



Article

Ferritin-Based Single-Electron Devices

Jacqueline A. Labra-Muñoz ^{1,2,*}, Arie de Reuver ², Friso Koeleman ², Martina Huber ¹ and Herre S. J. van der Zant ^{2,*}

- Department of Physics, Huygens-Kamerlingh Onnes Laboratory, Leiden University, Niels Bohrweg 2, 2300 RA Leiden, The Netherlands; huber@physics.leidenuniv.nl
- ² Kavli Institute of Nanoscience, Delft University of Technology, Orentzweg 1, 2628 CJ Delft, The Netherlands; a.r.dereuver@gmail.com (A.d.R.); f.m.koeleman@student.tudelft.nl (F.K.)
- $* \quad Correspondence: j.labramunoz@tudelft.nl \ (J.A.L.-M.); h.s.j.vanderzant@tudelft.nl \ (H.S.J.v.d.Z.)$

Abstract: We report on the fabrication of single-electron devices based on horse-spleen ferritin particles. At low temperatures the current vs. voltage characteristics are stable, enabling the acquisition of reproducible data that establishes the Coulomb blockade as the main transport mechanism through them. Excellent agreement between the experimental data and the Coulomb blockade theory is demonstrated. Single-electron charge transport in ferritin, thus, establishes a route for further characterization of their, e.g., magnetic, properties down to the single-particle level, with prospects for electronic and medical applications.

Keywords: ferritin; Coulomb blockade; single-electron transport; nanoelectronics



Citation: Labra-Muñoz, J.A.; de Reuver, A.; Koeleman, F.; Huber, M.; van der Zant, H.S.J. Ferritin-Based Single-Electron Devices. *Biomolecules* **2022**, *12*, 705. https://doi.org/10.3390/ biom12050705

Academic Editor: Vladimir N. Uversky

Received: 22 April 2022 Accepted: 14 May 2022 Published: 15 May 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/licenses/by/4.0/).

1. Introduction

Ferritin, the iron storage protein, has drawn the attention of the scientific community in recent decades. Its unique structure consists of a spherical shell composed of 24 peptide subunits ([1]). In mammals, two types of subunits are present: a heavy (21 kDa) and a light (19 kDa) chain ([2]). The heavy chain is responsible for the oxidation of Fe(II) to Fe(III). The light chain fosters nucleation and the storage of Fe(III) as a mineral mixture consisting mainly of ferrihydrite, storing iron in a nontoxic form ([1]). Dysfunctional ferritin has been related to neurodegenerative diseases for different reasons. First, it allows unwanted egress of iron from the ferritin, which increases the available amount of free iron that is toxic to cells, causing oxidative stress injury ([1,3]). Second, the ferritin core's composition in the brain of patients affected by these diseases has been found to be altered ([4,5]). The mechanism leading to dysfunctional ferritin remains unclear; therefore, a better understanding of the physical properties of individual ferritin particles is vital.

Published studies on the electrical characterization of ferritin have mostly been performed on ferritin networks and monolayers ([6–23]). Almost 20 years ago, Xu et al. [7] reported that holoferritin networks, i.e., ferritin with iron core, are 5–15 times more conductive than the networks of the iron-depleted version, apoferritin ([7]). Although Xu et al. reported S-shaped current vs. voltage (*IV*) characteristics, other phenomena have been observed, such as switching ([10]) and negative differential resistance ([17]). The charge transport through ferritin monolayers has been studied by Kumar et al. [6], who reported different transport mechanisms including direct tunneling, sequential tunneling, and hopping, depending on the ferritin iron content. Ferritin multilayers have been studied as well ([20]), finding highly correlated electron transport, consistent with electron transport in a quantum dot. Bera et al. [14] reported similar *IV*s for six layers of holo- and apoferritin. Targeted applications range from multilayered gate dielectrics [8], to metal-insulator nanocomposites [13], high-efficiency solar energy conversion [11], bio-nanobatteries [12], to p-n junctions [15], among others. From a medical perspective, Holovchenko et al. [21] showed that the average conductance of Alzheimer's ferritin networks is about two orders

Biomolecules **2022**, 12, 705 2 of 9

of magnitude lower than that of control ferritin, suggesting the use of ferritin resistance measurements as a diagnostic tool.

