

## Article

# Simultaneous Chemical and Sensory Analysis of Domestic Cat Urine and Feces with Headspace Solid-Phase Microextraction and GC-MS-Olfactometry

Chumki Banik <sup>1</sup>, Jacek A. Koziel <sup>1,\*</sup> and James Z. Li <sup>2</sup>
<sup>1</sup> Department of Agriculture and Biosystems Engineering, Iowa State University, 4350 Elings Hall, Ames, IA 50011, USA; cbanik@iastate.edu

<sup>2</sup> Nestle Purina, 1 CheckerBoard Sq., St. Louis, MO 63164, USA; James.li2@rd.nestle.com

\* Correspondence: koziel@iastate.edu; Tel.: +1-515-294-4206

**Abstract:** The association between humans and cats (*Felis catus*) is well known. This domestic animal is also known for its malodorous urine and feces. The complexity of the odorous urine and feces impacts human life by triggering the human sensory organ in a negative way. The objective of this research was to identify the volatile organic chemicals (VOCs) and associated odors in cat urine and feces using gas chromatography–mass spectrometry and simultaneous sensory analysis of fresh and aged samples. The solid-phase microextraction (SPME) technique was used to preconcentrate the VOCs emitted from urine or feces samples. Twenty-one compounds were identified as emitted from fresh urine, whereas 64 compounds were emitted from fresh feces. A contrasting temporal impact was observed in the emission of VOCs for urine and feces. On aging, the emission increased to 34 detected chemicals for stale urine, whereas only 12 chemicals were detected in stale feces. Not all compounds were malodorous; some compounds had a pleasant hedonic smell to the human nose. Although trimethylamine, low-molecular-weight organic acids, and ketones were contributors to the odor to some extent, phenolic compounds and aromatic heterocyclic organic N compounds generated the most intense odors and substantially contributed to the overall malodor, as observed by this study. This work might be useful to formulate cat urine and feces odor remediation approaches to reduce odor impacts.

**Keywords:** feline; smell; odor; SPME; GC-MS-O; VOCs; feline



**Citation:** Banik, C.; Koziel, J.A.; Li, J.Z. Simultaneous Chemical and Sensory Analysis of Domestic Cat Urine and Feces with Headspace Solid-Phase Microextraction and GC-MS-Olfactometry. *Separations* **2021**, *8*, 15. <https://doi.org/10.3390/separations8020015>

Academic Editor:

Bárbara Socas-Rodríguez

Received: 31 December 2020

Accepted: 27 January 2021

Published: 31 January 2021

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Copyright:** © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

The companionship between humans and cats (*Felis catus*) is more than 8000 years old [1]. The market research statistics by the American Veterinary Medical Association counted 74 million domestic cats in the USA in a report presented in 2012 [2]. This popular companion of humans builds importance in human life as a family member. While cat owners love their cats, they have a less positive relationship with their cats' litterboxes due to several factors, including the smell of urine and feces.

The potent odor of domestic cat urine causes a continually growing research interest (Table 1). Improved separation and identification techniques were used over the decades to report compounds with a catty smell. One of the responsible specific amino acids, feline, excreted by the Felidae family does not have a specific odor, but the degradation products of feline are odorous. Fractionation and separation of feline and its derivatives were done using paper chromatographic techniques and spot tests in earlier studies [3]. Using GC–MS total ion chromatogram of the cat urine headspace analysis, Miyazaki et al. (2006) identified a total of 25 compounds in male domestic cat urine [4]. They reported the urinary protein Cauxin to be involved in feline production. Feline, the sulfur-containing amino acid, is carried in the cat bloodstream as 3-methylbutanol-glutathione (MBG) [5]. An increase in testosterone concentration can increase the free feline in the male and female

cat [6] because testosterone increases the production of MBG and shifts the distribution of MBG metabolites towards the generation of free felinine. In addition to felinine, several organic chemicals can be emitted from cat urine and feces, depending on the age and sex-related factors of cats.

Cat urine and feces contain several volatile and nonvolatile compounds that help to recognize sex and species [7]. These volatile compounds emitted through urine and feces also act as chemical signaling in mammals to define their territory, dominance, and reproduction [8,9]. Stray and domestic cats also use urine as chemical signaling and to bury their feces around the home range [10], and that odor of cat urine and feces can be annoying to humans. Research articles have been published that are focused more on the odorous components in cat urine and less on feces, although both waste products are putrid. The concentration of VOCs emitted by cat feces can significantly differ with cat age and sex irrespective of their food diet or habitat, such as 1-butanol in feces found significantly in lower concentrations in female cats, and indoles and phenols such as odorous compounds can increase with the age of male cats. Moreover, the aging of the cat urine and feces emits odorous chemicals.

A recent study by Suzuki et al., 2019, reported a significant impact of time (fresh and 24 h) on VOC released from the same urine sample, and the reason was provided to be the degradation of VOCs by bacteria in urine, urinary enzymatic reaction, or oxidation [11]. Fresh cat urine does not emit a strong odor and can be described as “ammonia-like” and “savory-like”; however, on interaction with soil, the bacteria can emit a cat urine smell described as “intensely fishy” [12]. These experiments were set to find the cat species chemical signaling for habituation–dishabituation and may not guide the resolution of odor issues for human annoyance. Simultaneous chemical identification and sensory analysis of the VOC data from cat excrete by the human nose is still limited; moreover, a simple technique and temporal data set are always in need to build the odor profile emitted from cat urine and feces. The water intake by cats can vary with their food diet, and the volume of water intake can end up in a release of different amounts of urine or feces samples [13].

It has only been a decade since scientists started using solid-phase microextraction (SPME) for biological sample VOC extraction. It is considered a noninvasive sampling device that extracts biomarkers for the early diagnosis of advanced or chronic diseases or for reporting impurity in food samples to assess food quality [14]. That study also reported that SPME is 10–50 times more efficient than any static headspace sampling. Moreover, the use of SPME is able to extract volatile organic compounds (VOCs) from a biological sample both *ex vivo* and *in vitro* for analysis using gas chromatography–mass spectrometry (GC–MS). This approach of sample extraction can reduce sample preparation steps and can extract the chemicals without modifying their original form. The use of SPME fibers has only recently been reported for marking fluid extraction and identification for *Panthera tigris* subspecies [8]. To our knowledge, there is no research published on odorous chemicals emitted by urine and fecal samples of domestic cat species using SPME–GC–MS–olfactometry (SPME–GC–MS–O) chemical and sensory analysis, especially in the context of smell development over time. Simultaneous chemical and sensory analyses can facilitate linking conventional chemical speciation with specific odors. Chemical identification of odor-causing chemicals can be aided by odor databases [15,16]. This current study was designed to find out the temporal behavior of odorous compounds emitted from cat urine and feces in a noninvasive way for the betterment of the human environment.

**Table 1.** Literature related to domestic cat urine and feces sample preparation for chemical and sensory analysis to identify and compare the chemical constituents of cat excretes.

Species	Sample Type	Sample Preparation	Chemical Analysis	Sensory Analysis	Identified Compounds	Study Purpose	Reference
Domestic Cat ( <i>Felis catus</i> )	Urine	Solvent extraction and derivatization	Paper chromatography	Not conducted	Felinine and amino acids	To separate felinine and its derivatives	Westall, R.G. 1953 [3]
Domestic Cat ( <i>Felis catus</i> )	Urine	Solvent extraction and membrane concentration	HPLC, GC-MS-headspace	Not conducted	Carboxylesterases cauxin, felinine, and felinine derivatives	To identify hydrolyzed products of cauxin and degradative products of felinine	Miyazaki, M. et al. 2006 [4]
Domestic Cat ( <i>Felis catus</i> )	Urine and blood	Solvent extraction and derivatization	HPLC	Not conducted	Felinine, N-acetylfelinine, creatinine, testosterone, and estradiol	To quantify felinine and NAcFel and report effects of testosterone and estradiol on free felinine, NAcFel, and c-glutamylfelinylglycine	Hendricks, W.H. et al. 2008 [6]
Domestic cat ( <i>Felis catus</i> )	Urine and soiled urine	Solvent extraction	UPLC-MS, GC-MS-O, and NMR	GC-MS-O	34 volatile and nonvolatile chemicals synthesized and reported and 14 odor attributes reported	To identify the key odorants in cat urine	Starkenmann, C. et al. 2014 [12]
Domestic Cat ( <i>Felis catus</i> )	Fresh feces	Gas sampling	GC-MS	Not conducted	24 volatile organic compounds	To determine sex and age using volatile compounds emitted from fecal samples	Uetake, K. et al. 2017 [10]
Domestic Cat ( <i>Felis silvestris catus</i> )	Anal sac secretions	Sorbed onto Tenax TA tube	TD-GC-MS	Olfactory of cats	10 free fatty acids	To determine behavioral bioassays via olfactory habituation–dishabituation	Miyazaki, T. et al. 2018 [17]
Domestic Cat ( <i>Felis silvestris catus</i> )	Fresh and up to 24 h aged cat urine	GC X GC-MS VOC preconcentrator	GC-MS	Olfactory of cats	36 compounds	To discriminate temporal changes and individual differences in urine	Suzuki, S. et al. 2019 [11]

HPLC: High-performance liquid chromatography; GC-MS: Gas chromatography; UPLC-MS: Ultra-performance liquid chromatography–mass spectrometry, NMR: Nucleic magnetic resonance.

