



HIV-HBV Coinfection—Current Challenges for Virologic Monitoring

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Abstract: HIV-HBV coinfected patients have higher rates of liver-related morbidity, hospitalizations, and mortality compared to HBV or HIV mono-infected ones. Clinical studies have shown an accelerated progression of liver fibrosis and an increased incidence of HCC, resulting from the combined action of HBV replication, immune-mediated hepatocytolysis, and HIV-induced immunosuppression and immunosenescence. Antiviral therapy based on dually active antiretrovirals is highly efficient, but late initiation, global disparities in accessibility, suboptimal regimens, and adherence issues may limit its impact on the development of end-stage liver disease. In this paper, we review the mechanisms of liver injuries in HIV-HBV coinfected patients and the novel biomarkers that can be used for treatment monitoring in HIV-HBV coinfected persons: markers that assess viral suppression, markers for liver fibrosis evaluation, and predictors of oncogenesis.

Keywords: HIV-HBV co-infection; public health; pathogenesis; biomarkers; RT-PCR; ddPCR; antiretroviral treatment

1. Introduction

The World Health Organization's sustainable development goals for 2030 aim to end the epidemic of AIDS, combat hepatitis and other communicable and sexually transmitted diseases by 2030 [1]. Coinfection with Human Immunodeficiency Virus (HIV) and hepatitis B virus (HBV) is a global public health problem, with a more severe outcome than HBV or HIV mono-infections, including an increased risk for liver-related morbidity and mortality [2]. The rates of significant clinical events, liver-related hospitalizations, as well as the incidence of hepatocellular carcinoma (HCC) are much higher in HIV-HBV coinfected patients than in HBV or HIV mono-infected ones, and liver mortality is still one of the leading causes of non-AIDS deaths in people living with HIV/AIDS (PLWH), including very young ones [2]. Various mathematical modeling studies using ordinary differential equation models, fractional order models, approaches based on modern evolutionary computational techniques, and Padé approximation have been used to improve the understanding of HIV and HBV viral dynamics and to predict clinical evolution and response to treatment [3–5].

Worldwide, it is estimated that 5–20% of PLWH are also chronically infected with HBV [6]. Extremely high prevalence rates of HBV coinfections in PLWH are reported in Central and Western Africa (median: 16.4%), as compared to very low prevalence rates reported in Eastern Asia (China: 0.4%) [7]. Both viruses have similar routes of transmission and can be acquired via parenteral, sexual, or materno-fetal (mainly perinatal) ways. It is estimated that 25% of PLWH can acquire HBV during adulthood (a pattern seen in



Citation: Ruta, S.; Grecu, L.; Iacob, D.; Cernescu, C.; Sultana, C. HIV-HBV Coinfection—Current Challenges for Virologic Monitoring. *Biomedicines* 2023, 11, 1306. https://doi.org/ 10.3390/biomedicines11051306

Academic Editor: Vincent Vieillard

Received: 21 March 2023 Revised: 21 April 2023 Accepted: 25 April 2023 Published: 28 April 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). areas with low HBV prevalence, such as the USA or Western Europe [8]), while 50–90% are co-infected at birth or in early childhood [2] (a pattern frequently reported in Africa and in countries from Eastern Europe before the introduction of universal immunization programs [2,6]).

Recent data report high global HIV-HBV coinfection rates in several vulnerable groups: people who inject drugs (11.8%), men having sex with men (6.1%), commercial sex workers, and heterosexual adults (6.1%) [7]. There are significant differences according to the geographic area [7,9–17], with spots of high prevalence (for men who have sex with men—Germany in Europe [18] or Taiwan in Asia [19], for commercial sex workers—Central African Republic in Africa [20]). Still, data are missing for many regions, mainly due to difficulties in reaching risk groups. The accessibility of harm reduction measures is an equally important determinant of HIV and HBV prevalence, and stigma remains an important obstacle in the development of effective policies for diagnosis, treatment, and prevention in vulnerable groups.

HIV-HBV coinfected patients must be treated for both infections using dually active antiretroviral drugs [21,22]. The first-line therapy for HIV-HBV coinfection is based on tenofovir, administered either as tenofovir disoproxil fumarate (TDF) or tenofovir alafenamide (TAF)—a prodrug of TDF which preserves its high antiviral potency and high barrier to resistance while leading to fewer renal and bone side effects [21]. Regimens based on lamivudine (3TC) or emtricitabine (FTC) monotherapy, once suggested as acceptable in resource-limited settings for patients with undetectable or low plasma HBV-DNA [23], are no longer recommended. Other regimens sparing 3TC/FTC/TDF or TAF must be avoided due to their risk of hepatitis reactivation.

Antiretroviral therapy (ART) has a major impact on the evolution of HIV-HBV coinfection, especially when initiated early in subjects without severe fibrosis, as it decreases both HBV and HIV viral loads and prevents flares of hepatitis reactivation due to immune reconstitution in patients with severe immunosuppression [24]. However, worldwide, only 16.7% of HBV mono-infected patients receive treatment [8], as access to dually active antiretrovirals has been limited until recently in low-income countries, where most at-risk populations remain untreated [25]. Although important progress has been made by scaling-up tenofovir-based ART for PLWH [7], important local and regional disparities in accessibility persist, and high proportions of coinfected patients had suboptimal treatments, which rapidly select resistant HBV strains, altering the long-term evolution of the liver disease.

These epidemiological and therapeutic data are concerning, and there is a continuous need for patient monitoring and testing. The aim of this review is to assess the diagnostic and therapeutic challenges in HIV-HBV coinfected persons and to bring a novel perspective on the current and future biomarkers used for virologic monitoring and treatment prioritization.

2. Pathogenesis of Liver Injuries in HIV-HBV Coinfected Patients

According to a large multicentric cohort study performed in the early 2000s, the rate of liver-related mortality was significantly higher in people with HIV-HBV coinfection, namely 14.2/1000 person-years, compared with 1.7/1000 person-years in HIV mono-infection and 0.8/1000 person-years in HBV mono-infection [8].

Although the rate of liver-induced deaths has decreased over time, liver fibrosis can still be observed in up to one-third of all cases, even in individuals who are undergoing long-term ART [26]. Additionally, the incidence rate of hepatocellular carcinoma in HIV-HBV coinfected patients can reach up to 2/1000 person-years [27,28]. Interestingly, the adherence to HCC screening protocols in HIV-HBV coinfected is significantly lower than in HIV-HCV coinfected patients and remains below 20% in all cases of HBV and/or HCV coinfection [29].



The specific pathogenic mechanisms of liver injuries in HIV-HBV coinfected patients include the combined action of HIV and HBV replication, immunosuppression, and host-related comorbidities or genetic factors (Figure 1) [2].

Figure 1. Mechanisms of liver injuries in HIV-HBV coinfected patients. The pathogenic mechanisms derive from the combined influence of HIV and HBV replication inside the liver, the associated immune distress and immune-mediated hepatocytolysis, and the additional host-related risk factors (comorbidities, genetic background, ART response). Created with BioRender.com (accessed on 26 April 2023), with a license to publish.

2.1. HIV-Related Factors

The progression of HBV-induced liver disease is accelerated by the coinfection with HIV [30]. HIV acts through multiple direct and indirect mechanisms to trigger or amplify liver injuries [17,31]. On the one hand, HIV can bind to CXCR4 receptors expressed on the hepatocytes and can persist in the liver even during ART treatment [32,33], destabilizing the cytokine environment and promoting inflammation [34,35], as well as liver fibrosis (already seen in 8–10% of HIV mono-infected patients) [36,37]. Moreover, HIV exerts both a direct and indirect effect on liver hepatic stellate cells [31], which are key players in both the hepatic immune responses, fibrogenesis, and possibly in carcinogenesis through transdifferentiation in- or regulation of- hepatic stem cells [38]. HIV enters hepatic stellate cells via a CD4-independent route and can be transferred through direct cell contact to intrahepatic lymphocytes, maintaining an active local replication. Viral antigens, such as gp120, can trigger or amplify a proinflammatory/profibrotic response of activated hepatic stellate cells [39]. In addition, HIV-infected T helper cells lose their ability to impede profibrotic signaling by NK cells [40,41]. An indirect effect of HIV infection on other cell populations, such as Kupffer cells, can further amplify liver injuries in coinfected patients [42,43]. These and other more specific mechanisms have also been involved in liver fibrosis development during HIV-HCV coinfection [44].

HIV- induced CD4 T lymphocyte depletion and systemic immune activation is followed by an inadequate innate and adaptive response, with the consecutive release of proinflammatory and profibrotic cytokines (TGF beta, IL-1, IL-6), increasing the risk of liver fibrosis. HIV-HBV coinfected patients with high HBV viral loads have slower rates of CD4+ T cell restoration under treatment [45] and maintain a higher risk of mortality even after early initiation of tenofovir-based therapy [46]. In addition, coinfected patients with severe immunosuppression and high HIV viral load can experience immune-mediated liver injuries, with hepatic flares, during the immune reconstitution syndrome, that can paradoxically develop 4–8 weeks after the initiation of antiretroviral treatment [47]. ART introduction may lead, in some cases, to the reactivation of chronic hepatitis or decompensation of liver cirrhosis [48].

2.2. HBV-Related Factors

Immunopathogenesis is the main mechanism of liver disease progression in chronic hepatitis B, where HBV intrahepatic replication can occur even in the absence of detectable viremia [49]. Exhausted or tolerant HBV-specific CD8+ T cells [50,51], as well as suppression of CD4+ T-cell responses, mediated by dendritic cell impairment [52] or by upregulation of PD-1 expression [53,54], and amplified by HIV-induced immunosuppression, can contribute to disease chronicization. HBV can also suppress TLR-mediated immune responses and promote TGF- β expression, therefore activating the proinflammatory cytokines and accentuating the cytokine imbalance produced by HIV infection, although this mechanism was not confirmed as a key player in liver disease progression [49,55]. Activation of hepatocyte Fas/FasL pathways following Hepatitis B surface antigen (HBsAg) production during HBV replication might also be important for liver disease progression [56].

2.3. Other Contributing Factors

Several comorbidities (especially diabetes, non-alcoholic fatty liver disease, nonalcoholic steatohepatitis), heavy alcohol use, and the baseline level of liver fibrosis contribute to the development of liver complications in HIV-HBV coinfected patients [31,32].

The prevalence of diabetes varies across different cohorts from the United States and Europe. Up to 10% of HIV-infected adults can develop diabetes versus up to 8.3% of the cases in the general population [57]. However, the risk of diabetes in adults with HBV or HCV coinfection remains unclear. Data from a large Canadian cohort showed a lower prevalence in coinfected subjects (8.1% in those HIV-HBV coinfected and 4.9% in HIV-HCV co-infected [57]).

Data on alcohol use and alcoholic liver disease in PLH is limited. Approximately 16–22% of individuals with HIV infection report an increased average of current or lifetime alcohol consumption vary [58]. According to Butt et al., the rate of alcohol use is higher in HIV-HCV coinfected individuals than in HIV-HBV coinfected ones (45% vs. 16.2%), but the rate of alcoholic liver disease in the latter population is unclear [59].

The rate of non-alcoholic fatty liver disease (NAFLD) in HIV mono-infected adults varies between 9 and 73%, while the prevalence in HIV non-infected adults reaches up to 25% [60]. Several studies indicate high percentages of NAFLD in HIV-HBV coinfected patients, varying from 15 to 73% on liver biopsies [61]. HIV-HBV coinfected patients with non-alcoholic steatohepatitis (NASH) display higher progression rates for both fibrosis progression, cirrhosis, and hepatocellular carcinoma compared with HIV mono-infection. This is probably due to various HBV-linked molecular mechanisms that modulate the expression of cholesterol synthesis genes and dysregulate the cellular signal transduction pathways affecting cell growth and apoptosis [62,63].

The influence of the genetic background on the progression of liver diseases remains debatable. Several mutations associated with a high risk of HCC were commonly found in HIV-HBV coinfected patients, possibly explaining the high rates of carcinogenesis [64]. Nonetheless, double mutations in the core region (A1762T and G1764A), overlapping with the oncogenic HBx protein-encoding region, that were associated with advanced liver disease and HCC in HBV mono-infected patients, are less common in HIV coinfected ones [65,66].