Studies focused on the electrical characterization of single ferritin are scarce. These studies are based on either scanning tunneling microscopy (STM) ([18,22,24]) or atomic force microscopy (AFM) ([7,25,26]) and are performed at room- or higher temperatures. Single holoferritin was found to be more conductive than apoferritin by AFM ([25]), as found in networks. In contrast, Rakshit et al. [24] obtained similar *IV* characteristics for apo- and holoferritin by STM. In this work, single horse-spleen ferritin particles are studied by trapping them in self-aligned nanogaps and measuring their electronic properties at low temperatures. We find that the devices are unstable at room temperature, but below 100 K they allow for the acquisition of reproducible data that establishes the Coulomb blockade as the main transport mechanism through them.

2. Results

We used commercial horse-spleen ferritin purchased from Sigma Aldrich. Figure 1a shows a schematic representation of ferritin. It consists of a mineral core surrounded by an organic shell with an outer diameter of 12 nm, and a shell thickness of 2 nm, approximately. From the same batch used in the conductance measurements, we determined the size distribution of ferritin cores through transmission electron microscopy (TEM); see Figure S2 for details. The analysis of 1502 particles showed that the cores' size varied from 4 to 8.6 nm, as shown in Figure S2b.

Self-aligned nanogaps were fabricated as reported in the Methods section, following published fabrication routes ([27,28]). Figure 1d shows a scanning electron microscopy (SEM) image of a self-aligned nanogap before ferritin deposition. The distance between the source and drain electrodes (gap length) varied between 8 and 25 nm, while the gap width was $10 \, \mu m$.

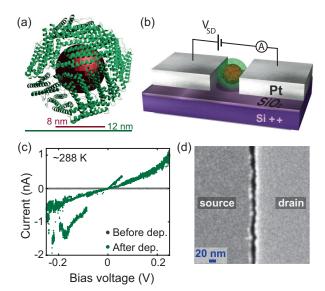


Figure 1. (a) Schematic representation of ferritin, based on the protein data base (PDB) of horse-spleen apoferritin (PDB ID: 2W0O [29]). The organic shell is in green; the mineral core occupying the internal cavity is in red. (b) Schematic circuit of a device containing one ferritin particle. (c) Electrical characterization of device A1 before (grey curve) and after (green curve) ferritin deposition, measured at room temperature, in vacuum. The grey curve indicates an open circuit, reflecting an empty device. The increase in current shown in the green curve indicates the capture of ferritin. In both cases, the current is measured by sweeping the voltage from negative to positive values, followed by sweeping from positive to negative values. (d) Scanning electron microscopy image of an empty device showing the gap of 9–19 nm size between source and drain electrodes.

Biomolecules **2022**. 12, 705 3 of 9

Prior to ferritin deposition, we recorded at room temperature and in a vacuum (10^{-4} mbar) the current vs. voltage (IV) characteristics of each electrode pair; some samples were also checked at helium temperature. Devices were disregarded that exhibited a current greater than the noise floor (2 pA) over the bias voltage range probed (± 1 V and ± 400 mV, for larger (11-25 nm) and smaller gaps (8-13 nm), respectively). Thus, we only selected open gaps for further studies with ferritin particles (see Figure 1c, grey curve).

Deposition was performed by drop-casting 2–4 μL of ferritin solution diluted 200 times (\sim 270 $\mu g/mL$) onto the electrodes, followed by immediate vacuum pumping. Figure 1b shows a schematic representation of a gap after ferritin deposition. We identified the presence of ferritin trapped within the gap by comparing the IV characteristics of the gap before and after deposition, measured in a vacuum. Figure 1c shows representative examples of IV curves, measured before (gray curve) and after (green curve) deposition at room temperature for device A1. After deposition, a clear increase in current was seen, but the IV was unstable with switches between higher and lower conductive states. Furthermore, hysteretic behavior was also often observed while sweeping the bias voltage up and down. This behavior was generally observed for other samples as well.

The ferritin-deposited devices were cooled down to temperatures close to $T=4.2~\rm K$. At these temperatures, the IV characteristics were stable, and the hysteresis was no longer present. Figure S3 summarizes the four behaviors detected after ferritin deposition, at temperatures close to 4.2 K. The behaviors were: highly conductive linear IVs (most likely ferritin aggregates); still open gaps (no ferritin trapped); tunneling; and Coulomb-blockade (CB)-like IVs. Figure S4 shows the reference measurements, using only the buffer solution; after three buffer depositions, no CB-like features were detected.

