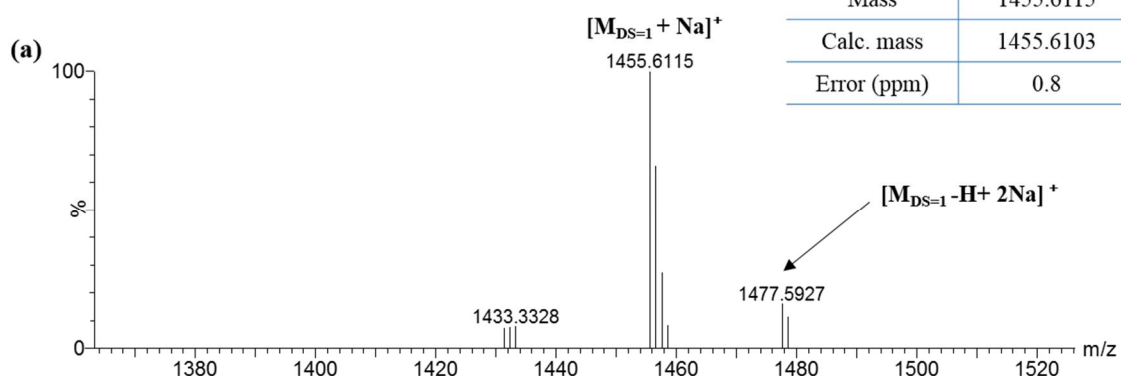
Figure S7: ESI⁺-HRMS (a), ¹H NMR (b) and ¹³C NMR spectra (c) of α -CD(C₉)₂OOH 7.

β -CD(C₉)₂OOH, 8

MWa: 1433.0 g/mol

Yield: 69%

[M+Na] ⁺	DS ₁
Formula	C ₆₀ H ₁₀₄ O ₃₈ Na
Mass	1455.6115
Calc. mass	1455.6103
Error (ppm)	0.8



(c) ¹³C NMR (151 MHz, Pyridine-d₅) spectrum showing chemical shift (ppm) versus relative intensity. The spectrum displays peaks corresponding to the carbons of the molecule, with chemical shifts ranging from 14.6 to 176.5 ppm. The peaks are assigned to C=O (176.5 ppm), C₁ (104.3 ppm), C₄ (85.4-82.7 ppm), C₂, C₃, C₅ (75.5-72.2 ppm), C₆ (62.5-61.2 ppm), CH₂ (alkyl chains) (35.4-22.9 ppm), and CH₃ (14.6 ppm).

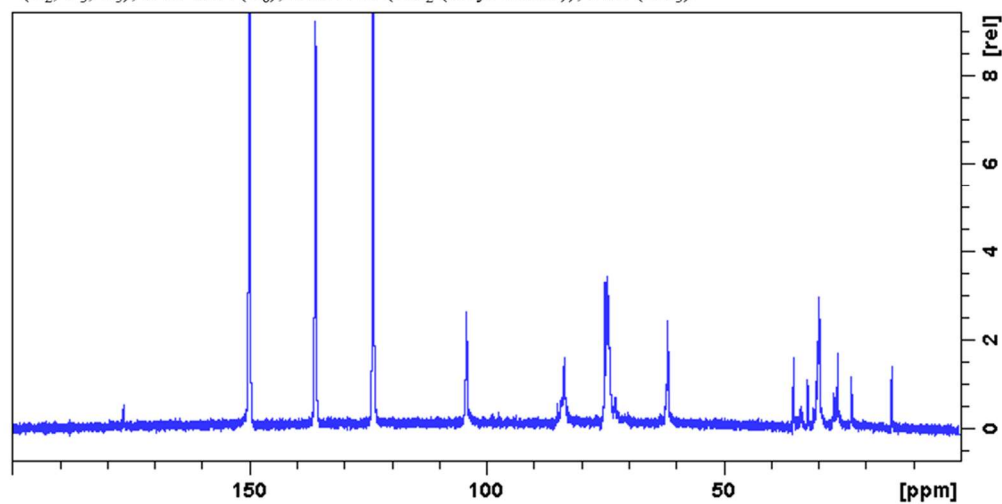


Figure S8: ESI⁺-HRMS (a), ¹H NMR (b) and ¹³C NMR spectra (c) of β -CD(C₉)₂OOH 8.

γ -CD(C₉)₂OOH, 9

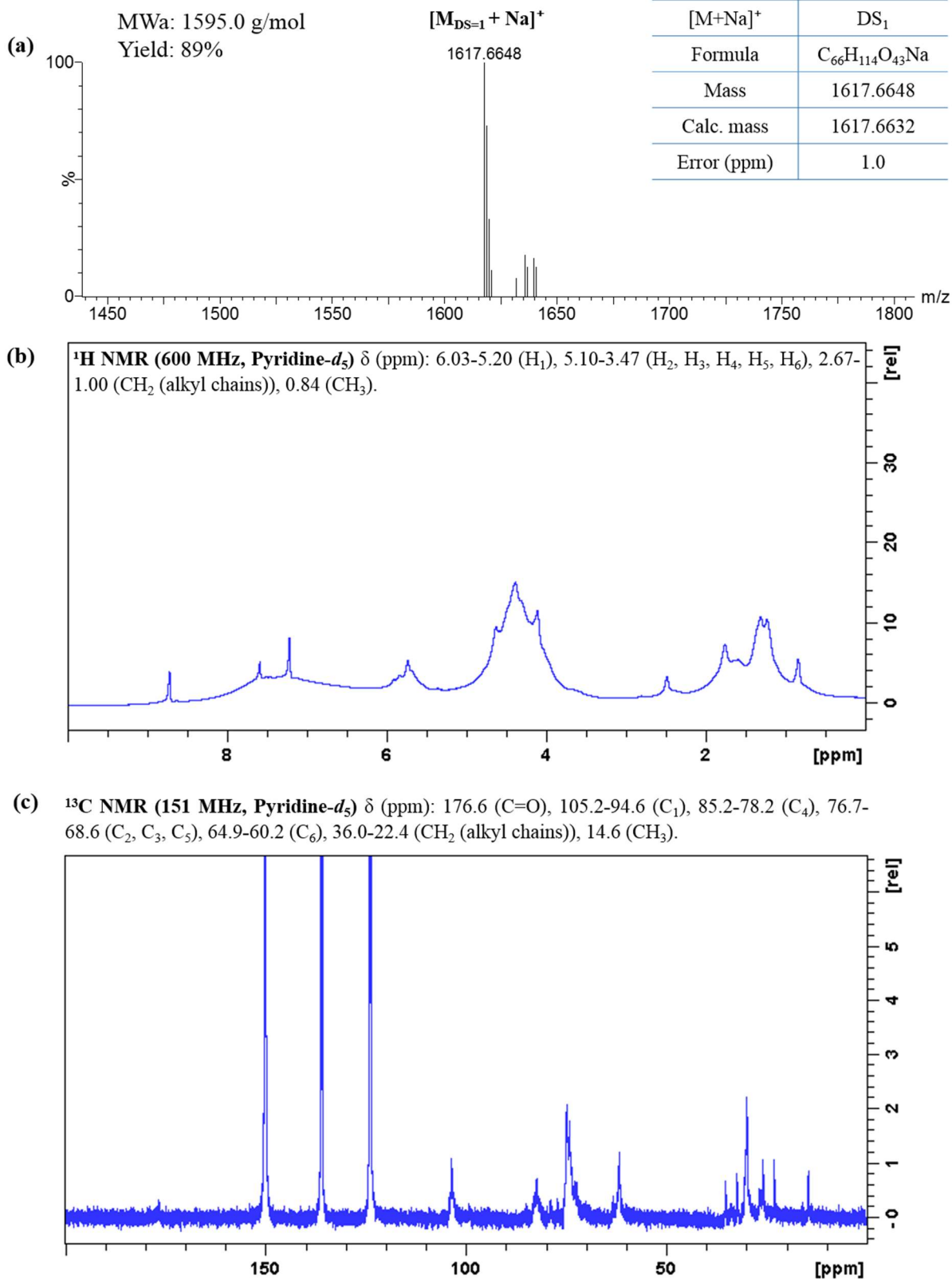


Figure S9: ESI⁺-HRMS (a), ¹H NMR (b) and ¹³C NMR spectra (c) of γ -CD(C₉)₂OOH 9.

