Supplementary Information.

Development of Nano-Antifungal therapy for systemic and endemic mycoses

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Figure S1. SEM-EDX analysis of Cul nanostructured materials. A) SEM micrograph of powdered Cul NMs. B) EDX analysis highlighting copper content. C) EDX analysis highlighting iodide content. The mass (wt) and atomic (at) percentages of the elements confirm the formation of Cul materials.



Figure S2. SEM micrographs illustrating the behavior of powdered CuI nanostructured materials at different doses in phosphate buffered medium in the presence (A^*-C^*) and absence (A-C) of *S.schenckii*.





Figure S3. UV-Vis characterization of Cu@AG colloidal suspension. The spectra shows the characteristic plasmon of Cu NPs.

Figure S4. Evaluation of the antifungal activity (*C. albicans*) of Cul@AG (A), chitosan (B) and Cul@Ch (C) at different doses.



Figure S5. Evaluation of the antifungal activity (*C. albicans*) of Cu@AG (A), growth from dilution (10⁻³) (B) at different doses.



Figure S6. Evaluation of the antifungal activity of Cu@AG to inhibit *C.albicans* growth. A) Viability of control group (decimal dilutions) under experimental conditions (Top row). Evaluation of the fungicidal activity of Cu@AG against *C. albicans* (drop dilution test) at differen NPs concentrations. B) Log reduction on the CFU/mL of *C. albicans* exposed to different amounts of Cu@AG.



Figure S7. Evaluation of the antifungal activity (*S. schenckii*) of Cul@AG at different concentrations and exposition of 5h. A) Fungal cell are viable in the growth controls at different serial decimal dilutions under experimental conditions. B) Evaluation of the antifungal activity of Cul@AG by the drop dilution method. Fungal growth is only observed for growth control. C–F) Evaluation of the antifungal activity of Cul@AG by the drop dilution by the pour plate method. Fungal growth is only observed for growth control. C–F) Evaluation of the antifungal activity of Cul@AG by the drop dilution method.



Figure S8. Evaluation of the antifungal activity (*S. schenckii*) of Cul@Ch at different concentrations and exposition of 5h. A) Fungal cell are viable in the growth controls at different serial decimal dilutions under experimental conditions. B) Evaluation of the antifungal activity of Cul@Ch by the drop dilution method. Fungal growth is only observed for growth control. C–F) Evaluation of the antifungal activity of Cul@Ch by the drop dilution by the pour plate method. Fungal growth is only observed for growth control (C).



Figure S9. Evaluation of the antifungal activity (*F. oxysporumi*) of Cul@AG at different concentrations and exposition of 20h. A) Fungal cell are viable in the growth controls at different serial decimal dilutions under experimental conditions. B–D) Evaluation of the antifungal activity of Cul@AG at different doses by the drop dilution method. Fungal growth is only observed for the fungi exposed to 75 μ g/mL of NMs.



Figure S10. Optical microscopy examination of CuI-*C. albicans* interaction at diferent CuI doses. A) Big aggregates of *C. albicans* are observed for control group. B–D) The aggregates decrease as the concentration of NMs increases (50, 75 and 100 μ g/mL respectively).



Figure S11. AFM analysis of the morphological changes in *C. albicans* due to exposure to Cul@AG (50 µg/mL x 1h). Formation of a pit in fungal cell is observed due to NMs penetration (red dotted circles).A– C are the low-magnification images, whereas D–F are the high magnification images.



Figure S12. AFM analysis of the morphological changes in *S. schenckii* due to exposure to Cul@Ch (50 μg/mL x 5h). A–C) Control group. The fungi maintain their typical morphology under test conditions, demonstrating the bioavailability of the MOs at the end of the experiment. D-I) The NMs interact closely with fungal cells resulting in their destruction.