The current study's objective is to use SPME fiber extraction to identify odorous VOCs emitted from fresh and aged urine and feces of domestic cat species. This SPME–GC–MS–O will simplify the chemical and sensory characterization of odorous compounds emitted from cat urine and feces. It may also answer the following question: Is it possible to predict odor intensity using GC–MS chemical analysis? This study might also help formulate cleaners and other remediation techniques to reduce cat urine and feces odor problems in the long term.

## 2. Materials and Methods

### 2.1. Cat Urine and Feces Collection

The cat urine and feces samples were collected at Nestlé Purina pet facility. The cat urine and feces samples were collected from a group of healthy cats ( $N > 10$ ). The urine and feces samples were immediately frozen after collection. The urine and feces samples in this study represent a typical healthy cat. Freely collected urine and feces samples were homogenized and immediately frozen at  $-20\text{ }^{\circ}\text{C}$  upon collection. The urine and feces samples were shipped in a cooler box with dry ice protection via next day air transport to the Iowa State University lab for analysis.

### 2.2. Sample Storage, Preparation, and Extraction

After receiving the samples from the sample collector, all urine and feces samples were stored at  $-20\text{ }^{\circ}\text{C}$  until they were analyzed. A week before analysis, they were moved to a freezer at  $-4\text{ }^{\circ}\text{C}$  and thawed on the analysis day in the morning at lab temperature ( $24\text{ }^{\circ}\text{C}$ ) for 3–4 h. Each sample was then weighed to approximately 1 g into an amber color 10 mL vial in duplicate using a disposable dropper or spatula. Temperature facilitates the organic molecules to move from the urine or feces to headspace and then SPME fiber coating. However, a high T can thermally decompose compounds such as trimethylamine (a potent odorant). Too high of a T could diminish the extraction efficiency of the SPME fiber coating. On the other hand, a low T might challenge the SPME extraction of the high-molecular-weight compounds. Thus, the temperature used in this experiment was close to body temperature. Considering these T factors, the aging of urine and feces was performed at room temperature, and the SPME extraction was performed at  $37\text{ }^{\circ}\text{C}$ .

We relied on previous methods developed in our lab (Soso and Koziel, 2016 [8]). For example, Soso and Koziel, 2016, [8] tested five types of SPME fiber coatings (75  $\mu\text{m}$  Carboxen polydimethylsiloxane (PDMS), 85  $\mu\text{m}$  Carboxen/PDMS, 65  $\mu\text{m}$  PDMS/(divinylbenzene) DVB, 50/30  $\mu\text{m}$  DVB/Carboxen/PDMS, and 100  $\mu\text{m}$  PDMS), two time intervals (1 h and 24 h), two sample sizes, and two Ts. Among the fiber coatings, DVB/Carboxen/PDMS was the least effective in extracting the characteristic smell of marking fluid of the Siberian tiger. However, the DVB/PDMS/Carboxen fiber was chosen because our objective was to record all the odors and chemicals emitted by the fresh and stale cat urine or feces samples rather than any specific chemical.

### 2.3. Multidimensional Gas Chromatography–Mass Spectrometry Olfactometry

All sample analyses of cat urine and feces were completed using the multidimensional gas chromatograph–mass spectrometer olfactometer (GC–MS–O; Microanalytics, Round Rock, TX, USA). All compounds emitted from the sample vial headspace were extracted using SPME fiber of 2 cm 50/30  $\mu\text{m}$  DVB/PDMS/Carboxen (57248-U, Supelco, Bellefonte, PA, USA). The samples were heated to  $37\text{ }^{\circ}\text{C}$  during extraction to enhance the emissions. A 50 min extraction time was used for all of the extractions, except an additional 10 min extraction was done for a fresh urine sample for comparison purposes. A schematic of the method is given in the appendix (Figure A1).

All the cat urine vials were kept at lab temperature ( $24\text{ }^{\circ}\text{C}$ ) for two weeks, and the feces samples were aged at lab temperature for one week for extraction of the aged urine and feces samples. These vial caps were left closed to avoid samples drying out and were opened for a couple of minutes of air every other day to avoid complete anaerobic

situations. On the analysis day, the vials were closed for the VOCs to equilibrate and accumulate for an hour under lab condition, and then, the vial was put on a hot plate set at 37 °C for 10 min before inserting the SPME fiber to extract the headspace VOCs for 50 min.

After extraction, the SPME fiber was loaded with odorants inserted into the 260 °C GC injector for thermal desorption of samples to the GC columns and separation and analysis using MS and an olfactometer. The GC–MS–O analysis was performed on an Agilent 6890 GC with a restrictor guard column, non-polar capillary column (BP-5, 30.0 m × 530 µm inner diameter × 0.5 µm thickness, SGE, Austin, TX, USA) and polar capillary column (BP-20, 30.0 m × 530 µm inner diameter × 0.5 µm thickness, SGE, Austin, TX, USA) connected in series. The sample flow was split 3:1 via an open split interface to an olfactometry port and mass spectrometer, respectively. The GC oven temperature was programmed at the initial 40 °C for 3 min, followed by ramping up to 240 °C at a rate of 7 °C/min, where it was maintained for 8.43 min. The quadrupole MS used an electron ionization mode with ionization energy of 70 eV during operation, and the full scan range was 34 to 350 *m/z*.

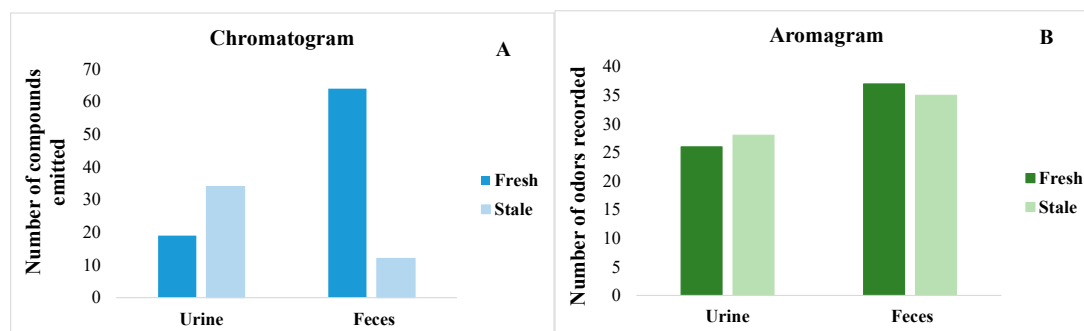
The odor event was detected by the panelist, and the aromagram peak is the intensity of the aroma event. The trained panelist recorded the start and end of the odor event, a description of the odor event, and the odor intensity. The odor intensity was evaluated on a 0–100% scale, where 0% means no odor and 100% means the strongest odor detected by the panelist. A humidified air was constantly delivered at a rate of 5.7 psi to the panelist's nose to reduce the dry out of the mucus membrane during the analysis. Aromagrams for odors were generated using Aromagram software (version 6.0, Microanalytics, Round Rock, TX, USA).

Analysis of the compounds and data files were generated from Agilent Chemstation software, and the peaks were identified using PBM-Benchtop software and matched using Wiley 7 and NIST database library.

### 3. Results

#### 3.1. Identification of Volatile Organic Compounds in Cat Urine and Feces Using GC–MS–O

The use of simultaneous sensory analyses (via GC–MS–olfactometry) enabled the detection of malodors that GC–MS could not detect (Figure 1). For example, the 10 min equilibration and 50 min SPME extraction of 1-week-old stale feces headspace had only 12 detectable compounds (via GC–MS), while the use of human nose enabled the detection of as many as 35 distinct odors (via GC–MS–olfactometry). However, for fresh feces, GC–MS detected as many as 64 compounds, but the human nose (via GC–MS–olfactometry) detected 37 distinct compounds. Tables 1 and 2 contain the odor descriptions for the chemical compounds matched with the NIST and Wiley7 chemical library. A list of the odor descriptions is provided in the Supplementary Materials (Tables S1–S6). The difference in the number of events occurs because a human nose is more sensitive compared to a chemical analyzer. Moreover, each chemical can possess a distinct aroma or aroma pattern and more than one chemical could have a similar aroma.