HP- α -CD(C₉)₂OOH, 10

$\overline{DS}$ (hydroxypropyl group): 3.9
 MWa: 1499.5 g/mol
 Yield: 82%

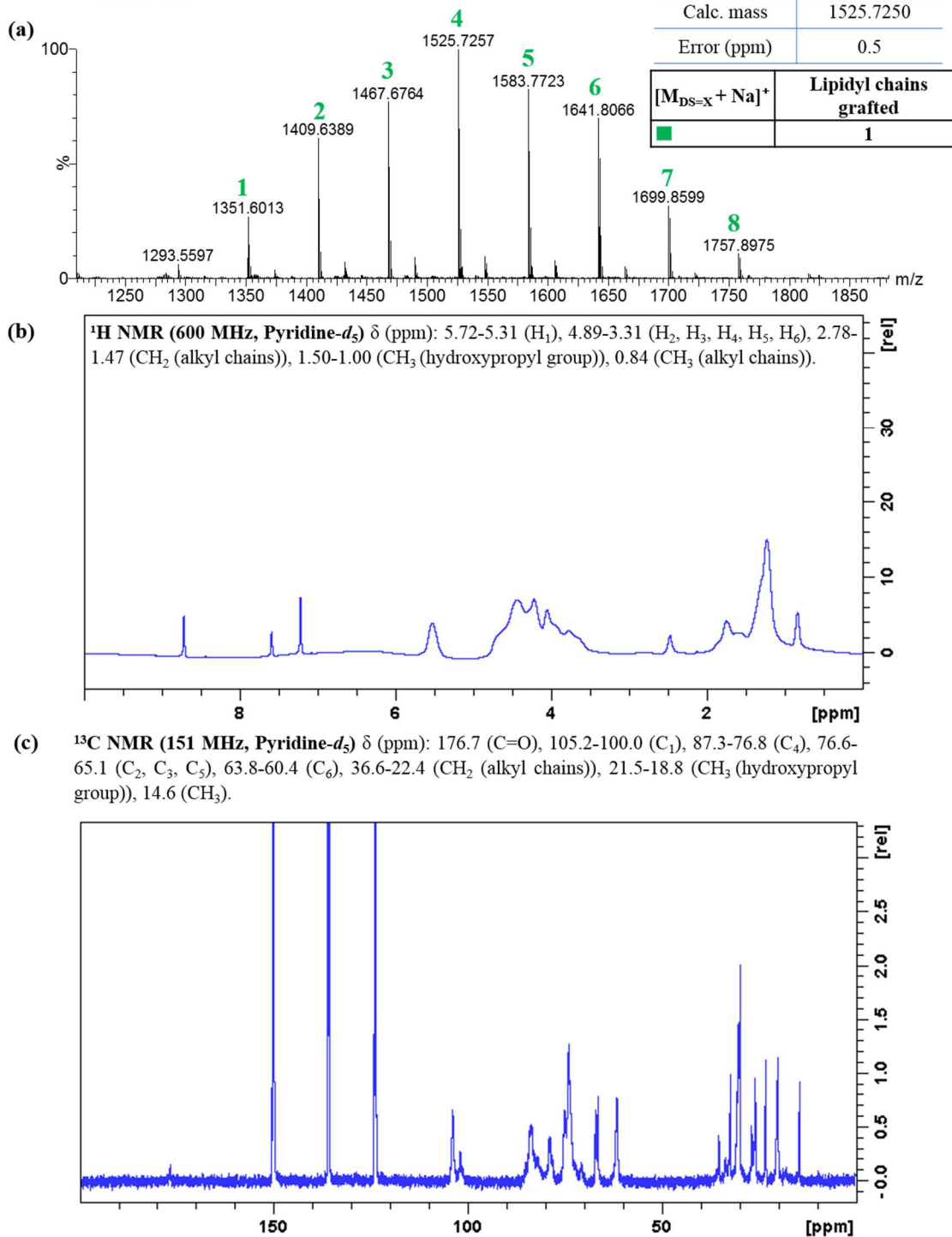


Figure S10: ESI⁺-HRMS (a), ¹H NMR (b) and ¹³C NMR spectra (c) of HP- α -CD(C₉)₂OOH 10.