Here, we focused on analyzing the CB-like IV characteristics, since they may represent devices with one particle trapped (see below). Figure 2 shows two typical CB-like IVs recorded at 4.2 or 5 K, displaying clear step-like features (light blue dots) or a single transport gap centered around zero bias (green dots). For clarity and to facilitate the simulations, these IV curves were either the descendant curves of the IV cycles, i.e., the current recorded from 50 mV to -50 mV, or the ascendant ones. In some cases, abrupt IV changes, especially in conductivity, were observed after thermal, voltaic, and temporal cycling (for details, see Supplementary Materials). An example of this effect is shown in Figure 2; device D1 (Figure 2g) was recorded first, at 4.2 K. The same device was measured at 5 K again (Figure 2a), after warming it up to 250 K. The current increased by three orders of magnitude, and the IV curve showed clear steps.

We used the orthodox Coulomb Blockade (CB) model ([30]) to describe the IV characteristics, measured at the lowest recorded temperature as shown in Figure 2. The model considered single-electron tunneling, without incorporating second-order processes such as tunneling through virtual states and cotunneling. A Matlab script was written with R_1 , R_2 , C_1 , C_2 , Q_0 , and T as the simulation parameters; the parameters are defined in the caption of Table 1. The temperature was taken to be the measured temperature near the sample and was not adjusted in the simulations. The black dashed lines shown in Figure 2 are the CB fits to the data; the simulation parameters are listed in Table 1. The consistency between the data and simulations is visible both in the IV and the differential conductance (dI/dV) plots. The excellent agreement indicates that the dominant transport channel is indeed through a single particle, although three-terminal measurements with a gate are needed to give a conclusive answer.

To further investigate the validity of the CB model, we measured IVs at different temperatures. Figure 3 shows IVs obtained at different temperatures, on device A1. At 4.2 K (blue light dots), the IVs present clear Coulomb-blockade steps. At 22 K (orange dots), steps are no longer visible, and the blockade part of the IVs is linear, i.e., the blockade has been lifted. The black dashed lines are the Coulomb-blockade fits to the data, using the same simulations parameters R_1 , R_2 , C_1 , C_2 , and Q_0 , while adjusting only the temperature.

Biomolecules **2022**, 12, 705 4 of 9

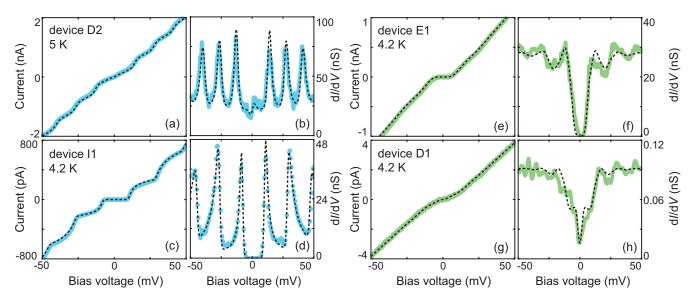


Figure 2. Experimental current—voltage (*IV*) characteristics (colored dots) and the corresponding calculated curves using the orthodox Coulomb blockade model (black dashed lines) acquired on four devices, displaying clear step-like features (light blue dots) or a single transport gap centered around zero bias (green dots). (**a**,**e**,**g**) Experimental *IV*s measured on, respectively, device D2, E1, and D1; obtained by sweeping the voltage from positive to negative values. (**c**) *IV* measured on device I1, obtained by sweeping the voltage from negative to positive values. The corresponding differential conductance is depicted in figures (**b**,**d**,**f**,**h**). The data were recorded at either 4.2 or 5.0 K. The parameters used for the simulations are presented in Table 1.

Table 1. Coulomb-blockade simulation parameters used to generate the four simulated IVs depicted in Figure 2. C_1 and C_2 are the junction capacitances on the left and right sides, respectively. R_1 and R_2 are the tunnel resistances on the left and right sides, respectively. Q_0 is the offset charge, and T is the temperature.