**Figure 1.** Comparison of the number of compounds detected in the headspace of fresh/stale urine and feces using GC–MS analysis ((A) chromatogram) and GC–MS–olfactometry analysis ((B) aromagram): the solid-phase microextraction (SPME) fiber was exposed for 50 min at 37 °C.

**Table 2.** Compounds identified in the headspace of fresh and stale cat urine on short and long exposures to the SPME fiber.

Compounds	Retention Time (min)	Odor Description	Odor Description Panelist	% Match with Nist and WILEY7	CAS #	Fresh Urine, 10 Min Exposure to SPME at 24 °C	Fresh Urine, 50 Min Exposure to SPME at 37 °C	Stale Urine, 50 Min Exposure to SPME at 37 °C	Ion (% Relative Intensity)
						MS Detector Response, Peak Area Counts (PACs), and Arbitrary Units			
Carbon disulfide	3.09			72	75-15-0			52,104	76(100), 58(10), 78(8), 44(5)
Acetone	3.13	Fruity <sup>b</sup> , Camphor	Soil, fruity	74	67-64-1	236,687	1,222,443	2,285,446	43(100), 58 (30), 42(8)
Propanal, 2-methyl-	3.38	Pungent <sup>a</sup> , malt		79	78-84-2			66,637	41(100). 43(70), 72(70), 40(40)
2-Butanone	4.00		Fruity	79	78-93-3	632,475	4,102,112	2,778,630	43(100), 72(30), 57(8), 42(5)
Butanal, 3 methyl-	4.62	Cocoa <sup>a</sup> , almond		93	590-86-3			378,438	41(100), 44(80), 43(78), 58(50), 39(45)
2-Butanone, 3methyl-	4.71		Sweet, chemical	72	563-80-4	516,109	2,132,140	1,126,561	43(100), 86(20), 41(18)
Silanol, trimethyl-	5.08			76	1066-40-6			381,502	75(100), 45(30), 47(12), 76(5)
2-Pentanone	5.4		grassy	72	107-87-9	1,448,327	7,777,565	2,602,556	43(100), 86(20), 41(12), 71(10)
2-Pentanone, 4-methyl	6.34			86	108-10-1		518,121	59,465	43(100), 58(40), 42(25), 57(25),85(22)
2-Pentanone, 3 methyl	6.64			83	565-61-7	1,846,692	7,433,319	1,662,796	43(100), 57(40), 41(35), 72(30)
Dimethyl, disulfide	7.07	Onion <sup>a</sup> , putrid		93	624-92-0		684,042		94(100), 45(50), 79(50), 46(25)
Pyrazine	8.04			83	290-37-9			747,153	80(100), 53(45), 58(40), 52(15), 51(10)
3-Pentanone 2,2-dimethyl	8.7		Burnt	58 <sup>#</sup>	564-04-5			121,125	57(100), 41(22), 114(10), 86(5)
3-Buten-1-ol, 3-methyl-	9.18			63	763-32-6			196,830	41(100), 56(90), 68(80), 86(25)
4-Heptanone	9.80			86	123-19-3	172,967	730,516		43(100), 71(90), 41(30), 114(20)
2-Propanol, 1-propoxy	10.2			50 <sup>#</sup>	1569-01-3			2,845,241	45(100), 43(95), 41(50), 73(48), 42(40)
Pyrazine, methyl-	10.36	Popcorn <sup>a</sup>	Sweet	93	109-08-0			329,275	94(100), 67(50), 40(20), 39(18), 53(12)
2-Heptanone	10.59			81	110-43-0	94,433	420,707		43(100), 58(75), 71(20), 114(7)
Prenol	10.63			74	556-82-1			226,665	71(100), 41(60), 39(55), 53(50), 67(30), 68(30)
3-Heptanone, 2-methyl-	11.03			74	13019-20-0	69,259	225,824	111,557	57(100), 43(75), 85(75), 41(60), 71(50)
Pyrazine, 2,5-dimethyl	12.25	Roast beaf <sup>a</sup> , medicine, cocoa		93	123-32-0			2,865,550	108(100), 42(70), 39(35), 40(20), 81(15)
Cyclohexane, ethyl-	13.26			59 <sup>#</sup>	1678-91-7	181,554	1,793,164	952,512	83(100), 55(70),112(45), 41(40), 56(40)
Limonene	13.7	Camphor, lemon, orange, citrus		97	138-86-3	142,286	1,182,493		68(100), 93(85), 67(80), 79(45)

Table 2. Cont.

Compounds	Retention Time (min)	Odor Description	Odor Description Panelist	% Match with Nist and WILEY7	CAS #	Fresh Urine, 10 Min Exposure to SPME at 24 °C	Fresh Urine, 50 Min Exposure to SPME at 37 °C	Stale Urine, 50 Min Exposure to SPME at 37 °C	Ion (% Relative Intensity)
Pyrrole	13.96			94	109-97-7			1,226,880	67(100), 39(48), 41(43), 40(31), 38(18)
Benzaldehyde	15.3		Sweet, fruity	95	100-52-7			1,331,202	106(100), 105(95), 77(90), 51(45), 50(30)
N-Acetyl pyrrole	15.43			83	609-41-6			2,069,470	67(100), 109(45), 43(32), 40(20), 39(20)
2-Methyl-4-decanone	17.60			70 NIST only		189,683	104,228		57(100), 85(95), 43(85), 95(85), 41(80)
Pyrimidine	17.86			50	289-95-2			497,501	80(100), 43(50), 123(30), 57(25)
Acetophenone	17.88	Must, flower, almond	smoke	88	98-86-2		104,228		105(100), 77(80), 51(30), 120(28)
Methoxy phenyl oxime	18.39			63		422,536	1,904,766	636,563	133(100), 151(60), 135(22)
5-Methyl-2 thiophenecarboxaldehyde	18.93		Plastic, burnt	74	13679-70-4			49,628	125(100), 126(88), 97(60), 53(20), 45(20)
3,3-dimethyl-4 thiapentan-1-ol	19.08			81				93,069	69(100), 41(88), 134(30), 39(30), 89(20), 56(20)
1,2-Ethanediol, 1-phenyl-	19.66			63	93-56-1			107,385	79(100), 107(95), 77(80), 51(40)
Benzyl alcohol	20.42	Sweet, flower		91	100-51-6			560,858	79(100), 108(40), 77(32), 94(30)
Dimethyl sulfone	20.46	Sulfur, burnt	Smoke, butter	71	67-71-0		158,852		79(100), 94(45), 45(20), 108(15)
Phenol	22.12	Phenol, Plastic, rubber		91	108-95-2			21,533	94(100), 66(30), 39(25), 65(20)
p-Cresol	23.37	Medicine, smoke	Smokey	93	106-44-5		228,817	651,731	107(100), 108(80), 77(20), 39(20)
Jasmone	24.13	Jasmine, flower		96	488-10-8			358,180	79(100), 164(80), 91(70), 110(60), 41(50), 149(50)
4-Hydroxy-2nonenoic acid	24.68	Minty		63	21963-26-8			53,921	84(100), 55(80), 43(50), 125(40), 41(40)
Butylated hydroxytoluene	25.61			82	128-37-0		41,207		205(100), 57(48), 220(20), 41(20)
p-Acetylaniline	25.9		Foul, urinous	94	99-92-3			102,642	120(100), 135(60), 92(48), 65(35), 43(10)
Indole	28.38	Burnt, mothball	Smokey, animal	93	120-72-9		60,593	155,847	117(100), 90(40), 89(39), 45(20)

# Below 60% library match considered as semi-confirmative. Odor verified with <sup>a</sup> Flavornet [15] and <sup>b</sup> Good Scent Company [16].



The refrigerated fresh urine produced 21 compounds and 26 odor events, whereas the 15-day-old urine produced as many as 34 compounds and the number of odor events was 28. Not all compounds could be classified as “malodors”. Some of the compounds had a “pleasant” hedonic smell, even in stale urine and feces samples. Phenols and indoles were among the most intense odors and were a substantial contributor to malodors.

### 3.2. Temporal Effect on Volatile Organic Compounds in Cat Urine

Exposure of the SPME fiber to the urine sample improved the accumulation of several chemical compounds. A faint odor was recorded by the panelist, but the description was missing at this low concentration. The fresh urine had a mild odor and, upon short SPME exposure, did not reveal much information on odorous compounds. A short exposure of the fiber to the urine sample extracted most of the low-molecular-weight VOCs that a long exposure time of SPME extracted. However, we were not able to detect the high-molecular-weight VOCs with short SPME extractions, and therefore, the high-molecular-weight VOCs are absent in the list (Table 2). Moreover, the intensity of the many chemicals was low in concentration (low Peak Area Count (PAC)) to distinguish the odor between different chemicals. The use of the GC–MS–olfactometer, however, sensed the odors of phenolic compounds at a short SPME exposure to the hedonic urine sample, although the odor intensity was low.