ART treatment. Historically, in HIV mono-infected patients, the first-generation nucleoside analogs reverse transcriptase inhibitors (NRTI)—stavudine (d4T) and didanosine (ddI) carried a risk of severe microsteatosis and lactic acidosis [67]. The older non-nucleoside reverse transcriptase inhibitors (NNRTI)—nevirapine and efavirenz, as well as several protease inhibitors, were also associated with severe liver injury. However, none of these antiretrovirals are currently used for HIV-HBV coinfected individuals. The recommended dual-acting regimens are associated with a favorable effect on liver disease [68], without liver toxicity over 10 years, although there have been signals of a potential increase in end-stage liver diseases and hepatocellular carcinoma in the D:A:D study associated with cumulative exposure to several drugs, including tenofovir [69]. Mitochondrial toxicity and lactic acidosis remain rare events, even though drug-induced hepatotoxicity is more frequently reported than in mono-infected patients, especially during severe immunosuppression [55]. An early initiation of antivirals, as recommended by the START trial, can reduce the incidence of serious non-AIDS events, including decompensated liver disease [70].

3. Virological Monitoring of Therapeutic Success in HIV-HBV Coinfection

Four classic biomarkers are currently used to define therapeutic success in chronic hepatitis B in both immunocompetent and immunosuppressed patients: (1) HBV-DNA suppression (usually less than 60–80 IU/mL, although the lower quantification limit may differ among clinical trials), (2) ALT normalization, (3) anti-HBe-seroconversion and (4) HBsAg loss [22]. In addition, a systematic evaluation of liver fibrosis (every 3 months during the first year after diagnosis and every 6–12 months thereafter) using noninvasive markers and/or liver biopsy and hepatic ultrasound results are recommended for both HBV mono-infected and HIV-coinfected patients [21]. Nevertheless, virological and serological factors associated with long-term HIV-HBV coinfection evolution patterns are insufficiently understood, and, as a consequence, there is a continuous need for patient monitoring and testing. A series of novel biomarkers are now tested in HIV-HBV patients for an accurate diagnosis of viral replication, prediction of liver fibrosis, and HCC development. We present a synthetic view of the utility of these biomarkers for treatment monitoring and patients' stratification according to the potential for severe evolution (Figure 2).



Figure 2. Biomarkers for treatment monitoring in HBV mono-infected and HIV-HBV coinfected patients. Three categories of biomarkers are useful for diagnosis and therapeutic follow-up: predictors of treatment efficacy (biomarkers used to assess viral replication), Biomarkers used to evaluate the degree of liver fibrosis, and those used to predict the risk of hepatocellular carcinoma (HCC). HBcrAg-

hepatitis B core–related antigen, qHBsAg- quantitative hepatitis B surface antigen, cccDNA- covalently closed circular DNA, miR—microRNA, APRI-AST to Platelet Ratio, FIB4-Index combining age, AST, ALT and PLT count levels, AFP-alpha-fetoprotein, AFP-L3-*Lens culinaris* agglutinin-reactive fraction of alpha-fetoprotein; PIVKA-II-Protein induced by vitamin K absence or antagonist-II, PAGE-B—index comprising platelets, age, sex, and HBV infection, aMAP-index comprising age, gender, albumin, bilirubin and PLT count levels, GALAD—model including gender, age, AFP, L3-AFP and Des-carboxy-prothrombin, onco-miR—oncogenic microRNA. Created with BioRender.com (accessed on 26 April 2023), with a license to publish.

3.1. Predictors of Treatment Efficacy—Biomarkers of Viral Replication 3.1.1. Covalently Closed Circular DNA (cccDNA)

cccDNA represents both the transcriptional template and the reservoir of HBV. Its lifelong persistence in an episomal form in the infected hepatocytes [71] and its potential for reactivation during immunosuppression, even in patients with presumed resolution of HBV infection, is the main obstacle for a complete cure of chronic hepatitis B. Monitoring the cccDNA level with ultrasensitive molecular biology techniques (Table 1) constitutes a promising biomarker for both mono- and coinfected patients, as a declining level has been associated with therapeutic success [71] and the probability of a functional cure. In HIV-HBV coinfected people with prolonged courses of dually active antiretrovirals, the transcription of cccDNA is diminished. The majority of these patients have undetectable levels of intra-hepatic cccDNA when assessed with the qPCR technique [72]. However, it is still detectable with droplet digital (ddPCR) assays [73] that allow the partition of PCR reactions into water-in-oil droplets and quantification of nucleic acid without a reference standard, with important advantages in terms of sensitivity, absolute quantification, and accuracy [74].

	Biomarker	Diagnostic Test	Significance in Chronic HBV Mono-Infection	Significance in HIV-HBV Coinfection		
1.	Predictors of treatment efficacy					
	cccDNA	q Rt PCR ddPCR RCA in liver biopsies	Viral reservoir, declining levels associated with therapeutic success [71]	Reduced transcription under prolonged ART [73]		
	Serum HBV-RNA	qRT-PCR RT-ddPCR [74]	Good correlation with cccDNA [75] indicates patients with cirrhosis who may safely discontinue HBV therapy.	Correlated with detectable HBeAg [76–79] and quantitative HBsAg [66,78,79] potential use to guide ART changes		
	HBcrAg	Chemiluminescence/EIA	Predict the evolution of liver disease and the risk of carcinogenesis [80,81]	Monitors the evolution and the clearance of HBeAg [82]		
	qHBsAg	EIA Chemiluminescence	receiving treatment with nucleoside analogs and peginterferon therapy [83–89]	Decreased with an increasing number of CD4+ T cells [90–92]		
2.	Predictors of liver fibrosis					
		Fibrosis indexes based on serum markers				
	Fib-4 score	Index that combines age, AST, ALT, and PLT count levels	Diagnosis and follow-up of liver fibrosis (≥F2); can indicate the need to start HBV treatment; potential use as predictors of mortality [93–95]	Indicated for the screening of liver fibrosis in cases with elevated transaminases [94]		
	APRI	AST to Platelet Ratio	APRI threshold of 0.7 is sensitive and specific for the detection of significant fibrosis	An APRI < 0.5 are reported accordant with Fibroscan results to exclude fibrosis in 96.8% [96]		

Table 1. New biomarkers in chronic HBV mono-infection and HIV-HBV coinfection.

Biomarker	Diagnostic Test	Significance in Chronic HBV Mono-Infection	Significance in HIV-HBV Coinfection		
	GGT, bilirubin,				
Fibrotest [®]	haptoglobin,	Diagnosis, staging, and	Good accuracy for the diagnosis		
ribiotest	α2-macroglobulin,	follow-up of liver fibrosis [94]	of liver fibrosis [97]		
	apolipoprotein A1				
	Imaging-based technique	s for the detection of liver fibrosis			
	FibroScan/Echosens, (Paris, France)	Good accuracy for the	To literate the disc literate		
Transignt glastography		follow up of liver fibrosis	staging and follow up of liver		
(TE)		including cases with	fibrosis in HIV-HBV coinfected		
(11)		normal/discrete ALT	patients [99,100].		
		elevations [97–99]	L [, ,)].		
		Predicts liver fibrosis			
VCTE	Vibration-controlled transient	accurately in patients with	Noninvasive tool to assess liver		
VCIE	elastography	chronic hepatitis, irrespective	fibrosis [32,52]		
		of the etiology [101]			
	MicroRNAs as potential sur	MicroRNAs as potential surrogate biomarkers for liver fibrosis			
		Upregulated. Associated with	Biomarker of severe liver disease		
miR-122	RT-PCR, $2^{-\Delta\Delta CT}$ method [102]	activity stage of fibrosis	more data needed for HIV-HBV		
		HBsAg and HBV DNA [103]	coinfection		
			In HIV infection negatively		
D 105	$\mathbf{PT} \mathbf{PCP} \mathbf{a} = \mathbf{A} \mathbf{A} \mathbf{C} \mathbf{T}$ (1)	Downregulated, correlates	correlate with HIV-RNA [105];		
mik-125	RI-PCR; 2 method	with HBV viral load and	more data is needed for HIV-HBV		
		necroninaninatory activity	coinfection		
		Downregulated in HBV	Over-expressed in HIV infection,		
miR-29	RT-PCR; $2^{-\Delta\Delta CT}$ method	infection [106]	control of viral replication		
3 Tumor markers			(mechanism unknown) [107]		
3. Tumor markers	Classic	tumor markers			
		Usually increase as HCC	No data [108]		
AFP/PIVKA-II/AFP-L3	EIA	develops, irrespective of the			
	IHC	etiology			
M2BPGi	Lectin-antibody sandwich	Marker for liver fibrosis stage	No data [109,110]		
	Statistical model that includes	Predicts early HCC			
CALAD	gender, age, AFP, L3-AFP, and	The association with	NI- Jaka [111]		
GALAD score	Des-carboxy-prothrombin	ultrasound and elastography	No data [111]		
	(DCP)	increases the performance			
	miRNAs as oncogenes HBV-encoded miRNA;				
HBV-miR-2	Deep sequencing technology	promotes tumoral cell growth	No data [112]		
		by suppressing apoptosis HBV-encoded miRNA:			
		promotes tumoral cell growth	No data [110]		
HBV-mik-3	Deep sequencing technology	by silencing PPM1A (Protein	No data [113]		
		phosphatase 1A)			
		Cellular miRNA; promotes			
		cell growth, tumorigenesis,	Similar activity in HIV-HBV		
M1K-181	RT-PCR; $2^{-\Delta\Delta CT}$ method	and decreasing of apoptosis; it	contected patients correlated		
		HBY gene of HBV [114]	with carcinogenesis [107]		
	Indexes based on biochemical markers as predictors of oncogenesis				

Table 1. Cont.

Biomarker	Diagnostic Test	Significance in Chronic HBV Mono-Infection	Significance in HIV-HBV Coinfection
aMAP	Index that comprises age, gender, albumin, bilirubin, and PLT count levels	A cutoff value of 50 is predictive of the risk of carcinogenesis [115]	Not yet tested
PAGE-B index	Index that comprises platelets, age, sex, and HBV infection	Carcinogenesis prediction in treated HBV-infected patients [116] Recommended by EASL guidelines to delay the HCC surveillance [117]	Not yet tested

Table 1. Cont.

Legend: HBcrAg—hepatitis B core-related antigen, qHBsAg—quantitative hepatitis B surface antigen, AFP—alpha-fetoprotein, PIVKA-II—protein induced by vitamin K absence or antagonist-II, AFP-L3-Lens culinaris agglutinin-reactive fraction of alpha-fetoprotein; M2BPGi—Mac-2-binding protein glycosylation isomer; qRT PCR—quantitative real-time PCR; ddPCR—digital droplet-polymerase chain reaction; RCA—rolling circle amplification; RT-ddPCR—reverse-transcription ddPCR; EIA—enzyme immunoassay, IHC—immunohistochemical analysis, LiPA—line probe assay; EASL—European Association for the Study of the Liver; HCC—hepatocellular carcinoma.

3.1.2. Serum HBV-RNA

Serum HBV-RNA (also reported as preC RNA, pgRNA, or 3.5 kb RNA) can be detected as pre-genomic HBV-RNA and total serum HBV-RNA (for variants that cause defects in RNA splicing). Pre-genomic RNA indicates transcription of cccDNA and can be used in HBV mono-infected patients to identify cases (especially those with cirrhosis) who may safely discontinue HBV therapy or as a marker for the effectiveness of therapeutic agents that target cccDNA [75]. Data on the therapeutical implications of HBV-RNA detection in coinfected patients remains limited and requires an ongoing follow-up due to multiple fluctuations, irrespective of the baseline level, even on ART [76,77]. In HBV mono-infected patients, strong correlations were reported between HBV-RNA and both HBV-DNA and HBcrAg levels, while in HBV-HIV coinfected patients, the statistical correlations between various markers of HBV replication are less powerful [78,79]. Of note, in HIV-HBV coinfected patients, irrespective of their treatment status, pgRNA and total HBV-RNA are not associated with CD4 T cell count or HIV RNA [78,79].

