

# Supplementary Materials

## Purification, Characterization, and Ferrous Oxidation Kinetics of Iron Oxidase from *Acidithiobacillus ferridurans*

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### Supplementary Text S1

**Text S1.** The regenerated and synthesized nucleotide gene sequence of the recombinant iron oxidase from *A. ferridurans* Riv11. The coding sequence of proteins is black and regenerated by codon optimization according to the amino acid sequence using the GenSmart™ Codon Optimization tool (<https://www.genscript.com.cn/tools/gensmart-codon-optimization>). The translation stop codon is in purple. The ligase sequences of the *NdeI* restriction site at 5'-terminal and the *HindIII* restriction site at 3'-terminal are in blue.

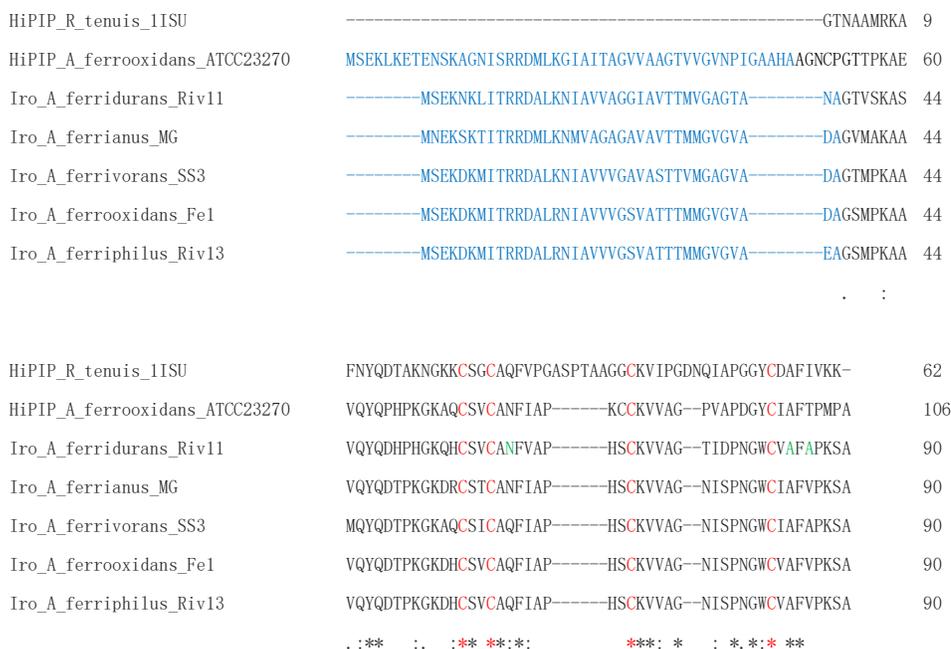
>The synthesized nucleotide sequence of the recombinant iron oxidase from *A. ferridurans* Riv11  
CATATGCATCACCACCACCACCATGGAACAGTATCTAAGGCGAGCGTCCAGTACCAGGATCATC  
CGCATGGTAAGCAACACTGCTCCGTGTGCGCGAACTTCGTGGCTCCGCACAGCTGCAAAGTTGT  
TGCCGGCACCATCGACCCGAATGGTTGGTGTGTGCGTTTGCGCCAAAAAGCGCATAAAGCTT

**Supplementary Table S1**

**Table S1.** Amino acid composition and property analysis of the recombinant iron oxidase from *A. ferridurans* Riv11. The hydrophobicity values of amino acid residue are from the Kyte and Doolittle hydrophobicity values [26]. The net charge values at pH 2.0 of amino acid residues are computed according to the pKa values in the initial pKa values for standard amino acids [27]. The total counts for amino acid residue counts and their buried counts and exposed counts do not include N-Terminal, C-Terminal or the [Fe<sub>4</sub>S<sub>4</sub>] cluster.