Device	C ₁ (aF)	C ₂ (aF)	R_1 (M Ω)	R_2 (M Ω)	Q ₀ (e)	T (K)
D2	12.0	12.0	0.4	22.0	-0.55	5.0
I1	0.8	8.7	8.6	38.2	-0.06	4.2
E1	9.5	9.2	17.8	17.8	-0.12	4.2
D1	12.0	12.0	5500.0	5500.0	-0.35	4.2

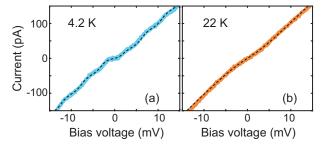


Figure 3. Experimental current–voltage characteristics acquired on device A1, at two different temperatures. (a) Data measured at 4.2 K (light blue dots). (b) Data acquired at 22 K (orange dots). The black dashed lines indicate the calculated curves using the orthodox Coulomb blockade model with parameters: $C_1 = 28$ aF, $C_2 = 34.7$ aF, $R_1 = 0.4$ M Ω , $R_2 = 0.08$ G Ω , $Q_0 = 0.15$ e.

In total, 22 devices displayed Coulomb-blockade like IVs. Figure S5 shows the experimental IVs and the CB fits of all data. The simulation parameters are collected in Table S1. Figure 4 summarizes the total capacitances and resistances used for the simulations. A broad dispersion was found in the parameters: the total capacitance varied over

Biomolecules **2022**, 12, 705 5 of 9

two orders of magnitude from 0.84 to 62.7 aF, while the total resistance varied over four orders of magnitude from 0.02 G Ω to 72 G Ω .

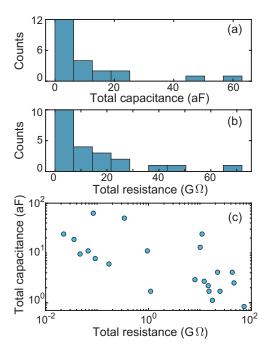


Figure 4. Total capacitances ($C = C_1 + C_2$) and total resistances ($R = R_1 + R_2$) calculated with the orthodox Coulomb blockade model for the 22 devices showing Coulomb-blockade at temperatures below 100 K. Table S1 contains the entire set of Coulomb blockade parameters used to model each device, at the lowest recorded temperature. (**a**) Total capacitance histogram. (**b**) Total resistance histogram. (**c**) Total capacitance plotted against the total resistance.

3. Discussion

To place the values of the fit parameters in a broader context, we compared them with values found in other experiments. For ferritin, a resistance of 2 G Ω was reported ([24]) by STM, but, a much higher resistance was reported by AFM (25 G Ω –1.8 T Ω) ([7,26,31]). Our experimental resistances were closer to that reported by STM and to the lower range obtained by AFM. Capacitance values have not been reported for single ferritin particles in the literature.

We note that the measured core distribution can lead to a sizable dispersion in the capacitances/resistances but cannot fully account for the observed dispersion in Table S1 and Figure 4. However, different geometrical arrangements are discussed below, which reproduce this broad dispersion. Figure 4c indicates that high capacitances were found for low resistances. This trend is consistent with geometrical considerations in which good electrical contacts (low resistances) imply large electrode-particle capacitances. More specifically, the particle could lie symmetrically in between the electrodes (case I; see Figure S7) or on top of them (case II). If the gap between the electrode is larger than the ferritin size, the particle could couple asymmetrically to the electrode if one side of the particle makes direct contact with one of the electrodes, while there is some space in between the particle and the other electrode. This results in two additional cases to be considered: asymmetric contact with the particle laying completely between the electrodes (case III) or on top of them (case IV).

For these four different cases we calculated the capacitances using a parallel plate model, by considering ϵ_r to be between 10 and 20 [32,33], see Supplementary Materials for details. The estimates are shown in Supplementary Materials, and the minimum estimate for the total capacitance (0.4 aF) was in agreement with the minimum experimental capacitance (0.84 aF). The maximum estimated capacitance (11.3 aF) was about five times lower than the maximum experimental capacitance (62.7 aF), although only two capacitances

Biomolecules **2022**, 12, 705 6 of 9

were found to be larger than 24 aF. Ways to account for this difference could be to consider a barrier thickness thinner than 2 nm or a higher dielectric constant.

Within the CB model, the total resistance (R) of an individual ferritin particle in a gap between source and drain contacts, comprised two tunnel resistances in series, from the ferritin core to each electrode, i.e., $R = R_1 + R_2$. Each of these resistances can be expressed as $R_{1,2} = R_0 \exp(\beta d)$, where R_0 is an effective contact resistance, d is the thickness of the dielectric, and β is the exponential distance decay factor. For β we take 0.26 A^{-1} , which is the average reported value for proteins ([34]).