HP- β -CD(C₉)₂OOH, **11**

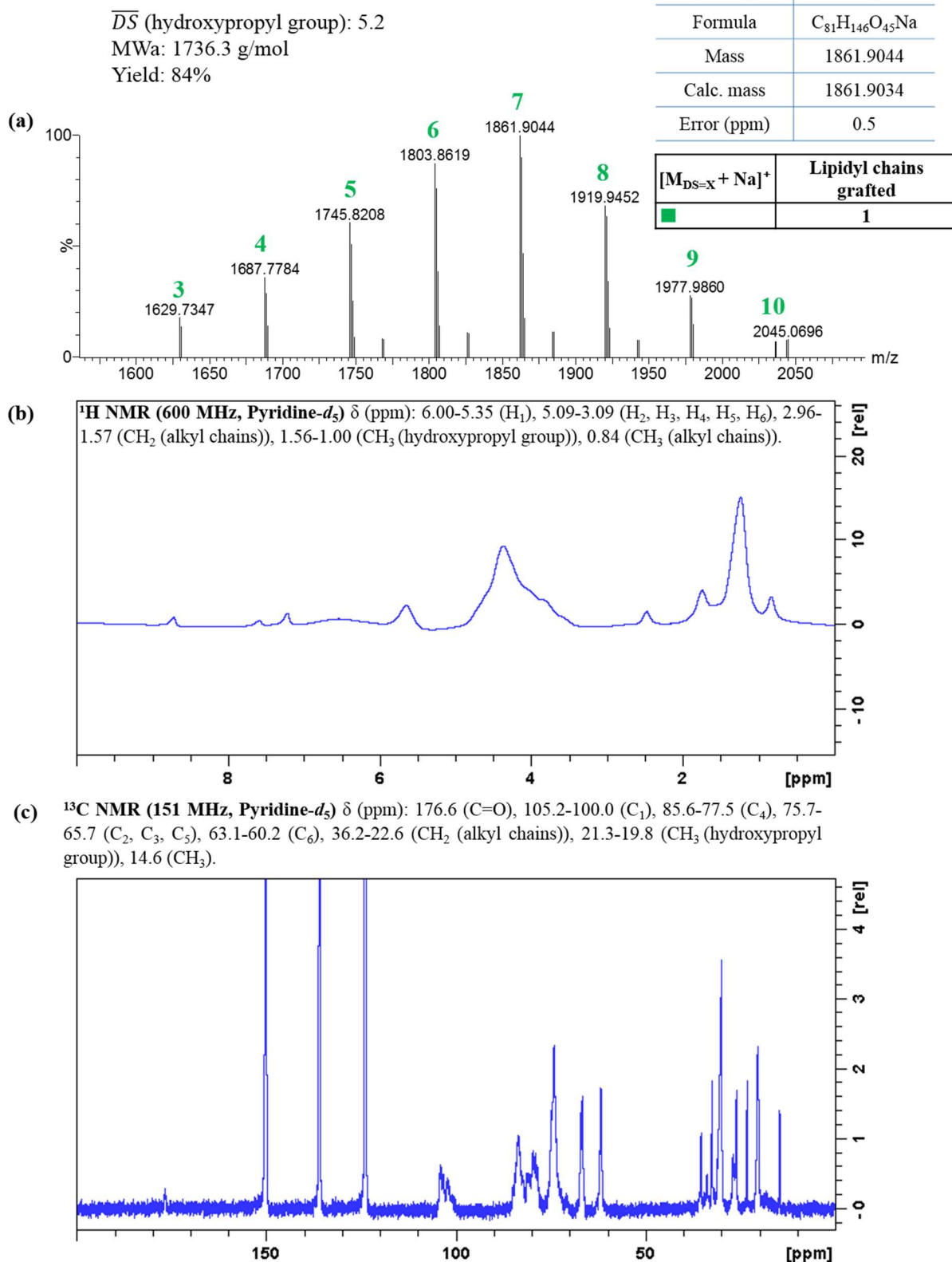
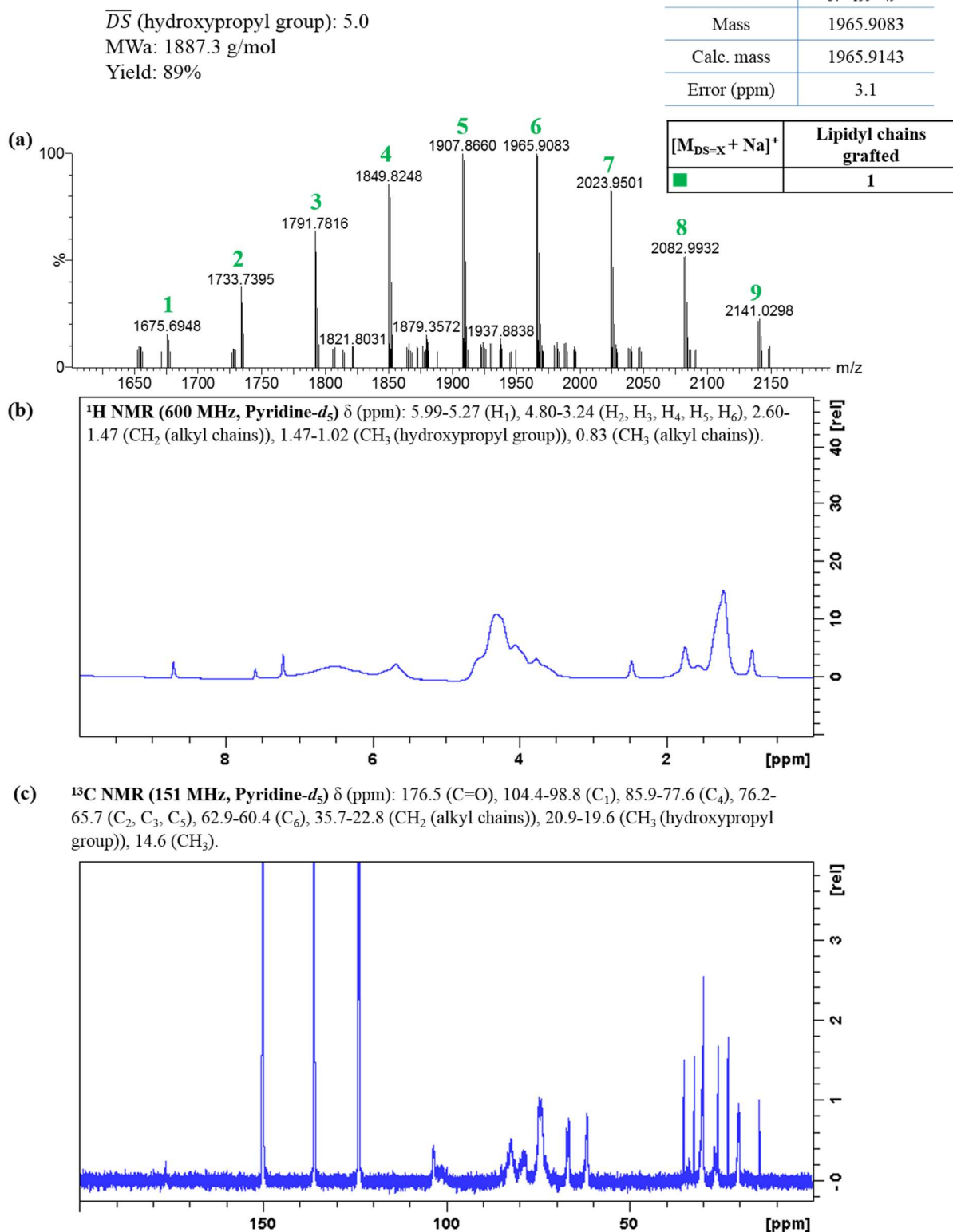


Figure S11: ESI⁺-HRMS (a), ¹H NMR (b) and ¹³C NMR spectra (c) of HP- β -CD(C₉)₂OOH **11**.

Figure S12: ESI⁺-HRMS (a), ¹H NMR (b) and ¹³C NMR spectra (c) of HP- γ -CD(C₉)₂OOH **12**.

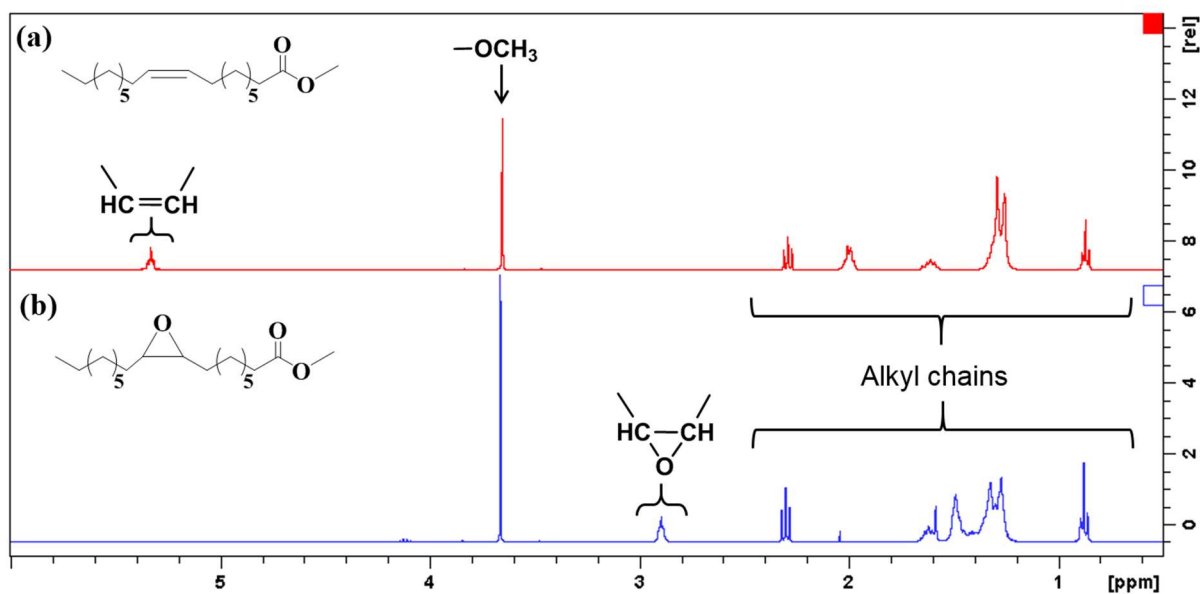


Figure S13: ^1H NMR spectra of methyl oleate **(a)** and of epoxidated methyl oleate **(b)** (400 MHz, CDCl_3 , 298 K).