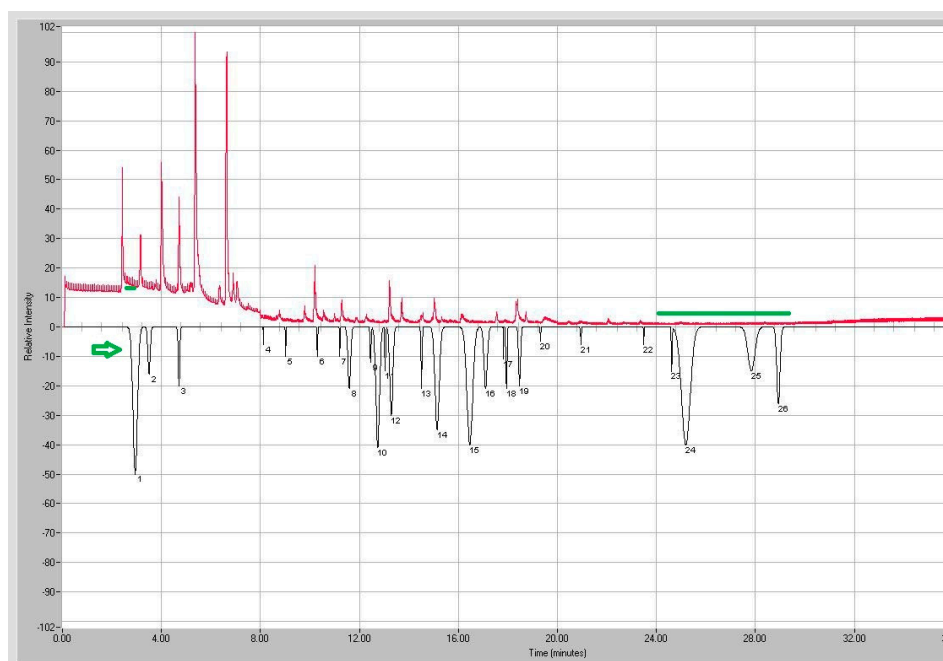
A dynamic aging temporal change was observed in the number of chemical compounds, odor events, and odor intensity observed in the headspace analysis of the urine and feces samples. Fresh cat urine has a weak odor intensity and odors described as urine, indole, and animal-like. Stale urine has many intense odorous compounds (Table 2). A foul smell was recorded by the panelist at retention time (RT) for 2.7 min, possibly of the compound trimethyl amine, although no compound was identified by the GC–MS. The identified compounds increased from 19 to 34, and it is worth mentioning that N-containing compounds like pyrazine; pyrrole; pyrimidine; and some other ketones, aldehyde, and alcohols emerged with the aging of the urine, whereas dimethyl disulfide-like malodor compounds were only present in fresh urine. However, the intensity of the odor for the compounds present in the aged urine was higher than the fresh urine, as noticed by increasing the PAC. The presence of 2-heptanone (“fruity” smell) and limonene (“mint-like” smell) was observed in fresh urine samples for both long and short extraction times that was missing in the aged urine samples; however, jasmone (“flowery” smell), a pleasant aroma, emerged in the aged urine sample and was absent in the fresh urine (Table 2).

The trace amount of odorous phenolic compounds had high odor intensity in both fresh and stale urine. The odor intensity of some compounds present in trace amount was substantially higher in the green line region in Figure 2; this odor intensity was also recorded in stale cat urine, as revealed by the aromagrams from GC–MS–olfactometry. It is quite evident that time and possibly temperature are factors in releasing odor to the atmosphere from cat urine or feces samples as both time and temperature are drivers to diffuse these VOCs and help to move from a source to a sensory organ.

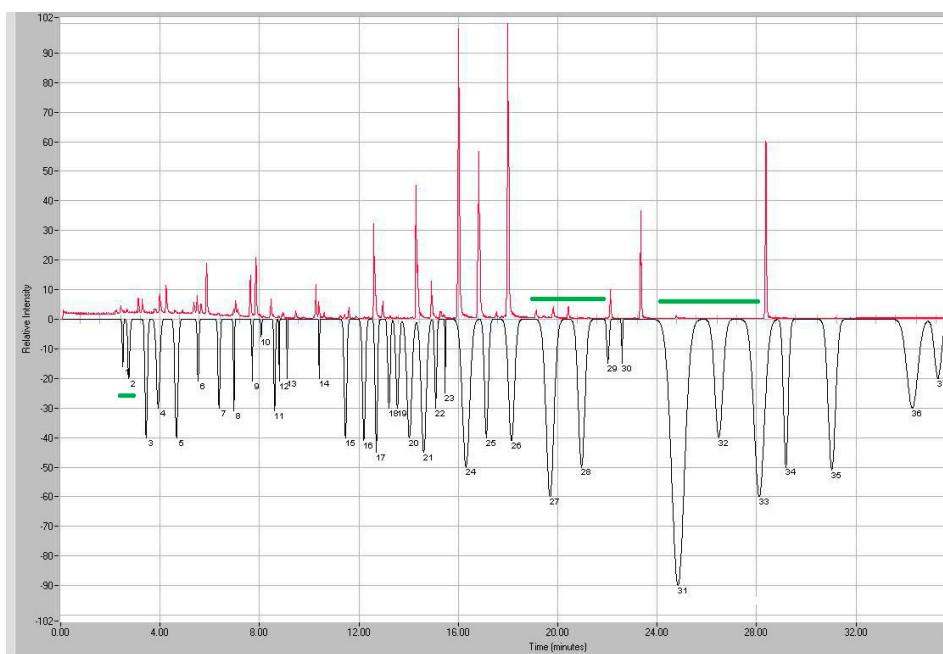
### 3.3. Temporal Effect on Volatile Organic Compounds in Cat Feces

Several volatile fatty acids (VFAs) and phenolic compounds contributed to overall cat feces odors. However, the phenolics appreciably contributed to the overall feces odor. The aged feces (1 week old) showed a significant drop in the number of emitted compounds in the headspace (Table 3), and therefore, the aging process was not further carried out. The VFAs that dominated in the cat feces were isobutyric, propanoic, butanoic, hexanoic, and acetic. Among the phenolics, phenol, p-cresol, and guaiacol, and the aromatic heterocyclic 1H-indole and 3-methyl indole were the contributors to overall odor. The stale cat feces had trimethylamine, a rotten fish-like odor that was not identified by GC–MS in fresh feces sample. However, a fish-like foul odor was recorded by the odor panelist (Figures 3 and 4). The observation supports the fact that a trace level of concentration can be sensed by living sensory organs.





**Figure 2.** An overlay of the aromagram and total ion chromatogram of the **fresh** cat urine exposed to SPME fiber for 50 min at 37 °C: an increase in the black signal height represents an increased intensity of odor, and the chromatogram is in red for total ion chromatogram (TIC) signal. The green zone shows that trace levels of the concentration of malodors can produce an intense olfactory response detected by the panelist using GC–MS–O.



**Figure 3.** An overlay of the aromagram and total ion chromatogram of the **fresh** cat feces exposed to the SPME fiber for 50 min at 37 °C: an increase in the black signal height represents an increased intensity of odor, and the chromatogram is in red for the total ion chromatogram (TIC) signal. The green zone shows that trace levels of the concentration of malodors can produce an intense olfactory response detected by the panelist using GC–MS–O.