Amino acid residue	Count	Buried count	Exposed count	Hydrophobicity	Charge at pH 7.0	Charge at pH2.0	Total charges
Ala (A)	7	0	7	1.8	0	0	0
Arg (R)	0	0	0	-4.5	+1	+1	0
Asn (N)	2	0	2	-3.5	0	0	0
Asp (D)	2	0	2	-3.5	-1	0	0
Cys (C)	4	4	0	2.5	-1	-1	-4
Gln (Q)	3	0	3	-3.5	0	0	0
Glu (E)	0	0	0	-3.5	-1	0	0
Gly (G)	4	1	3	-0.4	0	0	0
His (H)	10	0	10	-3.2	0	+1	+10
Ile (I)	1	1	0	4.5	0	0	0
Leu (L)	0	0	0	3.8	0	0	0
Lys (K)	4	0	4	-3.9	+1	+1	+4
Met (M)	1	0	1	1.9	0	0	0
Phe (F)	2	0	2	2.8	0	0	0
Pro (P)	4	0	4	-1.6	0	0	0
Ser (S)	5	0	5	-0.8	0	0	0
Thr (T)	2	0	2	-0.7	0	0	0
Trp (W)	1	0	1	-0.9	0	0	0
Tyr (Y)	1	0	1	-1.3	0	0	0
Val (V)	7	1	6	4.2	0	0	0
N-Terminal	1	0	1		+1	+1	+1
C-Terminal	1	0	1		-1	0	0
[Fe <sub>4</sub> S <sub>4</sub> ]	1	1	0		+3	+3	+3
<b>Total</b>	<b>60</b>	<b>7</b>	<b>53</b>				<b>+14</b>

## Supplementary Figure S1



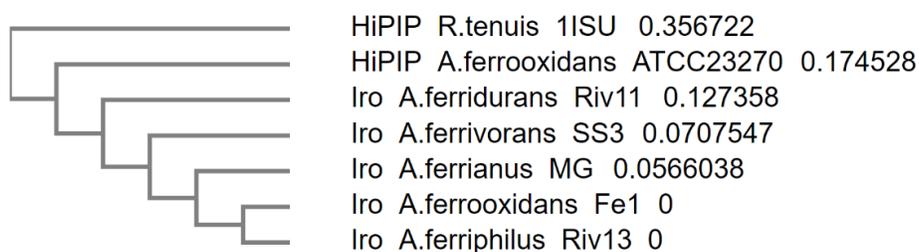
**Figure S1. The multiple sequence alignment for the wild proteins of iron oxidase and HiPIP from various sources.** HiPIP: high redox potential iron sulfur protein; Iro: iron oxidase. All the sequences can be obtained from NCBI Genbank (<https://www.ncbi.nlm.nih.gov/>) and their accession codes are as follows: Iro A. ferridurans Riv11:WP\_215878501;Iro A. ferrianus MG:WP\_239058863; Iro A. ferrivorans SS3:AEM47577;Iro A. ferrooxidans Fe1:CAA40594;Iro A. ferriphilus Riv13:WP\_215860796;HiPIP A. ferrooxidans ATCC23270:ACK79288;HiPIP R. tenuis(1ISU):P33678. The signal peptide sequece are in blue. Residues conserved in all sequences are marked with an \*. Residues not conserved in all sequences but conserved in some sequences are marked with (:), based on the degree of conservation. The cysteine residues ligating the [Fe<sub>4</sub>S<sub>4</sub>] cluster are red. The key residues for interaction with ferrous ion are in blue.

## Supplementary Figure S2

1: HiPIP_R_tenuis_1ISU	100.00	35.19	37.04	40.74	42.59	42.59	42.59
2: HiPIP_A_ferrooxidans_ATCC23270	35.19	100.00	54.44	55.56	56.67	53.33	53.33
3: Iro_A_ferridurans_Riv11	37.04	54.44	100.00	68.89	73.33	74.44	74.44
4: Iro_A_ferrianus_MG	40.74	55.56	68.89	100.00	77.78	80.00	78.89
5: Iro_A_ferrivorans_SS3	42.59	56.67	73.33	77.78	100.00	86.67	85.56
6: Iro_A_ferrooxidans_Fe1	42.59	53.33	74.44	80.00	86.67	100.00	98.89
7: Iro_A_ferriphilus_Riv13	42.59	53.33	74.44	78.89	85.56	98.89	100.00