3.1.3. Hepatitis B Core-Related Antigen (HBcrAg)

HBcrAg contains three products encoded by the precore/core gene, which share an identical 149 amino acid sequence [80]. HBeAg—the soluble peptide derived from the precore protein by proteolysis; HbcAg—the viral nucleocapsid that remains in the hepatocyte; and p22 cr—a 22-kDa precore protein present in non-infectious HBV DNAnegative particles [81,82]. HBcrAg might be used in mono- and coinfected patients to identify those (a) who can safely discontinue nucleoside analogues therapy; (b) at risk for HBV reactivation during therapy; (c) at risk for carcinogenesis. A decrease in the HBcrAg level can be associated with undetectable levels of intra-hepatic cccDNA, followed by HBsAg loss—a major goal for a functional cure in chronic hepatitis B infection [80]. In HBV mono-infected patients, the association of quantitative HBcrAg with antiHBc antibody titers is used as a predictor of the level of intrahepatic cccDNA [69], especially during immune-modulatory therapies [44,73,74]. In HIV-HBV coinfected patients undergoing tenofovir therapy, the same combination can be used to monitor the kinetics and clearance of HBeAg [75].

3.1.4. Quantitative HBsAg

The quantitative determination of HBsAg was correlated with intrahepatic replication. The level of HBsAg (expressed in IU/mL) is a widely available assay currently used to predict sustained virological response and achieve a functional cure for hepatitis B [82]. In HBV mono-infected patients, its clinical utility was mainly correlated with peginterferon

treatment monitoring and liver fibrosis evaluation. In HbeAg-positive patients, an HBsAg level < 1000 IU/mL combined with an HBV-DNA level < 2000 IU/mL indicates minimal current viral activity [83]. Conversely, in spontaneous HBeAg seroconverters, with HBV-DNA level < 2000 IU/mL, an HBsAg level higher than 1000 IU/mL is associated with an increased risk of HBeAg-negative hepatitis, with active viral replication [83].

However, in treatment-naïve HBe positive patients, lower levels of HBsAg have been associated with higher levels of liver fibrosis [84] and are considered an indicator of an increased duration of liver disease and immune-mediated clearance of infected cells. These findings were not confirmed in Asian patients [84,85].

Recently, genotype-specific cutoffs of the HBsAg level have been proposed to rule out cirrhosis in HBeAg-positive patients [86]. The utility of qHBsAg testing for HIV-HBV coinfected patients is currently unclear.

3.2. Predictors of Liver Fibrosis

In efficiently treated HBV mono-infected patients, liver fibrosis remains stationary, and a certain degree of regression can occur in time. Information about the on-treatment evolution of liver fibrosis in HIV-HBV patients is incomplete and sometimes discordant. During the last decade, the role of liver biopsy became less prominent, as there was a continuous development of noninvasive tests that can accurately detect advanced stages of liver fibrosis and cirrhosis and predict liver-related outcomes of HIV-HBV infections.

3.2.1. Fibrosis Indexes Based on Serum Biomarkers

APRI and FIB-4 scores, which utilize routine laboratory parameters, are very convenient and suitable noninvasive scores for liver fibrosis [118,119]. According to the most recent European Association for the Study of the Liver (EASL) recommendations, the regular calculation of simple, noninvasive fibrosis scores (FIB-4 or APRI) in populations at risk of liver fibrosis is recommended in combination with transient elastography (TE) for the stratification and linkage to care infections [120]; worth mentioning that neither TE nor noninvasive scores alone are considered appropriate to diagnose or exclude significant fibrosis.

FIB-4 score is based on a combination of age, AST, ALT, and PLT count levels and was first used for HIV-HCV coinfected patients. A lower limit of less than 1.45 indicates the absence of advanced fibrosis detected by liver biopsy (negative predictive value of 90%; sensitivity of 70%), while an upper limit higher than 3.25 indicates advanced fibrosis (with a positive predictive value of 65% and specificity of 97%) [121]. In subsequent studies, FIB-4 demonstrated a relatively high diagnostic value in patients with chronic hepatitis B when the threshold for advanced fibrosis was set at more than 2.0 (sensitivity of 69% and specificity of 95%) [122].

APRI index (AST to Platelet Ratio Index) is another noninvasive, inexpensive, and affordable score, tested initially on HCV-infected patients to assess the level of liver fibrosis; an APRI threshold of 0.7 was sensitive and specific for detection of significant fibrosis detected by liver biopsy (sensitivity of 77% and specificity of 72%, respectively), while for cirrhosis detection, an APRI cutoff of 1.0 was associated with 76% sensitivity and 72% specificity [123,124]. Subsequent studies have shown a good accuracy of the APRI score for HBV-induced cirrhosis [125,126].

Recent studies have shown that these scores are reliable noninvasive methods for the assessment of nonsignificant versus significant liver fibrosis in HIV-HBV coinfected patients also [127,128]. A FIB-4 value < 1.5 or APRI < 0.5 are concordant with Fibroscan results and can exclude fibrosis in 94.4% and 96.8% of cases [96].

Fibrometer score combines several markers (age, platelet count, prothrombin index, aspartate aminotransferase, alpha2-macroglobulin, hyaluronate, urea). The score's sensitivity and specificity in predicting the advanced stages of fibrosis in patients with viral hepatitis are reported to be higher than those of the APRI test [129]. In HIV-HBV coinfected

patients, the score is used as a noninvasive marker for the outcome of liver fibrosis after antiviral therapy [23].

Supplementary indexes, based on different inflammatory and anti-inflammatory cytokines, are now tested for HIV-HBV infection, as serum interleukins and interferon-gamma inducible protein-10 have been previously found dysregulated in both hepatitis B, C, HIV, or HIV-HCV coinfection [130–132], playing an important role in chronic hepatic inflammation and fibrogenesis.

3.2.2. Imaging-Based Techniques for Detection of Liver Fibrosis

Recent studies using noninvasive tests for liver fibrosis assessment show that approximately 20–30% of HIV-HBV patients on long-term ART present with advanced liver fibrosis, with concordant results despite the variation of cutoffs. Data concerning advanced liver disease in HBsAg-positive PLH is further supported by studies using serological biomarkers and liver biopsies; the percentage of treatment-experienced patients displaying significant liver fibrosis in various studies can be seen in Table 2 [24,133].

Table 2. Prevalence of liver fibrosis in HIV-HBV infected patients on ART.

Author	Study Group	Assessment Method	Median ART Duration (Years)	Median CD4 T Cell Count	Undetectable HIV- RNA/Undetectable HBV DNA (%)	Prevalence of Liver Fibrosis
Iacob, D., 2022 [127]	212 HIV infected, 101 HIV-HBV coinfected	APRI and Fib-4 scores	13	369	68%/46%	10.8% at baseline, 11.3% at 5 years follow up
Sterling, R.K., 2018 [133]	114 HIV-HBV patients	Liver Histology, APRI and Fib-4 scores	14	568	77.9%/57.9%	37% significant fibrosis (Ishak ≥ 2) 24% advanced fibrosis (Ishak ≥ 3)
Maida, I., 2006 [134]	37 HIV-HBV patients	Transient elastography	3.3	490	89%/70%	57% no or mild fibrosis 13% significant fibrosis (F3) 11% advanced fibrosis (F4)
Audsley, J., 2016 [135]	70 HIV-HBV patients, of which 20 co-infected with HCV	Transient elastography	10	381	74.6%/74.6%	35.7% significant fibrosis (≥F3)
Boyd, A., 2017 [24]	148 HIV-HBV patients, of which 12 co-infected with HDV 19 co-infected with HCV	Fibrometer score and liver biopsy	5.7	420	53.4%/17.8%	31% F3–F4 fibrosis
Miailhes, P., 2011 [136]	59 patients (46 patients on cART)	Transient elastography, liver biopsy, and Fibrotest	Not specified	397	85%/78%	33.8% F3–F4 fibrosis
Stockdale, A.J., 2015 [98]	106 HIV-HBV patients on ART and 15 ART-naive	Transient elastography	3.75	571	67%/49.1%	2.7% F3–F4 fibrosis

Transient elastography (TE), an easily-performed and rapid test, is the most validated imaging-based technique in HBV infection [98]. It gives rapid information for clinical use, with a sensitivity of approximately 78% and a specificity of 81–82% for predicting significant fibrosis in HBV-infected patients, compared to liver biopsy [137,138]. TE has been increasingly used to document the stage of liver fibrosis in HIV-HBV coinfected patients. Studies on its accuracy have shown a good concordance with liver biopsy [100].

VCTE (Vibration-controlled transient elastography) predicts liver fibrosis accurately in patients with chronic hepatitis, irrespective of the etiology. In a large meta-analysis, VCTE showed good diagnostic performance in predicting significant fibrosis in patients with chronic hepatitis B, compared to liver biopsy (sensitivity between 71.2 and 99%, and specificity between 73.9 and 94% in different studies) [101].

These noninvasive methods can be used in HIV-HBV coinfected patients for longitudinal monitoring at shorter intervals. Nevertheless, limited data are available on the optimal cutoff in both naive and treated patients. Combining TE or VCTE with other serological markers might increase their diagnosis accuracy [120].

Serum markers and imaging-based techniques have both advantages over liver biopsy (including low risks, affordable cost, and better acceptance) and several disadvantages (including lower diagnostic accuracy for the prediction of early stages of fibrosis and inconsistent results in the presence of other comorbidities that can induce liver inflammation or fibrosis).

ART is generally associated with improving liver stiffness or improved liver fibrosis scores on liver biopsies, yet a subset of patients can display the progression of liver fibrosis [24]. Although sometimes this can be explained by suboptimal treatment adherence, there are patients with undetectable HIV-RNA levels who continue to display a low-level HBV viremia [139], often without persistent elevation of liver enzymes. In these cases, other conditions, including immune activation and additive metabolic factors [132], can play a role in the evolution of liver disease and viral replication [140]. The management of these patients is particularly challenging, and specific intervention must be directed to lifestyle modifications (a balanced diet, increased physical activity, elimination of additional risk factors such as alcohol intake and drug usage).

3.2.3. MicroRNAs, as Potential Surrogate Biomarkers for Liver Fibrosis

Several microRNA species are constantly dysregulated during both HIV infection and chronic hepatitis B: miR-122, miR-210, and miR-181 expressions are increased, while miR-29 and miR-125 expressions are decreased [106]. The expression and the activity of these short transcripts in HBsAg-positive PLH are not very well understood, and their potential use as surrogate biomarkers for liver fibrosis is worth to be studied [141].

miR-122, a therapeutic target for HCV infection, is also directly correlated with increased necroinflammatory activity and advanced liver fibrosis [103] in HBV mono-infection. The serum levels of miR-122 are over-expressed in ART-treated HIV-1 patients before the development of liver disease [104], suggesting its potential clinical utility as a biomarker of severe liver disease evolution in coinfected patients.

miR-125, miR-124. The serum level of miRNA-125b is down-regulated and correlates with HBV viral load and is used either alone or combined with miRNA-124. It has the potential to discriminate between different grades of liver necroinflammation [142].

miR-29. Even though in HBV infection this transcript is reported to be downregulated, in treatment-experienced coinfected patients, miR-29 is upregulated, and associated with the virological and immunological markers of HIV-1 infection [143]. This is probably due to its involvement in the control of viral replication and immune responses [107].

The development of prediction scores, combining different miRNAs, can be an interesting approach to define the outcome of the HIV-HBV coinfected patients.

3.3. Tumor Markers and Predictors of Oncogenesis

3.3.1. Classic Tumor Markers

The incidence of HCC is rising worldwide, being one of the most important causes of cancer death globally. In patients with chronic hepatitis B, antivirals that maintain HBV suppression can prevent or delay the evolution towards cirrhosis and HCC, but monitoring using abdominal ultrasound is still recommended in high-risk populations.

Several tumor biomarkers have been proposed, but generally failed in the early detection of HCC [117].