**Table 3.** Compounds identified in the headspace of fresh and stale cat feces.

Compounds	Retention Time (min)	Odor Description Published Work	Description by Panelist	% Match Library	CAS #	Fresh Feces *	Stale Feces ** Short Equilibrium	Stale Feces ^^ Long Equilibrium	Ion (% Relative Intensity)
MS Detector Response, Peak Area Counts (PACs), and Arbitrary Units									
Trimethylamine	2.76	Fish <sup>a</sup>	Foul, fishy	83	75-50-3		1,395,179	1,237,586	58(100), 59(40), 42(32), 57(5)
Acetone	3.14	Fruity <sup>b</sup> , Camphor	Chemical, sweet	63	67-64-1	2,036,156			43(100), 58(30), 42(5)
Acetic acid, methyl ester	3.28			80	79-20-9	1,774,983			43(100), 74(30), 59(10)
2-Butanone	4.01			70	78-93-3	4,100,865			43(100), 72(20), 57(5), 42(5)
Methyl propionate	4.26		butter	88	554-12-1	4,300,327			57(100), 88(40), 59(30), 45(5)
Butanal, 3-methyl-	4.62	Fruity <sup>a</sup> , nutty		68	590-86-3			172,874	43(100), 39(62), 44(60), 58(35), 71(20), 86(20)
Butanal, 2-methyl-	4.70			59 <sup>#</sup>	96-17-3			64,220	41(100), 57(75), 58(60)
Butanoic acid, methyl ester	4.90			74	623-42-7	474,339			43(100), 71(55), 87(40), 41(40), 59(30)
Propanoic acid, ethyl ester	5.49			85	105-37-3	2,831,917			57(100), 75(18), 74(15), 102(15), 45(10)
n-Propyl acetate	5.66			76	109-60-4	1,738,458			43(100), 61(40), 73(20), 42(10), 41(8)
Butanoic acid, methyl ester	5.88			95	623-42-7	8,965,644			74(100), 43(90), 71(70), 41(40), 87(30)
Propanoic acid, 2-methyl-, ethyl ester	6.3	Citrus <sup>b</sup> , fruity, buttery	Herbaceous	81	97-62-1	348,148			43(100), 71(40), 41(30), 116(20)
2-Pentanone, 3-methyl-	6.6			63	565-61-7	121,781			43(100), 57(40), 41(35), 72(30)
3-Octene, (E)-	6.7			75	14919-01-8	127,564			41(100), 55(98), 70(40), 112(40)
Butanoic acid, 2-methyl-, methyl ester	6.96		Mint	86	868-57-5	439,900			88(100), 57(80), 41(50), 85(25)
Methyl isovalerate	7.04			88	556-24-1	1,341,790			74(100), 43(40), 59(35), 85(30), 41(25)
1-Butanol	7.13			72	71-36-3	612,423			56(100), 41(70), 43(40), 42(30), 55(20)
Butanoic acid, ethyl ester	7.62			95	105-54-4	6,640,131			71(100), 43(80), 88(55), 41(30), 60(20)
Propanoic acid, propyl ester	7.86			90	106-36-5	9,397,738			57(100), 75(50), 43(20), 87(10)

Table 3. Cont.

Compounds	Retention Time (min)	Odor Description Published Work	Description by Panelist	% Match Library	CAS #	Fresh Feces *	Stale Feces ** Short Equilibrium	Stale Feces ^^ Long Equilibrium	Ion (% Relative Intensity)
MS Detector Response, Peak Area Counts (PACs), and Arbitrary Units									
Acetic acid, butyl ester	8.16			72	123-86-4	416,938			43(100), 56(40), 73(20), 41(19), 61(15)
Methyl valerate	8.48		Foul	93	624-24-8	4,637,137			74(100), 85(38), 57(35), 43(30), 41(30)
Butanoic acid, 2- methyl-, ethyl ester	8.77		Floral	93	7452-79-1	498,859			57(100), 102(70), 41(40), 85(35), 74(20)
Butanoic acid, 3- methyl-, ethyl ester	8.96			85	108-64-5	242,635			43(100), 88(68), 41(60), 71(50), 85(45)
1-Pentanol	9.46			76	71-41-0	1,917,299			42(100), 55(85), 41(70), 70(60)
Acetoin	9.76			63	513-86-0	584,798			43(100), 45(60), 70(15), 55(10), 88(8)
2-Propanol, 1-propoxy-	10.19			83	1569-01-3	125,926			45(100), 43(90), 73(42), 41(30), 59(28)
Butanoic acid, propyl ester	10.27		Chemical, sweet	95	105-66-8	7,807,628			71(100), 43(70), 89(60), 41(30), 42(20)
Pentanoic acid, ethyl ester	10.40			95	539-82-2	3,496,629			88(100), 85(95), 57(70), 60(40), 101(30)
2-Heptanone	10.52	Soap <sup>a,b</sup> , Fruity, sweet, cheese		79	110-43-0	55,224			43(100), 58(75), 71(10), 114(7)
Propanoic acid, butyl ester	10.6			76	590-01-2	1,373,814			57(100), 56(35), 75(30), 41(20)
Heptanal	10.8	Citrus <sup>a</sup> , fat, rancid		83	111-71-7	68,145			70(100), 44(97), 41(82), 43(75), 55(60)
Acetic acid, pentyl ester	10.9			79	628-63-7	246,282	88,580		43(100), 70(40), 61(25), 55(22), 42(20)
Butanoic acid, 2-methyl-, propyl ester	11.42			70	37064-20-3	836,785			57(100), 103(80), 85(85), 41(60), 42(45)
Butanoic acid, 3-methyl-, propyl ester	11.6	Bitter <sup>b</sup> , sweet, apple fruity		90	557-00-6	3,074,917			85(100), 103(68), 41(60), 43(59), 57(58)
1-Hexanol	11.9	Resin <sup>a</sup> , flower, green	Floral, banana	72	111-27-3	812,614			56(100), 43(62), 55(50), 41(48), 42(40)
Pentanoic acid, 4-methyl-, ethyl ester	12.11			56 #	25415-67-2	61,216			88(100), 101(70), 43(60), 99(55), 55(35)
Propanoic acid, pentyl ester	12.21			79	624-54-4	528,515			57(100), 70(80), 43(45), 55(40), 41(25)
Pyrazine, 2,6-dimethyl-	12.39	Cocoa <sup>a</sup> , meat		88	108-50-9	490,854	49,044		108(100), 42(55), 39(35), 40(30),

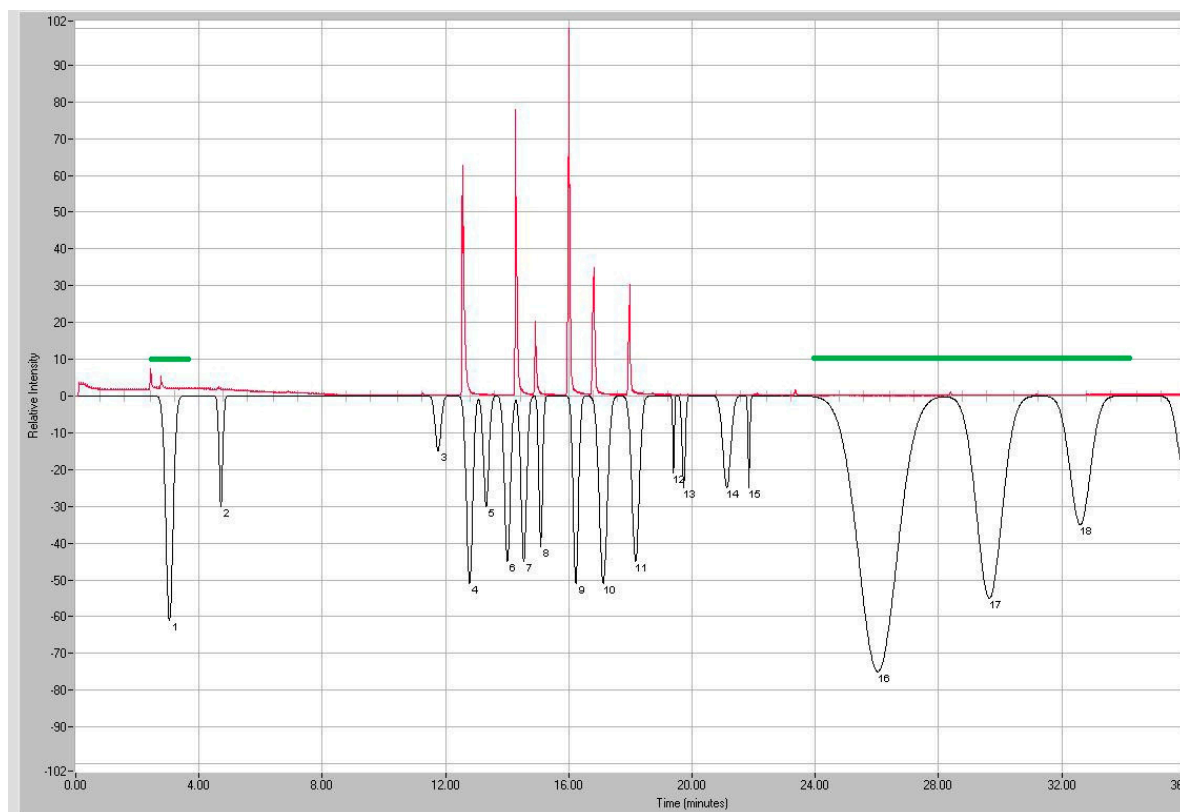
Table 3. Cont.