The calculated resistances in this model are again listed in the SI. We found that the maximum experimental resistance (72 G Ω) was within the range of our estimates (0.17–126.55 G Ω). The minimum experimental resistance (0.02 G Ω) was somewhat lower than our minimum estimate. A possible reason is that β is smaller than what we assumed for ferritin. For example, if we consider $\beta = 0.21 \,\text{Å}^{-1}$, the minimum resistance (case I) is 0.06 G Ω , which is on the same order of magnitude as the smaller experimental resistance.

In conclusion, we demonstrated that individual ferritin particles can be trapped in self-aligned nanogaps. The observed instability at room temperature was suppressed at low temperature, and clear Coulomb-blockade effects were observed including Coulomb staircases. Excellent fits of the current–voltage characteristics to the Coulomb blockade model strongly indicate that transport is through an individual particle. Estimates based on different arrangements of the particle in between the electrodes and reported values for transport through proteins reproduced the capacitance and resistance values determined from the CB model. Thus, the experiments showed that ferritin particles act as a model CB system, and in a next step, it would be interesting to relate the transport characteristics to specific ferritin properties. This, for example, includes a better understanding of the unstable behavior at elevated temperatures and of the magnetic properties of the core. The latter can be studied using a three-terminal device with a gate in combination with magnetic field measurements.

4. Methods

4.1. Ferritin Purity Assessment

Commercial horse-spleen ferritin purchased from Sigma Aldrich (Cat. No. 270-40, Lot: 08E1805) of 54 mg/mL protein concentration was used with no further purification. For the conductance measurements, the ferritin solution was diluted by a factor of 200 using a buffer solution (0.15 M NaCl and 10 mM Tris, pH 8.0, 0.1% sodium azide). The final solution had a protein concentration of 270 μ g/mL with >95% purity as assessed by sodium dodecyl sulfate–polyacrylamide gel electrophoresis (Supplementary Materials, Figure S1c). The homogeneity of the solution was assessed by the dynamic light scattering technique (DLS), which is reported in Supplementary Materials (Figure S1a,b).

4.2. Ferritin Size Statistics

The ferritin mineral cores were examined without further staining in a JEM-1400 transmission electron microscope (TEM). To further inspect the protein shell structure, uranyl acetate (2% solution) was used. For more details, refer to Supplementary Materials (Figure S2). A home-made Matlab script was used for particle detection and size statistics.

4.3. Device Fabrication

The devices were fabricated as follows. On top of a doped Si/SiO_2 substrate, the first electrode was defined by e-beam lithography (EBL) and evaporation of 5 nm of titanium (adhesive layer) and subsequently 25 nm of platinum. On top of the platinum layer, a 20 nm chromium layer was deposited. Upon oxidation chromium expands its size. In this manner, chromium oxide acted as a shadow mask of a few nanometers near the edge of the first electrode. The thickness of the chromium layer determined the expansion and, thus, the size of the gap. A second EBL cycle defined the second electrodes, by depositing 5 nm

Biomolecules **2022**. 12, 705 7 of 9

of titanium and 20 nm of platinum. In the final step, the chromium layer was etched away (wet-etch step) to reveal the underlying nanogaps.

4.4. Electrical Measurements

The electrical measurements of most of the devices were performed in a variable temperature insert (VTI) with TU Delft home-built low-noise electronics. The noise was about 1–2 pA at room temperature and below 1 pA at 4.2 K. A 1-K pot allowed measurements below 4.2 K for devices J1, J2, and K1. Device H1 was measured in a dilution refrigerator equipped with the same TUDelft home-built electronics and a similar noise level.

4.5. Coulomb-Blockade Fits

The *IV*s obtained showing CB-like features were fitted to the CB orthodox theory, using a home-made Matlab script that is available upon request. The optimal fit was determined by the user, i.e., by visual inspection.

Supplementary Materials: See the Supplementary Materials for more details of this study regarding the ferritin purity assessment and size statistics, reference measurements, all data fitted to the CB model, charge-offset variations, and estimates of the capacitances and resistances. The following supporting information can be downloaded at: https://www.mdpi.com/article/10.3390/biom120 50705/s1, Figure S1: Purity assessment of horse-spleen ferritin; Figure S2: TEM characterization of horse-spleen ferritin; Figure S3: *IV* types at low temperatures; Figure S4: Reference measurements; Figure S5: Low temperature data and CB fits; Figure S6: Charge offset variations; Table S1: CB parameters used for the simulations of 22 devices.