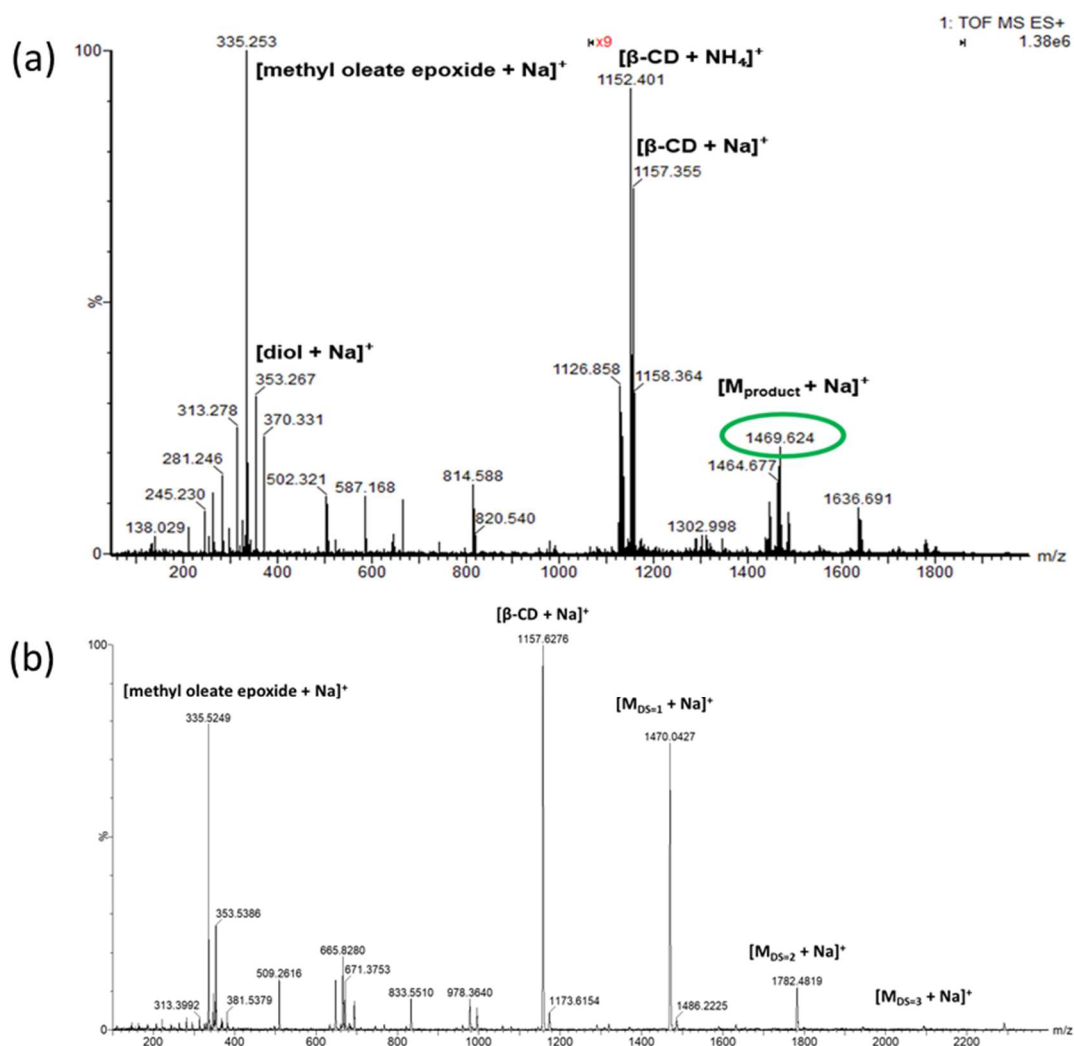


Figure S14: ESI⁺-MS spectra of the ball milling reaction medium of the methyl oleate epoxide opening with the β -CD assisted by APTS (a) and H_2SO_4 (b). The spectra highlight the presence of the expected grafted species and also the starting materials (epoxide, β -CD).

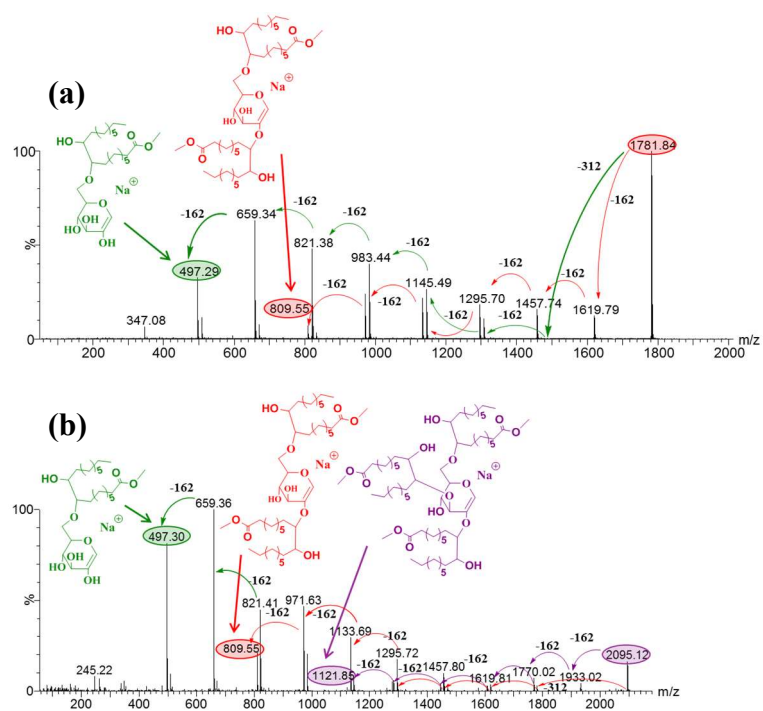


Figure S15: ESI⁺-MS/MS spectra of the [M+Na]⁺ ions of DS=2 **(a)** and DS=3 **(b)** species of β -CD(C₉)₂OOME **1** obtained from ball milling.

$[M_{DS-X} + Na]^+$ (X = number of HP substituents)	Lipidyl chains grafted
■	1
■	2
■	3

$$DS = \frac{1 \times \sum I_1 + 2 \times \sum I_2 + 3 \times \sum I_3}{\sum I_i} = 1.3$$

$$MW_a = MW_{a_{HP-\beta-CD}} + DS \times MW_{epoxide}$$

$$MW_a = 1490.5 + 1,3 \times 312.5 = 1892.5 \text{ g/mol}$$

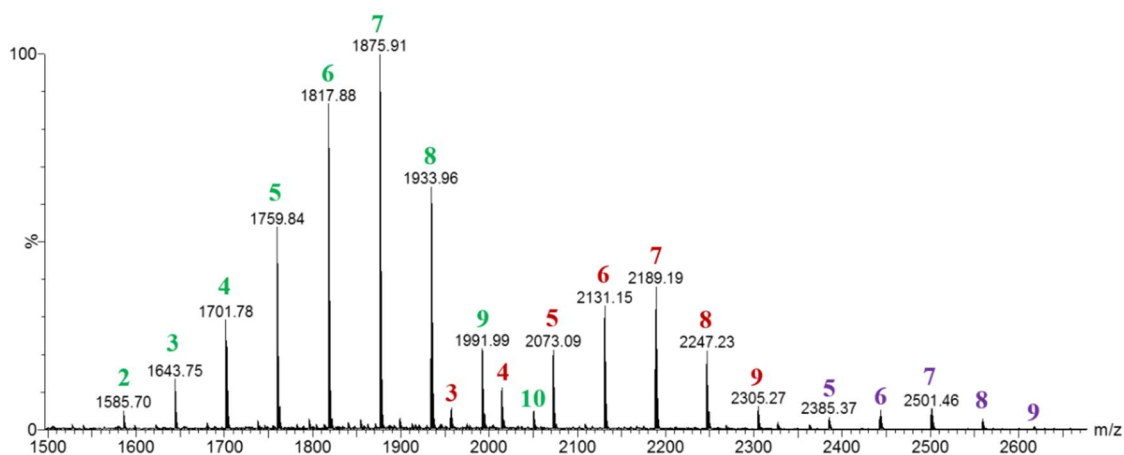


Figure S16: ESI⁺-MS spectrum of the of HPβ-CD(C₉)₂OOME 5 obtained from ball milling, showing mono (green), di (red) and tri (blue) grafted species.