AFP (alpha-fetoprotein) is the classic marker used for HCC risk monitoring. According to the American Association for Study of Liver Diseases (AASLD) guidelines, an AFP level higher than 20 ng/mL indicates the need for follow-up [144,145], but due to its low diagnostic accuracy, it must be used in combination with other markers for HCC surveillance. An alternative biomarker-AFP-L3 (*Lens culinaris* agglutinin-reactive fraction of alpha-fetoprotein or highly sensitive -hs-AFP-L3) [146] can be tested using an increased sensitivity immunoassay, alone or in association with PIVKA-II (Protein induced by vitamin K absence or antagonist-II). The levels of these two proteins are usually increasing as HCC develops and progresses to portal vein invasion [108]. According to EASL guidelines,

while PIVKA-II use has not been standardized until now [117]. Other biomarkers, such as Mac-2-binding protein glycosylation isomer (M2BPGi), a simple and reliable indicator of liver fibrosis in HBV mono-infection, used for prediction of hepatocarcinogenesis in patients treated with nucleoside analogs [109,110], have not yet been evaluated in HIV-HBV coinfection.

AFP-L3 is suboptimal in terms of cost-effectiveness for routine surveillance of early HCC,

3.3.2. Indexes Based on Biochemical Markers as Predictors of Oncogenesis

Different statistical models can generate indexes with higher specificity and predictive value for HCC evaluation and monitoring.

A recent score-*aMAP* (combining age, gender, albumin, bilirubin, and PLT count levels) was developed to assess the risk for carcinogenesis in patients with chronic hepatitis B and C, with a cutoff of 50 associated with a negative predictive value of 99.3–100% (sensitivity of 85.7–100%), irrespective of etiology [115].

PAGE-B index (comprising platelets, age, and gender) can predict the 5-year HCC risk in Caucasian HBV-infected patients with or without NA treatment [116,120]. EASL guidelines suggests that patients with a low PAGE-B score might even not require HCC surveillance [117].

GALAD score (calculated using gender, age, AFP, L3-AFP, and Des-carboxy-prothrombin) is a statistical model used for the early prediction of HCC development in patients with chronic hepatitis B. The association of the GALAD score with imaging-based techniques can generate better information for high-risk patient monitoring [111].

Still, none of these scores have been validated in HIV-HBV coinfected patients [147].

3.3.3. Genetic Biomarkers

As appropriate biomarkers for early detection of HCC remain scarce, the addition of genetic biomarkers, such as somatic mutations in Telomerase Reverse Transcriptase (TERT) gene can further improve the diagnosis. Using liquid biopsy, several TERT promoter mutations (C228T and C250T) are frequently detected both in cirrhosis and HCC [148], and can be used to stratify patients with premalignant status for continuous monitoring.

Cell-free DNA (cfDNA—the nucleic acid released by cancerous cells that can be detected in peripheral blood) has occurred as a novel predictive biomarker for HCC. The methylation status of several genes, such as HOXA1, TSPLY5, and PFKP from cfDNA, has a good predictive value for hepatic oncogenesis [149,150].

3.4. Viral and Host miRNAs That Can Act as Oncogenes

HBV-encoded miRNAs. HBV-miR-2 is preserved among different HBV subtypes [106] and can act as an oncogene, promoting the development of HBV-related HCC and tumoral cell growth by suppressing apoptosis and stimulating migration and invasion [112]. HBV-miR-3, highly expressed in HBV chronic infection, is also considered an emerging factor in tumorigenesis, as it silences PPM1A (Protein phosphatase 1A) and promotes tumoral cell proliferation [113].

MiR-181, a cellular miRNA that has important roles in cell growth promotion, tumorigenesis, and apoptosis inhibition [114], is an epigenetic target for the HBV HBx gene. Its expression is upregulated in both HBV mono-infected and HIV-HBV coinfected patients, correlated with carcinogenesis [107].

New biomarkers, such as composite indexes of microRNAs, can help to better understand and predict the progression to liver fibrosis/cirrhosis and/or hepatocellular carcinoma. Such additional biomarkers have been proposed for the long-term follow-up of HBV mono-infected patients. Their relevance for the HIV-HBV coinfection and their value for routine clinical practice must be attentively appraised.

4. Conclusions

HIV-HBV coinfection remains a major public health problem. High rates of liver fibrosis, liver-related morbidity, hospitalizations, and mortality due to hepatocellular carcinoma represent important clinical challenges, especially in vulnerable, sometimes neglected populations. The combination between HIV and HBV replication, immune dysregulations, and immune-mediated hepatocellular injuries led to a more severe evolution compared to each of the mono-infection. Although the early initiation of antiretroviral therapy has an undeniably favorable impact on the evolution of HIV-HBV coinfection, access to highly potent dually active antiretrovirals remains imbalanced between different geographic regions. When therapy is delayed, the chronic immune activation and profound immunosuppression linked to HIV infection progression towards AIDS, with a reduced number of interferon-gamma-producing HBV-specific CD8+ T cells, can increase the risk of severe evolution [151,152].

Therefore, long-term monitoring of the active viral replication, the size and activity of the viral reservoir, and the progression of liver fibrosis using affordable and accurate tests is essential in order to prevent end-stage liver disease.

New noninvasive biomarkers reviewed in this manuscript are clinically relevant for HIV-HBV coinfected patients' stratification, according to their risk for severe evolution and their response to therapy.

Author Contributions: Individual contributions are as follows: Conceptualization: S.R. and C.C.; methodology, C.S. and D.I.; validation, S.R.; writing—original draft preparation, C.S., L.G., D.I. and S.R.; writing—review and editing, C.S. and S.R.; supervision, S.R. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Acknowledgments: Publication of this paper was supported by the University of Medicine and Pharmacy Carol Davila, through the institutional program Publish not Perish.

Conflicts of Interest: The authors declare no conflict of interest.

References

- Sustainable Development Goals (SDGs): Goal 3. Target 3.3: By 2030, End the Epidemics of AIDS, Tuberculosis, Malaria and Neglected Tropical Diseases and Combat Hepatitis, Water-Borne Diseases and Other Communicable Diseases [Poster]. Available online: https://apps.who.int/iris/handle/10665/208281 (accessed on 20 March 2023).
- Ganesan, M.; Poluektova, L.Y.; Kharbanda, K.K.; Osna, N.A. Human immunodeficiency virus and hepatotropic viruses comorbidities as the inducers of liver injury progression. *World J. Gastroenterol.* 2019, 25, 398–410. [CrossRef]
- 3. Naik, P.A.; Zu, J.; Owolabi, K.M. Modeling the mechanics of viral kinetics under immune control during primary infection of HIV-1 with treatment in fractional order. *Phys. A Stat. Mech. Appl.* **2020**, *5*45, 123816. [CrossRef]
- Farman, M.; Tabassum, M.F.; Naik, P.A.; Akram, S. Numerical treatment of a nonlinear dynamical Hepatitis-B model: An evolutionary approach. *Eur. Phys. J. Plus* 2020, 135, 941. [CrossRef]
- 5. Approximate Solution of a Nonlinear Fractional-Order HIV Model Using Homotopy Analysis Method. Available online: https://global-sci.org/intro/article_detail/ijnam/20349.html (accessed on 20 April 2023).

- European Centre for Disease Prevention and Control. Public Health Guidance on HIV, Hepatitis B and C Testing in the EU/EEA—An Integrated Approach. Stockholm: ECDC. 2018. Available online: https://www.ecdc.europa.eu/sites/default/ files/documents/hiv-hep-testing-guidance_0.pdf (accessed on 2 February 2023). [CrossRef]
- Platt, L.; French, C.E.; McGowan, C.R.; Sabin, K.; Gower, E.; Trickey, A.; McDonald, B.; Ong, J.; Stone, J.; Easterbrook, P.; et al. Prevalence and burden of HBV co-infection among people living with HIV: A global systematic review and meta-analysis. *J. Viral Hepat.* 2020, 27, 294–315. [CrossRef]
- 8. Hutin, Y.; Nasrullah, M.; Easterbrook, P.; Nguimfack, B.D.; Burrone, E.; Averhoff, F.; Bulterys, M. Access to Treatment for Hepatitis B Virus Infection—Worldwide, 2016. *MMWR. Morb. Mortal. Wkly. Rep.* **2018**, *67*, 773–777. [CrossRef]
- Thio, C.L.; Seaberg, E.C.; Skolasky, R.; Phair, J.; Visscher, B.; Muñoz, A.; Thomas, D.L. HIV-1, hepatitis B virus, and risk of liver-related mortality in the Multicenter Cohort Study (MACS). *Lancet* 2002, 360, 1921–1926. [CrossRef]
- 10. Sherman, K.E.; Peters, M.G.; Thomas, D. Human immunodeficiency virus and liver disease: A comprehensive update. *Hepatol. Commun.* **2017**, *1*, 987–1001. [CrossRef]
- Pinchoff, J.; Tran, O.C.; Chen, L.; Bornschlegel, K.; Drobnik, A.; Kersanske, L.; Fuld, J. Impact of hepatitis B on mortality and specific causes of death in adults with and without HIV co-infection in NYC, 2000–2011. *Epidemiol. Infect.* 2016, 144, 3354–3364. [CrossRef]
- 12. Burns, G.S.; Thompson, A.J. Viral hepatitis B: Clinical and epidemiological characteristics. *Cold Spring Harb. Perspect. Med.* **2014**, 4, a024935. [CrossRef]
- Poynard, T.; Lebray, P.; Ingiliz, P.; Varaut, A.; Varsat, B.; Ngo, Y.; Norha, P.; Munteanu, M.; Drane, F.; Messous, D.; et al. Prevalence of liver fibrosis and risk factors in a general population using non-invasive biomarkers (FibroTest). *BMC Gastroenterol.* 2010, 10, 40. [CrossRef]
- 14. Roșca, A.; Iacob, D.; Ene, L.; Temereanca, A.; Grancea, C.; Sultana, C.; Achim, C.L.; Ruță, S. Liver function in a cohort of young HIV-HBV co-infected patients on long-term combined antiretroviral therapy. *Farmacia* **2020**, *68*, 42–47. [CrossRef]
- 15. Wandeler, G.; Mulenga, L.; Vinikoor, M.J.; Kovari, H.; Battegay, M.; Calmy, A.; Cavassini, M.; Bernasconi, E.; Schmid, P.; Bolton-Moore, C.; et al. Liver fibrosis in treatment-naïve HIV-infected and HIV/HBV co-infected patients: Zambia and Switzerland compared. *Int. J. Infect. Dis.* **2016**, *51*, 97–102. [CrossRef]
- 16. Mallet, V.; Vallet-Pichard, A.; Pol, S. The impact of human immunodeficiency virus on viral hepatitis. *Liver Int.* **2011**, *31*, 135–139. [CrossRef] [PubMed]
- 17. Thio, C.L.; Smeaton, L.; Saulynas, M.; Hwang, H.; Saravan, S.; Kulkarni, S.; Hakim, J.; Nyirenda, M.; Iqbal, H.S.; Lalloo, U.G.; et al. Characterization of HIV-HBV coinfection in a multinational HIV-infected cohort. *AIDS* **2013**, *27*, 191–201. [CrossRef] [PubMed]
- Haussig, J.M.; Nielsen, S.; Gassowski, M.; Bremer, V.; Marcus, U.; Wenz, B.; Bannert, N.; Bock, C.T.; Zimmermann, R. A large proportion of people who inject drugs are susceptible to hepatitis B: Results from a bio-behavioural study in eight German cities. *Int. J. Infect. Dis.* 2018, *66*, 5–13. [CrossRef] [PubMed]
- Hsieh, M.-H.; Tsai, J.-J.; Hsieh, M.-Y.; Huang, C.-F.; Yeh, M.-L.; Yang, J.-F.; Chang, K.; Lin, W.-R.; Lin, C.-Y.; Chen, T.-C.; et al. Hepatitis C Virus Infection among Injection Drug Users with and without Human Immunodeficiency Virus Co-Infection. *PLoS* ONE 2014, 9, e94791. [CrossRef]
- Longo, J.D.D.; Simaleko, M.M.; Diemer, H.S.C.; Grésenguet, G.; Brücker, G.; Belec, L. Risk factors for HIV infection among female sex workers in Bangui, Central African Republic. *PLoS ONE* 2017, *12*, e0187654. [CrossRef] [PubMed]
- EACS European AIDS Clinical Society. EACS Guidelines 11.0. 2021. Available online: https://www.eacsociety.org/media/final2 021eacsguidelinesv11.0_oct2021.pdf (accessed on 3 March 2022).
- 22. European Association for the Study of the Liver. EASL 2017 Clinical Practice Guidelines on the management of hepatitis B virus infection q. J. Hepatol. 2017, 67, 370–398. [CrossRef]
- Price, H.; Dunn, D.; Pillay, D.; Bani-Sadr, F.; de Vries-Sluijs, T.; Jain, M.K.; Kuzushita, N.; Mauss, S.; Núñez, M.; Nüesch, R.; et al. Suppression of HBV by Tenofovir in HBV/HIV Coinfected Patients: A Systematic Review and Meta-Analysis. *PLoS ONE* 2013, 8, e68152. [CrossRef]
- Boyd, A.; Bottero, J.; Miailhes, P.; Lascoux-Combe, C.; Rougier, H.; Girard, P.-M.; Serfaty, L.; Lacombe, K. Liver fibrosis regression and progression during controlled hepatitis B virus infection among HIV-HBV patients treated with tenofovir disoproxil fumarate in France: A prospective cohort study. J. Int. AIDS Soc. 2017, 20, 21426. [CrossRef]
- Ozaras, R.; Corti, G.; Ruta, S.; Lacombe, K.; Mondelli, M.U.; Irwing, W.L.; Puoti, M.; Khalighi, A.; Santos, M.L.; Harxhi, A.; et al. Differences in the availability of diagnostics and treatment modalities for chronic hepatitis B across Europe. *Clin. Microbiol. Infect.* 2015, 21, 1027–1032. [CrossRef]
- 26. Van Welzen, B.J.; Smit, C.; Boyd, A.; I Lieveld, F.; Mudrikova, T.; Reiss, P.; E Brouwer, A.; Hoepelman, A.I.M.; E Arends, J. Decreased All-Cause and Liver-Related Mortality Risk in HIV/Hepatitis B Virus Coinfection Coinciding With the Introduction of Tenofovir-Containing Combination Antiretroviral Therapy. *Open Forum Infect. Dis.* 2020, 7, ofaa226. [CrossRef] [PubMed]
- Sun, J.; Althoff, K.N.; Jing, Y.; Horberg, M.A.; Buchacz, K.; Gill, M.J.; Justice, A.C.; Rabkin, C.S.; Goedert, J.J.; Sigel, K.; et al. Trends in Hepatocellular Carcinoma Incidence and Risk Among Persons With HIV in the US and Canada, 1996–2015. *JAMA Netw. Open* 2021, 4, 2037512. [CrossRef]
- Nina Kim, H.; Newcomb, C.W.; Carbonari, D.M.; Roy, J.A.; Torgersen, J.; Althoff, K.N.; Kitahata, M.M.; Rajender Reddy, K.; Lim, J.K.; Silverberg, M.J.; et al. Risk of Hepatocellular Carcinoma with Hepatitis B Viremia among HIV/Hepatitis B Virus-Coinfected Persons in North America. *Hepatology* 2021, 74, 1190–1202. [CrossRef]