Compounds	Retention Time (min)	Odor Description Published Work	Description by Panelist	% Match Library	CAS #	Fresh Feces *	Stale Feces ** Short Equilibrium	Stale Feces ^^ Long Equilibrium	Ion (% Relative Intensity)
MS Detector Response, Peak Area Counts (PACs), and Arbitrary Units									
2-Heptanone 5-methyl-	12.52	sour <sup>a</sup>	Herbaceous, grassy, earthy	81	18217-12-4	127,769	159,486,602	74,310,817	43(100), 58(40), 71(38), 70(25), 41(20)
Acetic acid	12.6		sour, nutty	96	64-19-7	33,638,253			43(100), 45(88), 60(60)
Pentanoic acid, propyl ester	12.95			68	141-06-0	4,421,968			85(100), 103(75), 57(70), 41(60)
Propanoic acid, pentyl ester	13.25		Grassy, soil	63	624-54-4	741,201			57(100), 70(40), 75(40), 43(40), 55(25)
5-Hepten-2-one-6-methyl-	13.52	Pungent <sup>a</sup> , rancid	Old cheese	95	110-93-0	228,324	9,605,716	66,087,807	43(100), 41(60), 108(40), 69(40), 39(28)
3-Octanol	14.08			72	589-98-0	171,440			59(100), 83(60), 55(60), 43(55), 44(48)
Butanoic acid, 3-methyl-, butyl ester	14.16			63	109-19-3	157,326			85(100), 57(85), 41(80), 103(72), 56(70)
Propanoic acid	14.31		Unpleasant, butter	93	79-09-4	54,686,812			74(100), 45(72), 73(60), 57(40)
Propanoic acid, 2-methyl-	14.93		Medicinal	85	79-31-2	11,986,638			43(100), 41(55), 73(42), 39(25), 88(10)
Benzaldehyde	15.30		Butter	93	100-52-7	2,215,240			106(100), 105(95), 77(95), 51(45), 50(25)
Butanoic acid	16.04	almond <sup>b</sup>		95	107-92-6	113,602,470	123,858,474	85,788,896	60(100), 73(40), 41(22), 40(20)
Butanoic acid, 3-methyl-	16.84		Sweet, fruity	83	503-74-2	76,585,646			60(100), 41(60), 74(42), 87(30)
2-Methyl-4-decanone	17.60			70 (NIST only)	6628-25-7	116,081			57(100), 85(95), 41(85), 95(85), 43(80), 113(30)
Acetophenone	17.88		Smokey	88	98-86-2	359,940			105(100), 77(80), 120(30), 51(28)
Pentanoic acid	18.02	Must <sup>a,b</sup> , flower, almond		76	109-52-4	108,458,001	52,809,697	24,373,275	60(100), 73(45), 41(20), 45(18)
Pentanoic acid, 4- methyl-	19.13		Butter, basmati rice, butter	85	646-07-1	3,286,558			57(100), 60(80), 41(75), 73(75), 55(62)
Hexanoic acid	19.81			83	142-62-1	3,822,922			60(100), 73(55), 41(32), 87(12)
o-Guaiacol	20.42		Woody, wild	95	90-05-1	4,755,984			109(100), 124(90), 81(70), 53(20)

Table 3. Cont.

Compounds	Retention Time (min)	Odor Description Published Work	Description by Panelist	% Match Library	CAS #	Fresh Feces *	Stale Feces ** Short Equilibrium	Stale Feces ^^ Long Equilibrium	Ion (% Relative Intensity)
MS Detector Response, Peak Area Counts (PACs), and Arbitrary Units									
Benzeneethanol	21.24			91	60-12-8	384,960			91(100), 92(50), 122(25), 65(20)
Benzene propanoic acid, methyl ester	21.93			93	103-25-3	210,723			104(100), 91(60), 164(30), 105(30)
Phenol	22.10	phenol <sup>a</sup>	Medicinal	96	108-95-2	10,114,684	2,023,698	315,371	94(100), 66(35), 65(25), 39(25)
2-Dodecanone	22.5			85	6175-49-1	77,879			58(100), 43(90), 71(35), 59(30), 41(22)
Benzenepropanoic acid, ethyl ester	23.01			85	2021-28-5	148,940			104(100), 91(45), 105(30), 107(28), 178(20)
p-Cresol	23.30	Smoke <sup>a</sup> , medicine	Medicinal	93	106-44-5	39,957,137	3,947,249	1,376,506	107(100), 108(80), 77(20), 39(20)
Phenol, 4-ethyl-	24.80	Must <sup>a</sup>	Foul, unpleasant	94	123-07-9	1,021,035	103,774		107(100), 122(30), 77(20), 106(8)
Butylated hydroxytoluene	25.10	-		96	128-37-0	182,612	52,706		205(100), 220(25), 57(15), 204(15)
Indole	28.38	Burnt <sup>a</sup>	Medicinal, unpleasant	96	120-72-9	69,332,928	195,879	810,599	117(100), 90(40), 89(39), 45(20)
Diethyl Phthalate	29.08	-		94	84-66-2	81,563			149(100), 177(23), 117(20), 150(10), 176(7)
Indole, 3-methyl-	29.2	Fecal <sup>a</sup>	Urinous, animal	71	83-34-1	78,683			130(100), 131(50), 149(20), 117(15), 77(10)

\* Fifty min exposure to SPME at 37 °C; \*\* 15 min equilibrium and 50 min exposure to SPME at 37 °C; ^^ 24 h equilibrium and 50 min exposure to SPME at 37 °C; odor verified with <sup>a</sup> Flavornet [15] and <sup>b</sup> Good Scent Company [16]. # Below 60% library match considered as semi-confirmative.



**Figure 4.** An overlay of the aromagram and total ion chromatogram of the **stale** cat feces, equilibrated for 24 h and exposed to the SPME fiber for 50 min at 37 °C: an increase in the black signal height represents an increased intensity of odor, and the chromatogram is in red for the total ion chromatogram (TIC) signal. The green zone shows that trace levels of the concentration of malodors can produce an intense olfactory response detected by the panelist using GC–MS–O.

The fresh feces on 50 min exposure to SPME fiber caused several ketones, aldehydes, esters, acids, and phenols to accumulate in the SPME fiber. Among these identified chemicals, most of the high-molecular-weight compounds have “smoke”, “medicinal”, “animal”, and “foul” smells, whereas the low-molecular-weight chemicals had more “chemical”, “sweet”, “fruity”, and “grassy” or “earthy” smells. From this, it is more obvious that the high-molecular-weight phenolic and N-aromatic heterocyclic compounds are the contributors to the overall smell of the fresh feces.

The compounds with high molar masses that appeared after a retention time of 20 min or higher had intense malodors. It is interesting to mention that, for stale feces, a short equilibration time of 10 min resulted in missing many low-molecular-weight compounds, and several high-molecular-weight compounds had high PAC-like p-cresol, phenol, and guaiacol compared to long equilibration times of 24 h. In stale feces, the primary contributors such as 3-methyl indole were missing, and phenolics had lower PAC (4-ethyl phenol, p-cresol, and phenol) than phenolics in fresh feces headspaces. The odor intensity of the many chemicals, including phenolic compounds, were similar in the stale cat feces, as revealed by the GC–MS–olfactometer. The aromagrams of feces samples show that the presence of odorants even in the below-the-detection-limit concentration for GC–MS can cause a considerable odor impact (the green line in Figures 3 and 4) problem. The aromagram also revealed that compounds not in the GC–MS data can still contribute to the overall odor, thus triggering the human nose to react to the odorants.



#### 4. Discussion

The focus of this study is reporting the malodorous VOCs in domestic cat urine and feces and the influence of aging urine and feces on the emitted VOCs. Domestic cats spray urine to mark their territory, and the odor is unpleasant to many people. The concentration of volatile compounds can differ by age and sex of the species [10]. This study reported several VOCs emitted from fresh cat urine and feces that were not reported in previous studies. The precursor of many sulfur-containing compounds, felinine, was not identified previously in research [6]. As reported by Miyazaki et al. (2018), the presence of a fishy odor in the headspace of domestic cat anal sac secretions suggesting that cat urine can produce distinct fishy odorous trimethylamine is also present in our findings of the distinctive fishy odor in the fresh and stale feces but not in fresh or aged urine headspace [17]. In contrast, Banik et al., 2020, reported the odor of trimethylamine released from a carpet contaminated with cat urine upon two weeks of aging; however, the “fishy” smell was absent in the emissions from fresh urine-treated carpets [18]. Feral male cats spray urine more often than females [19] and use unburied feces as scent-marking [20], and the released volatiles in both cases could be offensive to humans; the odor becomes even worse with time, although the VOC concentrations become trace levels in feces sample with time (Figure 4). This current study recorded the odor description of emitted VOCs that were never reported before for aged cat urine and feces studies. In addition to phenolics as a major odor contributor, indoles are prominent in stale urine, and organic acids and indoles are prominent in fresh urine odor. Our work supports that 2,5-dimethyl pyrazine and other pyrazines are the product of cat urine aging [11]. Additionally, this work also reports that longer (e.g., 50 min) exposure of the SPME to the aged urine headspace allowed the pyrazines, pyrrole, and pyrimidine to be absorbed onto the fiber and to be detectable in GC-MS.