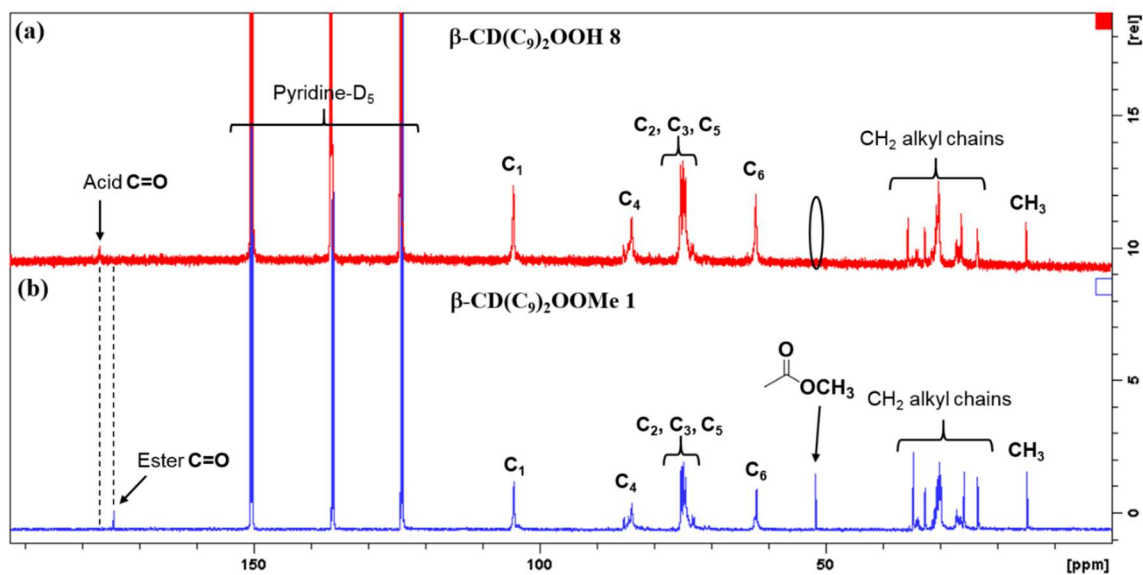


Figure S17: ¹³C NMR spectra of β-CD(C₉)₂OOH 8 (a) and of β-CD(C₉)₂OOME 1 (b) (151 MHz, pyridine-D₅, 298 K).

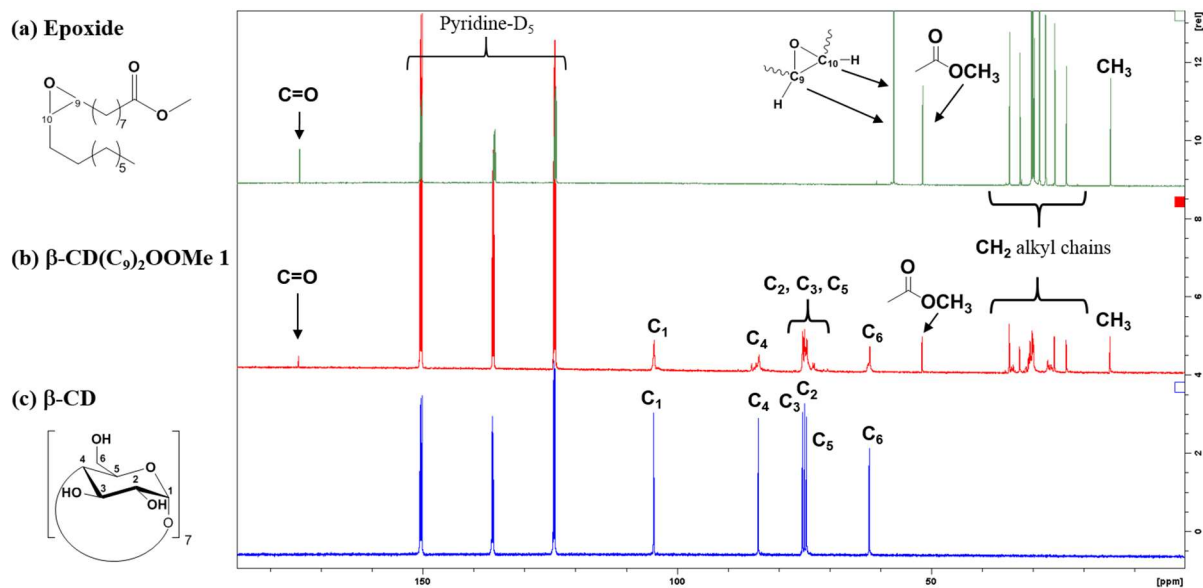


Figure S18: ¹³C NMR spectra of methyl oleate epoxide **(a)**, β -CD(C₉)₂OOME **1 (b)** and β -CD native **(c)** (151 MHz, pyridine-D₅, 298 K).