- 29. Willemse, S.; Smit, C.; Sogni, P.; Sarcletti, M.; Uberti-Foppa, C.; Wittkop, L.; Raben, D.; Monforte, A.D. Low compliance with hepatocellular carcinoma screening guidelines in hepatitis B/C virus co-infected HIV patients with cirrhosis. *J. Viral Hepat.* **2019**, 26, 1224–1228. [CrossRef] [PubMed]
- 30. Corcorran, M.A.; Kim, N. Chronic hepatitis B and HIV coinfection. Top. Antivir. Med. 2023, 31, 14–22.
- Parvez, M.K. HBV and HIV co-infection: Impact on liver pathobiology and: Therapeutic approaches. World J. Hepatol. 2015, 7, 121–126. [CrossRef]
- North, T.W.; Higgins, J.; Deere, J.D.; Hayes, T.L.; Villalobos, A.; Adamson, L.; Shacklett, B.L.; Schinazi, R.F.; Luciw, P.A. Viral Sanctuaries during Highly Active Antiretroviral Therapy in a Nonhuman Primate Model for AIDS. J. Virol. 2010, 84, 2913–2922. [CrossRef]
- Hong, F.; Saiman, Y.; Si, C.; Mosoian, A.; Bansal, M.B. X4 Human Immunodeficiency Virus Type 1 gp120 Promotes Human Hepatic Stellate Cell Activation and Collagen I Expression through Interactions with CXCR4. *PLoS ONE* 2012, 7, e33659. [CrossRef]
- Debes, J.D.; Bohjanen, P.R.; Boonstra, A. Mechanisms of accelerated liver fibrosis progression during HIV infection. J. Clin. Transl. Hepatol. 2016, 4, 328–335. [CrossRef]
- Yu, Y.; Gong, R.; Mu, Y.; Chen, Y.; Zhu, C.; Sun, Z.; Chen, M.; Liu, Y.; Zhu, Y.; Wu, J. Hepatitis B Virus Induces a Novel Inflammation Network Involving Three Inflammatory Factors, IL-29, IL-8, and Cyclooxygenase-2. *J. Immunol.* 2011, 187, 4844–4860. [CrossRef] [PubMed]
- 36. Maggi, P.; Altizio, S.; Di Biagio, A.; Nicolini, L.; Volpe, A.; Tancorre, T.; Leone, A.; Bellacosa, C.; Ladisa, N.; Angarano, G. Prevalence and Risk Factors for Significant Liver Fibrosis in Patients with HIV Infection. *In Vivo* 2015, 29, 771–775. [PubMed]
- Lombardi, R.; Lever, R.; Smith, C.; Marshall, N.; Rodger, A.; Bhagani, S.; Tsochatzis, E. Liver test abnormalities in patients with HIV mono-infection: Assessment with simple noninvasive fibrosis markers. *Ann. Gastroenterol.* 2017, 30, 349–356. [CrossRef] [PubMed]
- Friedman, S.L. Hepatic stellate cells: Protean, multifunctional, and enigmatic cells of the liver. *Physiol. Rev.* 2008, 88, 125–172.
 [CrossRef]
- Tuyama, A.C.; Hong, F.; Saiman, Y.; Wang, C.; Ozkok, D.; Mosoian, A.; Chen, P.; Chen, B.K.; Klotman, M.E.; Bansal, M.B. Human Immunodeficiency Virus (HIV)-1 Infects Human Hepatic Stellate Cells and Promotes Collagen I and Monocyte Chemoattractant Protein-1 Expression: Implications for the Pathogenesis of HIV/Hepatitis C Virus–Induced Liver Fibrosis. *Hepatology* 2010, 52, 612. [CrossRef] [PubMed]
- Kitahata, M.M.; Drozd, D.R.; Crane, H.M.; Van Rompaey, S.E.; Althoff, K.N.; Gange, S.J.; Klein, M.B.; Lucas, G.M.; Abraham, A.G.; Lo Re, V.; et al. Ascertainment and verification of end-stage renal disease and end-stage liver disease in the north american AIDS cohort collaboration on research and design. *AIDS Res. Treat.* 2015, 2015, 923194. [CrossRef]
- Gange, S.J.; Kitahata, M.M.; Saag, M.S.; Bangsberg, D.R.; Bosch, R.J.; Brooks, J.T.; Calzavara, L.; Deeks, S.G.; Eron, J.J.; Gebo, K.A.; et al. Cohort Profile: The North American AIDS Cohort Collaboration on Research and Design (NA-ACCORD). *Int. J. Epidemiol.* 2007, *36*, 294–301. [CrossRef]
- 42. Zhang, L.; Bansal, M.B. Role of Kupffer Cells in Driving Hepatic Inflammation and Fibrosis in HIV Infection. *Front. Immunol.* **2020**, *11*, 1086. [CrossRef]
- Klein, M.B.; Althoff, K.N.; Jing, Y.; Lau, B.; Kitahata, M.; Lo Re, V.; Kirk, G.D.; Hull, M.; Kim, H.N.; Sebastiani, G.; et al. Risk of End-Stage Liver Disease in HIV-Viral Hepatitis Coinfected Persons in North America from the Early to Modern Antiretroviral Therapy Eras. *Clin. Infect. Dis.* 2016, 63, 1160–1167. [CrossRef]
- 44. Gobran, S.T.; Ancuta, P.; Shoukry, N.H. A Tale of Two Viruses: Immunological Insights into HCV/HIV Coinfection. *Front. Immunol.* **2021**, *12*, 726419. [CrossRef]
- 45. Anderson, M.; Gaseitsiwe, S.; Moyo, S.; Thami, K.P.; Mohammed, T.; Setlhare, D.; Sebunya, T.K.; Powell, E.A.; Makhema, J.; Blackard, J.T.; et al. Slow CD4+ T-Cell Recovery in Human Immunodeficiency Virus/Hepatitis B Virus-Coinfected Patients Initiating Truvada-Based Combination Antiretroviral Therapy in Botswana. *Open Forum Infect. Dis.* 2016, 3, ofw140. [CrossRef] [PubMed]
- 46. Kouame, G.M.; Boyd, A.; Moh, R.; Badje, A.; Gabillard, D.; Ouattara, E.; Ntakpe, J.B.; Emième, A.; Maylin, S.; Chekaraou, M.A.; et al. Higher Mortality Despite Early Antiretroviral Therapy in Human Immunodeficiency Virus and Hepatitis B Virus (HBV)-Coinfected Patients with High HBV Replication. *Clin. Infect. Dis.* 2018, 66, 112–120. [CrossRef] [PubMed]
- Crane, M.; Oliver, B.; Matthews, G.; Avihingsanon, A.; Ubolyam, S.; Markovska, V.; Chang, J.J.; Dore, G.J.; Price, P.; Visvanathan, K.; et al. Immunopathogenesis of Hepatic Flare in HIV/Hepatitis B Virus (HBV)–Coinfected Individuals after the Initiation of HBV-Active Antiretroviral Therapy. J. Infect. Dis. 2009, 199, 974–981. [CrossRef] [PubMed]
- Chang, M.L.; Liaw, Y.F. Hepatitis B flares in chronic hepatitis B: Pathogenesis, natural course, and management. J. Hepatol. 2014, 61, 1407–1417. [CrossRef] [PubMed]
- Ming, D.; Yu, X.; Guo, R.; Deng, Y.; Li, J.; Lin, C.; Su, M.; Lin, Z.; Su, Z. Elevated TGF-β1/IL-31 pathway is associated with the disease severity of Hepatitis B virus-related liver cirrhosis. *Viral Immunol.* 2015, 28, 209–216. [CrossRef]
- Kakimi, K.; Isogawa, M.; Chung, J.; Sette, A.; Chisari, F.V. Immunogenicity and Tolerogenicity of Hepatitis B Virus Structural and Nonstructural Proteins: Implications for Immunotherapy of Persistent Viral Infections. J. Virol. 2002, 76, 8609–8620. [CrossRef] [PubMed]