Author Contributions: Conceptualization, J.A.L.-M. and H.S.J.v.d.Z.; data curation, J.A.L.-M., A.d.R. and F.K.; formal analysis, J.A.L.-M., A.d.R. and F.K.; funding acquisition, M.H. and H.S.J.v.d.Z.; investigation, J.A.L.-M. and H.S.J.v.d.Z.; methodology, J.A.L.-M. and A.d.R.; project administration, J.A.L.-M. and H.S.J.v.d.Z.; software, J.A.L.-M. and A.d.R.; supervision, J.A.L.-M. and H.S.J.v.d.Z.; visualization, J.A.L.-M. and H.S.J.v.d.Z.; writing—original draft, J.A.L.-M.; writing—review and editing, J.A.L.-M., M.H. and H.S.J.v.d.Z. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by NWO, through Nanofront project number NF17SYN05.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Data supporting the findings of this paper are available in 4TU.ResearchData, see doi:10.4121/19306865.

Acknowledgments: The authors thank Damian Bouwmeester for their assistance mounting the setup and electronics, as well as valuable discussions. Greatly appreciated is the financial support provided by NWO, through Nanofront project number NF17SYN05. We thank Lionel Ndamba for their contribution concerning the protein-purity characterization.

Conflicts of Interest: The authors declare no competing interests.

References

- 1. Arosio, P.; Elia, L.; Poli, M. Ferritin, cellular iron storage and regulation. *IUBMB Life* 2017, 69, 414–422. https://doi.org/10.1002/iub.1621.
- 2. Finazzi, D.; Arosio, P. Biology of ferritin in mammals: An update on iron storage, oxidative damage and neurodegeneration. *Arch. Toxicol.* **2014**, *88*, 1787–1802. https://doi.org/10.1007/s00204-014-1329-0.
- 3. Friedman, A.; Arosio, P.; Finazzi, D.; Koziorowski, D.; Galazka-Friedman, J. Ferritin as an important player in neurodegeneration. *Park. Relat. Disord.* **2011**, *17*, 423–430. https://doi.org/10.1016/j.parkreldis.2011.03.016.
- 4. Quintana, C.; Bellefqih, S.; Laval, J.; Guerquin-Kern, J.; Wu, T.; Avila, J.; Ferrer, I.; Arranz, R.; Patiño, C. Study of the localization of iron, ferritin, and hemosiderin in Alzheimer's disease hippocampus by analytical microscopy at the subcellular level. *J. Struct. Biol.* **2006**, *153*, 42–54. https://doi.org/https://doi.org/10.1016/j.jsb.2005.11.001.
- 5. Quintana, C.; Gutiérrez, L. Could a dysfunction of ferritin be a determinant factor in the aetiology of some neurodegenerative diseases? *Biochim. Biophys. Acta* (*BBA*)—*Gen. Subj.* **2010**, 1800, 770–782. https://doi.org/https://doi.org/10.1016/j.bbagen.2010.04.012.

Biomolecules **2022**, 12, 705 8 of 9

6. Kumar, K.S.; Pasula, R.R.; Lim, S.; Nijhuis, C.A. Long-Range Tunneling Processes across Ferritin-Based Junctions. *Adv. Mater.* **2016**, *28*, 1824–1830. https://doi.org/https://doi.org/10.1002/adma.201504402.

