Table S1. Bacterial strains used in this study

Strain	Serotype	Source	Verotoxin production
***E. coli* CECT 4782**	O157:H7	Human stool from the outbreak of hemorrhagic colitis	stx1, stx2
***E. coli* CECT 4783**	O157:H7	Raw hamburger meat implicated in hemorrhagic colitis outbreak	stx1, stx2
***E. coli* CECT 4267**	O157:H7	Human stool from the outbreak of hemorrhagic colitis	stx1, stx2
***E. coli* CECT 5947**	O157:H7	-	Gene stx2 has been replaced
***E. coli* NCTC 12900**	O157:H7	-	NT
***E. coli* CECT 352**	O127a:K63(B8):H-	-	EPEC
***E. coli* CECT 504**	O141:K85(B):H4	Swine edema	ND
***E. coli* CECT 515T**	O1:K1(L1):H7	Human urine -cystitis	ND
***E. coli* CECT 533**	O103:K-:H-	-	ND
***E. coli* CECT 727**	O111:K58(B4):H-	Infantile gastroenteritis	EPEC
***E. coli* CECT 730**	O55:K59(B5):H-	-	ND
***E. coli* CECT 736**	O28a,28c:K73(B18):H-	Faeces	ND
***E. coli* CECT 740**	O125a,125b:K70(B15):H19	Human gastroenteritis	ND
***E. coli* CECT 744**	O158:K-:h23	Faeces of an infant with diarrhea	ND
***E. coli* CECT 832**	O111:K58(B4):H-	Infantile gastroenteritis	ND
***E. coli* CECT 4537**	O10:K5(L5):H4	Human peritonitis	ND
***E. coli* CECT 4555**	O97:K-:H-	-	ND
***E. coli* CECT 434**	O6:KN	Human gastroenteritis	ND
E. coli* N5	ND	Bovine faeces	ND
***E. coli* ATCC 29425 (K12)**	OR:H48:K-	-	ND
E. coli* K12 Δ*impA	-	Laboratory collection	-
E. coli* ER1100A Δ*entF	-	Laboratory collection	-
***Listeria monocytogenes* CECT 5873**	-	-	-
***Salmonella enterica* SGSC 2476**	-	-	-
***Shigella dysenteriae* ATCC 11335**	-	-	-
***Staphylococcus aureus* CECT 86**	-	-	-
***Staphylococcus epidermidis* CECT 4184**	-	-	-

NT – non-toxigenic *E. coli*; EPEC – enteropathogenic *E. coli* (epidemiologically implicated as pathogens, but virulence mechanism is not related to the excretion of enterotoxins); ND – Not determined; * - Isolates; SGSC – *Salmonella* Genetic Stock Centre; ATCC – American Type Culture Collection; NCTC - National Collection of Type Cultures; CECT - Spanish Type Culture Collection.

Table S2. Primers used in this study

Target colicin gene	Primers sequence (5′ to 3′)		Tm (°C)	Amplicon size (bp)
For PCR studies				
Ia; Ib	Fw	GCAGACACGGAATGACAGGGC	68	406
	Rv	GCCGTAACTTATCCCATTCAGC	66	
E1-E3; E6; E8-E9	Fw	CCTTATGATGATAAGGGGCAGG	66	576
	Rv	CCAGCTCAGATTGTGCAGCAGC	70	
E7	Fw	CCGAGAACCAATGGCTGCTGG	68	476
	Rv	CCTGGGTTCTAGTCTTGGGCG	68	
5; 10	Fw	CCAGAGTTGCAGGRGAGC	59	658
	Rv	CATTAATGCCACAATTTTTGCC	60	
B	Fw	CCAAAGGCTATAAGGGCCGAGC	70	709
	Rv	CCCGAAATCCAGGAAGATGGCG	70	
M	Fw	GGTTACTTCTGTAACGCCG	58	495
	Rv	GCCTTGTGAGCGACTCTCC	62	
For qPCR studies				
B	Fw	CCACACTTAATACCAGGTCCGG	57	124
	Rv	CCGATGACAGTGCCAGTAGTGG	59	
M	Fw	CCAAACATGTGTCTTCAGGC	52	140
	Rv	GGGTGAAGAACCAGATTTCG	52	
E	Fw	GGGCGCGCATAGCACAAGTGG	60	124
	Rv	GCTACCGGAACCACCACCC	58	
GAPDH	Fw	GCCTCTTTTTCGGCGTAAACC	54	129
	Rv	CCCCAAAAATTCTCGCCTACC	54	

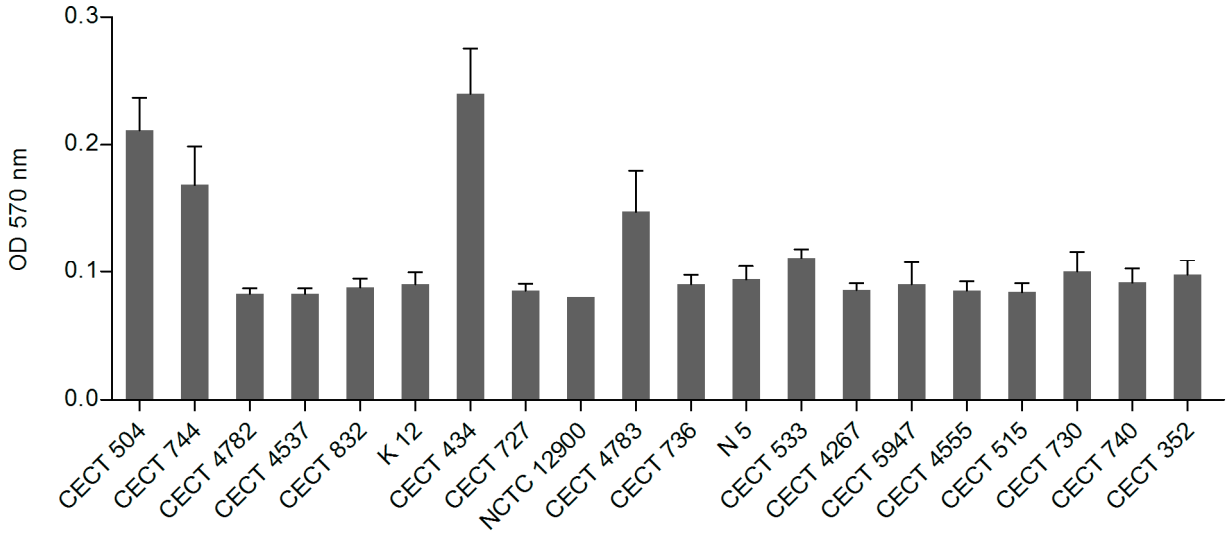

Figure S1. Single-species biofilm formation of *E. coli*. Biofilms were grown in 96-well microtiter plates for 24 h in LB and LB supplemented with 0.25 % (w/v) glucose media. The biomass amount was assessed by the crystal violet method as described in the methods section of this manuscript.