The use of SPME sampling has allowed easy preconcentrating of sample without the use of a solvent or derivatization of the VOCs responsible for the nuisance to human sensory organs. The volatile compound types and the relative PACs emitted by urine and feces significantly changed over time, even under laboratory conditions. The total number of compounds increased for the aged urine sample and decreased for the aged feces samples. Many esters and acidic VOCs dropped in number for the aged feces, either degraded to other compounds by the microbial community present in the sample, oxidized, or lost to the atmosphere during the aging process [11].

A number of phenol, alcohol, aldehyde, N-compounds (amines, pyrazines, and indoles), S-compounds (dimethyl disulfide and dimethyl sulfone), ketones, and acids reported in this study were emitted from the urine of lion, tiger, and domestic cat species as reported in previous studies (Table 4) [4,8,21]. Highly odorous gases emitted from urine or feces pose likely low inhalation risks. The human nose is very sensitive to many impactful odorants emitted from feces and urine in general. The nuisance odor experienced by homeowners or visitors can be attributed to stale urine. The aged urine is difficult to clean and completely remove from carpets and subflooring materials. Some homeowners experience these stale urine-like smells from floor areas that linger for years even after the cats are removed and no longer contributing urine. Once the carpet is contaminated with cat urine, many of the VOCs emitted are different compared to the VOCs emitted initially by urine itself; however, several VOCs emitted from the contaminated carpet are common to those reported here [18].

**Table 4.** Some literature recorded the VOCs emitted from urine, feces, and fruits of different biological species commonly found that are also in the current study.

Compounds	CAS #	Lion Urine (Soso and Koziel, 2017) [21]	Tiger Urine (Soso and Koziel, 2016) [8]	Cat Urine (Miyazaki, 2006) [4]	Swine Manure (Lo et al., 2006) [22]	Grapes (Rice et al., 2019) [23]	Cat Urine-Carpet (Banik et al., 2020) [18]	Current Study
Phenol	108-95-2	X	X		X		X	X
p-Cresol	106-44-5	X	X	X	X			X
Phenol, 4-ethyl-	123-07-9				X			X
3-Octanol	589-98-0				X			X
1-Butanol	71-36-3		X		X	X		X
1-Hexanol	111-27-3		X		X	X	X	X
Benzyl alcohol	100-51-6					X		X
3-Buten-1-ol, 3-methyl-	763-32-6			X				X
Benzene ethanol	60-12-8	X	X		X			X
Dimethyl disulfide	624-92-0	X			X			X
Dimethyl sulfone	67-71-0			X				X
Benzaldehyde	100-52-7	X	X	X	X	X		X
Butanal, 3-methyl-	590-86-3	X	X					X
Isobutyraldehyde	78-84-2					X		X
Trimethyl amine	75-50-3	X	X					X
2-Dodecanone	6175-49-1				X			X
2-Heptanone	110-43-0		X		X		X	X
2-Pentanone	107-87-9	X					X	X
3-methyl 2-pentanone	565-61-7				X			X
2-Butanone	78-93-3	X	X	X	X	X		X
Acetone	67-64-1	X	X	X	X	X		X
5-Hepten-2-one, 6-methyl-	110-93-0				X		X	X
Acetophenone	98-86-2				X	X		X
Jasmone	488-10-8			X	X			X
Indole	120-72-9	X	X		X			X
3-methylindole	95-20-5				X			X
Pyrazine, 2,6 dimethyl -	108-50-9	X	X					X
Acetic acid	64-19-7			X	X	X		X
Butanoic acid	107-92-6			X	X			X
Pentanoic acid, 2-methyl-	79-31-2				X			X
Butanoic acid, 3-methyl-	503-74-2		X		X			X
Pentanoic acid	109-52-4				X	X		X
Propanoic acid	79-09-4				X			X
Hexanoic acid	142-62-1					X		X
Butanoic acid, 3- methyl	503-74-2			X				X
Carbon disulfide	75-15-0					X		X
Butanoic acid, ethyl ester	105-54-4					X		X

CAS # = Chemical Abstracts Service; X represents the VOCs common to this study and previously published work.

Many VOCs emitted from the cat feces samples, as reported in this study, were also reported to be emitted from swine manure samples [22]. In addition, a few VOCs extracted by SPME and reported in this study are found to be common in the SPME extraction from cold-hardy grape samples [23]. The characteristic odor compound 2,5 dimethyl pyrazine from lion and cat urine indicates the evolutionary similarities between animals. Chemicals such as phenol and p-cresol in human urine are important biomarkers, and they are challenging to measure due to their presence at a low detection limit [24]. However, the use of GC–MS–O can verify their presence by their distinct “ruinous” or “barnyard” odor even at trace levels (Figures 3 and 4). These observations also indicate the fact that SPME in a combination of GC–MS–O for the headspace extraction is better suited for assessing odor than only GC–MS. Overall, SPME reduces the sampling time for volatile or semi-volatile compound determination compared to a traditional sampling technique [25], and this technology can be developed more for future assessment of quantitative analysis of these odorous constituents characteristic to the overall smell of urine or feces sample.

Several VOCs were common in urine and feces samples, such as acetone, 2-butanone, 3-methyl butanal, 3 methyl 2-pentanone, phenol, p-cresol, and indole. Chemicals such as phenol and indole are commonly produced by cat species irrespective of their sex and age [8,10]. N-heterocycles like pyrimidine and pyrroles were only recorded in the urine after aging but not in the fresh nor stale feces. Similarly, o-guaiacol, 4-ethyl phenol, and 2-methyl indole were only present in the feces samples but not in the urine. This reveals that some chemicals possibly carry urine and feces samples’ characteristic VOCs and substantially contribute to their specific odor. This research reported chemicals emitted from domestic cat fresh urine and feces to the vial headspace and upon aging. However, there are limitations to this work as the research did not test different SPME fiber coatings and extraction regimes (time (T)). There is still plenty of opportunities to explore specific questions, e.g., the effectiveness of cleaning products for cat urine and the mitigation of malodors from cat litter boxes.

**Supplementary Materials:** The following are available online at <https://www.mdpi.com/2297-8739/8/2/15/s1>, Table S1: Aroma description as recorded by the panelist for fresh cat urine at 10 min extraction, Table S2: Aroma description as recorded by the panelist for fresh cat urine at 50 min extraction, Table S3: Aroma description as recorded by the panelist for stale cat urine at 50 min extraction, Table S4: Aroma description as recorded by the panelist for fresh feces at 50 min extraction, Table S5: Aroma description as recorded by the panelist for stale feces at 15 min equilibrium and 50 min extraction, Table S6: Aroma description as recorded by the panelist for stale cat feces at 24 h equilibrium and 50 min extraction.

**Author Contributions:** Conceptualization, J.A.K. and J.Z.L.; methodology, J.A.K., J.Z.L., and C.B.; formal analysis, C.B.; investigation, C.B.; resources, J.A.K.; data curation, C.B.; writing—original draft preparation, C.B.; writing—review and editing, J.A.K. and J.Z.L.; visualization, C.B.; supervision, J.A.K.; project administration, J.A.K.; funding acquisition, J.A.K. All authors have read and agreed to the published version of the manuscript.

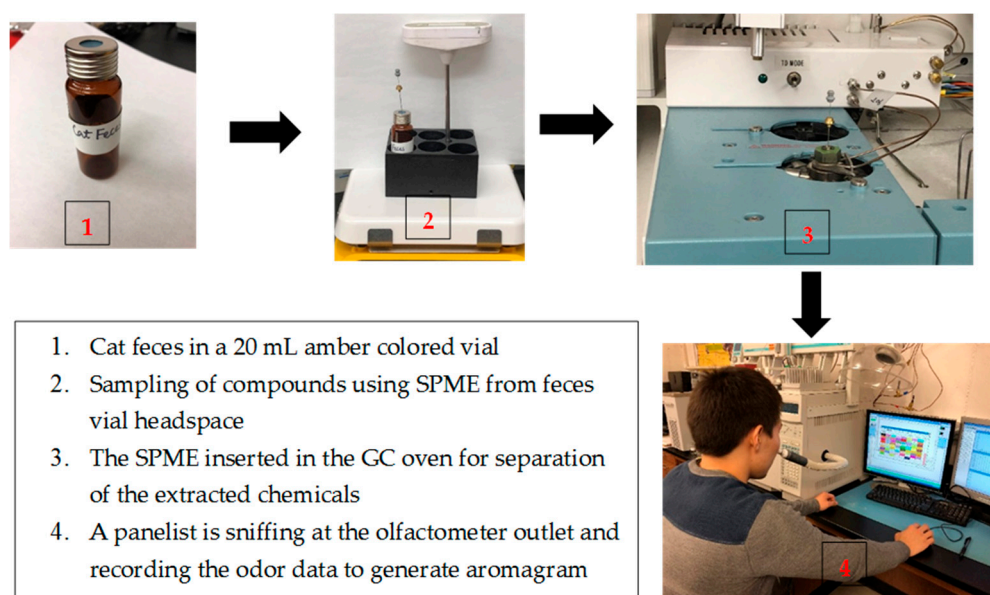
**Funding:** This research was funded by Nestlé Purina Pet Care Company. Partial support came from the Iowa Agriculture and Home Economics Experiment Station, Ames, Iowa, project no. IOW05556 (Future Challenges in Animal Production Systems: Seeking Solutions through Focused Facilitation) sponsored by Hatch Act and State of Iowa funds.