- 7. Xu, D.; Watt, G.D.; Harb, J.N.; Davis, R.C. Electrical Conductivity of Ferritin Proteins by Conductive AFM. *Nano Lett.* **2005**, 5,571–577. https://doi.org/10.1021/nl048218x.
- 8. Kim, B.J.; Ko, Y.; Cho, J.H.; Cho, J. Organic Field-Effect Transistor Memory Devices Using Discrete Ferritin Nanoparticle-Based Gate Dielectrics. *Small* **2013**, *9*, 3784–3791. https://doi.org/https://doi.org/10.1002/smll.201300522.
- 9. Yamamoto, S.I.; Kobayashi, K.; Yamada, H.; Yoshioka, H.; Uraoka, Y.; Fuyuki, T.; Yamashita, I. Electrical characterisitics of ferritin cores investegated by Kelvin Probe force microscopy. *J. Phys. Conf. Ser.* **2008**, *100*, 052004. https://doi.org/10.1088/1742-6596/100/5/052004.
- 10. Zhang, C.; Shang, J.; Xue, W.; Tan, H.; Pan, L.; Yang, X.; Guo, S.; Hao, J.; Liu, G.; Li, R.W. Convertible resistive switching characteristics between memory switching and threshold switching in a single ferritin-based memristor. *Chem. Commun.* **2016**, 52, 4828–4831. https://doi.org/10.1039/C6CC00989A.
- 11. Erickson, S.; Smith, T.; Moses, L.; Watt, R.; Colton, J. Non-native Co-, Mn-, and Ti-oxyhydroxide nanocrystals in ferritin for high efficiency solar energy conversion. *Nanotechnology* **2015**, *26*, 015703. https://doi.org/10.1088/0957-4484/26/1/015703.
- 12. Watt, G.; Kim, J.W.; Zhang, B.; Miller, T.; Harb, J.; Davis, R.; Choi, S. A Protein-Based Ferritin Bio-Nanobattery. *J. Nanotechnol.* **2012**, 2012, 516309. https://doi.org/10.1155/2012/516309.
- 13. Bera, S.; Kolay, J.; Banerjee, S.; Mukhopadhyay, R. Nanoscale On-Silico Electron Transport via Ferritins. *Langmuir* **2017**, 33, 1951–1958, https://doi.org/10.1021/acs.langmuir.6b04120.
- 14. Bera, S.; Kolay, J.; Pramanik, P.; Bhattacharyya, A.; Mukhopadhyay, R. Long-range Solid-state Electron Transport through Ferritin Multilayers. *J. Mater. Chem. C* **2019**, *7*, 9038–9048. https://doi.org/10.1039/C9TC01744E.
- 15. Mulyana, Y.; Uenuma, M.; Okamoto, N.; Ishikawa, Y.; Yamashita, I.; Uraoka, Y. Creating Reversible p–n Junction on Graphene through Ferritin Adsorption. *ACS Appl. Mater. Interfaces* **2016**, *8*, 8192–8200. https://doi.org/10.1021/acsami.5b12226.
- 16. Shin, H.J.; Shin, K.M.; Lee, J.W.; Kwon, C.H.; Lee, S.H.; Kim, S.I.; Jeon, J.H.; Kim, S.J. Electrocatalytic characteristics of electrodes based on ferritin/carbon nanotube composites for biofuel cells. *Sens. Actuators B Chem.* **2011**, *160*, 384–388. https://doi.org/https://doi.org/10.1016/j.snb.2011.07.066.
- 17. Kolay, J.; Bera, S.; Rakshit, T.; Mukhopadhyay, R. Negative Differential Resistance Behavior of the Iron Storage Protein Ferritin. *Langmuir* **2018**, 34, 3126–3135. https://doi.org/10.1021/acs.langmuir.7b04356.
- 18. Rakshit, T.; Mukhopadhyay, R. Tuning Band Gap of Holoferritin by Metal Core Reconstitution with Cu, Co, and Mn. *Langmuir* **2011**, 27, 9681–9686. https://doi.org/10.1021/la202045a.
- 19. Rakshit, T.; Bera, S.; Kolay, J.; Mukhopadhyay, R. Nanoscale solid-state electron transport via ferritin: Implications in molecular bioelectronics. *Nano-Struct. Nano-Objects* **2020**, 24, 100582. https://doi.org/https://doi.org/10.1016/j.nanoso.2020.100582.
- 20. Rourk, C.; Huang, Y.; Chen, M.; Shen, C. Indication of Strongly Correlated Electron Transport and Mott Insulator in Disordered Multilayer Ferritin Structures (DMFS). *Materials* **2021**, *14*, 4527. https://doi.org/10.3390/ma14164527.