- Reignat, S.; Webster, G.J.M.; Brown, D.; Ogg, G.S.; King, A.; Seneviratne, S.L.; Dusheiko, G.; Williams, R.; Maini, M.K.; Bertoletti, A. Escaping high viral load exhaustion: CD8 cells with altered tetramer binding in chronic hepatitis B virus infection. *J. Exp. Med.* 2002, 195, 1089–1101. [CrossRef]
- Xu, D.; Fu, J.; Jin, L.; Zhang, H.; Zhou, C.; Zou, Z.; Zhao, J.-M.; Zhang, B.; Shi, M.; Ding, X.; et al. Circulating and Liver Resident CD4 + CD25 + Regulatory T Cells Actively Influence the Antiviral Immune Response and Disease Progression in Patients with Hepatitis B. J. Immunol. 2006, 177, 739–747. [CrossRef]
- Li, M.; Sun, X.-H.; Zhu, X.-J.; Jin, S.-G.; Zeng, Z.-J.; Zhou, Z.-H.; Yu, Z.; Gao, Y.-Q. HBcAg induces PD-1 upregulation on CD4+T cells through activation of JNK, ERK and PI3K/AKT pathways in chronic hepatitis-B-infected patients. *Lab. Investig.* 2012, 92, 295–304. [CrossRef]
- 54. Liang, X.S.; Zhou, Y.; Li, C.Z.; Wan, M. Bin Natural course of chronic hepatitis B is characterized by changing patterns of programmed death type-1 of CD8-positive T cells. *World J. Gastroenterol.* **2010**, *16*, 618–624. [CrossRef]
- 55. Wu, J.; Meng, Z.; Jiang, M.; Pei, R.; Trippler, M.; Broering, R.; Bucchi, A.; Sowa, J.P.; Dittmer, U.; Yang, D.; et al. Hepatitis B virus suppresses toll-like receptor-mediated innate immune responses in murine parenchymal and nonparenchymal liver cells. *Hepatology* **2009**, *49*, 1132–1140. [CrossRef] [PubMed]
- Jing, Z.-T.; Liu, W.; Wu, S.-X.; He, Y.; Lin, Y.-T.; Chen, W.-N.; Lin, X.-J.; Lin, X. Hepatitis B Virus Surface Antigen Enhances the Sensitivity of Hepatocytes to Fas-Mediated Apoptosis via Suppression of AKT Phosphorylation. J. Immunol. 2018, 201, 2303–2314. [CrossRef] [PubMed]
- Hernandez-Romieu, A.C.; Garg, S.; Rosenberg, E.S.; Thompson-Paul, A.M.; Skarbinski, J. Is diabetes prevalence higher among HIV-infected individuals compared with the general population? Evidence from MMP and NHANES 2009–2010. BMJ Open Diabetes Res. Care 2017, 5, 304. [CrossRef]
- Ferguson, T.F.; Rosen, E.; Carr, R.; Brashear, M.; Simon, L.; Theall, K.P.; Ronis, M.J.; Welsh, D.A.; Molina, P.E. Associations of Liver Disease with Alcohol Use among People Living with HIV and the Role of Hepatitis C: The New Orleans Alcohol Use in HIV Study. *Alcohol Alcohol.* 2020, 55, 28–36. [CrossRef]
- Butt, Z.A.; Wong, S.; Rossi, C.; Binka, M.; Wong, J.; Yu, A.; Darvishian, M.; Alvarez, M.; Chapinal, N.; Mckee, G.; et al. Concurrent Hepatitis C and B Virus and Human Immunodeficiency Virus Infections Are Associated with Higher Mortality Risk Illustrating the Impact of Syndemics on Health Outcomes. *Open Forum Infect. Dis.* 2020, 7, ofaa347. [CrossRef]
- 60. Soti, S.; Corey, K.E.; Lake, J.E.; Erlandson, K.M. NAFLD and HIV: Do Sex, Race, and Ethnicity Explain HIV-Related Risk? *Curr. HIV/AIDS Rep.* **2018**, *15*, 212. [CrossRef]
- Woreta, T.A.; Chalasani, N. Fatty Liver Disease in Human Immunodeficiency Virus–Hepatitis B Virus Coinfection: A Cause for Concern. *Clin. Infect. Dis.* 2020, 73, e3286–e3287. [CrossRef]
- 62. Suliman, I.; Abdelgelil, N.; Kassamali, F.; Hassanein, T.I. The Effects of Hepatic Steatosis on the Natural History of HBV Infection. *Clin. Liver Dis.* **2019**, 23, 433–450. [CrossRef]
- 63. Villa, G.; Owusu, D.; Smith, C.; Azumah, M.; Abdullahi, A.; Phillips, S.; Sayeed, L.; Austin, H.; Chadwick, D.; Phillips, R.O.; et al. Liver steatosis and fibrosis in people with human immunodeficiency virus in West Africa and the relationship with hepatitis B virus coinfection. *Hepatol. Commun.* **2022**, *6*, 3036. [CrossRef]
- 64. Li, K.-W.; Kramvis, A.; Liang, S.; He, X.; Chen, Q.-Y.; Wang, C.; Yang, Q.-L.; Hu, L.-P.; Jia, H.-H.; Fang, Z.-L. Higher prevalence of cancer related mutations 1762T/1764A and PreS deletions in hepatitis B virus (HBV) isolated from HBV/HIV co-infected compared to HBV-mono-infected Chinese adults. *Virus Res.* 2017, 227, 88–95. [CrossRef]
- Audsley, J.; Littlejohn, M.; Yuen, L.; Sasadeusz, J.; Ayres, A.; Desmond, C.; Spelman, T.; Lau, G.; Matthews, G.V.; Avihingsanon, A.; et al. HBV mutations in untreated HIV-HBV co-infection using genomic length sequencing. *Virology* 2010, 405, 539–547. [CrossRef] [PubMed]
- Ryom, L.; Cotter, A.; De Miguel, R.; Béguelin, C.; Podlekareva, D.; Arribas, J.R.; Marzolini, C.; Mallon, P.G.M.; Rauch, A.; Kirk, O.; et al. 2019 update of the European AIDS Clinical Society Guidelines for treatment of people living with HIV version 10.0. *HIV Med.* 2020, 21, 617. [CrossRef] [PubMed]
- 67. Mokrzycki, M.H.; Harris, C.; May, H.; Laut, J.; Palmisano, J. Lactic acidosis associated with stavudine administration: A report of five cases. *Clin. Infect. Dis.* **2000**, *30*, 198–200. [CrossRef] [PubMed]
- 68. Duarte-Rojo, A.; Heathcote, E.J. Efficacy and safety of tenofovir disoproxil fumarate in patients with chronic hepatitis B. *Therap. Adv. Gastroenterol.* **2010**, *3*, 107–119. [CrossRef] [PubMed]
- 69. Chamroonkul, N.; Bansal, M.B. HIV and the liver. Nat. Rev. Gastroenterol. Hepatol. 2019, 16, 1–2. [CrossRef]
- Dharan, N.J.; Neuhaus, J.; Rockstroh, J.K.; Peters, L.; Gordin, F.; Arenas-Pinto, A.; Emerson, C.; Marks, K.; Hidalgo, J.; Sarmento-Castro, R.; et al. Benefit of Early versus Deferred Antiretroviral Therapy on Progression of Liver Fibrosis among People with HIV in the START Randomized Trial. *Hepatology* 2019, 69, 1135–1150. [CrossRef]
- Nassal, M. HBV cccDNA: Viral persistence reservoir and key obstacle for a cure of chronic hepatitis B. *Gut* 2015, 64, 1972–1984. [CrossRef]
- Lai, C.L.; Wong, D.; Ip, P.; Kopaniszen, M.; Seto, W.K.; Fung, J.; Huang, F.Y.; Lee, B.; Cullaro, G.; Chong, C.K.; et al. Reduction of covalently closed circular DNA with long-term nucleos(t)ide analogue treatment in chronic hepatitis B. *J. Hepatol.* 2017, 66, 275–281. [CrossRef]

- 73. Lebossé, F.; Inchauspé, A.; Locatelli, M.; Miaglia, C.; Diederichs, A.; Fresquet, J.; Chapus, F.; Hamed, K.; Testoni, B.; Zoulim, F. Quantification and epigenetic evaluation of the residual pool of hepatitis B covalently closed circular DNA in long-term nucleoside analogue-treated patients. *Sci. Rep.* 2020, *10*, 21097. [CrossRef]
- 74. Li, H.; Bai, R.; Zhao, Z.; Tao, L.; Ma, M.; Ji, Z.; Jian, M.; Ding, Z.; Dai, X.; Bao, F.; et al. Application of droplet digital PCR to detect the pathogens of infectious diseases. *Biosci. Rep.* **2018**, *38*, BSR20181170. [CrossRef]
- Liu, Y.; Jiang, M.; Xue, J.; Yan, H.; Liang, X. Serum HBV RNA quantification: Useful for monitoring natural history of chronic hepatitis B infection. BMC Gastroenterol. 2019, 19, 53. [CrossRef]
- 76. Chung, R.T.; King, W.C.; Ghany, M.G.; Lisker-Melman, M.; Hinerman, A.S.; Khalili, M.; Sulkowski, M.; Jain, M.K.; Choi, E.Y.K.; Nalesnik, M.A.; et al. A Prospective Cohort Study of Novel Markers of Hepatitis B Virus Replication in Human Immunodeficiency Virus Coinfection. *Clin. Gastroenterol. Hepatol.* 2022, *21*, 125–135. [CrossRef] [PubMed]
- 77. Hawkins, C.; Kang, M.; Bhattacharya, D.; Cloherty, G.; Kuhns, M.; Matining, R.; Thio, C.; Samaneka, W.; Chinula, L.; Mulinda, N.; et al. Hepatitis B surface antigen and hepatitis B RNA changes in HIV/hepatitis B virus co-infected participants receiving hepatitis B virus-active antiretroviral therapy. *AIDS* 2022, *36*, 975–984. [CrossRef] [PubMed]
- 78. Inoue, T.; Tanaka, Y. Novel biomarkers for the management of chronic hepatitis B. Clin. Mol. Hepatol. 2020, 26, 261–279. [CrossRef]
- 79. Xu, L.; Li, X.; Lu, L.; Liu, X.; Song, X.; Li, Y.; Han, Y.; Zhu, T.; Cao, W.; Li, T. HBV pgRNA profiles in Chinese HIV/HBV coinfected patients under pre- and posttreatment: A multicentre observational cohort study. *J. Viral Hepat.* **2022**, *29*, 616–626. [CrossRef]
- 80. Inoue, T.; Tanaka, Y. The role of hepatitis B core-related antigen. *Genes* **2019**, *10*, 357. [CrossRef]
- 81. Locarnini, S.; Zoulim, F. Molecular genetics of HBV infection. Antivir. Ther. 2010, 15, 3–14. [CrossRef]
- Kimura, T.; Ohno, N.; Terada, N.; Rokuhara, A.; Matsumoto, A.; Yagi, S.; Tanaka, E.; Kiyosawa, K.; Ohno, S.; Maki, N. Hepatitis B Virus DNA-negative Dane Particles Lack Core Protein but Contain a 22-kDa Precore Protein without C-terminal Arginine-rich Domain*. J. Biol. Chem. 2005, 280, 21713–21719. [CrossRef]
- Tseng, T.C.; Liu, C.J.; Yang, W.T.; Chen, C.L.; Yang, H.C.; Su, T.H.; Wang, C.C.; Kuo, S.F.T.; Liu, C.H.; Chen, P.J.; et al. Hepatitis B surface antigen level complements viral load in predicting viral reactivation in spontaneous HBeAg seroconverters. *J. Gastroenterol. Hepatol.* 2014, 29, 1242–1249. [CrossRef]
- Martinot-Peignoux, M.; Asselah, T.; Marcellin, P. HBsAg quantification to optimize treatment monitoring in chronic hepatitis B patients. *Liver Int.* 2015, 35, 82–90. [CrossRef]
- Li, Q.; Li, W.; Lu, C.; Huang, Y.; Chen, L. Serum hepatitis B surface antigen levels predict insignificant fibrosis and non-cirrhosis in hepatitis B e antigen positive patients with normal or mildly elevated alanine transaminase levels. *Oncotarget* 2017, *8*, 86463–86470. [CrossRef] [PubMed]
- 86. Sonneveld, M.J.; Hansen, B.E.; Brouwer, W.P.; Chan, H.L.Y.; Piratvisuth, T.; Jia, J.D.; Zeuzem, S.; Chien, R.N.; De Knegt, R.J.; Wat, C.; et al. Hepatitis B Surface Antigen Levels Can Be Used to Rule Out Cirrhosis in Hepatitis B e Antigen-Positive Chronic Hepatitis B: Results From the SONIC-B Study. J. Infect. Dis. 2022, 225, 1967–1973. [CrossRef] [PubMed]
- 87. Limothai, U.; Chuaypen, N.; Poovorawan, K.; Chotiyaputta, W.; Tanwandee, T.; Poovorawan, Y.; Tangkijvanich, P. Reverse transcriptase droplet digital PCR vs reverse transcriptase quantitative real-time PCR for serum HBV RNA quantification. *J. Med. Virol.* **2020**, *92*, 3365–3372. [CrossRef]
- Vachon, A.; Osiowy, C. Novel Biomarkers of Hepatitis B Virus and Their Use in Chronic Hepatitis B Patient Management. *Viruses* 2021, 13, 951. [CrossRef] [PubMed]
- 89. Dezanet, L.N.C.; Maylin, S.; Gabassi, A.; Rougier, H.; Miailhes, P.; Lascoux-Combe, C.; Chas, J.; Girard, P.M.; Delaugerre, C.; Lacombe, K.; et al. Kinetics of hepatitis B core-related antigen and anti-hepatitis B core antibody and their association with serological response in human immunodeficiency virus-hepatitis B coinfection. *J. Infect. Dis.* **2021**, *221*, 1826–1837. [CrossRef]
- Chevaliez, S.; Hézode, C.; Bahrami, S.; Grare, M.; Pawlotsky, J.M. Long-term hepatitis B surface antigen (HBsAg) kinetics during nucleoside/nucleotide analogue therapy: Finite treatment duration unlikely. J. Hepatol. 2013, 58, 676–683. [CrossRef]
- Chen, C.H.; Lu, S.N.; Hung, C.H.; Wang, J.H.; Hu, T.H.; Changchien, C.S.; Lee, C.M. The role of hepatitis B surface antigen quantification in predicting HBsAg loss and HBV relapse after discontinuation of lamivudine treatment. *J. Hepatol.* 2014, 61, 515–522. [CrossRef]
- 92. Jaroszewicz, J.; Reiberger, T.; Meyer-Olson, D.; Mauss, S.; Vogel, M.; Ingiliz, P.; Payer, B.A.; Stoll, M.; Manns, M.P.; Schmidt, R.E.; et al. Hepatitis B Surface Antigen Concentrations in Patients with HIV/HBV Co-Infection. *PLoS ONE* **2012**, *7*, e43143. [CrossRef]
- 93. Terrault, N.A.; Bzowej, N.H.; Chang, K.M.; Hwang, J.P.; Jonas, M.M.; Murad, M.H. AASLD guidelines for treatment of chronic hepatitis B. *Hepatology* **2016**, *63*, 261–283. [CrossRef]
- 94. World Health Organization. *Guidelines for the Prevention, Care and Treatment of Persons with Chronic Hepatitis B Infection;* World Health Organization: Geneva, Switzerland, 2015.
- Mao, W.; Sun, Q.; Fan, J.; Lin, S.; Ye, B. AST to Platelet Ratio Index Predicts Mortality in Hospitalized Patients With Hepatitis B-Related Decompensated Cirrhosis. *Medicine* 2016, 95, e2946. [CrossRef]
- 96. Yang, R.; Gui, X.; Ke, H.; Yu, X.; Yan, Y.; Xiong, Y. Accuracy of FIB-4 and APRI scores compared to transient elastography for liver fibrosis in patients with HIV and HBV co-infection. *Int. J. STD AIDS* **2023**, *34*, 18–24. [CrossRef] [PubMed]
- Bottero, J.; Lacombe, K.; Guéchot, J.; Serfaty, L.; Miailhes, P.; Bonnard, P.; Wendum, D.; Molina, J.M.; Lascoux-Combe, C.; Girard, P.M. Performance of 11 biomarkers for liver fibrosis assessment in HIV/HBV co-infected patients. *J. Hepatol.* 2009, 50, 1074–1083. [CrossRef] [PubMed]