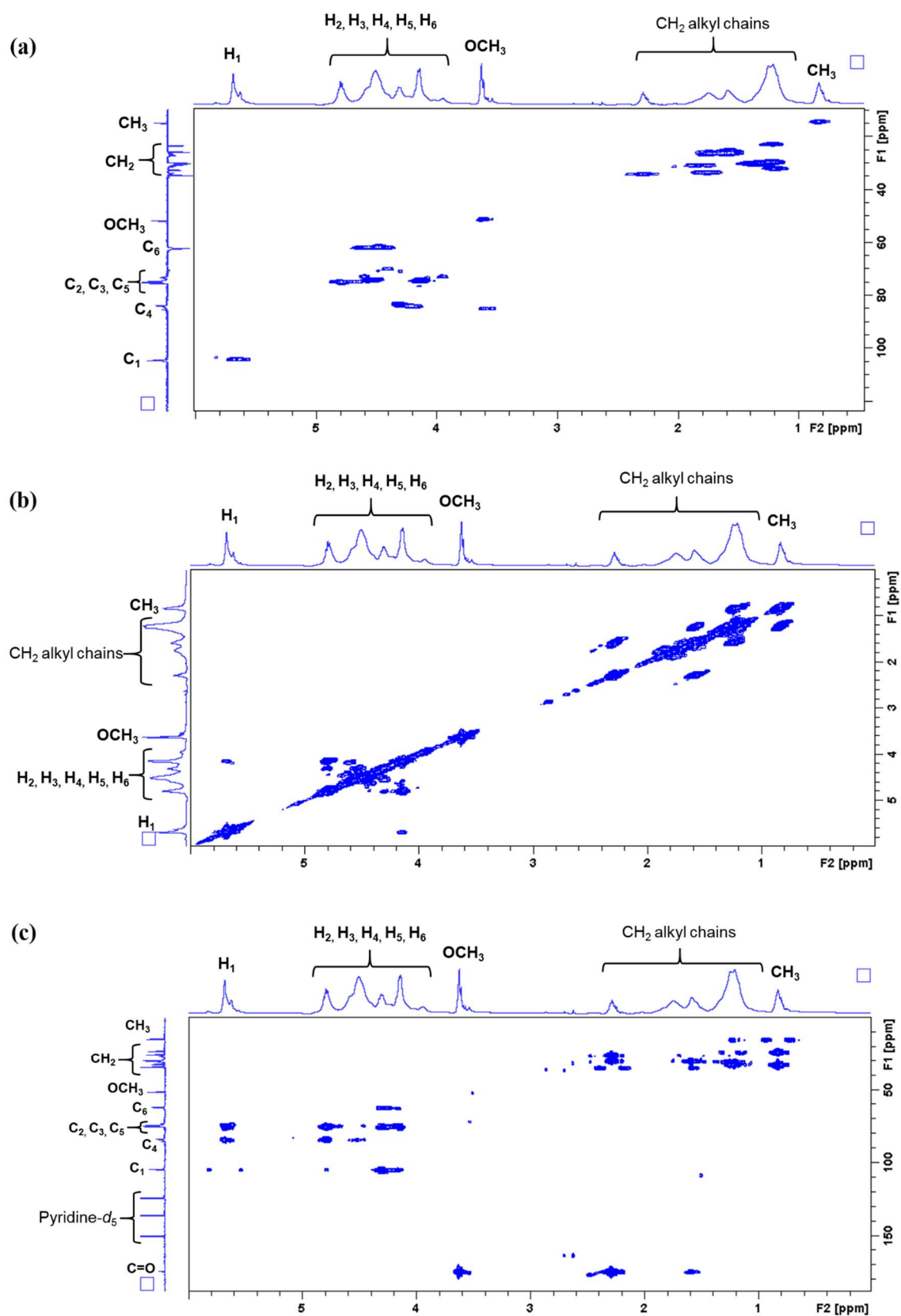
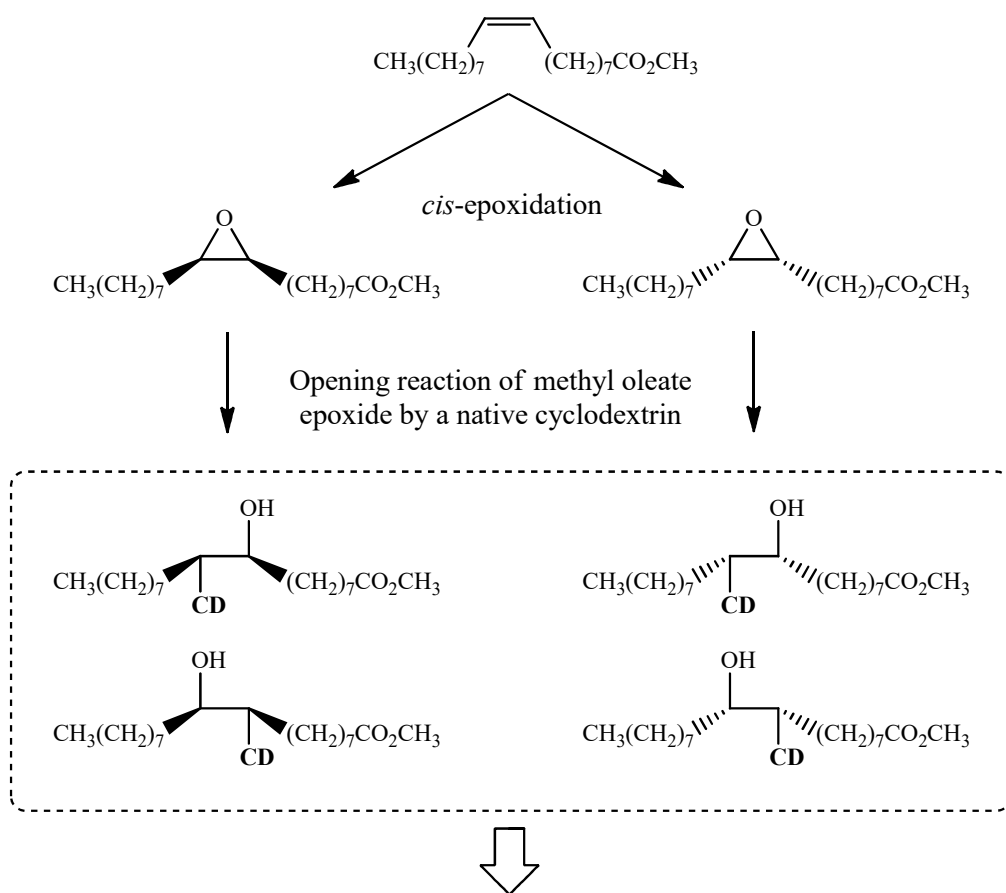


Figure S19: 2D NMR spectra of 1: HSQC (a), COSY (b) and HMBC (c) (600 MHz, pyridine-*D*₅, 298 K).



12 isomers because of the 3 different possibilities for the **CD** moiety:

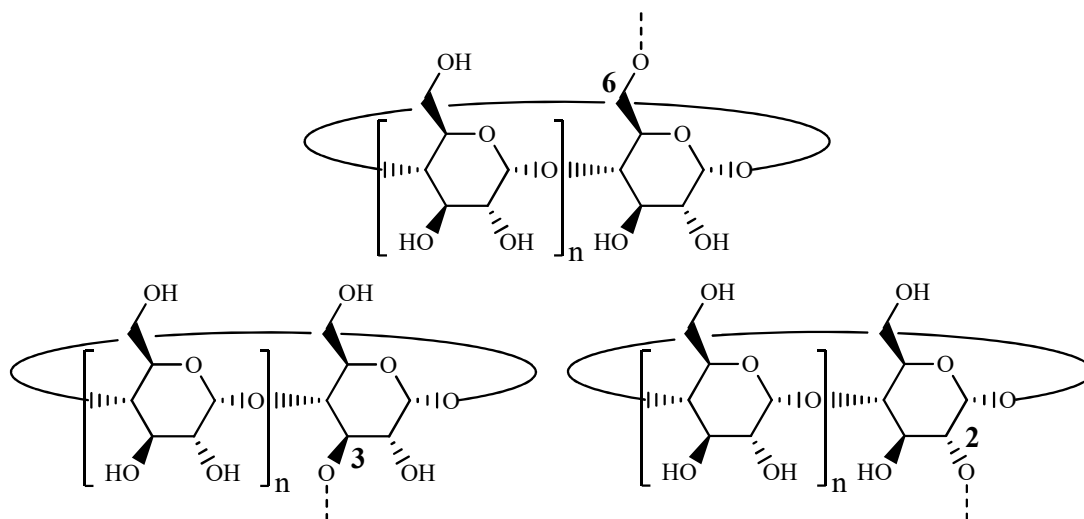
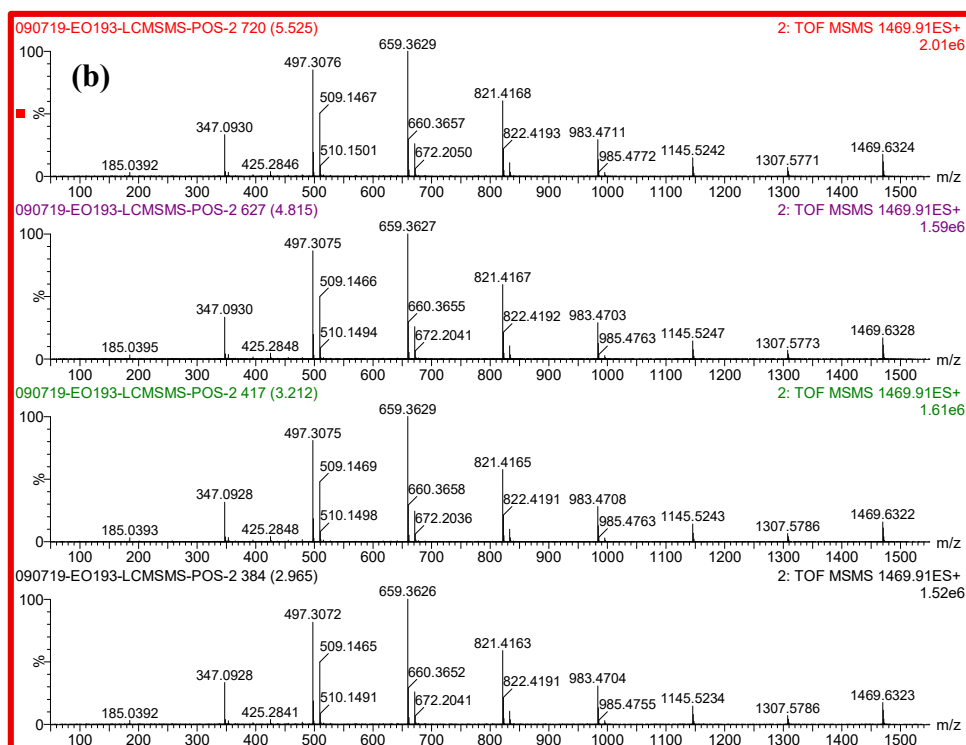
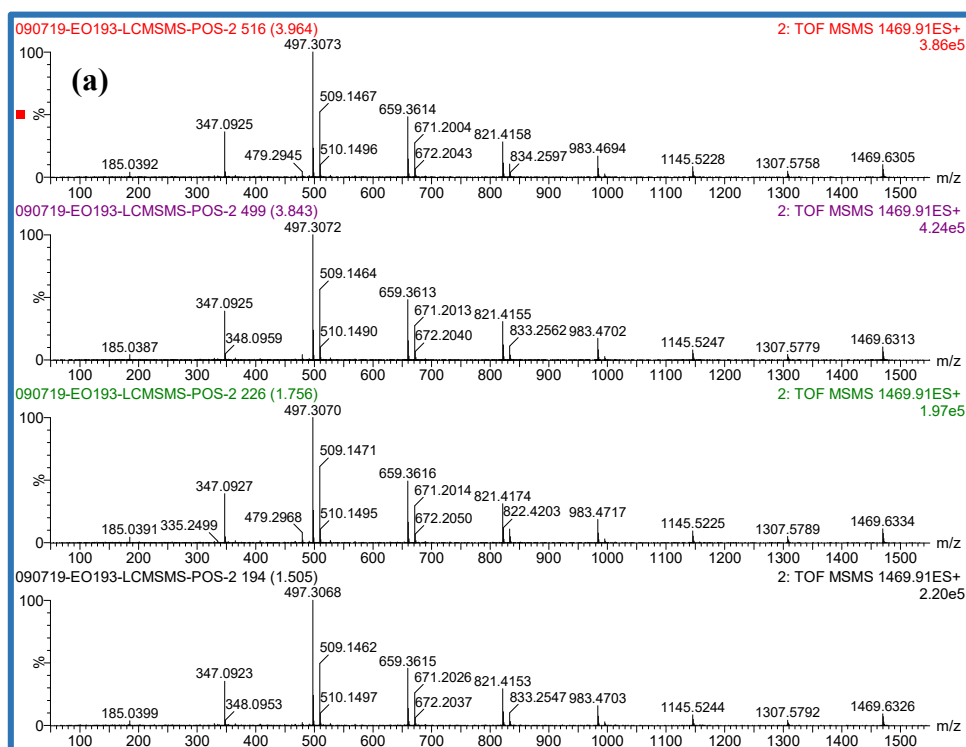


Figure S20: Opening reaction of epoxide by free cyclodextrin.



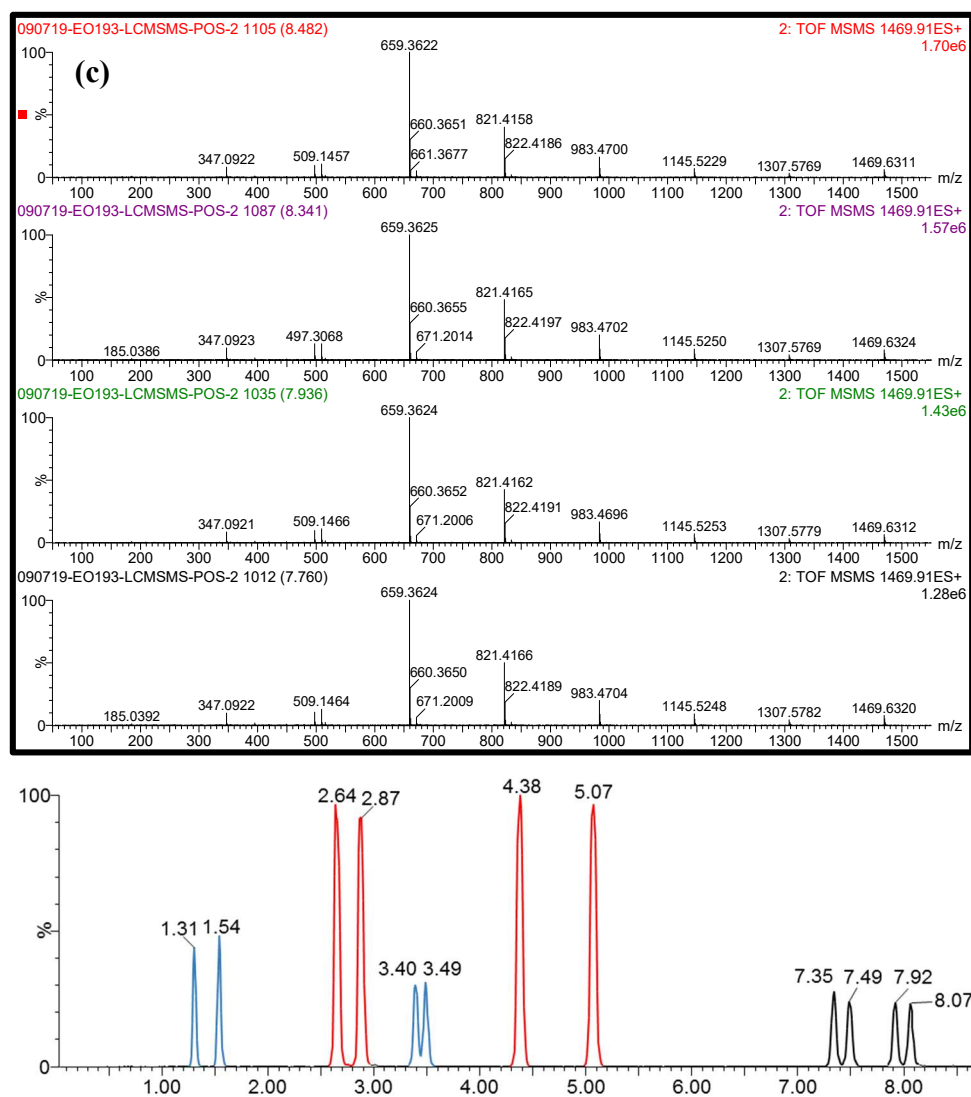


Figure S21: ESI⁺-MS/MS (95 eV) spectra of the twelve DS=1 isomers ([M+Na]⁺ m/z 1469.63) of **1** allowing the distinction of 3 groups of regioisomers. **(a)** Blue (*R*_t 3.49, 3.40, 1.54 and 1.31 min), **(b)** Red (*R*_t 5.07, 4.38, 2.87 and 2.64 min) and **(c)** Black (*R*_t 8.07, 7.92, 7.49 and 7.35 min).