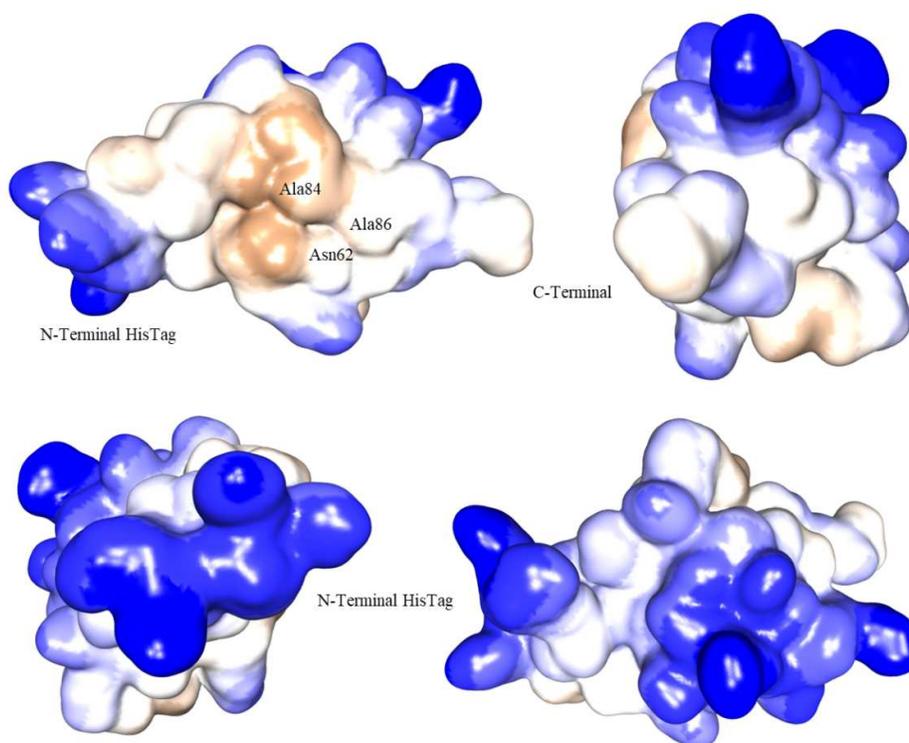
**Figure S2. The percent identity matrix for the sequences of iron oxidase and HiPIP from various sources.** The data were created by Clustal2.1 (<https://www.ebi.ac.uk/Tools/msa/clustalo/>) according to the multiple sequence alignment in Figure S1.

### Supplementary Figure S3



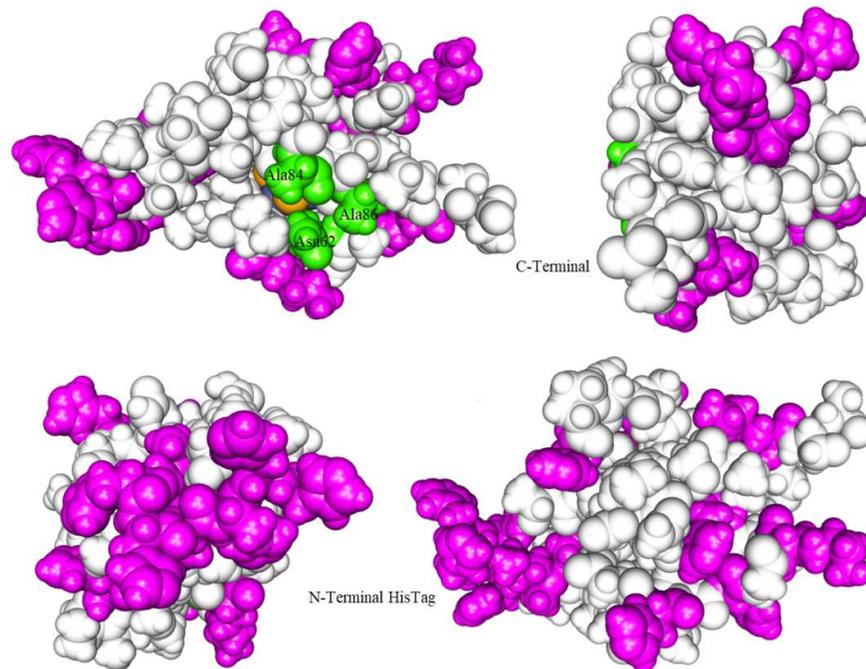
**Figure S3.** The phylogenetic tree for the sequences of iron oxidase and HiPIP from various sources. The data were created by Clustal2.1 (<https://www.ebi.ac.uk/Tools/msa/clustalo/>) according to the multiple sequence alignment in Figure S1 and the percent identity matrix in Figure S2.

### Supplementary Figure S4



**Figure S4.** The hydrophobic surface analysis of iron oxidase from *A. ferridurans*. The hydrophobic property of the molecular surface was rendered according to the level, with a hydrophobicity spectrum ranging from the most hydrophobicity in brown to the most hydrophilicity in blue.

### Supplementary Figure S5



**Figure S5.** The surface charge analysis of recombinant iron oxidase from *A. ferridurans*. The positively charged residues are in purple, the contact residues are in green and the other residues in the protein are in white.