**Data Availability Statement:** The original contributions presented in the study are included in the article and the Supplementary Materials; further inquiries can be directed to the corresponding author.

**Acknowledgments:** The authors would like to thank Baitong Chen, Jisoo Wi, Myeongseong Lee, and Zhanibek Meirirhanuly for technical support in the laboratory.

**Conflicts of Interest:** The authors declare no conflict of interest.

## Appendix A



**Figure A1.** Schematic representation of the cat urine, feces, and soiled litter sample preparation and analysis.

## References

- Morris, D. *Cat World: A Feline Encyclopedia*; Penguin Reference: New York, NY, USA, 1997; ISBN 0670100064.
- American Veterinary Medical Association. U.S. Pet Ownership Statistics. Available online: <https://www.avma.org/resources-tools/reports-statistics/us-pet-ownership-statistics> (accessed on 31 December 2020).
- Westall, R.G. The amino acids and other ampholytes of urine 2. The isolation of a new Sulphur-containing amino acid from cat urine. *Biochem. J.* **1953**, *55*, 244–248. [[CrossRef](#)] [[PubMed](#)]
- Miyazaki, M.; Yamashita, T.; Suzuki, Y.; Saito, Y.; Soeta, S.; Taira, H.; Suzuki, A. A major urinary protein of the domestic cat regulates the production of Felinine, a putative pheromone precursor. *Chem. Biol.* **2006**, *13*, 1071–1079. [[CrossRef](#)] [[PubMed](#)]
- Rutherford, K.J.; Rutherford, S.M.; Moughan, P.J.; Hendricks, W.H. Isolation and characterization of a felinine containing peptide from the blood of the domestic cat (*Felis catus*). *J. Biol. Chem.* **2002**, *277*, 114–119. [[CrossRef](#)] [[PubMed](#)]
- Hendriks, W.H.; Rutherford-Markwick, K.J.; Weidgraaf, K.; Ugarte, C.; Rogers, R. Testosterone increases urinary free felinine, N-acetylfelinine and methylbutanolglutathione excretion in cats (*Felis catus*). *J. Anim. Physiol. Anim. Nutr.* **2008**, *92*, 53–62. [[CrossRef](#)] [[PubMed](#)]
- Miyazaki, M.; Miyazaki, T.; Nishimura, T.; Hojo, W.; Yamashita, T. The chemical basis of species, sex and individual recognition using feces in the domestic cat. *J. Chem. Ecol.* **2018**, *44*, 364–373. [[CrossRef](#)]
- Soso, S.B.; Koziel, J.A. Analysis of odorants in marking fluid of Siberian tiger (*Panthera tigris altaica*) using simultaneous sensory and chemical analysis with headspace solid-phase microextraction and multidimensional gas chromatography-mass spectrometry-olfactometry. *Molecules* **2016**, *21*, 834. [[CrossRef](#)]
- Miyazaki, M.; Nishimura, T.; Hojo, W.; Miyazaki, T.; Laine, R.A.; Yamashita, T. Potential use of domestic cat (*Felis catus*) urinary extracts for manipulating the behavior of free-roaming cats and wild small fields. *Appl. Anim. Behav. Sci.* **2017**, *196*, 52–60. [[CrossRef](#)]
- Uetake, K.; Abumi, T.; Suzuki, T.; Hisamatsu, S.; Fukuda, M. Volatile faecal components related to sex and age in domestic cats (*Felis catus*). *J. Appl. Anim. Res.* **2017**, *46*, 766–770. [[CrossRef](#)]
- Suzuki, C.; Miyazaki, T.; Yamashita, T.; Miyazaki, M. GC X GC-MS-based volatile profiling of male domestic cat urine and olfactory abilities of cats to discriminate temporal changes and individual differences in urine. *J. Chem. Ecol.* **2019**, *45*, 579–587. [[CrossRef](#)]
- Starkemann, C.; Niclass, Y.; Cayeux, I.; Brauchli, R.; Gagnon, A. Odorant volatile sulfur compounds in cat urine: Occurrence of (+/-)-3,7-dimethyloct-3-sulfanyl-6-en-1-ol and its cysteine conjugate precursor. *Flavour Frag. J.* **2014**, *30*, 91–100. [[CrossRef](#)]
- Funaba, M.; Uchiyama, A.; Takahashi, K.; Kanako, M.; Yamamoto, H.; Namikawa, K.; Iriki, T.; Hatano, Y.; Abe, M. Evaluation of effects of dietary carbohydrate on formation of struvite crystals in urine and macromineral balance in clinically normal cats. *Am. J. Vet. Res.* **2004**, *65*, 138–142. [[CrossRef](#)] [[PubMed](#)]
- Bojko, B.; Reyes-Garcés, N.; Bessonneau, V.; Gorynski, K.; Mousavi, F.; Silva, E.A.S.; Pawliszyn, J. Solid-phase microextraction in metabolomics. *Trends Anal. Chem.* **2014**, *61*, 168–180. [[CrossRef](#)]
- Flavornet. Available online: <http://flavornet.org/flavornet.html> (accessed on 31 December 2020).

16. Good Scent Company. Available online: <http://www.thegoodscentcompany.com/search.html> (accessed on 31 December 2020).
17. Miyazaki, T.; Nishimura, T.; Yamashita, T.; Miyazaki, M. Olfactory discrimination of anal sac secretions in the domestic cat and the chemical profiles of the volatile compounds. *J. Ethol.* **2018**, *36*, 99–105. [[CrossRef](#)] [[PubMed](#)]
18. Banik, C.; Koziel, J.; Flickinger, E. Volatile compounds emitted from the cat urine contaminated carpet before and after treatment with marketed cleaning products: A simultaneous chemical and sensory analysis. *Data* **2020**, *5*, 88. [[CrossRef](#)]
19. Natoli, E. Behavioural responses of urban feral cats to different types of urine marks. *Behaviour* **1985**, *94*, 234–243. [[CrossRef](#)]
20. Ishida, Y.; Shimizu, M. Influence of social rank on defecating behaviours in feral cats. *J. Ethol.* **1998**, *16*, 15–21. [[CrossRef](#)]
21. Soso, S.B.; Koziel, J.A. Characterizing the scent and chemical composition of *Panther leo* marking fluid using solid-phase microextraction and multidimensional gas chromatography-mass spectrometry-olfactometry. *Sci. Rep.* **2017**, *7*, 5137–5152. [[CrossRef](#)]
22. Lo, Y.C.; Koziel, J.A.; Cai, L.; Hoff, S.J.; Jenks, W.S.; Xin, H. Simultaneous chemical and sensory characterization of volatile organic compounds and semi-volatile compounds emitted from swine manure using solid phase microextraction and multidimensional gas chromatography-mass spectrometry-olfactometry. *J. Environ. Qual.* **2008**, *37*, 521–534. [[CrossRef](#)]
23. Rice, S.; Maurer, D.L.; Fennell, A.; Dharmadhikari, M.; Koziel, J.A. Evaluation of volatile metabolites emitted in-vivo from cold-hardy grapes during ripening using SPME and GC-MS: A proof of concept. *Molecules* **2019**, *24*, 536. [[CrossRef](#)]
24. King, R.A.; May, B.L.; Davies, D.A.; Bird, A.R. Measurement of phenol and p-cresol in urine and feces using vacuum microdistillation and high-performance liquid chromatography. *Anal. Biochem.* **2009**, *384*, 27–33. [[CrossRef](#)]
25. Risticvic, S.; Chen, Y.; Kudlejova, L.; Vatinno, R.; Baltensperger, B.; Stuff, J.R.; Hein, D.; Pawliszyn, J. Protocol for the development of automated high-throughput SPME-GC methods for the analysis of volatile and semivolatile constituents in wine samples. *Nat. Protoc.* **2010**, *5*, 162–176. [[CrossRef](#)] [[PubMed](#)]