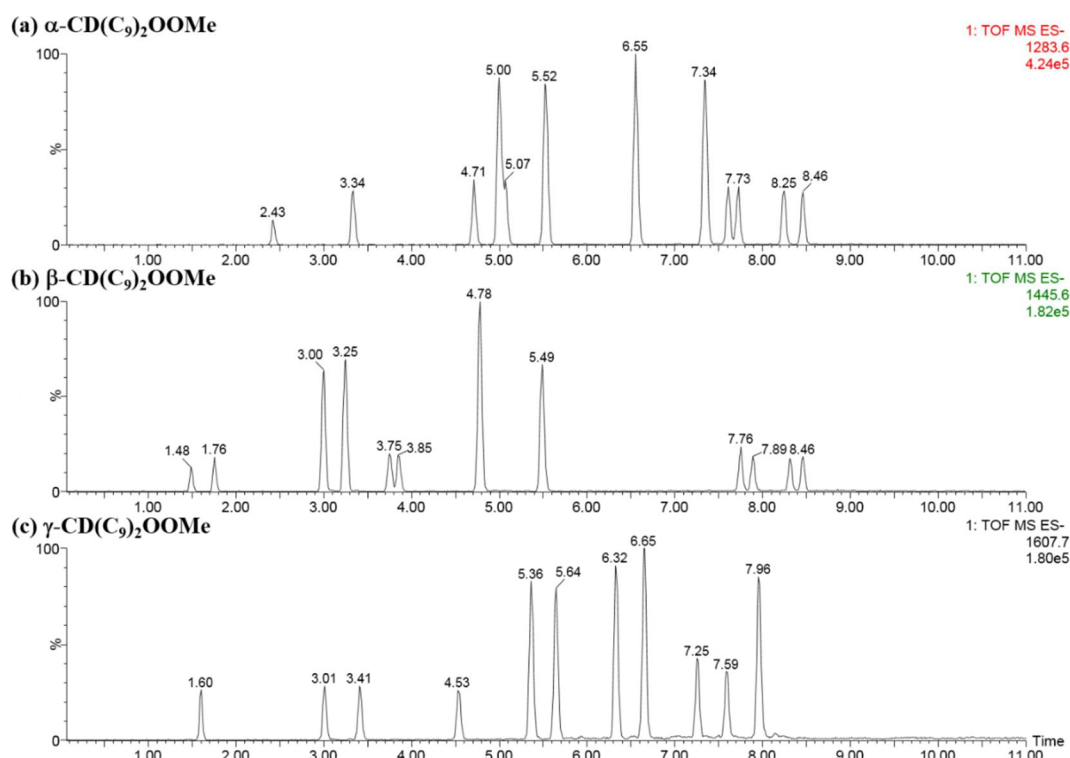


Figure S22: [M-H]⁺ reconstituted ion chromatograms of **2** : DS=1 m/z 1283.58 (**a**) **1** DS=1 m/z 1145.63 (**b**) and **3** DS=1 m/z 1607.68 (**c**).

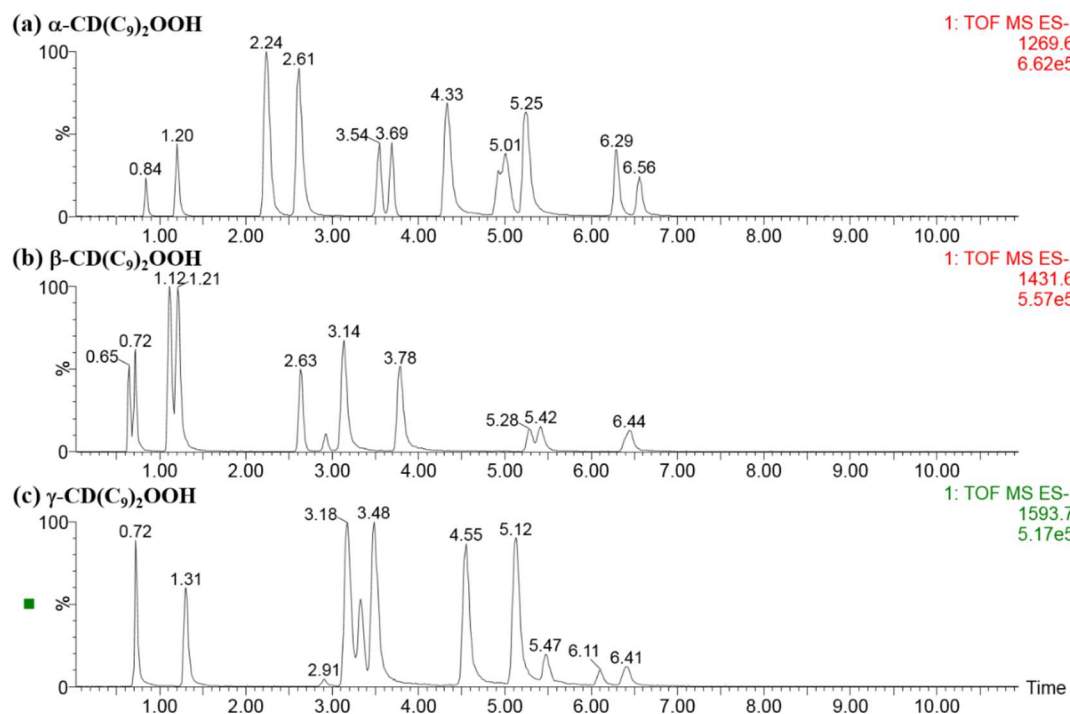


Figure S23 : [M-H]⁺ reconstituted ion chromatograms of **7** : DS=1 m/z 1269.56 (**a**), **8** DS=1 m/z 1431.61 (**b**) and **9** DS=1 m/z 1594.67 (**c**).

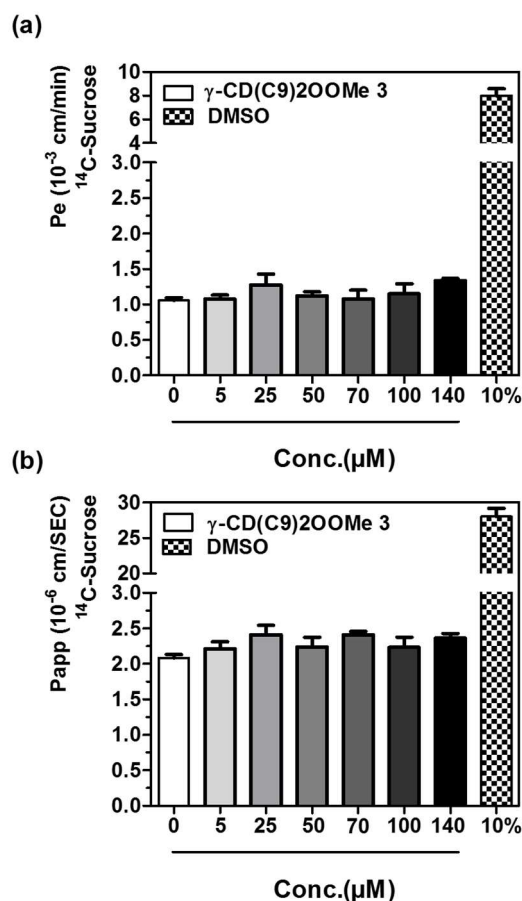


Figure S24: ^{14}C -Sucrose permeability (Pe) and apparent permeability (Papp) assessment in two in vitro models treated with different concentrations of 3. DMSO 10% was used as positive control of biological barrier disruptions. Bars represent the mean of 6 filters + standard error of the mean. (a) represents the Pe of ^{14}C -Sucrose in the BLECs model and (b) represents the Papp of ^{14}C -Sucrose in the intestinal Caco-2 model

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