- 21. Holovchenko, A. Electrical Characterization of Nanoparticle and Protein Networks. Ph.D. Thesis, Delft University of Technology, Delft, The Netherlands, 2017. https://doi.org/https://doi.org/10.4233/uuid:4b1f83ad-df5c-4f7f-9dde-d3ea2a6b79ec.
- 22. Preisinger, M.; Krispin, M.; Rudolf, T.; Horn, S.; Strongin, D. Electronic structure of nanoscale iron oxide particles measured by scanning tunneling and photoelectron spectroscopies. *Phys. Rev. B* **2005**, *71*, 165409. https://doi.org/10.1103/PhysRevB.71.165409.
- 23. Kumagai, S.; Yoshii, S.; Matsukawa, N.; Nishio, K.; Tsukamoto, R.; Yamashita, I. Self-aligned placement of biologically synthesized Coulomb islands within nanogap electrodes for single electron transistor. *Appl. Phys. Lett.* **2009**, *94*, 083103. https://doi.org/10.1063/1.3085767.
- 24. Rakshit, T.; Banerjee, S.; Mukhopadhyay, R. Near-Metallic Behavior of Warm Holoferritin Molecules on a Gold(111) Surface. *Langmuir* **2010**, *26*, 16005–16012. https://doi.org/10.1021/la101776m.
- 25. Rakshit, T. Differentiating Holo- and Apoferritin by AFM approach. J. Surf. Sci. Technol. 2015, 31, 165–170.
- 26. Choi, S.H.; Kim, J.W.; Chu, S.H.; Park, Y.; King, G.C.; Lillehei, P.T.; Kim, S.J.; Elliott, J.R. Ferritin-templated quantum dots for quantum logic gates. In *Smart Structures and Materials 2005: Smart Electronics, MEMS, BioMEMS, and Nanotechnology*; Varadan, V.K., Ed.; International Society for Optics and Photonics, SPIE: Bellingham, WA, USA, 2005; Volume 5763, pp. 213–232. https://doi.org/10.1117/12.625319.
- 27. Fursina, A.; Lee, S.; Sofin, R.G.S.; Shvets, I.V.; Natelson, D. Nanogaps with very large aspect ratios for electrical measurements. *Appl. Phys. Lett.* **2008**, 92, 113102. https://doi.org/10.1063/1.2895644.
- 28. Labra-Muñoz, J.; Konstantinović, Z.; Balcells, L.; Pomar, A.; van der Zant, H.S.J.; Dulić, D. Trapping and electrical characterization of single core/shell iron-based nanoparticles in self-aligned nanogaps. *Appl. Phys. Lett.* **2019**, *115*, 063104. https://doi.org/10.1063/1.5094352.
- 29. de Val, N.; Declercq, J.P.; Lim, C.K.; Crichton, R.R. Structural analysis of haemin demetallation by L-chain apoferritins. *J. Inorg. Biochem.* **2012**, 112, 77–84. https://doi.org/https://doi.org/10.1016/j.jinorgbio.2012.02.031.
- 30. Claeson, T.; Delsing, P.; Haviland, D.; Kuzmin, L.; Likharev, K.K., Correlated Single Electron Tunneling In Ultrasmall Junctions. In *Nonlinear Superconductive Electronics and Josephson Devices*; Costabile, G., Pagano, S., Pedersen, N.F., Russo, M., Eds.; Springer: Boston, MA, USA, 1991; pp. 197–228. https://doi.org/10.1007/978-1-4615-3852-3_16.
- 31. Axford, D.; Davis, J. Electron flux through apo-and holoferritin. *Nanotechnology* **2007**, *18*, 145502. https://doi.org/10.1088/0957-4484/18/14/145502.

Biomolecules **2022**, 12, 705 9 of 9

32. Laghaei, R.; Kowallis, W.; Evans, D.G.; Coalson, R.D. Calculation of Iron Transport through Human H-chain Ferritin. *J. Phys. Chem. A* **2014**, *118*, 7442–7453. https://doi.org/https://doi.org/10.1021/jp500198u.

- 33. Li, L.; Li, C.; Zhang, Z.; Alexov, E. On the Dielectric "Constant" of Proteins: Smooth Dielectric Function for Macromolecular Modeling and Its Implementation in DelPhi. *J. Chem. Theory Comput.* **2013**, *9*, 2126–2136. https://doi.org/10.1021/ct400065j.
- 34. Amdursky, N.; Marchak, D.; Sepunaru, L.; Pecht, I.; Sheves, M.; Cahen, D. Electronic Transport via Proteins. *Adv. Mater.* **2014**, 26, 7142–7161. https://doi.org/https://doi.org/10.1002/adma.201402304.