- 98. Stockdale, A.J.; Phillips, R.O.; Beloukas, A.; Appiah, L.T.; Chadwick, D.; Bhagani, S.; Bonnett, L.; Sarfo, F.S.; Dusheiko, G.; Geretti, A.M. Liver Fibrosis by Transient Elastography and Virologic Outcomes after Introduction of Tenofovir in Lamivudine-Experienced Adults with HIV and Hepatitis B Virus Coinfection in Ghana. *Clin. Infect. Dis.* 2015, *61*, 883–891. [CrossRef]
- Ferraioli, G.; Filice, C.; Castera, L.; Choi, B.I.; Sporea, I.; Wilson, S.R.; Cosgrove, D.; Dietrich, C.F.; Amy, D.; Bamber, J.C.; et al. WFUMB Guidelines and Recommendations for Clinical Use of Ultrasound Elastography: Part 3: Liver. *Ultrasound Med. Biol.* 2015, 41, 1161–1179. [CrossRef] [PubMed]
- 100. Van de Putte, D.F.; Blom, R.; van Soest, H.; Mundt, M.; Verveer, C.; Arends, J.; de Knegt, R.E.; Mauser-Bunschoten, E.; van Erpecum, K. Impact of Fibroscan on management of chronic viral hepatitis in clinical practice. *Ann. Hepatol.* 2011, 10, 469–476. [CrossRef]
- 101. Liang, X.E.; Chen, Y.P. Clinical application of vibration controlled transient elastography in patients with chronic hepatitis B. *J. Clin. Transl. Hepatol.* **2017**, *5*, 368–375. [CrossRef]
- Livak, K.J.; Schmittgen, T.D. Analysis of relative gene expression data using real-time quantitative PCR and the 2^{-ΔΔCT} method. *Methods* 2001, 25, 402–408. [CrossRef]
- 103. Waidmann, O.; Bihrer, V.; Pleli, T.; Farnik, H.; Berger, A.; Zeuzem, S.; Kronenberger, B.; Piiper, A. Serum microRNA-122 levels in different groups of patients with chronic hepatitis B virus infection. *J. Viral Hepat.* **2012**, *19*, e58–e65. [CrossRef]
- 104. Murray, D.D.; Suzuki, K.; Law, M.; Trebicka, J.; Neuhaus Nordwall, J.; Johnson, M.; Vjecha, M.J.; Kelleher, A.D.; Emery, S. Circulating MIR-122 and MIR-200a as biomarkers for fatal liver disease in ART-treated, HIV-1-infected individuals. *Sci. Rep.* 2017, 7, 10934. [CrossRef]
- Mantri, C.K.; Pandhare Dash, J.; Mantri, J.V.; Dash, C.C.V. Cocaine Enhances HIV-1 Replication in CD4+ T Cells by Down-Regulating MiR-125b. *PLoS ONE* 2012, 7, e51387. [CrossRef]
- 106. Loureiro, D.; Tout, I.; Narguet, S.; Benazzouz, S.M.; Mansouri, A.; Asselah, T. miRNAs as Potential Biomarkers for Viral Hepatitis B and C. *Viruses* **2020**, *12*, 1440. [CrossRef]
- 107. Yousefpouran, S.; Mostafaei, S.; Manesh, P.V.; Iranifar, E.; Bokharaei-Salim, F.; Nahand, J.S.; Mirzaei, H.; Taran, M.; Babaei, F.; Sayad, B.; et al. The assessment of selected MiRNAs profile in HIV, HBV, HCV, HIV/HCV, HIV/HBV Co-infection and elite controllers for determination of biomarker. *Microb. Pathog.* 2020, 147, 104355. [CrossRef]
- 108. Toyoda, H.; Kumada, T.; Tada, T.; Sone, Y.; Kaneoka, Y.; Maeda, A. Tumor Markers for Hepatocellular Carcinoma: Simple and Significant Predictors of Outcome in Patients with HCC. *Liver Cancer* **2015**, *4*, 126–136. [CrossRef]
- Chuaypen, N.; Chittmittraprap, S.; Pinjaroen, N.; Sirichindakul, B.; Poovorawan, Y.; Tanaka, Y.; Tangkijvanich, P. Serum Wisteria floribunda agglutinin-positive Mac-2 binding protein level as a diagnostic marker of hepatitis B virus-related hepatocellular carcinoma. *Hepatol. Res.* 2018, 48, 872–881. [CrossRef]
- Kim, S.U.; Heo, J.Y.; Kim, B.K.; Park, J.Y.; Kim, D.Y.; Han, K.H.; Ahn, S.H.; Kim, H.S. Wisteria floribunda agglutinin-positive human Mac-2 binding protein predicts the risk of HBV-related liver cancer development. *Liver Int.* 2017, 37, 879–887. [CrossRef] [PubMed]
- 111. Yang, J.D.; Addissie, B.D.; Mara, K.C.; Harmsen, W.S.; Dai, J.; Zhang, N.; Wongjarupong, N.; Ali, H.M.; Ali, H.A.; Hassan, F.A.; et al. GALAD Score for Hepatocellular Carcinoma Detection in Comparison to Liver Ultrasound and Proposal of GALADUS Score. Cancer Epidemiol. Biomark. Prev. 2019, 28, 531. [CrossRef]
- 112. Yao, L.; Zhou, Y.; Sui, Z.; Zhang, Y.; Liu, Y.; Xie, H.; Gao, H.; Fan, H.; Zhang, Y.; Liu, M.; et al. HBV-encoded miR-2 functions as an oncogene by downregulating TRIM35 but upregulating RAN in liver cancer cells. *EBioMedicine* 2019, 48, 117–129. [CrossRef] [PubMed]
- Chavalit, T.; Nimsamer, P.; Sirivassanametha, K.; Anuntakarun, S.; Saengchoowong, S.; Tangkijvanich, P.; Payungporn, S. Hepatitis B Virus-Encoded MicroRNA (HBV-miR-3) Regulates Host Gene PPM1A Related to Hepatocellular Carcinoma. *MicroRNA* 2019, 9, 232–239. [CrossRef]
- 114. Zou, C.; Li, Y.; Cao, Y.; Zhang, J.; Jiang, J.; Sheng, Y.; Wang, S.; Huang, A.; Tang, H. Up-regulated MicroRNA-181a induces carcinogenesis in Hepatitis B virus-related hepatocellular carcinoma by targeting E2F5. *BMC Cancer* 2014, 14, 97. [CrossRef] [PubMed]
- 115. Fan, R.; Papatheodoridis, G.; Sun, J.; Innes, H.; Toyoda, H.; Xie, Q.; Mo, S.; Sypsa, V.; Guha, I.N.; Kumada, T.; et al. aMAP risk score predicts hepatocellular carcinoma development in patients with chronic hepatitis. *J. Hepatol.* 2020, *73*, 1368–1378. [CrossRef]
- 116. de Macedo Costa, A.P.; da Silva, M.A.C.N.; Castro, R.S.; de Oliveira Sampaio, A.L.; Júnior, A.M.A.; da Silva, M.C.; Ferreira, A.d.S.P. PAGE-B and REACH-B Predicts the Risk of Developing Hepatocellular Carcinoma in Chronic Hepatitis B Patients from Northeast, Brazil. *Viruses* 2022, 14, 732. [CrossRef]
- Galle, P.R.; Forner, A.; Llovet, J.M.; Mazzaferro, V.; Piscaglia, F.; Raoul, J.L.; Schirmacher, P.; Vilgrain, V. EASL Clinical Practice Guidelines: Management of hepatocellular carcinoma. *J. Hepatol.* 2018, 69, 182–236. [CrossRef] [PubMed]
- 118. Jin, W.; Lin, Z.; Xin, Y.; Jiang, X.; Dong, Q.; Xuan, S. Diagnostic accuracy of the aspartate aminotransferase-to-platelet ratio index for the prediction of hepatitis B-related fibrosis: A leading meta-analysis. *BMC Gastroenterol.* **2012**, *12*, 14. [CrossRef]
- Kim, B.K.; Kim, D.Y.; Park, J.Y.; Ahn, S.H.; Chon, C.Y.; Kim, J.K.; Paik, Y.H.; Lee, K.S.; Park, Y.N.; Han, K.H. Validation of FIB-4 and comparison with other simple noninvasive indices for predicting liver fibrosis and cirrhosis in hepatitis B virus-infected patients. *Liver Int.* 2010, 30, 546–553. [CrossRef] [PubMed]

- Berzigotti, A.; Tsochatzis, E.; Boursier, J.; Castera, L.; Cazzagon, N.; Friedrich-Rust, M.; Petta, S.; Thiele, M. EASL Clinical Practice Guidelines on non-invasive tests for evaluation of liver disease severity and prognosis—2021 update. J. Hepatol. 2021, 75, 659–689. [CrossRef]
- 121. Sterling, R.K.; Lissen, E.; Clumeck, N.; Sola, R.; Correa, M.C.; Montaner, J.; Sulkowski, M.S.; Torriani, F.J.; Dieterich, D.T.; Thomas, D.L.; et al. Development of a simple noninvasive index to predict significant fibrosis in patients with HIV/HCV coinfection. *Hepatology* 2006, 43, 1317–1325. [CrossRef]
- Yin, Z.; Zou, J.; Li, Q.; Chen, L. Diagnostic value of FIB-4 for liver fibrosis in patients with hepatitis B: A meta-analysis of diagnostic test. Oncotarget 2017, 8, 22944. [CrossRef] [PubMed]
- 123. Wai, C.T.; Greenson, J.K.; Fontana, R.J.; Kalbfleisch, J.D.; Marrero, J.A.; Conjeevaram, H.S.; Lok, A.S.F. A simple noninvasive index can predict both significant fibrosis and cirrhosis in patients with chronic hepatitis C. *Hepatology* 2003, *38*, 518–526. [CrossRef] [PubMed]
- 124. Lin, Z.H.; Xin, Y.N.; Dong, Q.J.; Wang, Q.; Jiang, X.J.; Zhan, S.H.; Sun, Y.; Xuan, S.Y. Performance of the aspartate aminotransferaseto-platelet ratio index for the staging of hepatitis C-related fibrosis: An updated meta-analysis. *Hepatology* 2011, 53, 726–736. [CrossRef]
- 125. Ma, J.; Jiang, Y.; Gong, G. Evaluation of seven noninvasive models in staging liver fibrosis in patients with chronic hepatitis B virus infection. *Eur. J. Gastroenterol. Hepatol.* **2013**, *25*, 428–434. [CrossRef]
- 126. Ucar, F.; Sezer, S.; Ginis, Z.; Ozturk, G.; Albayrak, A.; Basar, O.; Ekiz, F.; Coban, S.; Yuksel, O.; Armutcu, F.; et al. APRI, the FIB-4 score, and Forn's index have noninvasive diagnostic value for liver fibrosis in patients with chronic hepatitis B. *Eur. J. Gastroenterol. Hepatol.* **2013**, 25, 1076–1081. [CrossRef] [PubMed]
- 127. Iacob, D.G.; Luminos, M.; Benea, O.E.; Tudor, A.M.; Olariu, C.M.; Iacob, S.A.; Ruta, S. Liver fibrosis progression in a cohort of young HIV and HIV/HBV co-infected patients: A longitudinal study using non-invasive APRI and Fib-4 scores. *Front. Med.* 2022, 9, 2163. [CrossRef]
- 128. Yang, R.; Gui, X.; Ke, H.; Xiong, Y.; Gao, S. Combination antiretroviral therapy is associated with reduction in liver fibrosis scores in patients with HIV and HBV co-infection. *AIDS Res. Ther.* **2021**, *18*, 98. [CrossRef]
- Calès, P.; Oberti, F.; Michalak, S.; Hubert-Fouchard, I.; Rousselet, M.C.; Konaté, A.; Gallois, Y.; Ternisien, C.; Chevallier, A.; Lunel, F. A novel panel of blood markers to assess the degree of liver fibrosis. *Hepatology* 2005, 42, 1373–1381. [CrossRef]
- 130. Su, Z.; Chen, J.; Zhang, J.; An, Y.; Liao, Y.; Wu, X.; Tao, C.; Wang, L.; Cai, B. Circulating IL-1β, IL-17, and IP-10 as Potential Predictors of Hepatitis B Virus Infection Prognosis. *J. Immunol. Res.* **2022**, 2022, 5202898. [CrossRef] [PubMed]
- 131. Sultana, C.; Erscoiu, S.M.; Grancea, C.; Ceausu, E.; Ruta, S. Predictors of Chronic Hepatitis C Evolution in HIV Co-Infected Patients From Romania. *Hepat. Mon.* **2013**, *13*, e8611. [CrossRef]
- 132. Lei, J.; Yin, X.; Shang, H.; Jiang, Y. IP-10 is highly involved in HIV infection. Cytokine 2019, 115, 97–103. [CrossRef]
- 133. Sterling, R.K.; Wahed, A.S.; King, W.C.; Kleiner, D.E.; Khalili, M.; Sulkowski, M.; Chung, R.T.; Jain, M.K.; Lisker-Melman, M.; Wong, D.K.; et al. Spectrum of Liver Disease in Hepatitis B Virus (HBV) Patients Co-infected with Human Immunodeficiency Virus (HIV): Results of the HBV-HIV Cohort Study. Am. J. Gastroenterol. 2018, 114, 746. [CrossRef] [PubMed]
- 134. Maida, I.; Soriano, V.; Castellares, C.; Ramos, B.; Sotgiu, G.; Martin-Carbonero, L.; Barreiro, P.; Rivas, P.; González-Lahoz, J.; Núñez, M. Liver fibrosis in HIV-infected patients with chronic hepatitis B extensively exposed to antiretroviral therapy with anti-HBV activity. *HIV Clin. Trials* 2006, 7, 246–250. [CrossRef]
- 135. Audsley, J.; Robson, C.; Aitchison, S.; Matthews, G.V.; Iser, D.; Sasadeusz, J.; Lewin, S.R. Liver fibrosis regression measured by transient elastography in human immunodeficiency virus (HIV)-hepatitis B virus (HBV)-coinfected individuals on long-term HBV-active combination antiretroviral therapy. *Open Forum Infect. Dis.* **2016**, *3*, ofw035. [CrossRef]
- 136. Miailhes, P.; Pradat, P.; Chevallier, M.; Lacombe, K.; Bailly, F.; Cotte, L.; Trabaud, M.A.; Boibieux, A.; Bottero, J.; Trepo, C.; et al. Proficiency of transient elastography compared to liver biopsy for the assessment of fibrosis in HIV/HBV-coinfected patients. *J. Viral Hepat.* 2011, *18*, 61–69. [CrossRef]
- 137. Li, Q.; Chen, L.; Zhou, Y. Diagnostic accuracy of liver stiffness measurement in chronic hepatitis B patients with normal or mildly elevated alanine transaminase levels. *Sci. Rep.* 2018, *8*, 5224. [CrossRef] [PubMed]
- 138. Qi, X.; An, M.; Wu, T.; Jiang, D.; Peng, M.; Wang, W.; Wang, J.; Zhang, C.; Li, Z.; Liu, F.; et al. Transient Elastography for Significant Liver Fibrosis and Cirrhosis in Chronic Hepatitis B: A Meta-Analysis. *Can. J. Gastroenterol. Hepatol.* 2018, 2018, 3406789. [CrossRef] [PubMed]
- 139. Matthews, G.V.; Seaberg, E.C.; Avihingsanon, A.; Bowden, S.; Dore, G.J.; Lewin, S.R.; Sasadeusz, J.; Revill, P.A.; Littlejohn, M.; Hoy, J.F.; et al. Patterns and causes of suboptimal response to tenofovir-based therapy in individuals coinfected with HIV and hepatitis B virus. *Clin. Infect. Dis.* **2013**, *56*, e87. [CrossRef]
- 140. Liu, C.; Wang, L.; Xie, H.; Zhang, L.; Wang, B.; Luo, C.; Wang, S.; Tang, M.; Fu, Z.; Ruan, H.; et al. The relationship between serum hepatitis B virus DNA level and liver histology in patients with chronic HBV infection. *PLoS ONE* 2018, 13, e0206060. [CrossRef] [PubMed]
- 141. Iacob, D.G.; Rosca, A.; Ruta, S.M. Circulating microRNAs as non-invasive biomarkers for hepatitis B virus liver fibrosis. *World J. Gastroenterol.* 2020, *26*, 1113–1127. [CrossRef]
- 142. Li, F.; Zhou, P.; Deng, W.; Wang, J.; Mao, R.; Zhang, Y.; Li, J.; Yu, J.; Yang, F.; Huang, Y.; et al. Serum microRNA-125b correlates with hepatitis B viral replication and liver necroinflammation. *Clin. Microbiol. Infect.* **2016**, *22*, 384.e1–384.e10. [CrossRef]

- 143. Rosca, A.; Anton, G.; Botezatu, A.; Temereanca, A.; Ene, L.; Achim, C.; Ruta, S. miR-29a associates with viro-immunological markers of HIV infection in treatment experienced patients. *J. Med. Virol.* **2016**, *88*, 2132–2137. [CrossRef]
- Marrero, J.A.; Kulik, L.M.; Sirlin, C.B.; Zhu, A.X.; Finn, R.S.; Abecassis, M.M.; Roberts, L.R.; Heimbach, J.K. Diagnosis, Staging, and Management of Hepatocellular Carcinoma: 2018 Practice Guidance by the American Association for the Study of Liver Diseases. *Hepatology* 2018, 68, 723–750. [CrossRef]
- 145. Reichl, P.; Mikulits, W. Accuracy of novel diagnostic biomarkers for hepatocellular carcinoma: An update for clinicians (Review). Oncol. Rep. 2016, 36, 613–625. [CrossRef]
- 146. Toyoda, H.; Kumada, T.; Tada, T. Highly sensitive Lens culinaris agglutinin-reactive α-fetoprotein: A new tool for the management of hepatocellular carcinoma. *Oncology* **2011**, *81*, 61–65. [CrossRef]
- 147. Wan, Q.; Anugwom, C.; Desalegn, H.; Debes, J.D. Hepatocellular carcinoma in Hepatitis B and Human Immunodeficiency Virus coinfection in Africa: A focus on surveillance. *Hepatoma Res.* 2022, *8*, 39. [CrossRef] [PubMed]
- 148. Cevik, D.; Yildiz, G.; Ozturk, M. Common telomerase reverse transcriptase promoter mutations in hepatocellular carcinomas from different geographical locations. *World J. Gastroenterol.* **2015**, *21*, 311. [CrossRef] [PubMed]
- Kisiel, J.B.; Dukek, B.A.; Kanipakam, R.V.S.R.; Ghoz, H.M.; Yab, T.C.; Berger, C.K.; Taylor, W.R.; Foote, P.H.; Giama, N.H.; Onyirioha, K.; et al. Hepatocellular Carcinoma Detection by Plasma Methylated DNA: Discovery, Phase I Pilot, and Phase II Clinical Validation. *Hepatology* 2019, 69, 1180–1192. [CrossRef]
- Chalasani, N.P.; Ramasubramanian, T.S.; Bhattacharya, A.; Olson, M.C.; Roberts, L.R.; Kisiel, J.B.; Reddy, K.R.; Lidgard, G.P.; Johnson, S.C.; Bruinsma, J.J. A Novel Blood-Based Panel of Methylated DNA and Protein Markers for Detection of Early-Stage Hepatocellular Carcinoma. *Clin. Gastroenterol. Hepatol.* 2021, 19, 2597–2605. [CrossRef] [PubMed]
- 151. Zhu, Z.; Qin, Y.; Liang, Q.; Xia, W.; Zhang, T.; Wang, W.; Zhang, M.; Jiang, T.; Wu, H.; Tian, Y. Increased HBV Coinfection and Decreased IFN-γ-Producing HBV-Specific CD8+ T Cell Numbers During HIV Disease Progression. *Front. Immunol.* 2022, 13, 861804. [CrossRef]
- Da Silva, C.M.; de Peder, L.D.; Silva, E.S.; Previdelli, I.; Pereira, O.C.N.; Teixeira, J.J.V.; Bertolini, D.A. Impact of HBV and HCV coinfection on CD4 cells among HIV-infected patients: A longitudinal retrospective study. J. Infect. Dev. Ctries. 2018, 12, 1009–1018. [CrossRef]

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